Pregnancy-induced maternal regulatory T cells, bona fide memory or maintenance by antigenic reminder from fetal cell microchimerism?

Jeremy M Kinder, Tony T Jiang, Dayna R Clark, Vandana Chaturvedi, Lijun Xin, James M Ertelt, and Sing Sing Way* Division of Infectious Diseases; Cincinnati Children's Hospital Medical Center; Cincinnati, OH USA

Keywords: T cells, immunological memory, regulatory T cells, pregnancy, microchimerism

*Correspondence to: Sing Sing Way; Email: singsing.way@cchmc.org Submitted: 01/17/2014

Revised: 01/27/2014

Accepted: 02/14/2014

Published Online: 02/19/2014

http://dx.doi.org/10.4161/chim.28241

Addendum to: Rowe JH, Ertelt JM, Xin L, Way SS. Pregnancy imprints regulatory memory that sustains anergy to fetal antigen. Nature 2012; 490:102-6; PMID:23023128; http://dx.doi. org/10.1038/nature11462

Long-term maintenance of immune components with defined specificity, without antigen is the hallmark feature of immunological memory. However, there are fundamental differences in how memory CD8+ compared with CD4+ T cells are maintained. After complete antigen elimination, CD8+ T cells can persist as a self-renewing numerically stable cell population, and therefore satisfy the most stringent definition of "memory." Comparatively, CD4+ T cell maintenance is considerably less stable, often requiring low-level antigen persistence or antigenic reminders. Recent studies show these basic memory features, classically ascribed to effector CD8+ and CD4+ T cells, extend to immune suppressive Foxp3⁺ regula**tory CD4+ T cells (Tregs). In particular, gestational expansion and postpartum retention of maternal Tregs with fetal specificity may explain the protective benefits of primary pregnancy on complications in subsequent pregnancy. Herein, the possibility of ongoing antigenic reminders from fetal cell microchimerism in postpartum maintenance of maternal Tregs with fetal specificity is considered.**

The mammalian immune system is endowed not only with efficient self, non-self discrimination, but also the ability to "remember" antigenic encounters. For immunologically foreign antigens, prior stimulation has the potential to prime long-term retention of "memory" immune cells with specificity to the inciting antigen. In turn, establishing the molecular and cellular requirements

for immunological memory has critical implications for developing more durable vaccines and other immune modulatory therapies.

Emerging studies highlight an interesting discordance in necessity for antigen persistence in maintaining longterm retention of CD8⁺ compared with CD4+ T cells with defined specificity.1-7 This is best illustrated by the dynamics of pathogen-specific CD8+ and CD4+ T cells after infection with viruses or other intracellular pathogens that do not cause persistence. While both T cell subsets expand robustly during acute infection, a numerically stable self-renewing pool of pathogen-specific CD8+ T cells is maintained indefinitely despite complete antigen elimination. By contrast, CD4+ T cells responding to the same acute infection undergo protracted, but stable contraction with an estimated half-life of 15 to 40 d.⁸⁻¹⁰ This discordance may reflect the necessity for each T cell subset in host defense. For acute infection with viruses or other intra-cytoplasmic pathogens (e.g., influenza A, lymphocytic choriomeningitis virus, or *Listeria monocytogenes*) where protection is conferred by CD8+ T cells, these cells are chosen for selective retention. Comparatively for pathogens that primarily cause persistent infection and reside within the phagocytic vacuole of infected cells thereby escaping detection or elimination by CD8+ T cells (e.g., *Mycobacterium tuberculosis*, *Leishmania major*, or *Salmonella* spp.), pathogen-specific CD4+ T cells play a more dominant protective role.11-13 Importantly however, while CD8+ T cell mediated protection against secondary

infection is maintained well after antigen elimination, protection by retained memory CD4+ T cells requires low-level antigen persistence. Accordingly for pathogens capable of establishing persistent infection, antigen elimination that occurs naturally or with adjunctive antimicrobials, accelerates contraction of pathogen-specific CD4+ T cells and overrides the protective benefits of prior infection.14-16 Therefore, unlike CD8+ T cells, the long-term maintenance of CD4+ T cell memory appear to require more frequent, if not constant, antigenic reminders.

