

Primary structure and gene organization of human hepatitis A virus

(human picornavirus/structural analysis/nucleotide sequence/sequence homology/polyprotein)

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ABSTRACT The RNA genome of human hepatitis A virus (HAV) was molecularly cloned. Recombinant DNA clones representing the entire HAV RNA were used to determine the primary structure of the viral genome. The length of the viral genome is 7478 nucleotides. An open reading frame starting at nucleotide 734 and terminating at nucleotide 7415 encodes a polyprotein of M_r 251,940. Comparison of the HAV nucleotide sequence with that of other picornaviruses has failed to reveal detectable areas of homology. However, a computer analysis of the putative amino acid sequence of HAV and poliovirus demonstrated the existence of short areas of homology in virion protein 3 (VP3) and throughout the carboxyl-terminal portion of the polyproteins. In addition, extensive protein structural homologies with poliovirus were detected.

Hepatitis A virus is a picornavirus belonging to the enterovirus genus. The viral RNA is 7.5 kilobases long, is of (+)-strand polarity, contains a poly(A) stretch at the 3' end, and may be covalently linked to a protein at the 5' terminus designated VPg (1-3). Evidence for virion proteins of M_r s 33,000, 29,000, and 27,000, designated VP1, VP2, and VP3, respectively, and additional peptides of M_r s 22,000 and 10,000 has been presented (4-6).

Two recent reports have described the molecular cloning of HAV-specific sequences (3, 7). We report here the molecular cloning and the nucleotide sequence of the entire HAV genome. The deduced amino acid sequence has been analyzed and compared to that of poliovirus. The sequence and structural homologies between these two human picornaviruses are discussed.

MATERIALS AND METHODS

Virus Isolation and Propagation. Several groups have reported the successful isolation and growth in tissue culture of HAV derived from both infected marmoset livers and human fecal samples (8-12). We have established our own isolate from the stools of a hepatitis A patient collected during an epidemic outbreak in Los Angeles (kindly provided by J. Rakela) according to published procedures (12). Viral RNA was extracted from virions by published procedures that included treatment with protease and extraction with phenol/chloroform followed by precipitation with ethanol (3).

Molecular Cloning. Details of the procedures and specific approaches used to obtain the entire HAV genome will be reported elsewhere. Briefly, 3'-proximal clones were obtained by screening recombinant cDNA libraries with the probes containing the following sequences (3):

HAV1 3' A-C-A-A-A-T-A-A-G-A-A-A-T-A-G-T-C-
A-T-T-T-A 5'

HAV2 3' A-T-G-T-C-T-G-A-A-T-T-T-A-G-A-A-T-A-C-
T-A-A-C-C-A-C-C 5'

Further portions of the genome were cloned by "walking" techniques and by using the 5'-terminal *Pst* I-*Pst* I fragment derived from clone pHAV1307 (kindly provided to us by J. Ticehurst, National Institutes of Health).

RESULTS AND DISCUSSION

Analysis of cDNA Clones. A set of four recombinant cDNA clones representing the whole HAV genome was selected for nucleotide sequencing studies. The four clones, termed pHAV16, pHAV1, pHAV8, and pHAV47, were mapped with restriction endonucleases, and the overlapping regions were confirmed by cross-hybridization studies (Fig. 1). The total length of the nonoverlapping portions of these clones added up to approximately 7.5 kilobases, the reported length of the HAV genome (3).

The nucleotide sequence of the entire HAV genome derived from the four cDNA clones is shown in Fig. 2. According to this sequence, the HAV genome is 7478 nucleotides long. The heteropolymeric sequence is followed by a poly(A) tract of undetermined length. A single open reading frame starts with an AUG triplet at nucleotide 734 and terminates with an UGA codon at nucleotide 7415. The polyprotein thus encoded is 2227 amino acids long, has a M_r of 251,940, and is divided into P1, P2, and P3 regions (13).

The sequence at the 3' end of the genome was determined from independent clones carrying the poly(A) 3'-terminal sequence. The sequence at the 5' end of the viral RNA was derived from five independent clones that carried a terminal *Pst* I-*Pst* I fragment of approximately 180 nucleotides. Of these five, three were found to be identical and contained the 5'-terminal sequence shown in Fig. 2.

