

# Complete Genome Sequence of *Paenibacillus polymyxa* SQR-21, a Plant Growth-Promoting Rhizobacterium with Antifungal Activity and Rhizosphere Colonization Ability

Shuqing Li,<sup>a,b,c</sup> Dongqing Yang,<sup>a,b,c</sup> Meihua Qiu,<sup>a,b,c</sup> Jiahui Shao,<sup>a,b,c</sup> Rong Guo,<sup>a,b,c</sup> Biao Shen,<sup>a,b,c</sup> Xihou Yin,<sup>d</sup> Ruifu Zhang,<sup>a,b,c</sup> Nan Zhang,<sup>a,b,c</sup> Qirong Shen<sup>a,b,c</sup>

National Engineering Research Center for Organic-Based Fertilizers, Nanjing Agricultural University, Nanjing, China<sup>a</sup>; Jiangsu Collaborative Innovation Center for Solid Organic Waste Utilization, Nanjing Agricultural University, Nanjing, China<sup>b</sup>; Key Laboratory of Plant Nutrition and Fertilization in Low-Middle Reaches of the Yangtze River, Ministry of Agriculture, Nanjing, Nanjing Agricultural University, Nanjing, China<sup>c</sup>; Department of Pharmaceutical Sciences, Oregon State University, Corvallis, Oregon, USA<sup>d</sup>

Here we report the complete genome sequence of a plant growth-promoting rhizobacterium (PGPR), *Paenibacillus polymyxa* SQR-21, which consists of one circular chromosome of 5,828,438 bp with 5,024 coding sequences (CDS). The data presented highlight multiple sets of functional genes associated with its plant-beneficial characteristics.

Received 13 March 2014 Accepted 24 March 2014 Published 10 April 2014

Citation Li S, Yang D, Qiu M, Shao J, Guo R, Shen B, Yin X, Zhang R, Zhang N, Shen Q. 2014. Complete genome sequence of *Paenibacillus polymyxa* SQR-21, a plant growth-promoting rhizobacterium with antifungal activity and rhizosphere colonization ability. Genome Announc. 2(2):e00281-14. doi:10.1128/genomeA.00281-14.

Copyright © 2014 Li et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 3.0 Unported license](http://creativecommons.org/licenses/by/3.0/).

Address correspondence to Nan Zhang, nanzhang@njau.edu.cn, or Qirong Shen, shenqirong@njau.edu.cn.

**P***Paenibacillus polymyxa* (formerly *Bacillus polymyxa*), a Gram-positive, sporulating bacterium, is considered to be a plant growth-promoting rhizobacterium (PGPR) (1, 2). Originally isolated from various environmental samples (3, 4), *P. polymyxa* has been widely applied in agriculture, industry, medicine, and environmental remediation (5–12). Thus far, only four *P. polymyxa* genomes, those of *P. polymyxa* E681 (NC\_014483) (13), *P. polymyxa* SC2 (NC\_014622) (14), *P. polymyxa* M1 (NC\_017542) (1), and *P. polymyxa* CR1 (NC\_023037) (15), have been completely sequenced. *P. polymyxa* SQR-21 was naturally isolated from watermelon rhizosphere. As an outstanding PGPR strain with the ability to produce various antibiotics (16, 17) and colonize rhizospheres (18), SQR-21 has been widely exploited in commercial biofertilizers for plant growth promotion and biological control of soilborne plant pathogens (19, 20). Here, we report the whole-genome sequence of SQR-21 in order to better elucidate its plant-beneficial characteristics and promote its agricultural applications.

Whole-genome sequencing of *P. polymyxa* SQR-21 was performed with Roche 454 sequencing technology. The constructed library was sequenced by the GS FLX Titanium series chemistry. A total of 1,000,000 sequence reads with average read lengths of 350 to 450 bp (resulting in up to 400 Mb of sequence data) were obtained, representing an average of 70-fold coverage of the genome. The reads were assembled into 19 scaffolds, and the 54 sequence gaps both within and between the scaffolds were filled by sequencing PCR products using an ABI 3730 capillary sequencer.

The complete genome of *P. polymyxa* SQR-21 is composed of a 5,828,438-bp circular chromosome, with a mean GC content of 45.64%. Based on the genomic data, 5,024 coding sequences (CDS) were predicted by GeneMark (21) and annotated by a BLAST tools search against the GenBank nonredundant protein database (NR), Kyoto Encyclopedia of Genes and Genomes (KEGG), and Clusters of Orthologous Groups

(COG). In addition, 111 tRNA loci and 13 rRNA operons were identified with the tRNAscan-SE (22) and RNAmmer 1.2 (23) servers, respectively.

