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## **β-2 adrenergic receptor gene methylation is associated with decreased asthma severity in Inner-City School Children**

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### **Abstract**

**Background**—Genetic variation in the β-2 adrenergic receptor gene (*ADRB2*) has been implicated in asthma severity and control with conflicting results. Epigenetic variation in the β-2 adrenergic receptor gene (*ADRB2*) may play an important role in asthma phenotype.

**Objective**—We aimed to evaluate whether DNA methylation of *ADRB2* is associated with asthma phenotypes in inner city school aged children.

**Methods**—Multiple CpG sites in the promoter region of *ADRB2* gene were analyzed in 177 children enrolled in the School Inner-City Asthma Study (SICAS). Blood or saliva-derived DNA was measured by bisulfite-polymerase chain reaction pyrosequencing assay. Average percent DNA methylation across the sites was evaluated for association with asthma severity (report of dyspnea, nighttime symptoms, rescue medication use, and baseline spirometry) and morbidity (school absences and unscheduled healthcare visits). Three clades composed of highly correlated methylation sites within the methylated segment of *ADRB2* were further analyzed.

**Results**—Methylation of individual sites generally ranged from 0 – 6% with average percent methylation across sites of 2.4%. Univariate analyses strongly favored the association of higher percent methylation with lower asthma severity measured by report of dyspnea. Furthermore, there was a non-significant trend toward less rescue medication use, nighttime symptoms, school absences, activity limitation due to asthma, and improved lung function measurements with

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increased methylation. Multivariate analysis demonstrated methylation of *ADRB2* gene significantly associated with less dyspnea (odds ratio (OR) 0.2, 95% confidence interval (CI), 0.1 – 0.6, P = 0.002). Each of the 3 clades of methylation sites showed a strong, but not statistically significant, effect on decreased dyspnea.

**Conclusions and Clinical Relevance**—DNA methylation in the *ADRB2* gene is associated with decreased asthma symptom severity, suggesting a role for methylation in asthma phenotypes.

## Introduction

Asthma is a complex disease whose manifestations are influenced by host characteristics and environmental exposures. Genotype – phenotype correlations in asthma have been inconsistent and likely belie the environmental contribution to phenotype, the genetic complexity of the disease and the influence of the myriad biological processes that influence the expression of genomic DNA.

It has long been suggested that the  $\beta$ -2 adrenergic receptor ( $\beta$ 2AR) plays an important role in the development of asthma. Szentivanyi proposed in 1968 that bronchial hyperresponsiveness is a result of partial beta-adrenergic blockade[1]. The merits of this hypothesis were supported by studies demonstrating reduced beta-adrenergic response, reduced  $\beta$ 2AR density and enhanced  $\beta$ 2AR downregulation in lymphocytes and human airway smooth muscle cells derived from asthmatics as compared to controls[2–7]. The  $\beta$ -2 adrenergic receptor has been mapped to chromosome 5q31-33, a region identified by several genome-wide and regional surveys as harboring an atopy or asthma-susceptibility locus[8–11]. Along with additional evidence demonstrating linkage with the 5q31-33 locus and bronchial hyperresponsiveness[12], these studies prompted the investigation of  $\beta$ 2AR as a candidate gene for asthma and its related phenotypes. Additionally,  $\beta$ 2-agonists are the mainstay of asthma therapy, but significant interindividual clinical differences in response to these medications have been noted[13]. The precise determinants of this variability are unclear, but genetic variation at the receptor level may be of importance. This series of observations have made the  $\beta$ 2AR one of the most extensively studied genes in asthma genetics and pharmacogenetics. An overview of this work illustrates the complexity of the  $\beta$ 2AR's biology, as well as the difficulties in elucidating the genetic determinants of complex disease and pharmacogenetics.

