



Published in final edited form as:

Exp Dermatol. 2014 April ; 23(4): 224–227. doi:10.1111/exd.12348.

Does Prostaglandin D₂ hold the cure to male pattern baldness?

Ashley Nieves, BA and Luis A. Garza, MD-PhD*

Abstract

Lipids in the skin are the most diverse in the entire human body. Their bioactivity in health and disease is underexplored. Prostaglandin D₂ has recently been identified as a factor which is elevated in the bald scalp of men with androgenetic alopecia and has the capacity to decrease hair lengthening. An enzyme which synthesizes it, prostaglandin D₂ synthase (PTGDS or lipocalin-PGDS) is hormone responsive in multiple other organs. PGD₂ has two known receptors, GPR44 and PTGDR. GPR44 was found to be necessary for the decrease in hair growth by PGD₂. This creates an exciting opportunity to perhaps create novel treatments for androgenetic alopecia which inhibit the activity of PTGDS, PGD₂ or GPR44. This review discusses the current knowledge surrounding PGD₂ and future steps needed to translate these findings into novel therapies for patients with androgenetic alopecia.

Introduction

Male pattern baldness, or androgenetic alopecia (AGA), is the cause of much anguish. Because hair is so tied to a person's physical appearance and sexual identity, its loss motivates a search for treatments. Current treatments are based on pathophysiological models which have not significantly gone beyond Aristotle's observation that "eunuchs do not go bald." Therefore modern science has extended this observation to demonstrate the importance of testosterone, and treatments against 5- α -reductase type II (finasteride) are currently a mainstay of treatment(1). However, serendipity has also shaped the alopecia armamentarium with the observation that minoxidil, originally used to treat hypertension, also causes hair growth, thus prompting its addition as a treatment for AGA(2).

Besides the above two therapies and, more successfully, hair transplantation, little else is effectively used to augment hair growth in AGA(3). This is partly because our knowledge of the pathogenesis is still meager and offers few rationale targets for therapy. Recently, however, work from our group and others have added to the understanding of AGA and suggested novel therapies(4–6). We have demonstrated that Prostaglandin D₂ is elevated in male balding scalp and that it functionally inhibits hair lengthening through its receptor GPR44, which directly suggests treatments for male pattern hair loss. This review will discuss both the advancements and future questions which must be addressed before we really understand whether these novel insights into AGA can be translated into meaningful interventions for patients.

One recent product to stimulate hair growth has been bimatoprost (Latisse). Bimatoprost is a prostaglandin analogue of Prostaglandin F_{2a} and is prescribed to augment the growth of eyelash hair, with extensive efforts to extend this for the treatment of scalp alopecias(7, 8). Latanoprost and Bimatoprost were both used to decrease ocular pressure in glaucoma and

*Corresponding author: Luis A. Garza, MD-PhD, Assistant Professor, Department of Dermatology, Johns Hopkins School of Medicine, Suite 204, Koch CRBII, 1550 Orleans Street, Baltimore, MD 21287, p 410-955-8662, f 410-614-0635, LAG@jhmi.edu.
http://www.hopkinsmedicine.org/dermatology/about_us/our_experts/garza_luis.html

were incidentally noted to cause hair lengthening(9–11). There exists a precedence for suspecting the importance of lipids in hair follicle function(12, 13), particularly in the context of the sebaceous gland and scarring alopecias (14–16). However, the discovery of the potency for latanoprost and bimatoprost to induce human hair lengthening remains some of the strongest evidence to implicate prostaglandins in hair follicle function.

The synthesis and metabolism of prostaglandins or eicosanoids is a very actively studied area, not only given its importance in skin biology such as inflammation(17). The cascade begins with membrane phospholipids which are metabolized by phospholipase into arachidonic acid. This lipid is the substrate for a host of different pathways including the leukotrienes, but also is a substrate for the cyclooxygenase enzymes (COX1/2 or PTGS ½). The cyclooxygenase enzymes produce PGH₂ which is converted into selectively different prostaglandin species by specific synthases. Multiple isoforms of synthases can act to create each individual prostaglandin species. For example, PGD₂ has at least 2 synthase enzymes using PGH₂ as a substrate(18). PGF_{2a} can also be synthesized from PGH₂ also through the enzyme PGH 9-11-endoperoxide reductase, but also from PGE₂ or PGD₂ via the enzymes PGE 9-ketoreductase and PGD 11-ketoreductase(19, 20) respectively. Prostaglandin synthesis enzymes are expressed in the skin diffusely including hair follicles(20, 21) and sebaceous glands(22). Complicating understanding of these pathways is that minutely different lipid compound substrates with very similar chemical structures can often still be metabolized by these same enzymes leading to distinct products with undetermined biologic significance.