While the memory features of CD4+ T cells has been best characterized for IFN-γ producing Th1 cells, other CD4+ effector lineages (e.g., Th2 or Th17 cells) appear to share a similar potential for longterm retention.7,10 By redirecting tools for tracking antigen-specific T cells, we and others have recently shown these memory features classically described for effector T cells also extend to immune suppressive regulatory CD4+ T cells (Tregs) identified by Foxp3 expression. Treg memory was first shown using transgenic mice where the model antigen, ovalbumin, could be inducibly expressed within the skin.17 Primary dermal stimulation with this surrogate self-antigen primed expansion and retention of ovalbumin specific Tregs that dampens the severity of localized autoimmune reactions when this antigen was re-expressed -30 d later. Likewise, a complementary study tracking Tregs after influenza A infection showed accelerated accumulation of virus-specific Tregs after secondary, compared with primary infection, which may be important for limiting pathological airway inflammation from over-exuberant effector CD8+ T cell activation.18 Our own studies investigating maternal Tregs with specificity to the immune-dominant I-Ab:2W1S_{55–68} peptide expressed as a surrogate fetal antigen during allogeneic pregnancy, showed Foxp3+ CD4+ T cells with this specificity progressively expand throughout gestation.¹⁹ Interestingly after delivery of the fetus and other gross products of conception, maternal Tregs with fetal specificity were maintained at markedly enriched levels; and these cells re-expand with accelerated tempo during

secondary pregnancy upon encounter with the same paternal-fetal antigen. Considering the necessity for expanded maternal Tregs in maintaining fetal tolerance during pregnancy,¹⁹⁻²⁴ these findings likely provide critical mechanistic insights for how primary pregnancy protects against complications stemming from fractured fetal tolerance in subsequent pregnancy.^{19,20,25,26} In turn, applied to the basic biology of CD4+ T cells, these findings together establish Foxp3+ Tregs, like effector T cells, can persist as memory immune cells.

Given the discordance in necessity for antigen persistence in sustaining long-term retention of CD8⁺ compared with CD4+ effector T cells with defined specificity, these findings also open up exciting new questions regarding whether retained Tregs reflects bona fide memory or maintenance in response to antigen persistence. In the case of Tregs with specificity for surrogate-self ovalbumin antigen within the skin, ongoing stimulation is unlikely since naive ovalbumin specific T cells failed to proliferate after adoptive transfer without induced antigen expression.17 Similarly, Tregs retained after influenza A infection are unlikely to reflect stimulation from residual viral antigen, since this pathogen is not known to cause persistent infection.18 However in each of these models, the longer-term durability of Tregs, with specificity to either self or pathogen, remain undefined since the impacts of secondary antigen challenge were reported at most ~35 d after silencing primary antigen stimulation.17,18 In our studies tracking maternal Tregs with surrogate fetal-2W1S specificity, enriched cells were maintained through at least 100 d postpartum despite progressively diminishing cell numbers.19 In particular, the postpartum decay kinetics of maternal Tregs with fetal specificity (estimated $t_{1/2}$ of 25 d) show striking similarity with effector CD4+ T cells primed by acute infection.

On the other hand and in sharp contrast to the tempo of antigen stimulation that occurs after acute infection conditions, retained maternal Tregs with fetal specificity are likely to have more frequent antigenic encounters from fetal cells that establish microchimerism, analogous

to low-level antigen stimulation in the later stages of persistent infection. Fetal cell microchimerism initiated during pregnancy and sustained postpartum probably occurs ubiquitously, but this phenomenon has become only recently widely appreciated with the use of molecular tools that allow these rare $(-1 \text{ in } 10^6)$ cells to be consistently identified.27-29 Accordingly, antigenic reminder from fetal cell microchimerism may be pivotal for sustaining memory among pregnancyinduced maternal Tregs. Moreover, if maternal CD4+ Treg memory is sustained by fetal cell microchimerism, it would be interesting to consider the necessity for comparable antigenic reminders in maintaining regulatory CD8+ T cells shown in other contexts.30-32 Along with the long-term maintenance of maternal cell microchimerism sustained by fetal Tregs in offspring,³³ this emerging body of evidence highlight remarkably potent and long-lived immunological programming that occurs naturally with the bidirectional transfer of cells and antigens between mother and fetus through in utero exposure.

Based on these findings, we propose important next steps are to more meticulously dissect the physiological milieu of pregnancy and in utero development that primes immunological tolerance and Treg memory. Taking cues from effector CD4+ T cell memory,^{1-7,34} this will likely include interrelated contributions from naive cell precursor frequency, primary expansion magnitude, antigen avidity, and response to cytokine growth factors, along with increased frequency of antigenic reminders. Furthermore, given the potential for Treg conversion into inflammatory cytokine producing effector T cells with the same specificity,^{35,36} microchimeric fetal cells also have the dangerous potential for sensitizing responses that may trigger autoimmunity.37-39 This is analogous to pathological responses to microchimeric maternal cells in offspring with various diverse autoimmune disorders including diabetes, 40 biliary atresia, 41 and dermatomyositis.42 Therefore, establishing the molecular signals that reinforce Treg differentiation stability are of equally high importance and priority.

