

Apigenin and Wogonin Regulate Epidermal Growth Factor Receptor Signaling Pathway Involved in MUC5AC Mucin Gene Expression and Production from Cultured Airway Epithelial Cells

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Background: We investigated whether wogonin and apigenin significantly affect the epidermal growth factor receptor (EGFR) signaling pathway involved in MUC5AC mucin gene expression, and production from cultured airway epithelial cells; this was based on our previous report that apigenin and wogonin suppressed MUC5AC mucin gene expression and production from human airway epithelial cells.

Methods: Confluent NCI-H292 cells were pretreated with wogonin or apigenin for 15 minutes or 24 hours and then stimulated with epidermal growth factor (EGF) for 24 hours or the indicated periods.

Results: We found that incubation of NCI-H292 cells with wogonin or apigenin inhibited the phosphorylation of EGFR. The downstream signals of EGFR such as phosphorylation of MEK1/2 and ERK1/2 were also inhibited by wogonin or apigenin.

Conclusion: The results suggest that wogonin and apigenin inhibits EGFR signaling pathway, which may explain how they inhibit MUC5AC mucin gene expression and production induced by EGF.

Keywords: Receptor; Epidermal Growth Factor; Mucins; Wogonin; Apigenin

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Introduction

Pulmonary mucus plays a pivotal role in defensive action against invading pathogenic microorganisms, chemicals and particles. Mucins are high molecular weight glycoproteins present in the airway mucus and produced by goblet cells in the surface epithelium as well as mucous cells in the submucosal gland. However, hypersecretion of airway mucus is one of the major symptoms associated with severe pulmonary diseases including asthma, chronic bronchitis, cystic fibrosis and bronchiectasis^{1,2}. Therefore, we suggest it is valuable to find the potential activity of regulating the excessive mucin production and/or secretion by the compounds derived from various medicinal plants. Flavonoids are a group of phenolic

phytochemicals, common in various medicinal plants, vegetables and fruits³. We have tried to investigate the possible activities of some flavonoids on mucin production (secretion) from airway epithelial cells. As a result of our trial, we previously reported that several compounds affected mucin production and/or secretion from airway epithelial cells⁴⁻⁷. According to folk medicine, *Scutellaria baicalensis* Georgi has been used for regulating airway allergic and inflammatory diseases⁸. Also, two of their components—wogonin and apigenin—were reported to have various biological effects including antiinflammatory effect⁹⁻¹⁵. In our previous study, wogonin inhibited tumor necrosis factor α -induced MUC5AC mucin gene expression and production through the inactivation of nuclear factor- κ B signaling in airway epithelial cells¹⁶. Also, we demonstrated that wogonin or apigenin inhibited epidermal growth factor (EGF)- or phorbol myristate acetate-induced MUC5AC mucin production and gene expression¹⁷. Expression of epidermal growth factor receptor (EGFR) is increased in asthmatic human airways¹⁸. EGF stimulates MUC5AC mucin gene expression and protein synthesis via binding to EGFR in a human airway epithelial cell line¹⁹. However, to the best of our knowledge, no other studies on these two compounds—wogonin and apigenin—on EGFR signaling pathway involved in MUC5AC mucin gene expression and production in human airway epithelial cells have been carried out. Therefore, in this study, we checked whether wogonin or apigenin affects the phosphorylation of EGFR, MEK1/2 and ERK1/2 in NCI-H292 cells, a human pulmonary mucoepidermoid cell line, which are frequently used for the purpose of studying the signaling pathway involved in airway mucin production and gene expression²⁰⁻²².

Materials and Methods

1. Materials

All the chemicals and reagents used in this study including wogonin (purity: 98.0%) and apigenin (purity: 95.0%) were purchased from Sigma (St. Louis, MO, USA) unless otherwise specified. Anti-actin antibodies were purchased from Santa Cruz Biotechnology (Santa Cruz, CA, USA). EGFR, p-EGFR, p-MEK1/2, and p-ERK1/2 antibodies were purchased from Cell Signaling Technology Inc. (Danvers, MA, USA).