The metabolism of prostaglandins is similarly complex. Two important proteins for prostaglandin metabolism include 15-PGDH and SLC02A1 or PGT. 15-PGDH is present in the hair follicle(23), catabolizes prostaglandins(24), and has been shown to be a tumor suppressor(25, 26) given PGE₂ pro-carcinogenic effects(27, 28). Similarly, SLC02A1 or PGT functions as a prostaglandin transporter which is important for the uptake and/or clearance of prostaglandins and when mutated in human syndromes includes skin symptoms(29). Given the plethora of potential chemical reactions on prostaglandins and the number of incompletely characterized lipid metabolizing enzymes in the human genome, it is very likely that many important pathways are yet to be defined.

Brain type lipocalin Prostaglandin D₂ synthase (PTGDS) in AGA

PTGDS role in hair biology was initially identified in an unbiased screen comparing bald versus haired scalp in male AGA patients, using the patient's own haired scalp as an internal control for every analysis(4). It was one of the most abundant transcripts in bald scalp compared to haired scalp. PTGDS is a prostaglandin synthase enzyme which acts downstream of the cyclooxygenase enzymes as mentioned above (30). PTGDS generates PGD₂(31–34). Therefore this array finding suggested that PGD₂, the product of PTGDS, would be elevated in bald scalp of men.

PTGDS was confirmed to be elevated in bald scalp by mRNA, protein and also its enzymatic product, PGD₂. Importantly, its increase was confirmed by mass spectrometry which has been shown to be superior to conventional ELISA based methods which use marginally specific antibodies for the selective identification of individual prostaglandin species(35). We showed that the normal expression of PTGDS matched the temporal and spatial expression of hair follicle areas which were lost in the dying phase of the hair cycle. PGD₂ levels peaked 7 fold higher than baseline levels immediately preceding catagen. PTGDS was expressed in the outer root sheath inferior to the bulge.

Having confirmed the correlation between PGD₂ and alopecia, the most critical question was its functional relevance. The next goal was functional efforts to identify the role of

PGD₂. All the above amounts to a correlation; the main difficulty of most array studies is that causality is not clear. Any items found to be elevated in bald scalp might be a product of that state rather than a cause of it. Therefore it was important to demonstrate that PGD₂ indeed did decrease hair lengthening in both man and mouse. Therefore, we tested the effect of PGD₂ on mouse models with inactivating mutations in each of the receptors for PGD₂—PTGDR(DP-1) or GPR44 (DP-2). While PTGDR was not required for PGD₂ to inhibit hair growth, GPR44 was required. Recent work from our group has shown that just as PGD₂ and GPR44 inhibit hair growth, they also inhibit hair follicle regeneration after wounding(36). Current work is now focused on demonstrating that in human follicles, already identified GPR44 inhibitors also can reverse the hair growth inhibition by PGD₂. Recent work has also confirmed that the hematopoietic type of PGD₂ synthase (HPGDS) also correlates with areas of hair loss(37).

Reasons why PTGDS involvement in AGA makes sense

All the above is good evidence for the importance of PGD₂ in the pathogenesis of AGA. The next question is the exact degree of importance. Is PGD₂ the dominant agent which is downstream of testosterone and inhibits hair growth? Or is it one of many agents and itself only a minor contributor to AGA. The following observations support that PGD₂ might be a dominant player, though clearly more work is required (see below).