HAV Map. The genome of the independent HAV isolate cloned in our laboratory has a restriction map substantially different from that of the isolate cloned by Ticehurst *et al.* (3). The observed differences may be due to single nucleotide changes that do not substantially alter the primary amino acid sequence and antigenic properties of the viral proteins. This notion is supported by a comparison of the 3'-terminal 500 nucleotides of our sequence with the homologous sequence reported by Ticehurst *et al.* (3). Only a single amino acid difference at position 2159 (methionine for isoleucine) is detected in this portion of the genome. In the same region there are 24 nucleotide substitutions that do not alter the amino acid sequence. Additional changes are present in the 3' noncoding region of the viral RNA. We conclude that

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Abbreviation: HAV, hepatitis A virus.

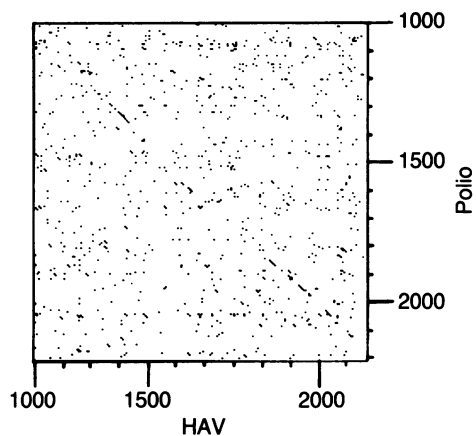


FIG. 3. Dot-matrix analysis of homology between the amino acid sequence of HAV and of poliovirus. The amino acid sequences of the two viruses were compared, starting at amino acid 1000. The computer program used a window of five amino acids with a filter of four matches out of five. Amino acids having similar properties were equated as follows: Thr=Ser; Tyr=Phe; Val=Leu=Ile; Asp=Glu; Lys=Arg; and Asn=Gln.

any detectable homology.

To verify the accuracy of the putative reading frame

throughout the HAV genome, two separate analyses of the sequence were conducted. First, the codon usage in the HAV open reading frame was examined and found to be consistent with that reported for poliovirus (15) and other human genes. In addition, the codon usage within the HAV sequence itself was checked by comparing consecutive blocks of 1,000 nucleotides through the entire genome. No inconsistencies were observed in this internal comparison, leading to the conclusion that the nucleotide sequence encodes the correct polypeptide in every part of the HAV genome, and no substantial areas of frameshift have been introduced by sequencing errors. Second, the amino acid sequences of HAV and poliovirus were compared. A single area of homology was detected within the amino-terminal 1000 amino acids of the two viruses. The amino acid sequence of this region from the two viruses is:

Poliovirus	Val-Pro-Trp-Ile-Ser-Asn-Ser-Thr-Xaa-Tyr-Arg
	507
HAV	Val-Pro-Trp-Ile-Ser-Asp-Xaa-Thr-Pro-Tyr-Arg
	418

The positions of this homology stretch in the protein sequence (poliovirus amino acid 508, HAV amino acid 418) indicates that HAV has undergone approximately a 300-nucleotide shift in the position of this sequence compared to

