

Pharmacological and toxicological effects of co-exposure of human gingival fibroblasts to silver nanoparticles and sodium fluoride

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Background: Silver nanoparticles (AgNPs) and fluoride (F) are pharmacological agents widely used in oral medicine and dental practice due to their anti-microbial/anti-cavity properties. However, risks associated with the co-exposure of local cells and tissues to these xenobiotics are not clear. Therefore, we have evaluated the effects of AgNPs and F co-exposure on human gingival fibroblast cells.

Methods: Human gingival fibroblast cells (CRL-2014) were exposed to AgNPs and/or F at different concentrations for up to 24 hours. Cellular uptake of AgNPs was examined by transmission electron microscopy. Downstream inflammatory effects and oxidative stress were measured by real-time quantitative polymerase chain reaction (PCR) and reactive oxygen species (ROS) generation. Cytotoxicity and apoptosis were measured by 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay and real-time quantitative PCR and flow cytometry, respectively. Finally, the involvement of mitogen-activated protein kinases (MAPK) was studied using Western blot.

Results: We found that AgNPs penetrated the cell membrane and localized inside the mitochondria. Co-incubation experiments resulted in increased oxidative stress, inflammation, and apoptosis. In addition, we found that co-exposure to both xenobiotics phosphorylated MAPK, particularly p42/44 MAPK.

Conclusion: A combined exposure of human fibroblasts to AgNPs and F results in increased cellular damage. Further studies are needed in order to evaluate pharmacological and potentially toxicological effects of AgNPs and F on oral health.

Keywords: nanoparticles, oxidative stress, apoptosis, inflammation, mitogen-activated protein kinases, matrix, metalloproteinases

Introduction

Despite the fact that demand for medical implants is growing worldwide, patients usually suffer from bacterial infections caused by these devices. In order to prevent such infections, the best approach is to deliver antimicrobial agents on the surface of the implant. Metallic silver has been used across civilizations for numerous medical conditions.¹⁻³ Compared to other nonessential heavy metals, silver shows the highest antimicrobial efficacy against microorganisms. In addition, microbes are unlikely to develop resistance against silver, because the metal attacks a broad range of targets in the microorganism. Over the past few years, nanotechnology has been used for many medical applications, including dental practice, with the development of silver nanoparticles (AgNPs) as a useful tool.^{2,3} It is known that smaller silver nanoparticles have more antimicrobial effectiveness than larger silver nanoparticles. The increased antimicrobial activity of the

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smaller nanoparticles is due to the fact that smaller particles penetrate more easily through cell membranes and cell walls, and that relative to larger nanoparticles, smaller particles have a greater surface area to volume ratio.^{1–5} Thus, silver compounds as nanoparticles have been widely used as dental restorative material, endodontic retrofill cements, dental implants, and as a caries inhibitory solution.^{2,3} Li et al⁴ found that incorporation of AgNPs into Scotchbond Multi-Purpose bonding system (3M, St Paul, MN, USA) exerts potent antibacterial activity without adversely affecting the microtensile dentin bond strength and fibroblast viability. Despite the effectiveness that AgNPs have shown in dental practice, the toxicity of AgNPs has not yet been fully evaluated.⁵

Fluoride (F) in various forms is the most popular active ingredient in dental products to prevent cavities as well as a commonly used agent in dental practice. On the other hand, F has been shown to be toxic, not only to the skeletal system, but also to the soft tissues by multiple mechanisms, such as enzyme activity inhibition, reactive oxygen and nitrogen species generation, impairment of the antioxidant defense system, induction of inflammation, and apoptosis.^{6–10}

As AgNPs and F may be used concomitantly in dental practice, we studied the effects of co-exposure to these xenobiotics on human gingival fibroblasts. It is known that xenobiotic interaction may increase or decrease the toxic effect of one or many substances.^{10–12} Therefore, the effects of AgNPs and F co-exposure on cell viability, oxidative stress, and inflammation were evaluated *in vitro*.

Materials and methods

Reagents

The silver nanoparticles (nanoparticle water dispersion: 2 nm, 2,000 ppm, colorless, and transparent) were obtained from US Research Nanomaterials (Houston, TX, USA). The fluoride solution was prepared using sodium fluoride (Sigma Aldrich, St Louis, MO, USA).

Cell culture

A CRL-2014 cell line was obtained from the American Type Culture Collection (ATCC-HBT-55) and maintained as a monolayer culture in T-75 cm² tissue culture flasks. The cells were grown in Dulbecco's Modified Eagle's Medium (Sigma Aldrich), a high glucose medium (4.5 g/L) containing sodium pyruvate (110 mg/L), and supplemented with 10% fetal bovine serum, 6 µg/mL penicillin-G, and 10 µg/mL streptomycin. Cells were maintained at 37°C in a humidified atmosphere of 95% O₂, 5% CO₂. When conflu-

ent, cells were detached enzymatically with trypsin-EDTA and sub-cultured into a new cell culture flask. The medium was replaced every 2 days.

Cell exposure to F or/and AgNPs

The concentrations of AgNPs and F used in the experiments were carefully chosen according to the results obtained from preliminary experiments and literature data.^{10,13,14} AgNPs ranging 1.5–5 nm are mostly used in dental practice. Therefore, we have carried out all experiments using 2 nm AgNPs.

The concentrations of AgNPs and F were selected based on the results of preliminary experiments. We found that F < 1 mM and AgNPs < 1.5 µg/mL did not induce significant effects in gingival fibroblasts. Therefore, the experiments were performed by combining the subthreshold concentrations of AgNPs and F to evaluate if co-exposure could lead to an enhanced toxicity effect.

The CRL-2014 cell line was treated with F (0.5; 1; 1.5 mM – 9.5; 19; 28.5 ppm, respectively) and/or AgNPs (0.5; 1.5; 2.5; 3.5 µg/mL) for 24 hours, except for Western blotting and PCR experiments (1 hour and 8 hours incubation time, respectively).