1. Prostaglandins are already known to modulate hair function.
 - a. Prostaglandins have already been shown to be important in hair follicle function(7), and even are clinically useful. The best examples are analogues of PGF₂a which have been shown in people to increase hair growth. In many ways reminiscent of the discovery of minoxidil, they were not rationally discovered but do give investigators a clue that the pathways which normally control hair lengthening involve prostaglandins. Also, mice overexpressing the COX2 enzyme develop alopecia(38, 39), though mice generally do not show AGA. Therefore the discovery of PTGDS on an unbiased screen is consistent with the above.
2. Prostaglandins often have opposing functions.
 - a. Prostaglandins typically control bodily functions in a “yin and yang” manner where they have opposing functions(30). This explains the observation that despite the involvement of prostaglandins in a multitude of organ systems, COX inhibitors like aspirin typically have very restricted clinical effects— because they will inhibit both system activators and system inhibitors with a net negligible effect. For example in the case of bronchial muscle tone, PGD₂ increases contraction while PGE₂ increases relaxation. In the case of hair, it is known that PGE₂ and PGF₂a stimulate hair growth, while PGD₂ inhibits hair growth(9, 10, 40). This discovery therefore fits current models of prostaglandin function.
3. PTGDS is hormonally responsive in other systems
 - a. PTGDS is present in seminal fluid and epididymis. PTGDS expression has been shown to decrease with castration and increase after androgen reintroduction(41–43). But PGDS is also estrogen responsive in some systems(44). Finally, although PGD₂ has not been tested, PGE₂ has been shown to induce male mating behavior circuits in the developing brain (45). Given low androgen receptor levels in keratinocytes (46–49), an effect of androgens on PTGDS expression might be indirect. Therefore it

is clear hormones frequently modulate prostaglandin levels which would be consistent with their role in AGA.

4. PGD₂ receptors are in the outer root sheath of the hair follicle.
 - a. Early reports are consistent with receptor's localization within among other areas to the upper and lower outer root sheath region as well as the dermal papilla (21). Therefore the location of receptors is appropriate to imagine a functional importance for hair follicle activity.
5. PGD₂ might explain sebaceous hyperplasia in AGA
 - a. Although controversial, PGD₂ and its non-enzymatic break down product 15-deoxy-delta 12,14-prostaglandin J₂ are published as ligands for PPARgamma(50, 51). PPARgamma is a known master transcription factor for adipose development and also supports sebaceous gland function(52). Therefore it is tempting to consider that the high levels of PGD₂ might explain the dramatic increase in sebaceous gland size in AGA. Whether the sebaceous gland hyperactivity is just a byproduct of AGA pathogenesis, or instead actively promotes alopecia is not fully known and also requires more study.
6. Lipid based pathways are underexplored area of biology
 - a. Given that lipid biology is comparatively understudied to genes and proteins, it is possible that it has been overlooked in this very common condition.
7. Minoxidil has effects on prostaglandins
 - a. In early literature on the mechanism of minoxidil's effect on reducing blood pressure, it was noted that it has the capacity to increase PGE₂, which has been shown to be reduced in AGA (53–55). Although not proven, minoxidil's effects on prostaglandins would be consistent with their aberrant regulation in AGA.
8. Recent genetic analysis show associations of AGA with GPR44 which approach significance.
 - a. Although not fully statistically significant, a recent GWAS dataset of AGA was analyzed for associations to the PGD₂ pathway(56). GPR44 was nominally significant (p=0.03). The authors point out that larger sample sets might improve this association.
9. Classic developmental pathways controlling hair morphogenesis likely will not be directly affected in a disease like AGA.
 - a. Many potent developmental pathways have been shown to modulate hair follicle function (wnts, shh for example(57)). However, if these pathways were directly modulated by androgens in AGA for example, the phenotype of AGA would likely be considerably broader. A more plausible model is that AGA will modulate upstream pathways which only indirectly modulate these very powerful developmental pathways. Prostaglandins might be that modulator factor.

Outstanding questions the field must solve

1. Replication by other groups.

are being designed for allergic diseases, their usefulness in AGA might be compromised if the necessary schedule and delivery are different for inhibiting GPR44 in the hair follicle. The larger the number of candidate compounds to test for GPR44 inhibition in the hair follicle, the greater likelihood of success. If existing candidates for GPR44 inhibition fail to prove useful, the search for new candidates will be more pertinent. Given the likely important locus of expression of GPR44 in keratinocytes, the development of keratinocyte reporter cell lines for GPR44 activity would allow for large scale screening of compounds to inhibit this pathway and provide a pipeline for potential small molecule candidates.

- b. If appropriate GPR44 inhibition can be achieved, the most important of all the above questions will address the reversibility of AGA in human subjects.

Conclusions and Perspective

An outline is emerging for a fundamental role of prostaglandins in modulating hair function. Inhibition of PTGDS might help male pattern hair loss, conceivably with agents as simple as selenium chloride(59). But considering this arena as much larger and given that the skin has one of the most diverse repertoire of lipids in the entire human body(60), the possibilities are vast for identifying bioactive lipids which will inhibit or promote hair growth. The tools are limited, but the task is simple: just find the needle in the haystack.