2. Cell culture

NCI-H292 cells, a human pulmonary mucoepidermoid carcinoma cell line, were purchased from the American Type Culture Collection (ATCC, Manassas, VA, USA) and cultured in RPMI 1640 supplemented with 10% fetal bovine serum (FBS) in the presence of penicillin (100 units/mL), streptomycin (100 μ g/mL), and HEPES (25 mM) at 37°C in a humidified

5% CO₂, 95% air water-jacketed incubator. For serum deprivation, at 60% confluence, cultures were washed twice with phosphate-buffered saline (PBS) and recultured in RPMI 1640 with 0.2% FBS for 24 hours.

3. Cell viability assay

NCI-H292 cells were seeded at a density of 2×10^5 /mL (0.1 mL/well) in 96-well microtiter plates and allowed to attach for 24 hours to keep the log phase growth at the time of drug treatment. Wogonin or apigenin was dissolved in dimethylsulfoxide (DMSO) and administered in RPMI 1640 supplemented with 10% FBS in the presence of penicillin (100 units/mL), streptomycin (100 μ g/mL), and HEPES (25 mM) (final concentrations of DMSO were under 0.5%). 0.5% DMSO alone did not affect the proliferation of cells. After incubation with the indicated drug concentrations for 72 hours, cell proliferation (viability) was determined using the sulforhodamine B (SRB) assay.

4. Treatment of cells with wogonin or apigenin

For measurement of mucin production, after 24 hours of serum deprivation, NCI-H292 cells were pretreated with varying concentrations of wogonin or apigenin for 30 minutes and treated with EGF (25 ng/mL) for 24 hours in serum-free RPMI 1640. Wogonin or apigenin was dissolved in DMSO and treated in culture medium (final concentrations of DMSO were 0.5%). The final pH values of these solutions were between 7.0 and 7.4. Culture medium and 0.5% DMSO did not affect mucin production from NCI-H292 cells. After 24 hours, cells were lysed with buffer solution containing 20 mM Tris, 0.5% NP-40, 250 mM NaCl, 3 mM EDTA, 3 mM EGTA, and protease inhibitor cocktail (Roche Diagnostics, Indianapolis, IN, USA) and collected to measure the production of MUC5AC protein (in 24-well culture plate). For western blot analysis, NCI-H292 cells (confluent in 150 mm culture dish) were pretreated with 20 μ M of wogonin or apigenin for 15 minutes or 24 hours and treated with EGF (25 ng/mL) for 24 hours or the indicated periods in serum-free RPMI 1640. Wogonin or apigenin was dissolved in DMSO and treated in culture medium (final concentrations of DMSO were 0.5%). Culture medium and 0.5% DMSO did not affect the phosphorylations of EGFR, MEK1/2, and ERK1/2 in NCI-H292 cells.

5. MUC5AC mucin analysis using enzyme-linked immunosorbent assay

MUC5AC protein was measured by using enzyme-linked immunosorbent assay. Cell lysates were prepared with PBS at 1:10 dilution, and 100 μ L of each sample was incubated at 42°C in a 96-well plate, until being dry. Plates were washed three times with PBS and blocked with 2% bovine serum

albumin for 1 hour at room temperature. Plates were again washed three times with PBS and then incubated with 100 μ L of 45M1, a mouse monoclonal MUC5AC antibody (1:200, NeoMarkers, Fremont, CA, USA) which was diluted with PBS containing 0.05% Tween 20 and dispensed into each well. After 1 hour, the wells were washed three times with PBS, and 100 μ L of horseradish peroxidase-goat anti-mouse IgG conjugate (1:3,000) was dispensed into each well. After 1 hour, plates were washed three times with PBS. Color reaction was developed with 3,3',5,5'-tetramethylbenzidine (TMB) peroxide solution and stopped with 1 N H₂SO₄. Absorbance was read at 450 nm.