Fluoride was dissolved in serum-free culture medium. Test solutions of AgNPs were also prepared in serum-free medium. Immediately before use, NP solutions were vortexed for 1 minute to prevent aggregation following the manufacturer's instructions. Controls were prepared with an equivalent volume of media with neither F nor AgNPs.

Cellular morphology

The morphology of CRL-2014 cells in the presence of xenobiotics was visualized by phase-contrast microscopy (ALTRA20 microscopy and Cell[^]A Acquisition software; Olympus, Tokyo, Japan).

Cellular uptake of AgNPs

Ultrathin sections of cells were analyzed using transmission electron microscopy (TEM) to reveal the uptake and distribution of NPs. Briefly, cells seeded into 12-well plates (1.5 × 10⁶ cells/well) were treated with AgNPs (1.5 µg/mL) for 24 hours. At the end of the incubation period, wells were washed with PBS to remove the excess of unbound NPs. Samples were first fixed in 2.5% glutaraldehyde in 0.1 M Sørensen's phosphate buffer (Riedel-deHaën AG, Seelze, Germany) for 2 hours at room temperature and then fixed in 1% osmium tetroxide (Oxkem Limited, Reading, UK) in Sørensen's phosphate buffer for 1 hour at room temperature. Subsequently,

the specimens were dehydrated in a graded ethanol series, at 30%, 50%, and 70%. To enhance the contrast en bloc, staining with 2% uranyl acetate (Agar Scientific, Essex, UK) in 70% ethanol was performed and then the dehydration was continued with 90% and 100% ethanol. When dehydration was complete, samples were transferred from 100% ethanol to acetone, from acetone to a mixture of one part acetone and one part epoxy resin (TAAB-Epon; Agar Scientific, Essex, UK) for 1 hour. To complete the resin infiltration, the samples were placed in 100% resin at 37°C for 2 hours. Finally samples were embedded in resin, and stored at 60°C for 24 hours until polymerization was complete.

For orientation purposes, sections from each sample were cut at 1 μ m, stained with toluidine blue, and examined by light microscopy (Leica DMLB; Leica Microsystems, Wetzlar, Germany).

From these survey sections, areas of interest were identified and ultrathin (80 nm) sections were obtained using a Leica EM UC6 ultramicrotome (Leica Microsystems). These sections were collected on 200-mesh thin bar copper grids, stained with uranyl acetate and lead citrate (Agar Scientific, Essex, UK), and examined by TEM (Tecnai G2 12 BioTWIN; FEI Company, Hillsboro, OR, USA) using an accelerating voltage of 120 kV. Images were recorded with a MagaView III camera (SIS, Münster, Germany).

Intracellular ROS generation

Briefly, the cells were seeded into 12-well plates at a concentration of 1×10^6 cells/well; 24 hours after seeding, cells were exposed to different concentrations of F and/or AgNPs in serum-free media as above, then collected and incubated with 40 μ M of carboxy- H_2 DCFDA (Molecular Probes, Eugene, OR, USA) for 15 minutes to assess ROS-mediated oxidation of DCFDA to the fluorescent compound 2,7-dichlorofluorescein DCF. The fluorescence of oxidized DCF was measured on a plate reader (FLUOstar OPTIMA; BMG Labtech, Aylesbury, UK) at excitation and emission wavelengths of 485 and 530 nm, respectively.

Lipid peroxidation

CRL-2014 cells were plated into 24-well plates at a density of 1.5×10^6 cells per well in complete medium. Pre-confluent cells were then exposed to different concentrations of F and/or AgNPs in serum-free medium as above for 24 hours at 37°C. Malondialdehyde (MDA), a measure of lipid peroxidation, was measured using an Oxiselect™ TBARS Assay kit (Cell Biolabs Inc., Cambridge, UK) following the manufacturer's

instructions. Fluorescent measurements were recorded on a plate reader (FLUOstar OPTIMA) at excitation and emission wavelengths of 485 and 530 nm, respectively. The concentration of MDA in test samples was calculated using MDA standards as the reference. The concentration of MDA in F and/or AgNPs-treated cells is presented as the fold increase of MDA production over the control (untreated cells).

Total antioxidant capacity

For determining cellular total antioxidant capacity, post-treatment cells were washed with PBS and suspended in 200 μ L of ice-cold lysis buffer and sonicated. The lysate was centrifuged at 10,000 rpm for 10 minutes, and the protein concentration of the supernatant fraction was determined by the Bradford method.¹³ The Trolox equivalent antioxidant activity was measured by assessing the ability of hydrogen-donating antioxidants to scavenge the radical cation generated by 2,2'-azino-bis(3-ethylbenzothiazoline)-6-sulfonic acid. Absorbance was recorded at 570 nm (FLUOstar, OPTIMA).

Cell viability was measured by MTT assay

CRL cells were seeded in triplicate at a density of 10^4 cells/100 μ L of cell culture medium into a 96-well. The next day, CRL-2014 cells were exposed to different concentrations of F and/or AgNPs as above for 24 hours. This assay evaluates mitochondrial activity (assesses cell growth and cell death) and is performed by adding a premixed optimized dye solution to culture wells. Absorbance was recorded at 570 nm (FLUOstar OPTIMA). Results from the treatment groups were calculated as a percentage of control values (unexposed cells) according to the following equation: % cytotoxicity = (experimental absorbance [abs] 570 nm of exposed cells/abs 570 nm of unexposed cells) $\times 100$.

Flow cytometry analysis of apoptosis

The Annexin V-FITC apoptosis detection kit (BD Pharmingen, San Jose, CA, USA) was used to detect apoptosis according to the manufacturer's instructions. Briefly, cells (5×10^6) were seeded into 25 cm^2 tissue culture flasks; 24 hours after seeding, cells were treated as described, then collected and washed twice with cold PBS. Five microliters of Annexin V and 2.5 μ L of PI (propidium iodide) were added to the cells, which were resuspended in binding buffer. The cells were gently shaken while incubating for 15 minutes at room temperature in the dark. Cells were further diluted with $1 \times$ binding buffer and analyzed using a BD FACSArray (BD Biosciences, San Jose, CA, USA). Ten thousand specific events were analyzed.