Nevertheless, applied to the devastating complications in human pregnancy that stem from underlying defects in fetal tolerance (preeclampsia, prematurity, miscarriage), basic investigation on the fundamental biology of CD4+ T cells and memory features for protective regulatory subsets provides renewed hope for new, more efficacious therapeutic approaches. In turn, given the striking parallels between Treg and effector CD4+ T cell memory, unraveling how maternal Treg memory is sustained will likely also provide critical insights for priming more durable effector T cells with pathogen specificity for augmenting host defense against infection.

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

Acknowledgments

This work was supported in part by the NIAID through awards R01-AI087830, R01-AI100934, and R21-AI112186. S.S.W. holds an Investigator in the Pathogenesis of Infectious Disease award from the Burroughs Wellcome Fund.

References

- 1. Seder RA, Ahmed R. Similarities and differences in CD4+ and CD8+ effector and memory T cell generation. Nat Immunol 2003; 4:835-42; PMID:12942084; http://dx.doi.org/10.1038/ni969
- 2. Williams MA, Holmes BJ, Sun JC, Bevan MJ. Developing and maintaining protective CD8+ memory T cells. Immunol Rev 2006; 211:146-53; PMID:16824124; http://dx.doi. org/10.1111/j.0105-2896.2006.00389.x
- 3. Prlic M, Williams MA, Bevan MJ. Requirements for CD8 T-cell priming, memory generation and maintenance. Curr Opin Immunol 2007; 19:315- 9; PMID:17433873; http://dx.doi.org/10.1016/j. coi.2007.04.010
- 4. Harty JT, Badovinac VP. Shaping and reshaping CD8+ T-cell memory. Nat Rev Immunol 2008; 8:107-19; PMID:18219309; http://dx.doi. org/10.1038/nri2251
- 5. Sprent J, Surh CD. Normal T cell homeostasis: the conversion of naive cells into memoryphenotype cells. Nat Immunol 2011; 12:478-84;
PMID:21739670; http://dx.doi.org/10.1038/ http://dx.doi.org/10.1038/ ni.2018
- 6. van Leeuwen EM, Sprent J, Surh CD. Generation and maintenance of memory CD4(+) T Cells. Curr Opin Immunol 2009; 21:167-72; PMID:19282163; http://dx.doi.org/10.1016/j.coi.2009.02.005
- 7. Pepper M, Jenkins MK. Origins of CD4(+) effector and central memory T cells. Nat Immunol 2011; 12:467-71; PMID:21739668; http://dx.doi. org/10.1038/ni.2038
- 8. Homann D, Teyton L, Oldstone MB. Differential regulation of antiviral T-cell immunity results in stable CD8+ but declining CD4+ T-cell memory. Nat Med 2001; 7:913-9; PMID:11479623; http:// dx.doi.org/10.1038/90950
- 9. Harrington LE, Janowski KM, Oliver JR, Zajac AJ, Weaver CT. Memory CD4 T cells emerge from effector T-cell progenitors. Nature 2008; 452:356- 60; PMID:18322463; http://dx.doi.org/10.1038/ nature06672
- 10. Pepper M, Linehan JL, Pagán AJ, Zell T, Dileepan T, Cleary PP, Jenkins MK. Different routes of bacterial infection induce long-lived TH1 memory cells and short-lived TH17 cells. Nat Immunol 2010; 11:83- 9; PMID:19935657; http://dx.doi.org/10.1038/ ni.1826
- 11. Reiley WW, Shafiani S, Wittmer ST, Tucker-Heard G, Moon JJ, Jenkins MK, Urdahl KB, Winslow GM, Woodland DL. Distinct functions of antigenspecific CD4 T cells during murine Mycobacterium tuberculosis infection. Proc Natl Acad Sci U S A 2010; 107:19408-13; PMID:20962277; http:// dx.doi.org/10.1073/pnas.1006298107
- 12. Winslow GM, Roberts AD, Blackman MA, Woodland DL. Persistence and turnover of antigenspecific CD4 T cells during chronic tuberculosis infection in the mouse. J Immunol 2003; 170:2046- 52; PMID:12574375
- 13. McSorley SJ, Cookson BT, Jenkins MK. Characterization of CD4+ T cell responses during natural infection with Salmonella typhimurium. J Immunol 2000; 164:986-93; PMID:10623848