6. Preparation of whole cell extract

After the treatment, media were aspirated and the cells were washed with cold PBS. The cells were collected by scraping and centrifuged in a microcentrifuge (3,000 rpm, 5 minutes, 4°C). The supernatant was discarded. The cells were mixed with RIPA buffer (25 mM Tris-HCl pH 7.6, 150 mM NaCl, 1% NP-40, 1% sodium deoxycholate, 0.1% sodium dodecyl sulfate) for 30 minutes with continuous agitation. The lysate was centrifuged in a microcentrifuge at 14,000 rpm for 15 minutes at 4°C. The supernatant was used immediately or stored at -80°C. Protein content in extract was determined by Bradford method.

7. Detection of proteins by western blot analysis

Whole cell extracts containing proteins (each 20–60 μ g as

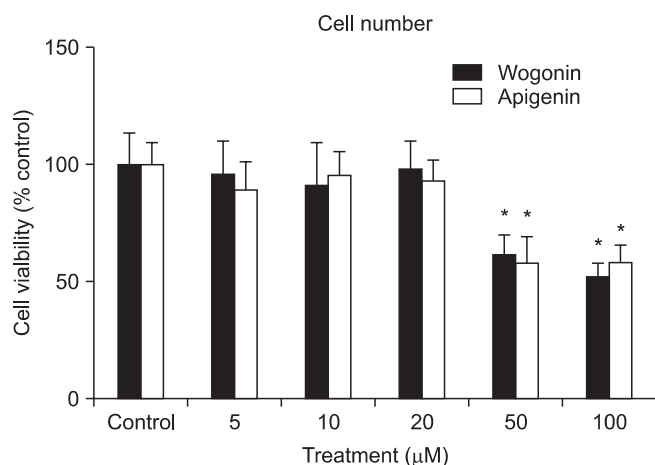


Figure 1. Effect of wogonin or apigenin on proliferation of NCI-H292 cells. NCI-H292 cells were incubated for 72 hours in the presence of varying concentrations of wogonin or apigenin. Cell viability was determined using the sulforhodamine B assay as described in Materials and Methods. Each bar represents a mean \pm standard error of mean of three independent experiments in comparison with the control set at 100%. *Significantly different from control ($p < 0.05$).

protein) were subjected to 7–15% sodium dodecyl sulfate polyacrylamide gel electrophoresis and then transferred onto the PVDF membrane. The blots were blocked using 5% skim milk and probed with appropriate primary antibody in blocking buffer overnight at 4°C. The membrane was washed with PBS and then probed with the secondary antibody conjugated with horseradish peroxidase (Calbiochem, San Diego, CA, USA). Immunoreactive bands were detected by an enhanced chemiluminescence kit (Pierce ECL western blotting substrate; Thermo Scientific, Waltham, MA, USA).

Results

1. Effect of wogonin or apigenin on proliferation of NCI-H292 cells

As can be seen in Figure 1, wogonin or apigenin did not affect the cell proliferation at the concentrations of 5, 10, and 20 μ M. However, 50 and 100 μ M of wogonin or apigenin significantly inhibited the proliferation of NCI-H292 cells. The numbers of cells in wogonin-treated cultures were 100 \pm 13%, 96 \pm 14%, 91 \pm 18%, 98 \pm 12%, 61 \pm 9%, and 52 \pm 6% for control, 5, 10, 20, 50, and 100 μ M, respectively. The numbers of cells in apigenin-treated cultures were 100 \pm 9%, 89 \pm 12%, 95 \pm 10%, 93 \pm 9%, 58 \pm 11%, and 58 \pm 8% for control, 5, 10, 20, 50, and 100 μ M, respectively.