Western blotting

Western blotting was used in order to investigate the mitogen-activated protein kinases (MAPK) pathways. Briefly, CRL-2014 cells were exposed to 1.5 $\mu\text{g}/\text{mL}$ AgNPs and/or 1 mM F for 60 minutes. Afterwards, conditioned media were discharged and attached cells rinsed (phosphate buffered saline [PBS] $\times 3$), detached, and homogenized. Following electrophoresis, proteins were transferred onto nitrocellulose membrane (Protran, Schleicher and Schuell; Dassel, Germany) and detected using rabbit anti-phosphorylated p38, p42/44, and JNK antibodies (Cell Signaling, Hitchin, UK). When appropriate, blots were stripped and analyzed for β -actin (Sigma Aldrich) as an internal control and for total p38, p42/44, and JNK (Cell Signaling).

Real-time quantitative polymerase chain reaction (PCR)

Cells were treated with different concentrations of F and/or AgNPs for 8 hours. Controls without F and AgNPs were also prepared. Following incubation, the cells were washed twice with PBS. DNA-free RNA was isolated using the Ambion[®] RiboPure[™] kit (Huntingdon, UK) according to the supplier's recommendations. Thereafter, isolated RNA in each sample was quantified using a NanoDrop[®] ND-1000 spectrophotometer (Fisher Scientific, Dublin, Ireland). An equal quantity of total DNA-free RNA from each sample was reverse-transcribed using a high-capacity cDNA reverse transcription kit (Applied Biosystems, Warrington, UK). Reverse transcription was performed using a Realplex² Mastercycler (Eppendorf, Cambridge, UK). Finally, an equal quantity of total complementary DNA (cDNA) of each sample was used to perform real-time quantitative PCR in duplicate, with predesigned Applied Biosystems TaqMan[®] gene expression assays for MMP-9, IL-8, IL-6, BAX, and Bcl-2 along with Applied Biosystems TaqMan[®] Universal PCR Master Mix. In all experiments, 18S ribosomal RNA (rRNA) was used as an internal control. Real-time quantitative PCR was performed using a Realplex² Mastercycler. The expression of each gene within each sample was normalized against 18S rRNA expression and expressed relative to the control sample using the formula $2^{-(\Delta\Delta C_t)}$, in which $\Delta\Delta C_t = (C_t \text{ mRNA} - C_t \text{ 18S rRNA}) \text{ sample} - (C_t \text{ mRNA} - C_t \text{ 18S rRNA}) \text{ control}$.

Zymography

Briefly, conditioned media of cells treated as above were collected after 24 hours and protein concentration assessed by the Bradford method.¹⁵ Briefly, samples were subjected to 10% sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) with copolymerized gelatin (0.2%), (Sigma

Chemical Co, St Louis, MO, USA). After electrophoresis, the gels were washed with 2% Triton X-100 (two times, 20 minutes each), and then incubated in development buffer (50 mmol/L Tris HCl, 200 mmol/L NaCl, 10 mmol/L CaCl_2 , and 1 mmol/L ZnCl_2 , pH = 7.5) at 37°C overnight. A conditioned medium of HT-1080 human fibrosarcoma cells was used as internal control. After incubation, gels were fixed and stained in 40% methanol, 10% acetic acid, and 0.1% (wt/v) Coomassie Brilliant Blue for 1 hour and then de-stained.

Statistical analysis

All data are presented as the mean \pm standard error of three independent experiments. Statistical analysis of the mean difference between multiple groups was determined by one-way analysis of variance (ANOVA) followed by Tukey–Kramer multiple comparison tests, and between two groups by two-tailed Student's *t*-tests. A *P*-value < 0.05 was considered to be statistically significant. All statistical analyses were performed using GraphPad Prism (v5.00 for Windows; GraphPad Software, San Diego, CA, USA).

Results

Uptake of AgNPs by CRL-2014 cells

TEM was used to study AgNPs-gingival fibroblast cell interactions and uptake. After 24 hours of incubation, AgNPs both

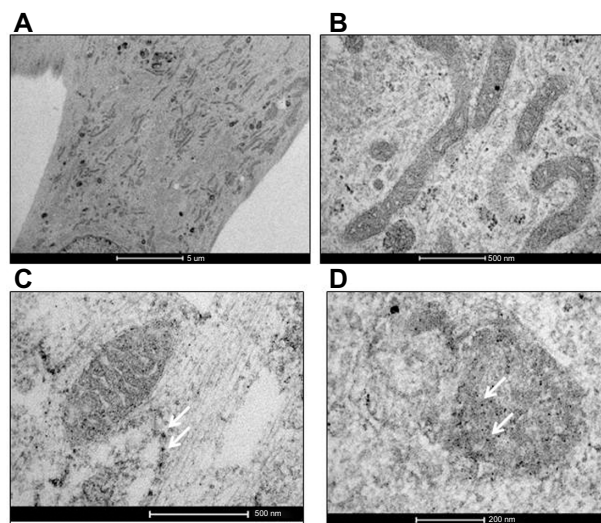


Figure 1 Uptake of AgNPs by CRL-2014 cells. (A) Low- and (B) high-power TEM images of untreated cells (controls); (C) low- and (D) high-power TEM images of treated cells. As shown by TEM, after exposure of cells to AgNPs, NPs were internalized and distributed within the cell (white arrows) (C). AgNPs were mainly found in the mitochondria (white arrows) (D). Some NPs agglomerates are found (C and D).

Notes: Original magnification in (A) is 4,200 \times , whereas original magnification in (B) is 43,000 \times ; original magnification in (C) is 43,000 \times , whereas original magnification in (D) is 87,000 \times .