The  $\beta$ 2AR is a G-protein coupled receptor present in respiratory epithelium, airway smooth muscle and lymphocytes, and is the principle target of beta-agonist bronchodilators. The gene encoding the receptor, *ADRB2*, has been extensively studied as a candidate gene for asthma susceptibility, asthma phenotypes, and response to medications[14, 15]. While the cumulative body of evidence has not supported *ADRB2* as an asthma susceptibility gene[16, 17], results have been mixed in regard to asthma severity[18], nocturnal asthma, airway hyperresponsiveness, lung function and response to medication. Inconsistencies between studies may be due to population differences in genetic background, epistatic gene-gene interactions, or physiologic conditions that alter the receptor expression and function[16]. Additionally, epigenetic variation affecting gene expression may be an important determinant of clinical asthma phenotype.

Epigenetics describes molecular factors and processes that regulate genome activity independent of DNA sequence. One such process is the methylation of cytosine of CpG dinucleotides, frequently resulting in transcriptional silencing of neighboring genes. Programmed methylation of CpG clusters (“CpG islands”) is critical for normal development and cellular differentiation, as well as X-chromosome inactivation and genetic imprinting. CpG methylation also results from environmental stimuli[19], including in utero and environmental tobacco smoke[20, 21]. In complex diseases like asthma, DNA methylation offers a potential mechanism for environmental modification of genetic responses, including those at the *ADRB2* locus. Indeed, a 407 base pair variably methylated CpG island overlaps the 5′ untranslated region and leading coding sequence of *ADRB2*, and may represent an important source of epigenetic regulation of this gene. A single previous study has shown a positive association of methylation of the *ADRB2* gene with asthma severity in a cohort of Caucasian children[22]. It is in this context that we herein describe the variation of DNA methylation in the *ADRB2* CpG island and its relationship to clinical asthma phenotypes in a cohort of extensively phenotyped school age inner city, ethnically and racially diverse children with asthma. We aimed to determine the effect of methylation at the *ADRB2* promoter region to asthma symptoms, morbidity, and lung function. Some of the results of this study have been previously reported in abstract form[23].

## Materials and Methods

### Study Population

This study was nested in the ongoing School Inner City Asthma Study (SICAS), an epidemiologic study of the effect of environmental exposures in school classrooms and asthma morbidity in inner city school children. Recruitment is summarized elsewhere[24]. Briefly, students with asthma were screened and recruited from entire urban elementary schools. Children who had a physician’s diagnosis of asthma, which has been shown to correlate with indices of disease[25, 26], AND cough, wheezing, dyspnea, or whistling in the chest in the previous 12 months were considered as students with asthma and were eligible for the study[24]. Eligibility requirements were modeled after other National Inner-City asthma studies[27]. Exclusion criteria included significant pulmonary disease other than asthma, significant cardiovascular disease requiring daily medication, use of beta-blocker medication, and an inability to complete study procedures. For the purposes of this study, only children who provided samples for DNA extraction were selected. We obtained 136 spit saliva-derived and 41 blood-derived DNA samples from 177 participants enrolled between July 2008 and July 2010. One subject had both blood and saliva obtained at the same visit. Complete baseline questionnaire data was available for all tested outcomes for each child.

### DNA Methylation

Blood and saliva-derived DNA were collected for evaluation of *ADRB2* methylation. CpG sites were selected from the *ADRB2* CpG island promoter region upstream from the transcriptional start site for high probability of affecting transcription, and thereby gene expression.

DNA methylation assays were performed by staff who were blinded to the study (EpigenDX, Worcester, MA), by bisulfate modification followed by PCR amplification and pyrosequencing. Percent methylation at each site was calculated as:  $C\% = C \text{ Peak} / (C \text{ Peak} + T \text{ Peak})$ . Correlation of in vitro prepared standards with expected methylation percentages demonstrated an r-squared of 0.8 or higher. In addition to assay quality control measures, duplicate samples for two subjects and one blood and saliva sample from the same subject, were compared. Three CpG sites were excluded due to greater than 50% variability from the mean between duplicate samples. The distributions of methylation at each position for saliva and blood-derived DNA were comparable. Therefore, the methylation profile at each locus is a combination of these sources. For analysis, the average percent methylation across the valid methylation sites was used as the predictor. Subsequently, this was divided into 3 clades of highly correlated sites for more specific investigation of methylation regions.