- 14. Uzonna JE, Wei G, Yurkowski D, Bretscher P. Immune elimination of Leishmania major in mice: implications for immune memory, vaccination, and reactivation disease. J Immunol 2001; 167:6967-74; PMID:11739516
- 15. Belkaid Y, Piccirillo CA, Mendez S, Shevach EM, Sacks DL. CD4+CD25+ regulatory T cells control Leishmania major persistence and immunity. Nature 2002; 420:502-7; PMID:12466842; http://dx.doi. org/10.1038/nature01152
- 16. Nelson RW, McLachlan JB, Kurtz JR, Jenkins MK. CD4+ T cell persistence and function after infection are maintained by low-level peptide:MHC class II presentation. J Immunol 2013; 190:2828- 34; PMID:23382562; http://dx.doi.org/10.4049/ jimmunol.1202183
- Rosenblum MD, Gratz IK, Paw JS, Lee K, Marshak-Rothstein A, Abbas AK. Response to self antigen imprints regulatory memory in tissues. Nature 2011; 480:538-42; PMID:22121024
- 18. Brincks EL, Roberts AD, Cookenham T, Sell S, Kohlmeier JE, Blackman MA, Woodland DL. Antigen-specific memory regulatory CD4+Foxp3+ T cells control memory responses to influenza virus infection. J Immunol 2013; 190:3438-46;
PMID:23467933; http://dx.doi.org/10.4049/ http://dx.doi.org/10.4049/ jimmunol.1203140
- 19. Rowe JH, Ertelt JM, Xin L, Way SS. Pregnancy imprints regulatory memory that sustains anergy to fetal antigen. Nature 2012; 490:102- 6; PMID:23023128; http://dx.doi.org/10.1038/ nature11462
- 20. Aluvihare VR, Kallikourdis M, Betz AG. Regulatory T cells mediate maternal tolerance to the fetus. Nat Immunol 2004; 5:266-71; PMID:14758358; http:// dx.doi.org/10.1038/ni1037
- 21. Rowe JH, Ertelt JM, Aguilera MN, Farrar MA, Way SS. Foxp3(+) regulatory T cell expansion required for sustaining pregnancy compromises host defense against prenatal bacterial pathogens. Cell Host Microbe 2011; 10:54-64; PMID:21767812; http:// dx.doi.org/10.1016/j.chom.2011.06.005
- 22. Somerset DA, Zheng Y, Kilby MD, Sansom DM, Drayson MT. Normal human pregnancy is associated with an elevation in the immune suppressive CD25+ CD4+ regulatory T-cell subset. Immunology 2004; 112:38-43; PMID:15096182; http://dx.doi. org/10.1111/j.1365-2567.2004.01869.x
- 23. Sasaki Y, Darmochwal-Kolarz D, Suzuki D, Sakai M, Ito M, Shima T, Shiozaki A, Rolinski J, Saito S. Proportion of peripheral blood and decidual CD4(+) CD25(bright) regulatory T cells in pre-eclampsia. Clin Exp Immunol 2007; 149:139-45; PMID:17459078; http://dx.doi. org/10.1111/j.1365-2249.2007.03397.x
- 24. Toldi G, Svec P, Vásárhelyi B, Mészáros G, Rigó J, Tulassay T, Treszl A. Decreased number of FoxP3+ regulatory T cells in preeclampsia. Acta Obstet Gynecol Scand 2008; 87:1229-33; PMID:19016357; http://dx.doi.org/10.1080/00016340802389470
- 25. Campbell DM, MacGillivray I, Carr-Hill R. Pre-eclampsia in second pregnancy. Br J Obstet Gynaecol 1985; 92:131-40; PMID:3970893; http:// dx.doi.org/10.1111/j.1471-0528.1985.tb01064.x
- 26. Trupin LS, Simon LP, Eskenazi B. Change in paternity: a risk factor for preeclamp-
sia in multiparas. Epidemiology 1996; sia in multiparas. Epidemiology 1996;
7:240-4; PMID:8728435; http://dx.doi. PMID:8728435; org/10.1097/00001648-199605000-00004
- 27. Bianchi DW, Zickwolf GK, Weil GJ, Sylvester S, DeMaria MA. Male fetal progenitor cells persist in maternal blood for as long as 27 years postpartum. Proc Natl Acad Sci U S A 1996; 93:705- 8; PMID:8570620; http://dx.doi.org/10.1073/ pnas.93.2.705
- 28. Gammill HS, Nelson JL. Naturally acquired microchimerism. Int J Dev Biol 2010; 54:531- 43; PMID:19924635; http://dx.doi.org/10.1387/ ijdb.082767hg