Abbreviations: AgNPs, silver nanoparticles; NPs, nanoparticles; TEM, transmission electron microscopy.

in the presence and absence of F were taken up, internalized, and distributed into CRL-2014 cells (Figure 1). They were mainly found in the mitochondria forming agglomerates (Figure 1).

AgNPs and F induce ROS generation and lipid peroxidation in CRL-2014 cells

As AgNPs were found in the mitochondria, we next studied endogenous ROS generation. We found that both AgNPs and F were able to induce ROS generation in a concentration-dependent manner. This effect was enhanced when cells were co-exposed to AgNPs and F (Figure 2A). Lipid peroxidation was also studied, as this degradation process could be initiated by ROS generation. Indeed, both AgNPs and F induced MDA production, an effect that was even greater when cells were co-exposed to AgNPs and F (Figure 2B).

AgNPs and F decrease total antioxidant capacity in CRL-2014 cells

As increased intracellular production of ROS is likely to impair antioxidant defenses, we then studied the total antioxidant capacity (TAC) of human gingival fibroblasts. We found that both AgNPs and F were able to reduce TAC, an effect that was enhanced when CRL-2014 cells were co-exposed to AgNPs and F (Figure 2C).

AgNPs and F co-exposure decreases viability of CRL-2014 cells

We then investigated if increased oxidative stress was associated with cell damage. AgNPs and F were found to reduce cell viability in a concentration-dependent manner (Figure 3A). This effect was enhanced when CRL-2014 cells were co-exposed to AgNPs and F (Figure 3A). Furthermore, we studied if decreased cellular viability could be linked

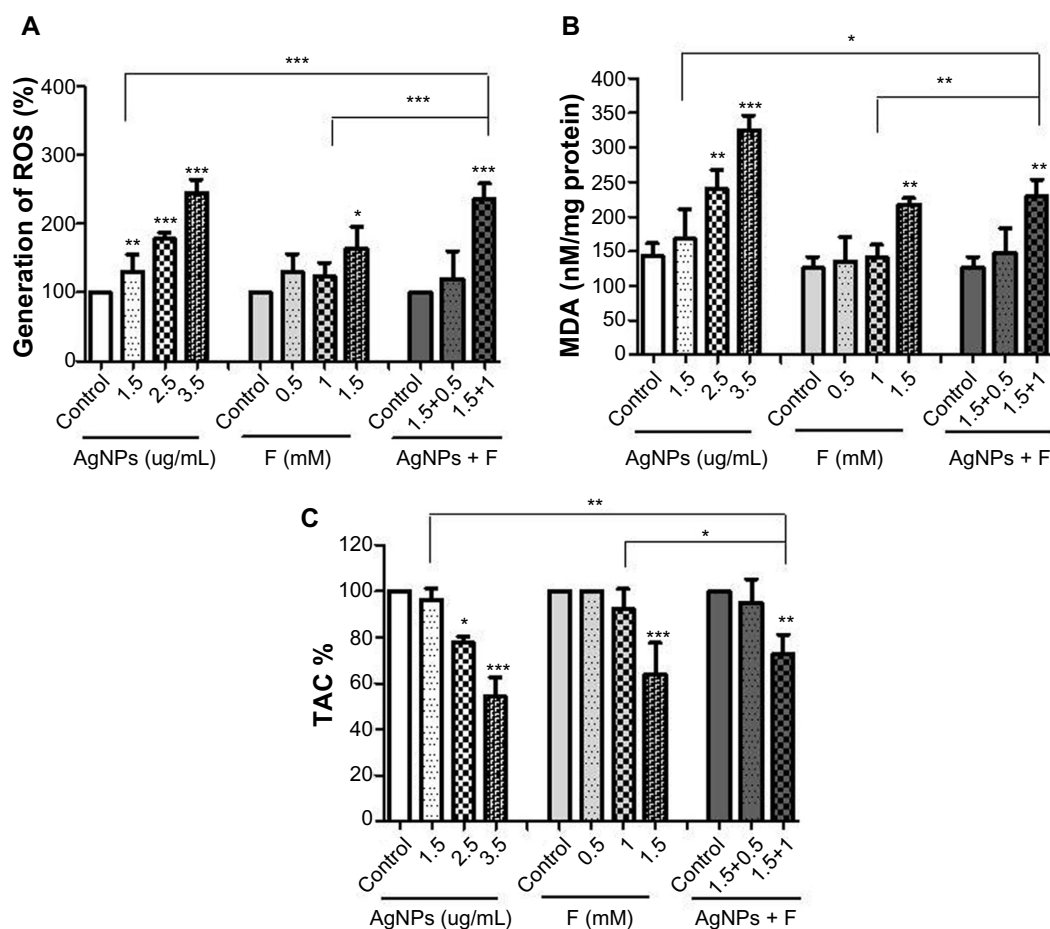


Figure 2 Induction of ROS and MDA along with reduction of TAC by AgNPs and F co-exposure in CRL-2014 cells. (A) CRL-2014 cells exposed simultaneously to both AgNPs and F produced significantly higher levels of ROS compared to control cells and AgNPs- and F-treated cells, respectively; (B) CRL-2014 cells exposed simultaneously to both AgNPs and F produced significantly higher levels of MDA compared to control cells and AgNPs- and F-treated cells, respectively; (C) CRL-2014 cells exposed simultaneously to both AgNPs and F produced significantly lower levels of TAC compared to control cells and AgNPs- and F-treated cells, respectively.

Notes: * $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$.

Abbreviations: AgNPs, silver nanoparticles; F, fluoride; MDA, malondialdehyde; ROS, reactive oxygen species; TAC, total antioxidant capacity.