Acknowledgments

This work was supported by R01AR064297 to LG from NIAMS/NIH. The authors wish to thank Drs. George Cotsarelis and Mayumi Ito for useful discussions on this topic. LG wrote the paper and AN designed the figure and edited the manuscript.

References

1. Paus R, Cotsarelis G. The biology of hair follicles. *N Engl J Med.* 1999; 341(7):491–7. Epub 1999/08/12. [PubMed: 10441606]
2. Price VH. Treatment of hair loss. *N Engl J Med.* 1999; 341(13):964–73. Epub 1999/09/25. [PubMed: 10498493]
3. Sawaya ME, Hordinsky MK. Advances in alopecia areata and androgenetic alopecia. *Adv Dermatol.* 1992; 7:211–26. discussion 27. Epub 1992/01/01. [PubMed: 1739581]
4. Garza LA, Liu Y, Yang Z, Alagesan B, Lawson JA, Norberg SM, et al. Prostaglandin D2 inhibits hair growth and is elevated in bald scalp of men with androgenetic alopecia. *Sci Transl Med.* 2012; 4(126):126ra34. Epub 2012/03/24 4/126/126ra34 [pii]. 10.1126/scitranslmed.3003122
5. Garza LA, Yang CC, Zhao T, Blatt HB, Lee M, He H, et al. Bald scalp in men with androgenetic alopecia retains hair follicle stem cells but lacks CD200-rich and CD34-positive hair follicle progenitor cells. *J Clin Invest.* 2011; 121(2):613–22. Epub 2011/01/06. 10.1172/JCI4447844478 [PubMed: 21206086]
6. Inui S, Itami S. Androgen actions on the human hair follicle: perspectives. *Exp Dermatol.* 2013; 22(3):168–71. Epub 2012/09/29. 10.1111/exd.12024 [PubMed: 23016593]
7. Khidhir KG, Woodward DF, Farjo NP, Farjo BK, Tang ES, Wang JW, et al. The prostamide-related glaucoma therapy, bimatoprost, offers a novel approach for treating scalp alopecias. *FASEB J.* 2013; 27(2):557–67. Epub 2012/10/30. fj.12-218156 [pii]. 10.1096/fj.12-218156 [PubMed: 23104985]
8. Woodward DF, Tang ES, Attar M, Wang JW. The biodisposition and hypertrichotic effects of bimatoprost in mouse skin. *Exp Dermatol.* 2013; 22(2):145–8. Epub 2013/01/03. 10.1111/exd.12071 [PubMed: 23278986]