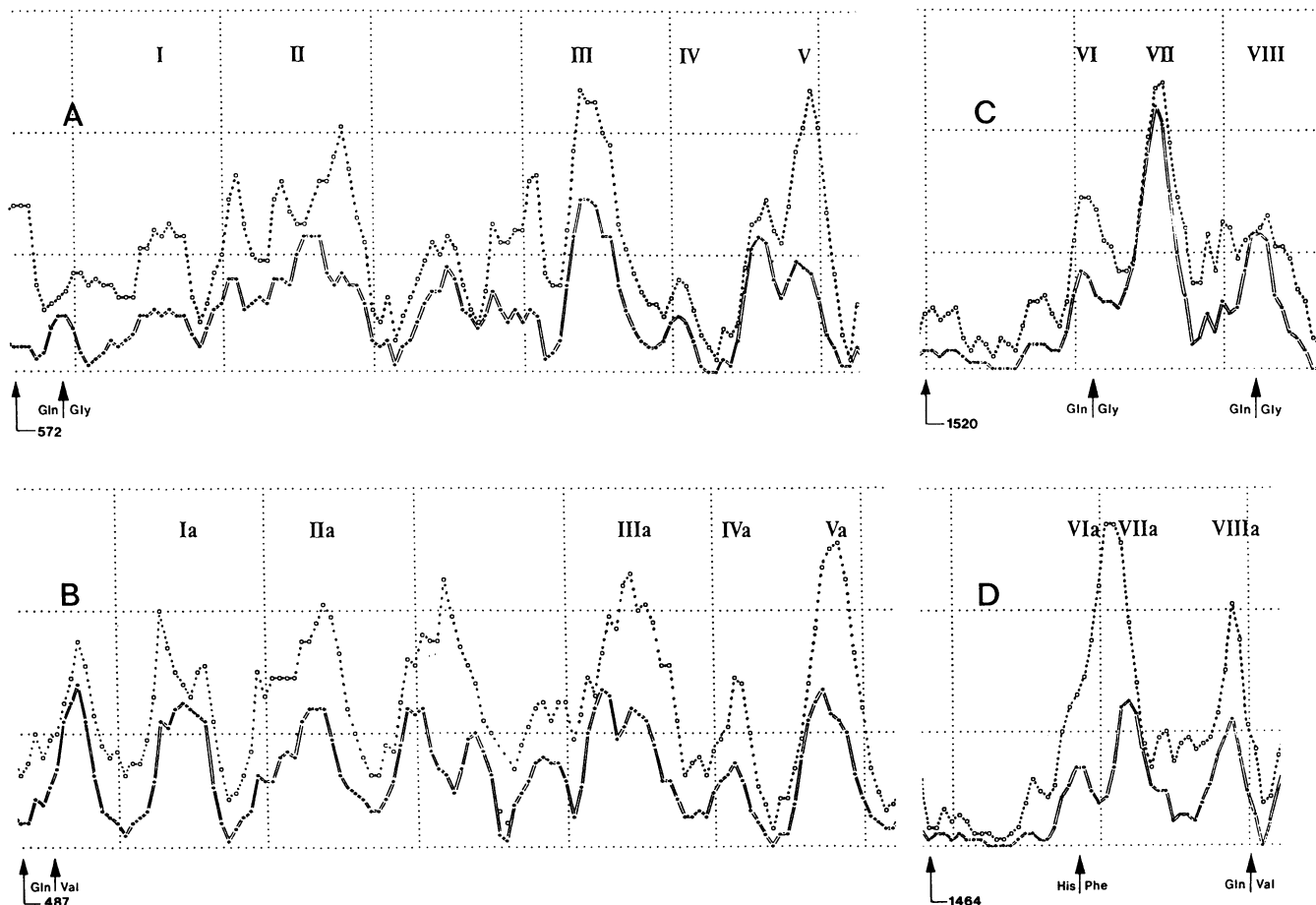


FIG. 4. Structural analysis of selected regions of HAV and poliovirus. The structural profile was computed by using a combination of structure and hydrophilicity. Three parameters were mixed: (i) the potential for helix or the β -sheet formation of the individual residues (19, 21) (values greater than 1.0 were scored as structure positive, and the higher one was selected; below 1.0, the lower value was selected), (ii) the hydrophilicity index of the residue (20), and (iii) the average hydrophilicity index of the four surrounding residues. Each parameter was normalized to affect the final curve evenly. Curves were corrected by averaging over seven residues., Curve showing the mixing of parameters ii and iii such that a peak represents a region of high hydrophilicity; = = =, curve showing the mixing of the three parameters such that a peak represents a region of high hydrophilicity and lack of structural features. Arrows with numbers indicate the position of the first amino acid shown on the viral polypeptide. Each dot of the grid represents one residue, and each grid space contains 20 residues. Known cleavage positions for poliovirus and putative ones for HAV are indicated by arrows and amino acid pairs. (A) Poliovirus VP1. (B) HAV VP1. (C) Poliovirus VPg. (D) HAV VPg.

poliovirus. Taking into account that both polyproteins start within a few nucleotides on the respective genomes, we conclude that the initial portion of region P1 in HAV has undergone a small deletion.