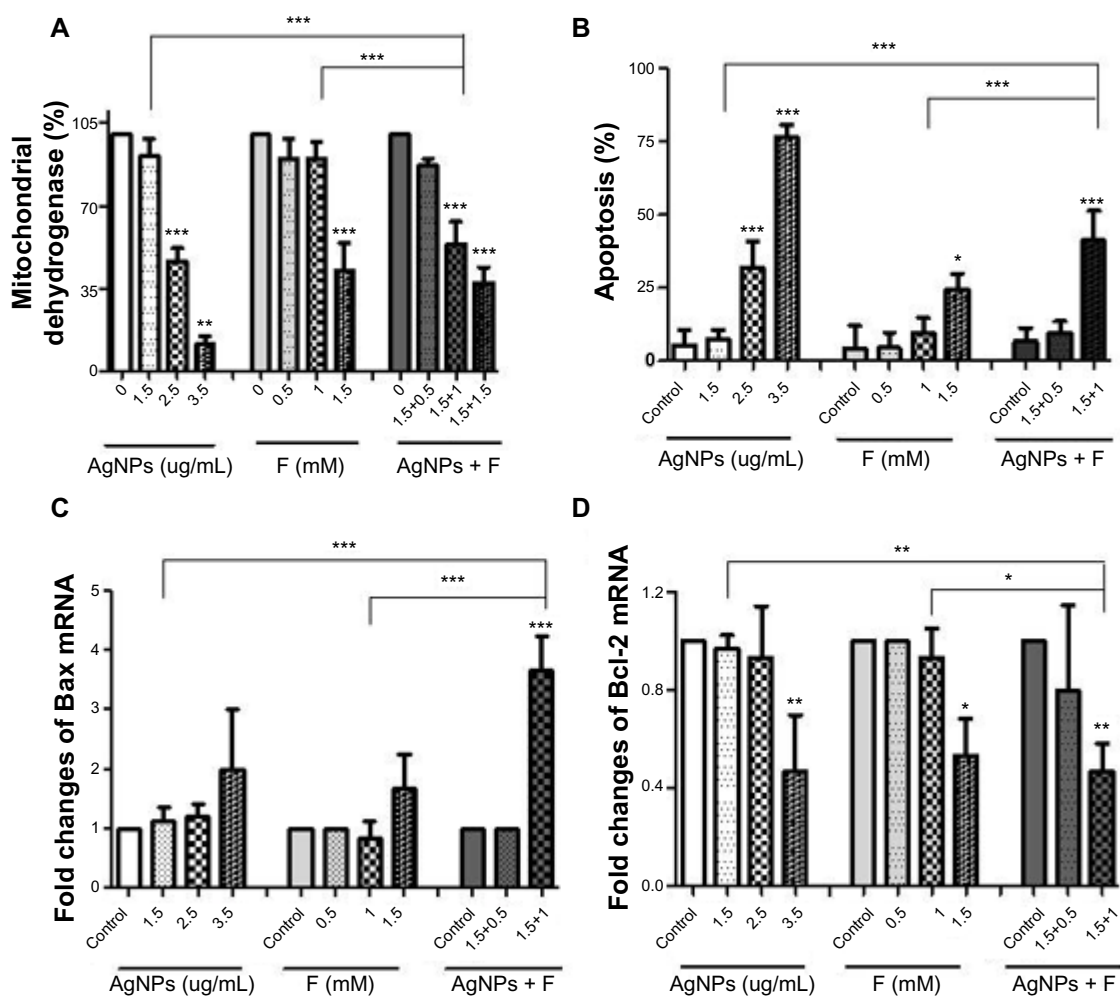


Figure 3 Effect of AgNPs and F co-exposure on cell survival. **(A)** Concentration-dependent toxicity of AgNPs and F on CRL-2014 cells. Cell viability was determined by the MTT assay. Co-exposure to both AgNPs and F significantly reduced cell survival compared to control cells and AgNPs- and F-treated cells, respectively; **(B)** increasing apoptotic effect of AgNPs and F on CRL-2014 cells. Apoptosis was determined by flow cytometry. Co-exposure to both AgNPs and F significantly increased apoptosis compared to control cells and AgNPs- and F-treated cells, respectively; **(C)** increasing levels of pro-apoptotic BAX gene. CRL-2014 cells exposed simultaneously to both AgNPs and F showed significantly higher level of Bax compared to controls and AgNPs- and F-treated cells, respectively; **(D)** effect of AgNPs and F on the expression level of anti-apoptotic Bcl-2 gene. Co-exposure to both AgNPs and F significantly reduced the level of Bcl-2 compared to control cells and AgNPs- and F-treated cells, respectively. **Notes:** * $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$.

Abbreviations: AgNPs, silver nanoparticles; F, fluoride; mRNA, messenger RNA.

to apoptosis. We found that co-exposure to AgNPs and F did increase apoptosis significantly in CRL-2014 cells (Figure 3B). Indeed, the pro-apoptotic BAX gene was upregulated (Figure 3C) in contrast to the anti-apoptotic Bcl-2 gene (Figure 3D).

We also used phase-contrast microscopy to evaluate the effects of AgNPs and F on cellular morphology. No significant morphological changes were detected upon incubation of cells with AgNPs or F separately (Figure 4A and B). However, when cells were co-exposed to AgNPs and F, the corresponding phase-contrast micrographs revealed cell shortening and irregular cell shapes, which could indicate cell toxicity (Figure 4C and D).

Activation of MAPK pathways and upregulation of inflammatory cytokines

Having implicated the release of ROS in AgNP-F-induced fibroblast cell damage and apoptosis, we studied signaling pathways that may mediate these reactions, such as MAPK proteins, pro-inflammatory cytokines (IL-6 and IL-8), and a downstream effector, matrix metalloproteinase-9 (MMP-9). Western blot analysis showed that F promoted phosphorylation of p42/44 MAPK, an effect that was enhanced when AgNPs were added (Figure 5). AgNPs and F separately were able to upregulate IL-6, IL-8, and MMP-9 at gene level. In addition, we found that upregulation of gene expression was higher when gingival fibroblasts were co-exposed to