9. Johnstone MA, Albert DM. Prostaglandin-induced hair growth. *Surv Ophthalmol.* 2002; 47 (Suppl 1):S185–202. Epub 2002/09/03 S0039625702003077 [pii]. [PubMed: 12204716]
10. Sasaki S, Hozumi Y, Kondo S. Influence of prostaglandin F2alpha and its analogues on hair regrowth and follicular melanogenesis in a murine model. *Exp Dermatol.* 2005; 14(5):323–8. Epub 2005/04/28. EXD270 [pii]. 10.1111/j.0906-6705.2005.00270.x [PubMed: 15854125]
11. Wolf R, Matz H, Zalish M, Pollack A, Orion E. Prostaglandin analogs for hair growth: great expectations. *Dermatol Online J.* 2003; 9(3):7. Epub 2003/09/04. [PubMed: 12952754]
12. Skolnik P, Eaglstein WH, Ziboh VA. Human essential fatty acid deficiency: treatment by topical application of linoleic acid. *Arch Dermatol.* 1977; 113(7):939–41. Epub 1977/07/01. [PubMed: 406855]
13. Menton DN. The effects of essential fatty acid deficiency on the skin of the mouse. *Am J Anat.* 1968; 122(2):337–55. Epub 1968/03/01. 10.1002/aja.1001220211 [PubMed: 4298148]
14. Gates AH, Karasek M. Hereditary Absence of Sebaceous Glands in the Mouse. *Science.* 1965; 148(3676):1471–3. Epub 1965/06/11. 148/3676/1471 [pii]. 10.1126/science.148.3676.1471 [PubMed: 17738154]
15. Zheng Y, Eilertsen KJ, Ge L, Zhang L, Sundberg JP, Prouty SM, et al. Scd1 is expressed in sebaceous glands and is disrupted in the asebia mouse. *Nat Genet.* 1999; 23(3):268–70. Epub 1999/11/05. 10.1038/15446 [PubMed: 10545940]
16. Sundberg JP, Boggess D, Sundberg BA, Eilertsen K, Parimoo S, Filippi M, et al. Asebia-2J (Scd1(ab2J)): a new allele and a model for scarring alopecia. *Am J Pathol.* 2000; 156(6):2067–75. Epub 2000/06/15. S0002-9440(10)65078-X [pii]. 10.1016/S0002-9440(10)65078-X [PubMed: 10854228]
17. Nicolaou A. Eicosanoids in skin inflammation. *Prostaglandins Leukot Essent Fatty Acids.* 2013; 88(1):131–8. Epub 2012/04/24. S0952-3278(12)00041-5 [pii]. 10.1016/j.plefa.2012.03.009 [PubMed: 22521864]
18. Urade Y, Watanabe K, Hayaishi O. Prostaglandin D, E, and F synthases. *J Lipid Mediat Cell Signal.* 1995; 12(2–3):257–73. [PubMed: 8777570]
19. Watanabe K. Prostaglandin F synthase. *Prostaglandins Other Lipid Mediat.* 2002; 68–69:401–7. Epub 2002/11/16.
20. Colombe L, Vindrios A, Michelet JF, Bernard BA. Prostaglandin metabolism in human hair follicle. *Exp Dermatol.* 2007; 16(9):762–9. Epub 2007/08/19. EXD586 [pii]. 10.1111/j.1600-0625.2007.00586.x [PubMed: 17697149]
21. Colombe L, Michelet JF, Bernard BA. Prostanoid receptors in anagen human hair follicles. *Exp Dermatol.* 2008; 17(1):63–72. Epub 2007/11/17. EXD639 [pii]. 10.1111/j.1600-0625.2007.00639.x [PubMed: 18005048]
22. Alestas T, Ganceviciene R, Fimmel S, Muller-Decker K, Zouboulis CC. Enzymes involved in the biosynthesis of leukotriene B4 and prostaglandin E2 are active in sebaceous glands. *J Mol Med (Berl).* 2006; 84(1):75–87. Epub 2006/01/03. 10.1007/s00109-005-0715-8 [PubMed: 16388388]
23. Michelet JF, Colombe L, Gautier B, Gaillard O, Benech F, Pereira R, et al. Expression of NAD+ dependent 15-hydroxyprostaglandin dehydrogenase and protection of prostaglandins in human hair follicle. *Exp Dermatol.* 2008; 17(10):821–8. Epub 2008/03/11. EXD706 [pii]. 10.1111/j.1600-0625.2008.00706.x [PubMed: 18328086]
24. Fincham N, Camp R. Novel prostaglandin dehydrogenase in rat skin. *Biochem J.* 1983; 212(1):129–34. Epub 1983/04/15. [PubMed: 6575778]
25. Tai HH, Chi X, Tong M. Regulation of 15-hydroxyprostaglandin dehydrogenase (15-PGDH) by non-steroidal anti-inflammatory drugs (NSAIDs). *Prostaglandins Other Lipid Mediat.* 2011; 96(1–4):37–40. Epub 2011/07/19. S1098-8823(11)00045-1 [pii]. 10.1016/j.prostaglandins.2011.06.005 [PubMed: 21763448]
26. Yan M, Rerko RM, Platzer P, Dawson D, Willis J, Tong M, et al. 15-Hydroxyprostaglandin dehydrogenase, a COX-2 oncogene antagonist, is a TGF-beta-induced suppressor of human gastrointestinal cancers. *Proc Natl Acad Sci U S A.* 2004; 101(50):17468–73. Epub 2004/12/03 0406142101 [pii]. 10.1073/pnas.0406142101 [PubMed: 15574495]