A comparison of the second half of HAV and poliovirus polyproteins (amino acids 1000–2200) is shown in Fig. 3. Here a number of homologous stretches were identified, indicating a higher degree of conservation in this part of the viral genome. Our data indicate that the portions of the HAV genome that, by analogy with poliovirus, are expected to code for the enzymes involved in viral replication are more highly conserved than the regions coding for the virion proteins and viral antigenic determinants. The finding of short regions of homology throughout the second part of the two genomes supports the notion that the HAV open reading frame is correctly identified by the nucleotide sequence.

Structural Comparison of HAV and Poliovirus Polyproteins. A number of recent reports on protein structure have indicated that proteins carrying out homologous functions but with little or no sequence homology do display considerable "structural" similarities. Among these, areas of high hydrophilicity devoid of structural features (i.e., α helices and β sheets) have been found to correspond often to major antigenic sites (13, 19, 20).

HAV and poliovirus polyproteins were compared by using a computer program that displays nonstructural and hydrophilic properties of polypeptides (Fig. 4). Highly similar profiles were obtained throughout the entire polyprotein. Examples of these similarities for regions of interest are shown in Fig. 4. The short region of amino acid homology between HAV and poliovirus (poliovirus, amino acid 507; HAV, amino acid 418) was used to match the two sequences.

Fig. 4A shows the amino-terminal portion of poliovirus VP1. The cleavage site between VP3 and VP1 is identified by the Gln-Gly amino acid pair. Fig. 4B represents the matched homologous region on HAV. In a position quite close to the Gln-Gly cleavage pair on poliovirus, a Gln-Val pair is found on HAV. We think that this is a likely candidate for the cleavage site between VP3 and VP1 of HAV. The overall structural and hydrophilicity profiles of the two proteins are quite similar. "Peaks" representing putative areas of structural homology have been identified with corresponding roman numerals. Areas III and V of poliovirus represent major antigenic sites of VP1 (22–25). One tentative conclusion that can be drawn from this analysis is that, if these structurally homologous regions serve similar functions in the two viruses, then areas IIIa and Va of HAV should represent potential major antigenic sites for this virus.

Another area of striking similarity is shown in Fig. 4C and D. These areas represent polio's VPg and the homologous HAV region. A long hydrophobic stretch (20 amino acids) precedes the Gln-Gly cleavage site in poliovirus (Fig. 4C). A similar region is found in HAV; by matching this region and areas VI–VIa in the two viruses, one can infer that the HAV VPg should start around the His-Phe residues shown in Fig. 4D. This is followed by areas VII–VIIa and VIII–VIIIa, also displaying a high degree of structural homology. The end of VPg is marked by a Gln-Gly pair in poliovirus and possibly by the Gln-Val pair in HAV. If these cleavage sites of HAV VPg are correct, the resulting protein is 23 amino acids long (poliovirus VPg is 22 amino acids long) and contains a tyrosine residue at position 1499 (see Fig. 2), which can mediate the protein nucleic acid linkage at the 5' end of the genomic RNA.

Using the same criteria of structural homology for comparison, we have attempted to map the rest of the HAV polyprotein with respect to the known poliovirus polyprotein-processing sites. Tentative boundaries for the P1, P2, and P3 regions have been identified and are shown in Fig. 2. Similarly, major putative cleavage sites are shown for the

virion structural and nonstructural proteins. All gene products identified for poliovirus appear to have homologs in HAV at very similar positions except for VP4, which as a consequence of the 100-amino acid shift in the VP1 position as discussed, may not be encoded as the first protein in P1 but rather as the last one, following VP1. In general, with the exception of one VPg cleavage site (His-Phe), potential cleavage sites have a glutamine/hydrophobic amino acid pair.

It is important to stress that all of the cleavage sites indicated in Fig. 2 are tentative and that, in the absence of specific protein size and sequence data, no firm identifications are possible. However, we believe that this kind of analysis provides us with good working hypotheses for future experimentation. For example, tentative identification of VP1 coding sequences and putative antigenic sites can now be experimentally tested by the use of synthetic peptides in a manner analogous to that used for poliovirus. In addition, expression of individual portions of the HAV genome in microorganisms may provide us with specific gene products for detailed immunological studies.

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