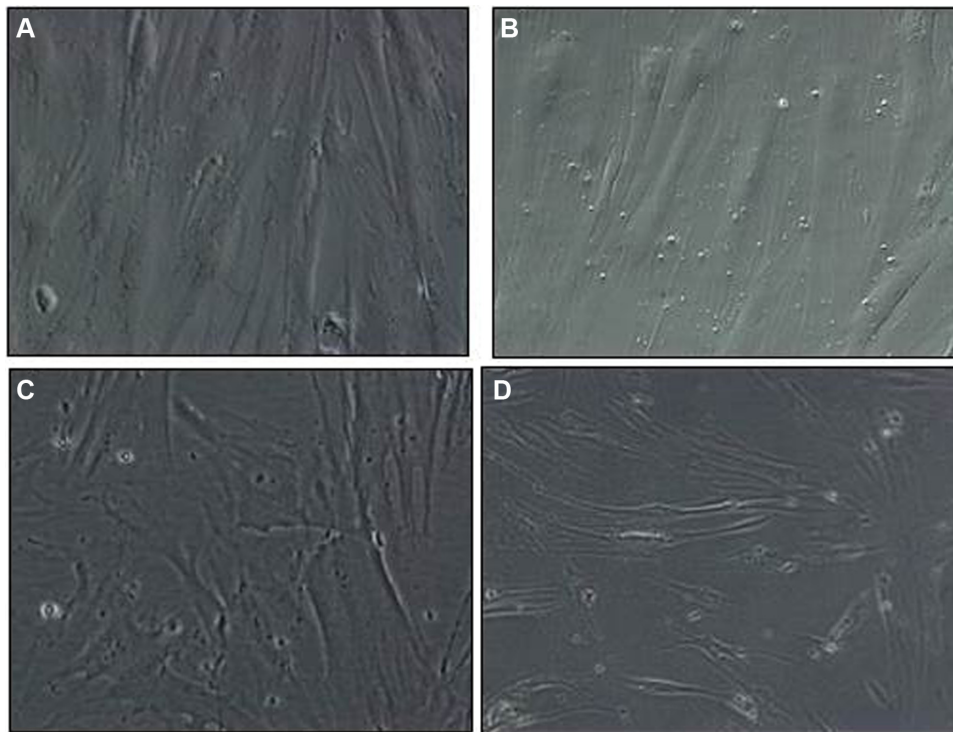


Figure 4 Morphological changes of CRL-2014 exposed to AgNPs and F by phase-contrast microscopy. Cells co-exposed to AgNPs (1.5 $\mu\text{g}/\text{mL}$) and F (1 mM) for 12 hours (C) and 24 hours (D) showed irregular cell shapes and cell shortening compared to AgNPs-treated cells (A) and F-treated cells (B).

Note: Original magnification was 20 \times .

Abbreviations: AgNPs, silver nanoparticles; F, fluoride.

AgNPs and F (Figure 5). Finally, co-exposure of CRL-2014 to AgNPs and F resulted in a significant release of MMP-9 into the cell culture supernatant (Figure 6).

Discussion

The major novel finding of this study is that AgNPs and F co-exposure of gingival fibroblasts results in enhanced oxidative stress, triggering a cascade of inflammatory reactions that lead to apoptosis and impairment of cell viability.

Fluoride has been widely used in dentistry because it is an effective caries prophylactic agent. Nowadays, AgNPs are also being introduced as therapeutic antimicrobial agents in dental practice, although some concerns have emerged regarding their toxicological effects.^{5,16}

Indeed, while the toxicological effects of both F and AgNPs had been previously investigated, our study is the first to analyze the outcome of pharmacological/toxicological interactions of a combination of two xenobiotics. Although positive interactions (additive or synergistic effects) were most likely to take place considering the individual profiles of the interacting agents, the opposite outcome, ie, antagonism had to be also considered. Our results clearly indicate that when used in combination,

AgNPs and F may cause increased gingival cytotoxicity. Indeed, we first showed that AgNPs were taken up by human gingival fibroblasts in the presence/absence of F. As AgNPs were internalized and mainly found in mitochondria that are vulnerable to oxidative stress,^{17,18} we studied the generation of ROS. Mitochondria dysfunction/damage by AgNPs has been previously found in human lung fibroblasts and liver cells.^{16,18} In physiology, ROS are produced during metabolic reactions and as this generation is potentially cytotoxic, they are generally neutralized by cell antioxidant systems (TAC). However, excessive ROS production may overwhelm the capacity of TAC leading to cellular dysfunction, and thus contributing to the pathogenesis of various human diseases.¹⁷

The results of studies examining the effects of AgNPs on TAC are not clear. Some researchers reported that AgNPs can act as free radical scavengers.^{19,20} In contrast, Chairuankitti et al²¹ found that the toxic effects of AgNPs on A549 cells are mediated via both ROS-dependent and ROS-independent mechanisms. Moreover, AshaRani et al¹⁶ showed that AgNPs caused mitochondrial dysfunction, namely the induction of ROS, which in turn set off DNA damage and chromosomal aberrations in normal human lung fibroblast cells.