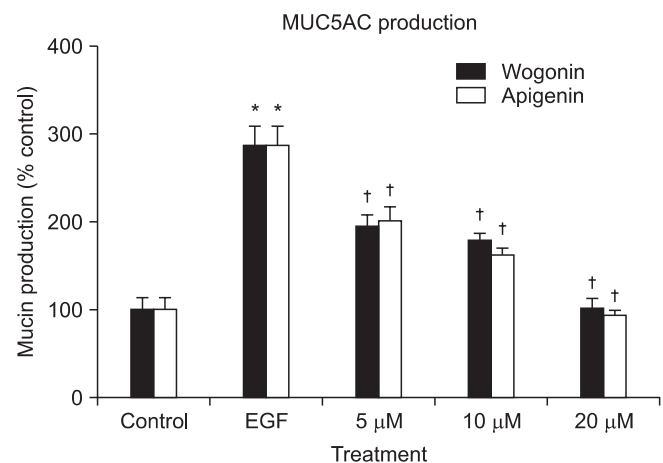


Figure 2. Effect of wogonin or apigenin on epidermal growth factor (EGF)-induced MUC5AC mucin production from NCI-H292 cells. NCI-H292 cells were pretreated with varying concentrations of wogonin or apigenin for 30 minutes and then stimulated with EGF (25 ng/mL) for 24 hours. Cell lysates were collected for measurement of MUC5AC mucin production by enzyme-linked immunosorbent assay. Three independent experiments were performed and the representative data were shown. Each bar represents a mean \pm standard error of mean of 3 culture wells in comparison with the control set at 100%. *Significantly different from control ($p < 0.05$). †Significantly different from EGF alone ($p < 0.05$).

2. Effect of wogonin or apigenin on EGF-induced MUC5AC production from NCI-H292 cells

As can be seen in Figure 2, wogonin significantly inhibited EGF-induced MUC5AC production from NCI-H292 cell. The amounts of mucin in the cells of wogonin-treated cultures were $100 \pm 13\%$, $286 \pm 23\%$, $195 \pm 13\%$, $179 \pm 7\%$, and $101 \pm 12\%$ for control, 25 ng/mL of EGF alone, EGF plus wogonin 5 μM , EGF plus wogonin 10 μM and EGF plus wogonin 20 μM , respectively. The amounts of mucin in the cells of apigenin-treated cultures were $100 \pm 13\%$, $286 \pm 23\%$, $201 \pm 16\%$, $162 \pm 8\%$, and $93 \pm 6\%$ for control, 25 ng/mL of EGF alone, EGF plus apigenin 5 μM , EGF plus apigenin 10 μM and EGF plus apigenin 20 μM , respectively.

3. Effect of wogonin or apigenin on the expression and phosphorylation of the EGFR in NCI-H292 cells

EGF (25 ng/mL, 24 hours) augmented the expression and phosphorylation of the EGFR. On the other hand, apigenin or wogonin significantly inhibited this expression and phos-

phorylation of EGFR, as shown by western blot analysis with the corresponding specific antibodies (Figure 3).

4. Effect of wogonin or apigenin on the phosphorylation of MEK1/2 in NCI-H292 cells

EGF increased the phosphorylation of MEK1/2 in NCI-H292 cells. However, pretreatment of wogonin or apigenin decreased the phosphorylation of MEK1/2 (Figure 4).

5. Effect of wogonin or apigenin on the phosphorylation of p44/42 mitogen-activated protein kinase in NCI-H292 cells

EGF increased the phosphorylation of p44/42 mitogen-activated protein kinase (MAPK) in NCI-H292 cells. However, pretreatment of wogonin or apigenin decreased the phosphorylation of p44/42 MAPK (Figure 5).

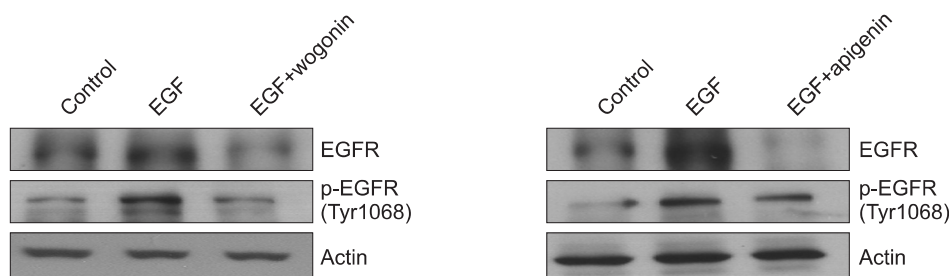


Figure 3. Effect of wogonin or apigenin on epidermal growth factor receptor (EGFR) expression and phosphorylation. NCI-H292 cells were pretreated with wogonin (20 μM) or apigenin (20 μM) for 15 minutes and then stimulated with epidermal growth factor (EGF; 25 ng/mL) for 24 hours. Whole cell extract was collected and western blot analysis of the cellular proteins with anti-EGFR and anti-phospho-EGFR (Tyr1068) antibodies was conducted. Three independent experiments were performed and the representative data were shown.