27. Ansari KM, Rundhaug JE, Fischer SM. Multiple signaling pathways are responsible for prostaglandin E2-induced murine keratinocyte proliferation. *Mol Cancer Res.* 2008; 6(6):1003–16. Epub 2008/06/24. 6/6/1003 [pii]. 10.1158/1541-7786.MCR-07-2144 [PubMed: 18567804]
28. Sung YM, He G, Hwang DH, Fischer SM. Overexpression of the prostaglandin E2 receptor EP2 results in enhanced skin tumor development. *Oncogene.* 2006; 25(40):5507–16. [PubMed: 16607275]
29. Sasaki T, Niizeki H, Shimizu A, Shiohama A, Hirakiyama A, Okuyama T, et al. Identification of mutations in the prostaglandin transporter gene *SLCO2A1* and its phenotype-genotype correlation in Japanese patients with pachydermoperiostosis. *J Dermatol Sci.* 2012; 68(1):36–44. Epub 2012/08/22. S0923-1811(12)00240-X [pii]. 10.1016/j.jdermsci.2012.07.008 [PubMed: 22906430]
30. Goodman, LS.; Gilman, A.; Hardman, JG.; Gilman, AG.; Limbird, LE. Goodman & Gilman's the pharmacological basis of therapeutics. 9. Vol. xxi. New York: McGraw-Hill, Health Professions Division; 1996. p. 1905[1] p. folded plate p
31. Eguchi N, Minami T, Shirafuji N, Kanaoka Y, Tanaka T, Nagata A, et al. Lack of tactile pain (allodynia) in lipocalin-type prostaglandin D synthase-deficient mice. *Proc Natl Acad Sci U S A.* 1999; 96(2):726–30. [PubMed: 9892701]
32. Matsuoka T, Hirata M, Tanaka H, Takahashi Y, Murata T, Kabashima K, et al. Prostaglandin D2 as a mediator of allergic asthma. *Science.* 2000; 287(5460):2013–7. [PubMed: 10720327]
33. Qu WM, Huang ZL, Xu XH, Aritake K, Eguchi N, Nambu F, et al. Lipocalin-type prostaglandin D synthase produces prostaglandin D2 involved in regulation of physiological sleep. *Proc Natl Acad Sci U S A.* 2006; 103(47):17949–54. [PubMed: 17093043]
34. Urade Y, Eguchi N. Lipocalin-type and hematopoietic prostaglandin D synthases as a novel example of functional convergence. *Prostaglandins & Other Lipid Mediators.* 2002; 68–69:375–82.
35. Bell-Parikh LC, Ide T, Lawson JA, McNamara P, Reilly M, FitzGerald GA. Biosynthesis of 15-deoxy-delta12,14-PGJ2 and the ligation of PPARgamma. *J Clin Invest.* 2003; 112(6):945–55. Epub 2003/09/17. 10.1172/JCI18012112/6/945 [PubMed: 12975479]
36. Nelson AM, Loy DE, Lawson JA, Katseff AS, Fitzgerald GA, Garza LA. Prostaglandin D2 Inhibits Wound-Induced Hair Follicle Neogenesis through the Receptor, Gpr44. *J Invest Dermatol.* 2013; 133(4):881–9. Epub 2012/11/30. jid2012398 [pii]. 10.1038/jid.2012.398 [PubMed: 23190891]
37. Larson AR, Zhan Q, Johnson E, Fragoso AC, Wan M, Murphy GF. A prostaglandin d-synthase-positive mast cell gradient characterizes scalp patterning. *J Cutan Pathol.* 2013;10.1111/cup.12286
38. Muller-Decker K, Leder C, Neumann M, Neufang G, Bayerl C, Schweizer J, et al. Expression of cyclooxygenase isozymes during morphogenesis and cycling of pelage hair follicles in mouse skin: precocious onset of the first catagen phase and alopecia upon cyclooxygenase-2 overexpression. *J Invest Dermatol.* 2003; 121(4):661–8. [PubMed: 14632179]
39. Bol DK, Rowley RB, Ho CP, Pilz B, Dell J, Swerdel M, et al. Cyclooxygenase-2 overexpression in the skin of transgenic mice results in suppression of tumor development. *Cancer Res.* 2002; 62(9):2516–21. [PubMed: 11980643]
40. Geng L, Hanson WR, Malkinson FD. Topical or systemic 16, 16 dm prostaglandin E2 or WR-2721 (WR-1065) protects mice from alopecia after fractionated irradiation. *Int J Radiat Biol.* 1992; 61(4):533–7. [PubMed: 1349335]
41. Zhu H, Ma H, Ni H, Ma XH, Mills N, Yang ZM. Expression and regulation of lipocalin-type prostaglandin d synthase in rat testis and epididymis. *Biol Reprod.* 2004; 70(4):1088–95. [PubMed: 14668211]
42. O'Shaughnessy PJ, Johnston H, Willerton L, Baker PJ. Failure of normal adult Leydig cell development in androgen-receptor-deficient mice. *J Cell Sci.* 2002; 115(Pt 17):3491–6. [PubMed: 12154079]
43. Zhu H, Ma H, Ni H, Ma XH, Mills N, Yang ZM. L-prostaglandin D synthase expression and regulation in mouse testis and epididymis during sexual maturation and testosterone treatment after castration. *Endocrine.* 2004; 24(1):39–45. [PubMed: 15249702]