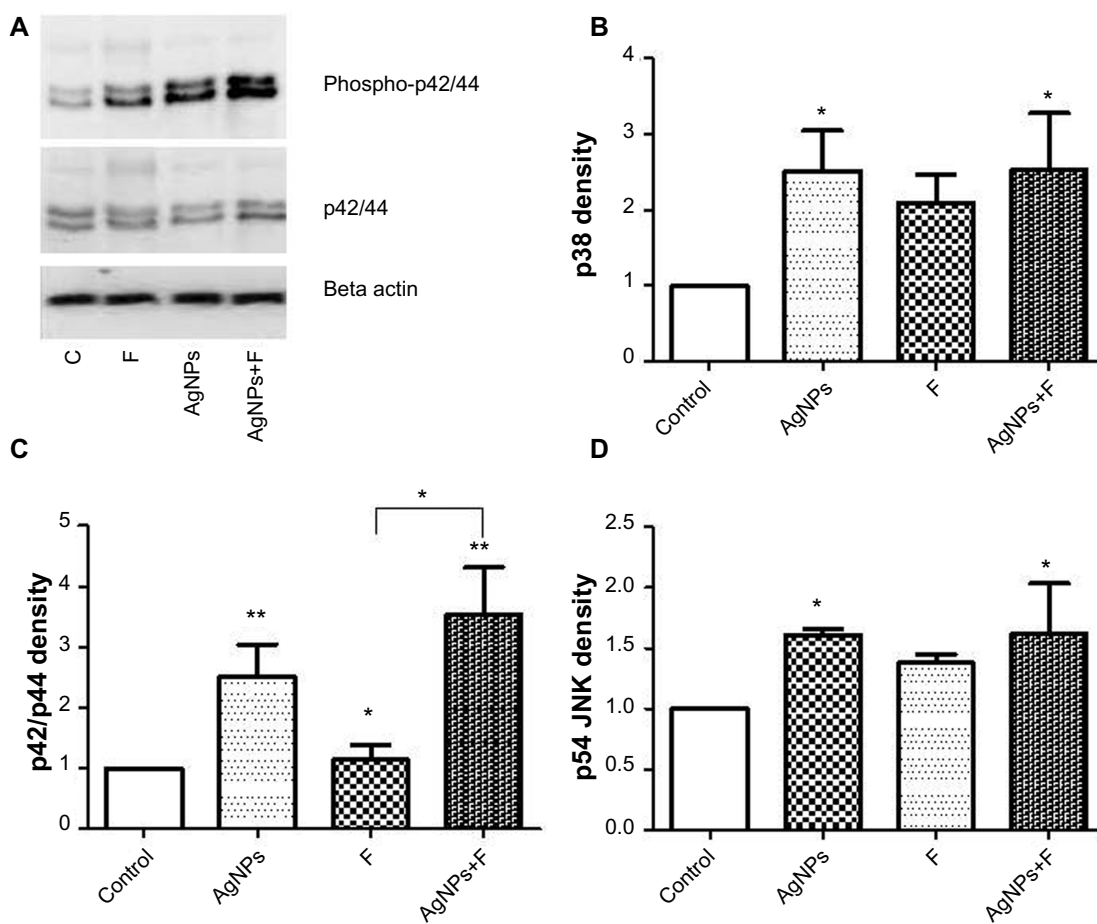


Figure 5 Effects of AgNPs and F on MAPK phosphorylation. (A) Representative Western blotting. Normalized densitometric values of phosphorylated p38 MAPK (B) p42/p44 MAPK (C), and JNK MAPK (D).

Notes: F was only able to induce p42/44 phosphorylation (* $P < 0.05$ vs control). However, when AgNPs were also added, phosphorylation of p42/44 MAPK was enhanced (* $P < 0.05$ vs F; ** $P < 0.01$ vs control).

Abbreviations: AgNPs, silver nanoparticles; F, fluoride; MAPK, mitogen-activated protein kinases; vs, versus.

In addition, some studies have demonstrated that free radicals play a key role in fluoride-induced toxicity.^{10,11,22} In the current study, we found that both AgNPs and F were able to induce ROS generation, but interestingly, this effect was significantly higher when CRL-2014 cells were exposed to both AgNPs and F. These data correlated well with the unfavorable effects of AgNPs and F on antioxidant cell defenses. Indeed, we found a significant reduction of TAC when cells were exposed to various concentrations of AgNPs and F. These results are consistent with our previous studies *in vivo*.¹¹

The cellular impact of unchecked increases in ROS levels is very significant. At high levels, ROS can lead to impaired physiological function through cellular damage of DNA, proteins, phospholipids, and other macromolecules.¹⁷ The cell membrane is one of the most susceptible sites to ROS damage. Lipid peroxidation can permanently impair fluidity and elasticity of the membrane, which can lead to cell rupture. Choi et al²³ demonstrated an increase in the levels of MDA,

a byproduct of cellular lipid peroxidation, in the liver of adult zebrafish after treatment with AgNPs. Moreover, F-induced lipid peroxidation was found in different cell culture, animal model, and epidemiological studies.^{11,24} Indeed, significant lipid peroxidation has also been reported in erythrocytes from children and adults suffering from endemic fluorosis.²⁵ In the present study we found that the levels of MDA were increased in human gingival fibroblasts treated with AgNPs and F. This effect was enhanced during co-exposure of cells to both xenobiotics. Again, these results correlated well with the overproduction of ROS. As ROS generation and lipid peroxidation could lead to cell death, we next studied cell viability. When cells were incubated with AgNPs (1.5 $\mu\text{g}/\text{mL}$) and F (1 mM) separately, no significant effects on cell viability were found. However, when cells were allowed to interact with the two xenobiotics at the indicated concentrations, cell viability was significantly reduced. Kleinsasser et al⁹ have previously found that F causes mucosal cell damage