*P. polymyxa* SQR-21 harbors four nonribosomal peptide synthetase (NRPS) gene clusters, including *pmx* and *fus* gene clusters responsible for biosynthesis of polymyxin and fusaricidin, respectively, which reveal high similarities to the published gene clusters (24–26). One polyketide synthetase (PKS) gene cluster, three hybrid PKS-NRPS gene clusters, and three gene clusters relevant to lantibiotic biosynthesis are also present in the SQR-21 genome. Several genes involved in plant growth promotion, including genes responsible for production of indole-3-acetic acid (IAA), 3-hydroxy-2-butanone (acetoin), and 2,3-butanediol, as well as phytase, were identified in the SQR-21 genome. In addition, the genome harbors a set of genes encoding extracellular enzymes involved in the degradation of plant-derived polysaccharides, including xylanase, glucanase, and chitinase. Given the strain specificity and application in agricultural production, the complete genome sequence of *P. polymyxa* SQR-21 provides useful information for both basic and applied research, which also facilitates the understanding of the functions and evolutions of the *Paenibacillus polymyxa* genome.

**Nucleotide sequence accession number.** The complete sequence of *Paenibacillus polymyxa* SQR-21 has been deposited in NCBI's GenBank under the accession number [CP006872](https://www.ncbi.nlm.nih.gov/nuccore/CP006872).

## ACKNOWLEDGMENTS

This research was financially supported by the Nature Science Foundation of China (31301845 and 31330069), the Chinese Ministry of Science and Technology (2013AA102802), and the Fundamental Research Funds for the Central Universities (KYZ201307). R.Z. and Q.S. were also supported by the 111 Project (B12009) and the Priority Academic Program Development (PAPD) of Jiangsu Higher Education Institutions.