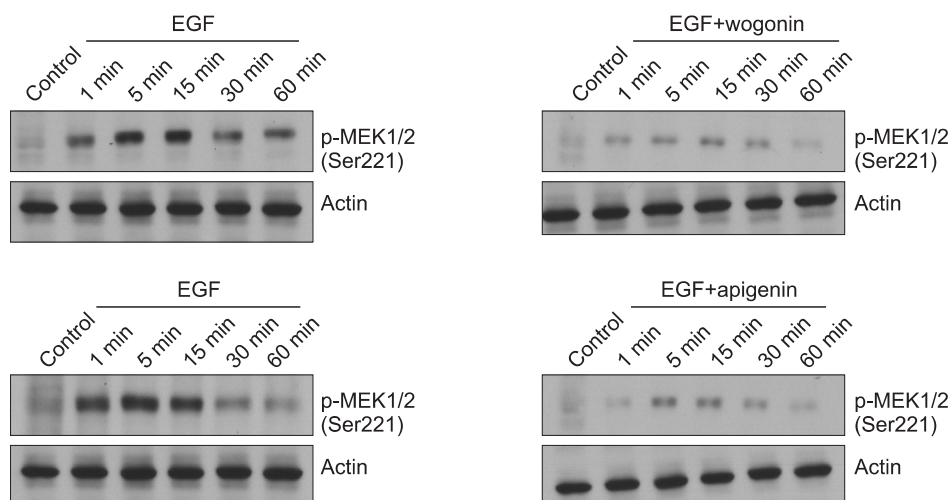


Figure 4. Effect of wogonin or apigenin on phosphorylation of MEK1/2. NCI-H292 cells were pretreated with wogonin (20 μM) or apigenin (20 μM) for 24 hours and then stimulated with epidermal growth factor (EGF; 25 ng/mL) for the indicated periods. Whole cell extract was collected and western blot analysis of the cellular proteins with anti-phospho MEK1/2 (Ser221) antibody was conducted. Three independent experiments were performed and the representative data were shown.

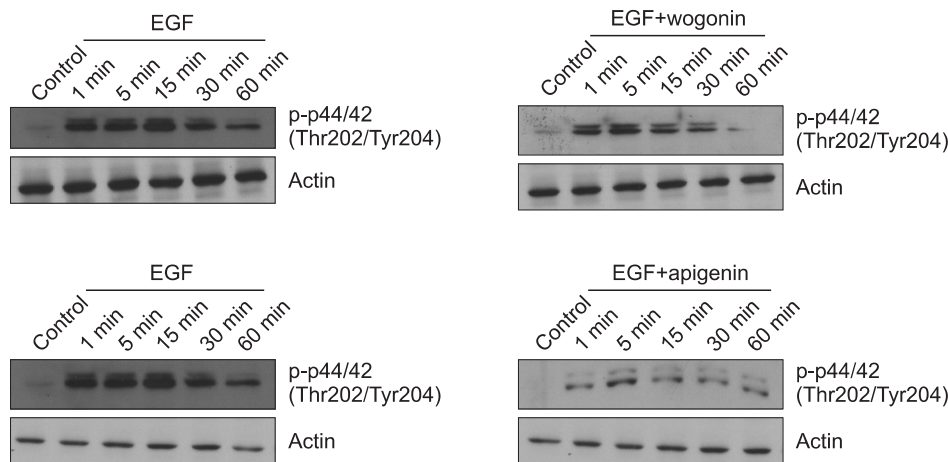


Figure 5. Effect of wogonin or apigenin on phosphorylation of p44/42. NCI-H292 cells were pretreated with wogonin (20 μM) or apigenin (20 μM) for 24 hours and then stimulated with epidermal growth factor (EGF; 25 ng/mL) for the indicated periods. Whole cell extract was collected and western blot analysis of the cellular proteins with anti-phospho p44/42 (Thr202/Tyr204) antibody was conducted. Three independent experiments were performed and the representative data were shown.