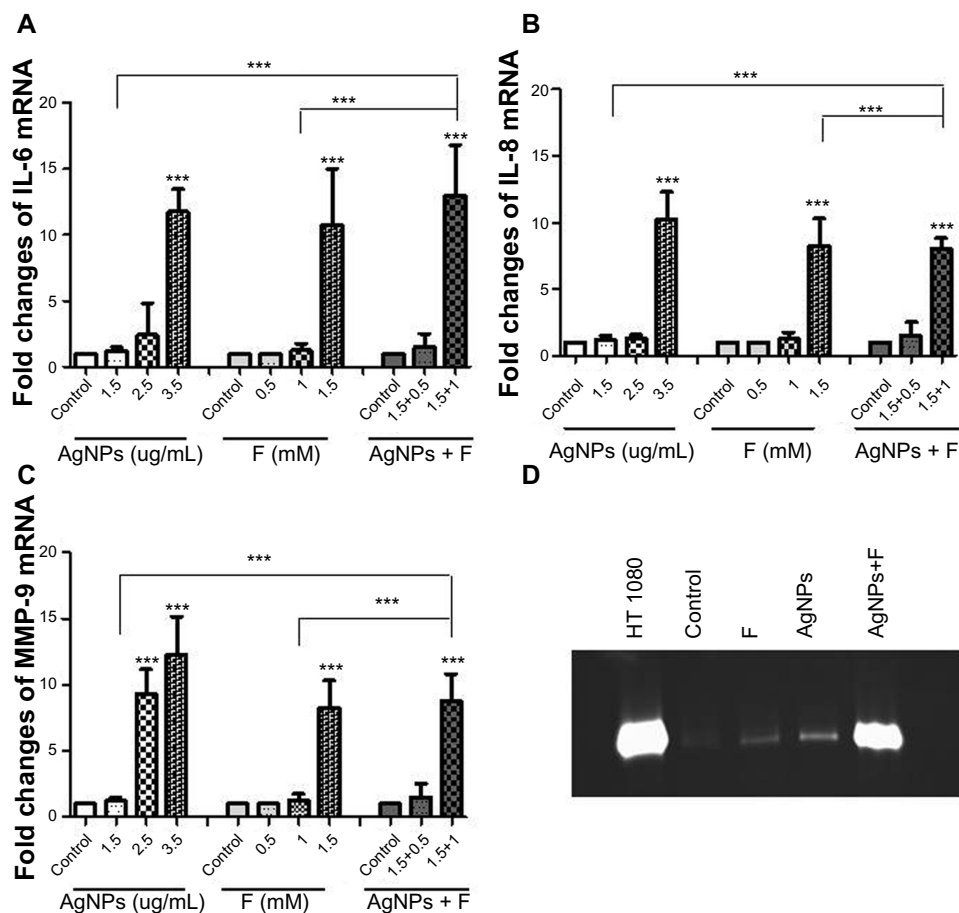


Figure 6 Effect of AgNPs and F on the expression level of inflammatory markers in CRL-2014 cells. The gene expression of IL-6 (A), IL-8 (B), and MMP-9 (C) was investigated. CRL-2014 cells exposed simultaneously to both AgNPs and F produced significantly higher levels of IL-6, IL-8, and MMP-9 compared to control cells and AgNPs- and F-treated cells, respectively ($***P < 0.001$); (D) representative zymogram showing the gelatinolytic activity of MMP-9 in the conditioned media from CRL-2014 exposed to AgNPs and F as in (A).

Note: HT1080 conditioned media was run as an internal control.

Abbreviations: AgNPs, silver nanoparticles; F, fluoride; mRNA, messenger RNA; MMP, matrix metalloproteinase-9.

in a concentration-dependent manner. Tissue necrosis after subgingival irrigation with a solution of F has also been demonstrated.⁸ Finally, AgNPs <20 nm increased cytotoxicity in human periodontal fibroblasts in a concentration- and time-dependent manner.⁵

This oxidant/antioxidant imbalance has previously been documented as an apoptotic mechanism of AgNP-mediated cytotoxicity.²⁶ Therefore, we next studied whether AgNP and F could induce apoptosis. Several studies have shown that millimolar levels of F can induce apoptosis in many cell types, including hepatic cells, epithelial lung cells, human leukemia HL-60 cells, and ameloblast-lineage cells.²⁷⁻³⁰ We have also shown that F increases apoptosis in hippocampal and osteoblast-like cells.¹⁰ However, other mechanisms can be involved in F-mediated cell death, such as endoplasmic reticulum stress.³¹ Furthermore, AgNPs are able to induce cell death through apoptotic mechanisms in cultured cells and animal tissues.^{18,32,33} In the present study, we have found

that co-exposure to AgNPs and F resulted in a significant increase in cell apoptosis.

As ROS generation could trigger the activation of different pathways involved in inflammation, we next studied the MAPK pathways. Generally, increased ROS production in a cell leads to the activation of p42/p44, JNKs, or p38 MAPK.^{34,35} Diverse cellular functions, ranging from cell survival to cell death, are regulated by MAPK. P42/p44 activation seems to promote survival in most cell types but it has also been reported that induction of apoptosis may be mediated via p42/p44.^{35,36} The importance of the p42/p44 MAPK pathway in mediating apoptosis has also been recently observed in other models of oxidant-induced apoptosis, including asbestosis, UV irradiation, and tumor necrosis factor.³⁴⁻³⁶ However, the other MAPK (JNK and p38) have been implicated primarily in the induction of apoptosis and inflammation after exposure to different agents.^{34,37} It has been found that AgNPs could activate p38 MAPK and JNK

in vitro.^{18,38} In addition, F could also induce the activation of JNK and p42/44 MAPK in osteoclast cells.^{6,7} Our studies show that both AgNPs and F significantly increased the phosphorylation of p42/p44 and this effect is enhanced during co-exposure to both xenobiotics. Therefore, the activation of different MAPK by AgNPs and F could be cell dependent. Finally, this activation could lead to the upregulation of different proinflammatory cytokines. Therefore, we studied whether AgNPs and F were able to induce the upregulation of IL-6, IL-8, and MMP-9. Interestingly, the upregulation of both cytokines and MMP-9 was found when CRL-2014 cells were exposed to both AgNPs and F; Vandana and Reddy³⁹ suggested that there is a strong association of periodontal disease in high-fluoride areas. It has also been found that high concentrations of F resulted in the upregulation of MMP-2 and MMP-9 in pro-osteoblast cells.³⁰ However, it has been also suggested that an oral hygiene regimen based on amine/stannous fluoride-containing toothpaste is effective in reducing plaque-associated gingivitis and gingival inflammation.⁴⁰ Of note, AgNPs cause inflammation in other cell types, such as liver cells both in vivo and in vitro.^{41,42}

In conclusion, we have shown that co-exposure of gingival fibroblasts to AgNPs and F amplifies the noxious effects of the individual xenobiotics. The mechanism of this action may depend on increased generation of ROS or lipid peroxidation along with a decrease in TAC that could lead to cell death and inflammation. Further studies are warranted to establish the safety profile of these agents for further clinical applications.

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Disclosure

CM is an SFI Stokes lecturer. The authors have no other conflicts of interest to report.

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