44. Otsuki M, Gao H, Dahlman-Wright K, Ohlsson C, Eguchi N, Urade Y, et al. Specific regulation of lipocalin-type prostaglandin D synthase in mouse heart by estrogen receptor beta. *Mol Endocrinol*. 2003; 17(9):1844–55. [PubMed: 12829806]
45. Amateau SK, McCarthy MM. Induction of PGE2 by estradiol mediates developmental masculinization of sex behavior. *Nat Neurosci*. 2004; 7(6):643–50. Epub 2004/05/25. nn1254 [pii]. 10.1038/nn1254 [PubMed: 15156148]
46. Choudhry R, Hodgins MB, Van der Kwast TH, Brinkmann AO, Boersma WJ. Localization of androgen receptors in human skin by immunohistochemistry: implications for the hormonal regulation of hair growth, sebaceous glands and sweat glands. *J Endocrinol*. 1992; 133(3):467–75. Epub 1992/06/01. [PubMed: 1613448]
47. Itami S, Kurata S, Sonoda T, Takayasu S. Interaction between dermal papilla cells and follicular epithelial cells in vitro: effect of androgen. *Br J Dermatol*. 1995; 132(4):527–32. Epub 1995/04/01. [PubMed: 7748741]
48. Pelletier G, Ren L. Localization of sex steroid receptors in human skin. *Histol Histopathol*. 2004; 19(2):629–36. Epub 2004/03/17. [PubMed: 15024720]
49. Jave-Suarez LF, Langbein L, Winter H, Praetzel S, Rogers MA, Schweizer J. Androgen regulation of the human hair follicle: the type I hair keratin hHa7 is a direct target gene in trichocytes. *J Invest Dermatol*. 2004; 122(3):555–64. Epub 2004/04/17. JID22336 [pii]. 10.1111/j.0022-202X.2004.22336.x [PubMed: 15086535]
50. Forman BM, Tontonoz P, Chen J, Brun RP, Spiegelman BM, Evans RM. 15-Deoxy-delta 12, 14-prostaglandin J2 is a ligand for the adipocyte determination factor PPAR gamma. *Cell*. 1995; 83(5):803–12. [PubMed: 8521497]
51. Kliewer SA, Lenhard JM, Willson TM, Patel I, Morris DC, Lehmann JM. A prostaglandin J2 metabolite binds peroxisome proliferator-activated receptor gamma and promotes adipocyte differentiation. *Cell*. 1995; 83(5):813–9. [PubMed: 8521498]
52. Trivedi NR, Cong Z, Nelson AM, Albert AJ, Rosamilia LL, Sivarajah S, et al. Peroxisome proliferator-activated receptors increase human sebum production. *J Invest Dermatol*. 2006; 126(9):2002–9. [PubMed: 16675962]
53. Michelet JF, Commo S, Billoni N, Mahe YF, Bernard BA. Activation of cytoprotective prostaglandin synthase-1 by minoxidil as a possible explanation for its hair growth-stimulating effect. *J Invest Dermatol*. 1997; 108(2):205–9. Epub 1997/02/01. [PubMed: 9008235]
54. Kvedar JC, Baden HP, Levine L. Selective inhibition by minoxidil of prostacyclin production by cells in culture. *Biochem Pharmacol*. 1988; 37(5):867–74. [PubMed: 3278714]
55. Messenger AG, Rundegren J. Minoxidil: mechanisms of action on hair growth. *Br J Dermatol*. 2004; 150(2):186–94. Epub 2004/03/05. 5785 [pii]. [PubMed: 14996087]
56. Heilmann S, Nyholt DR, Brockschmidt FF, Hillmer AM, Herold C, Maan C, et al. No genetic support for a contribution of prostaglandins to the aetiology of androgenetic alopecia. *Br J Dermatol*. 2013; 169(1):222–4. Epub 2013/03/02. 10.1111/bjd.12292 [PubMed: 23448296]
57. Millar SE. Molecular mechanisms regulating hair follicle development. *J Invest Dermatol*. 2002; 118(2):216–25. Epub 2002/02/14 1670 [pii]. 10.1046/j.0022-202x.2001.01670.x [PubMed: 11841536]
58. Chaurand P, Norris JL, Cornett DS, Mobley JA, Caprioli RM. New developments in profiling and imaging of proteins from tissue sections by MALDI mass spectrometry. *J Proteome Res*. 2006; 5(11):2889–900. Epub 2006/11/04. 10.1021/pr060346u [PubMed: 17081040]
59. Lee B, Hirst JJ, Walker DW. Prostaglandin D synthase in the prenatal ovine brain and effects of its inhibition with selenium chloride on fetal sleep/wake activity in utero. *J Neurosci*. 2002; 22(13):5679–86. [PubMed: 12097519]
60. Nicolaidis N. Skin lipids: their biochemical uniqueness. *Science*. 1974; 186(4158):19–26. Epub 1974/10/04. [PubMed: 4607408]