## REFERENCES

- Niu B, Rueckert C, Blom J, Wang Q, Borriis R. 2011. The genome of the plant growth-promoting rhizobacterium *Paenibacillus polymyxa* M-1 contains nine sites dedicated to nonribosomal synthesis of lipopeptides and polyketides. *J. Bacteriol.* 193:5862–5863. <http://dx.doi.org/10.1128/JB.005806-11>.
- Timmusk S. 2003. Mechanism of action of the plant growth promoting bacterium *Paenibacillus polymyxa*. Ph.D. thesis. Uppsala University, Uppsala, Sweden.
- Lal S, Romano S, Chiarini L, Signorini A, Tabacchioni S. 2012. The *Paenibacillus polymyxa* species is abundant among hydrogen-producing facultative anaerobic bacteria in Lake Averno sediment. *Arch. Microbiol.* 194:345–351. <http://dx.doi.org/10.1007/s00203-011-0763-0>.
- McSpadden Gardener BB. 2004. Ecology of *Bacillus* and *Paenibacillus* spp. in agricultural systems. *Phytopathology* 94:1252–1258. <http://dx.doi.org/10.1094/PHYTO.2004.94.11.1252>.
- Lal S, Tabacchioni S. 2009. Ecology and biotechnological potential of *Paenibacillus polymyxa*: a minireview. *Indian J. Microbiol.* 49:2–10. <http://dx.doi.org/10.1007/s12088-009-0008-y>.
- Raza W, Wang Y, Shen QR. 2008. *Paenibacillus polymyxa*: antibiotics, hydrolytic enzymes and hazard assessment. *J. Plant Pathol.* 90:419–430.
- Niu B, Vater J, Rueckert C, Blom J, Lehmann M, Ru JJ, Chen XH, Wang Q, Borriis R. 2013. Polymyxin P is the active principle in suppressing phytopathogenic *Erwinia* spp. by the biocontrol rhizobacterium *Paenibacillus polymyxa* M-1. *BMC Microbiol.* 13:137. <http://dx.doi.org/10.1186/1471-2180-13-137>.
- Anand R, Grayston S, Chanway C. 2013. N<sub>2</sub>-fixation and seedling growth promotion of lodgepole pine by endophytic *Paenibacillus polymyxa*. *Microb. Ecol.* 66:369–374. <http://dx.doi.org/10.1007/s00248-013-0196-1>.
- Haggag WM, Timmusk S. 2008. Colonization of peanut roots by biofilm-forming *Paenibacillus polymyxa* initiates biocontrol against crown rot disease. *J. Appl. Microbiol.* 104:961–969. <http://dx.doi.org/10.1111/j.1365-6722.2007.03611.x>.
- Yu B, Sun J, Bommarreddy RR, Song L, Zeng AP. 2011. Novel (2R,3R)-2,3-butenediol dehydrogenase from potential industrial strain *Paenibacillus polymyxa* ATCC 12321. *Appl. Environ. Microbiol.* 77:4230–4233. <http://dx.doi.org/10.1128/AEM.02998-10>.
- He Z, Kisla D, Zhang L, Yuan C, Green-Church KB, Yousef AE. 2007. Isolation and identification of a *Paenibacillus polymyxa* strain that coproduces a novel lantibiotic and polymyxin. *Appl. Environ. Microbiol.* 73: 168–178. <http://dx.doi.org/10.1128/AEM.02023-06>.
- Ariza A, Eklof JM, Spadiut O, Offen WA, Roberts SM, Besenmatter W, Friis EP, Skjøt M, Wilson KS, Brumer H, Davies G. 2011. Structure and activity of *Paenibacillus polymyxa* xyloglucanase from glycoside hydrolase family 44. *J. Biol. Chem.* 286:33890–33900. <http://dx.doi.org/10.1074/jbc.M111.262345>.
- Kim JF, Jeong H, Park SY, Kim SB, Park YK, Choi SK, Ryu CM, Hur CG, Ghim SY, Oh TK, Kim JJ, Park CS, Park SH. 2010. Genome sequence of the polymyxin-producing plant-probiotic rhizobacterium *Paenibacillus polymyxa* E681. *J. Bacteriol.* 192:6103–6104. <http://dx.doi.org/10.1128/JB.00983-10>.
- Ma M, Wang C, Ding Y, Li L, Shen D, Jiang X, Guan D, Cao F, Chen H, Feng R, Wang X, Ge Y, Yao L, Bing X, Yang X, Li J, Du B. 2011. Complete genome sequence of *Paenibacillus polymyxa* Sc2, a strain of plant growth-promoting rhizobacterium with broad-spectrum antimicrobial activity. *J. Bacteriol.* 193:311–312. <http://dx.doi.org/10.1128/JB.01234-10>.
- Eastman AW, Weselowski B, Nathoo N, Yuan Z-C. 2014. Complete genome sequence of *Paenibacillus polymyxa* CR1, a plant growth-promoting bacterium isolated from the corn rhizosphere exhibiting potential for biocontrol, biomass degradation, and biofuel production. *Genome Announc.* 2(1):e01218-13. <http://dx.doi.org/10.1128/genomeA.01218-13>.
- Li S, Zhang R, Wang Y, Zhang N, Shao J, Qiu M, Shen B, Yin X, Shen Q. 2013. Promoter analysis and transcription regulation of fus gene cluster responsible for fusaricidin synthesis of *Paenibacillus polymyxa* SQR-21. *Appl. Microbiol. Biotechnol.* 97:9479–9489. <http://dx.doi.org/10.1007/s0253-013-5157-6>.
- Raza W, Yang XM, Wu HS, Wang Y, Xu YC, Shen QR. 2009. Isolation and characterisation of fusaricidin-type compound-producing strain of *Paenibacillus polymyxa* SQR-21 active against *Fusarium oxysporum* f. sp. *nevium*. *Eur. J. Plant Pathol.* 125:471–483. <http://dx.doi.org/10.1007/s10658-009-9496-1>.
- Ling N, Zhang WW, Tan SY, Huang QW, Shen QR. 2012. Effect of the nursery application of bioorganic fertilizer on spatial distribution of *Fusarium oxysporum* f. sp. *niveum* and its antagonistic bacterium in the rhizosphere of watermelon. *Appl. Soil Ecol.* 59:13–19. <http://dx.doi.org/10.1016/j.apsoil.2012.05.001>.
- Ling N, Xue C, Huang QW, Yang XM, Xu YC, Shen QR. 2010. Development of a mode of application of bioorganic fertilizer for improving the biocontrol efficacy to *Fusarium* wilt. *BioControl* 55:673–683. <http://dx.doi.org/10.1007/s10526-010-9290-1>.
- Wu HS, Yang XN, Fan JQ, Miao WG, Ling N, Xu YC, Huang QW, Shen QR. 2009. Suppression of fusarium wilt of watermelon by a bio-organic fertilizer containing combinations of antagonistic microorganisms. *BioControl* 54:287–300. <http://dx.doi.org/10.1007/s10526-008-9168-7>.
- Borodovsky M, Mills R, Besemer J, Lomsadze A. 2003. Prokaryotic gene prediction using GeneMark and GeneMark.hmm. *Curr. Protoc. Bioinformatics* 1:4.5.1–4.5.16. <http://dx.doi.org/10.1002/0471250953.bi0405s01>.
- Lowe TM, Eddy SR. 1997. tRNAscan-SE: a program for improved detection of transfer RNA genes in genomic sequence. *Nucleic Acids Res.* 25: 955–964. <http://dx.doi.org/10.1093/nar/25.5.0955>.
- Lagesen K, Hallin P, Rødland EA, Staerfeldt HH, Rognes T, Ussery DW. 2007. RNAmmer: consistent and rapid annotation of ribosomal RNA genes. *Nucleic Acids Res.* 35:3100–3108. <http://dx.doi.org/10.1093/nar/gkm160>.
- Choi SK, Park SY, Kim R, Kim SB, Lee CH, Kim JF, Park SH. 2009. Identification of a polymyxin synthetase gene cluster of *Paenibacillus polymyxa* and heterologous expression of the gene in *Bacillus subtilis*. *J. Bacteriol.* 191:3350–3358. <http://dx.doi.org/10.1128/JB.01728-08>.
- Choi SK, Park SY, Kim R, Lee CH, Kim JF, Park SH. 2008. Identification and functional analysis of the fusaricidin biosynthetic gene of *Paenibacillus polymyxa* E681. *Biochem. Biophys. Res. Commun.* 365:89–95. <http://dx.doi.org/10.1016/j.bbrc.2007.10.147>.
- Li J, Jensen SE. 2008. Nonribosomal biosynthesis of fusaricidins by *Paenibacillus polymyxa* PKB1 involves direct activation of a D-amino acid. *Chem. Biol.* 15:118–127. <http://dx.doi.org/10.1016/j.chembiol.2007.12.014>.