- 29. Khosrotehrani K, Johnson KL, Cha DH, Salomon RN, Bianchi DW. Transfer of fetal cells with multilineage potential to maternal tissue. JAMA 2004; 292:75-80; PMID:15238593; http://dx.doi. org/10.1001/jama.292.1.75
- 30. Robb RJ, Lineburg KE, Kuns RD, Wilson YA, Raffelt NC, Olver SD, Varelias A, Alexander KA, Teal BE, Sparwasser T, et al. Identification and expansion of highly suppressive CD8(+)FoxP3(+) regulatory T cells after experimental allogeneic bone marrow transplantation. Blood 2012; 119:5898- 908; PMID:22538855; http://dx.doi.org/10.1182/ blood-2011-12-396119
- 31. Fleissner D, Hansen W, Geffers R, Buer J, Westendorf AM. Local induction of immunosuppressive CD8+ T cells in the gut-associated lymphoid tissues. PLoS One 2010; 5:e15373; PMID:20975955; http:// dx.doi.org/10.1371/journal.pone.0015373
- 32. Uss E, Rowshani AT, Hooibrink B, Lardy NM, van Lier RA, ten Berge IJ. CD103 is a marker for alloantigen-induced regulatory CD8+ T cells. J Immunol 2006; 177:2775-83; PMID:16920912
- 33. Mold JE, Michaëlsson J, Burt TD, Muench MO, Beckerman KP, Busch MP, Lee TH, Nixon DF, McCune JM. Maternal alloantigens promote the development of tolerogenic fetal regulatory T cells in utero. Science 2008; 322:1562-5; PMID:19056990; http://dx.doi.org/10.1126/science.1164511
- 34. Gratz IK, Truong HA, Yang SH, Maurano MM, Lee K, Abbas AK, Rosenblum MD. Cutting Edge: memory regulatory t cells require IL-7 and not IL-2 for their maintenance in peripheral tissues. J Immunol 2013; 190:4483-7; PMID:23543753; http://dx.doi. org/10.4049/jimmunol.1300212
- 35. Bailey-Bucktrout SL, Martinez-Llordella M, Zhou X, Anthony B, Rosenthal W, Luche H, Fehling HJ, Bluestone JA. Self-antigen-driven activation induces instability of regulatory T cells during an inflammatory autoimmune response. Immunity 2013; 39:949-62; PMID:24238343; http://dx.doi. org/10.1016/j.immuni.2013.10.016
- 36. Komatsu N, Okamoto K, Sawa S, Nakashima T, Oh-Hora M, Kodama T, Tanaka S, Bluestone JA, Takayanagi H. Pathogenic conversion of Foxp3(+) T cells into TH17 cells in autoimmune arthritis. Nat Med 2014; 20:62-8; PMID:24362934; http:// dx.doi.org/10.1038/nm.3432
- 37. Adams KM, Nelson JL. Microchimerism: an investigative frontier in autoimmunity and transplantation. JAMA 2004; 291:1127-31; PMID:14996783; http://dx.doi.org/10.1001/jama.291.9.1127
- 38. Stevens AM. Microchimeric cells in systemic lupus erythematosus: targets or innocent bystanders? Lupus 2006; 15:820-6; PMID:17153857; http:// dx.doi.org/10.1177/0961203306070068
- 39. Mosca M, Curcio M, Lapi S, Valentini G, D'Angelo S, Rizzo G, Bombardieri S. Correlations of Y chromosome microchimerism with disease activity in patients with SLE: analysis of preliminary data. Ann Rheum Dis 2003; 62:651-4; PMID:12810428; http://dx.doi.org/10.1136/ard.62.7.651
- 40. Nelson JL, Gillespie KM, Lambert NC, Stevens AM, Loubiere LS, Rutledge JC, Leisenring WM, Erickson TD, Yan Z, Mullarkey ME, et al. Maternal microchimerism in peripheral blood in type 1 diabetes and pancreatic islet beta cell microchimerism. Proc Natl Acad Sci U S A 2007; 104:1637- 42; PMID:17244711; http://dx.doi.org/10.1073/ pnas.0606169104
- 41. Suskind DL, Rosenthal P, Heyman MB, Kong D, Magrane G, Baxter-Lowe LA, Muench MO. Maternal microchimerism in the livers of patients with biliary atresia. BMC Gastroenterol 2004; 4:14; PMID:15285784; http://dx.doi. org/10.1186/1471-230X-4-14
- 42. Reed AM, Picornell YJ, Harwood A, Kredich DW. Chimerism in children with juvenile dermatomyositis. Lancet 2000; 356:2156-7; PMID:11191546; http://dx.doi.org/10.1016/S0140-6736(00)03500-5