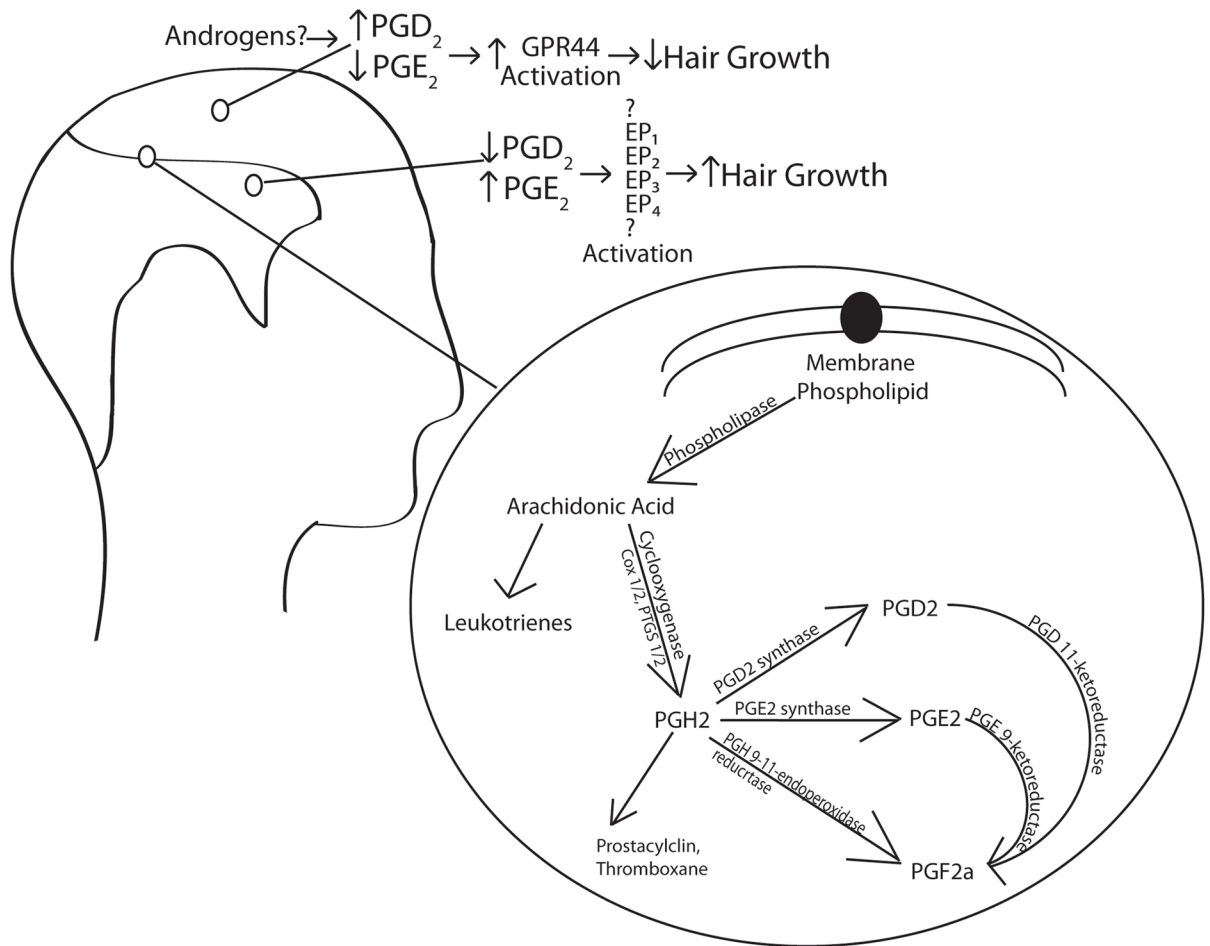


Figure 1. Schematic of PGD2 pathway in alopecia
 Depicted is our current understanding of prostaglandin synthesis and changes in alopecia.