Discussion

As aforementioned in introduction, we demonstrated in our previous study that wogonin or apigenin inhibited EGF-induced MUC5AC mucin protein production and gene expression¹⁷. In the present study, we tried again to clarify the effects of wogonin and apigenin on production of airway MUC5AC mucin and got the same result (Figure 2). Based on this result, we investigated whether wogonin or apigenin at the concentration of 20 μM affects the EGFR signaling cascade. EGF activates EGFR signaling cascade and MUC5AC expression in NCI-H292 cells. EGFR serves a central role as a primary regulator of epithelial function, transducing extracellular signals from its activating ligands into intracellular signaling cascades through dimerization and transphosphorylation catalyzed by the intrinsic tyrosine kinase²³. To induce EGFR activation, two different pathways have been reported. First, the binding of its ligands to EGFR activates the intrinsic receptor tyrosine kinase and induces tyrosine phosphorylation²⁴. Second, tyrosine phosphorylation of EGFR can be activated by a ligand-independent mechanism called transactivation. EGFR transactivation is known to occur with various stimuli such as H₂O₂²⁵, ultraviolet light, osmotic stress²⁶, and growth hormone²⁷. Depending on the triggering stimuli, the signal transduction pathways which control mucin transcription have distinct pathways^{1,28}. EGFR activation also plays a major role in modulating mucus production in airway epithelium. In this study, EGF, an endogenous ligand of the EGFR, was selected as a direct activator of this pathway based on previous studies in cultured human airway epithelial NCI-H292 cells^{21,29}. It was found that EGFR is constitutively expressed in NCI-H292 cells, as faint band observed in western blot analysis with anti-EGFR monoclonal antibody in the control group (Figure 3). Being consistent with the notion that the overexpression of MUC5AC is the consequence of the activation of the EGFR signaling cascade, Mata et al.³⁰ also found that preincubation with EGFR tyrosine kinase inhibitors prevented

the EGF-induced augmentation of the MUC5AC gene expression and protein production. EGF increases the protein tyrosine kinase activity of its receptor and thereby activates other kinase cascades such as MAPKs including p38 and p44/42 MAPKs³¹. As can be seen in results, EGF (25 ng/mL, 24 hours) augmented the expression and phosphorylation of the EGFR and 20 μM of wogonin or apigenin significantly inhibited this expression and phosphorylation (Figure 3). There was no cytotoxic effect of wogonin or apigenin at the concentration of 20 μM, based on the data from cell viability test using SRB assay (Figure 1). Also, EGF increased the phosphorylation of MEK1/2 in NCI-H292 cells. However, pretreatment of wogonin or apigenin decreased the phosphorylation of MEK1/2 (Figure 4). As expected, it was found that an early activation of p44/42 MAPK was observed as well as phosphorylation of EGFR after exposure to EGF for 24 hours (Figure 5). Mata et al.³⁰ reported that inhibition of p38 and p44/42 MAPKs with the selective inhibitors SB20202190 and PD98059 abrogated the EGF-induced MUC5AC gene expression. In our result, EGF increased the phosphorylation of p44/42 MAPK in NCI-H292 cells. However, pretreatment of wogonin or apigenin decreased the phosphorylation of p44/42 MAPK (Figure 5). Taken together, the result from this study suggests a possibility of using wogonin or apigenin as a new efficacious mucoregulator for inflammatory pulmonary diseases, although further studies are essential.

In conclusion, Wogonin or apigenin inhibits EGFR signaling pathways, which may explain how they inhibit MUC5AC mucin gene expression and production induced by EGF.

Acknowledgements

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