### Measurement of Lung Function

At the baseline clinic visit, pre- and post-bronchodilator spirometry (Koko spirometer, nSpire Health, inc., Longmont, CO, USA) was performed according to ATS guidelines[28] and compared to predicted outcomes from the NHANES III[29]. Sixty two subjects had only pre-bronchodilator spirometry measured and therefore do not have a measure of bronchodilator response. Each spirometry session was reviewed for acceptability and repeatability according to ATS guidelines[28, 30].

### Measurement of Asthma Severity and Morbidity

Baseline caregiver questionnaire recorded clinical asthma symptoms and activities of daily life concurrent with DNA sampling. We used 4 week recall of asthma symptoms and rescue medication use to evaluate the severity of asthma symptoms and impairment each subject suffered immediately to the time of sample attainment. Specifically, caregivers were asked to report whether: (1) the child had wheeze or difficulty breathing when exercising, (2) wheezing during the day when not exercising, (3) waking up at night with wheezing or difficulty breathing,(4) missed any days of school and (5) missed any daily activities due to his/her asthma over the previous 4 weeks. In order to quantify asthma symptom severity in the context of the impairment domain of NHLBI criteria[31], several variables were derived from categorical questions regarding the frequency of shortness of breath, nighttime symptoms, and rescue medication use. Since the responses were not identical to characterizations in the guidelines, some categorizations were approximated. The characterization of symptom severity was as follows: (1) A response for shortness of breath (termed 'dyspnea' herein) was deemed positive if it occurred greater than twice weekly over the previous 4 weeks, (2) frequent nighttime awakening due to asthma was defined as one or more interrupted nights in the previous 4 weeks, and (3) frequent rescue medication use was defined as use of rescue medication greater than or equal to twice per week over the previous 4 weeks.

The number of unscheduled healthcare visits for asthma, which included emergency department visits and unscheduled clinic or doctor's office visits, and estimated number of days absent from school due to asthma over the previous 12 months were also obtained by caregiver questionnaire.

## Statistical Analysis

Univariate and multivariate regression approaches were used to evaluate the relationships between average % methylation and asthma outcomes. While the inclusion of sample type in the univariate and multivariate analyses had little effect on the outcomes, sample type was included as a covariate to minimize potential differential tissue effect. Relationships from univariate analysis between the primary predictor and outcomes of interest that met a threshold association of  $\alpha = 0.0035$  based on the Bonferroni correction for multiple testing ( $P = 0.05 \div 14 \text{ comparisons} = 0.0035$ ) were carried forward to multivariate regression analysis. Multivariate analysis also evaluated the more specific clades of highly associated (correlation  $>0.5$ ) methylation sites in order to more specifically understand the methylation site effect on clinical outcome. Age, race and gender were included in all models as potential confounders. Additional potential confounders were evaluated in each statistical model and were retained if associated with outcomes of interest at a level of  $\alpha = 0.05$  and had reasonable likelihood to be associated with methylation. In this regard, the following variables were considered: family income, environmental tobacco smoke exposure, family history (parents, siblings, grandparents) of asthma, preterm birth, physician diagnosis of hayfever/allergic rhinitis and physician diagnosis of eczema. Logistic, linear, and Poisson regression was used for respective binomial, Gaussian, and Poisson distributed outcomes. SAS version 9.2 (SAS Institute Inc., Cary, NC) was used for all statistical analysis. We performed post-hoc analyses to assess the possibility of genotype-specific methylation at amino acid position 16 (Arg16Gly).

Several steps were taken to assess the impact of including blood and saliva in a combined analysis. First, a stratified analysis limited to saliva-derived DNA or blood-derived DNA was performed for the significant associations. Second, a separate analysis of the residuals versus outcomes when sample type was included in the regression was undertaken to evaluate the confounding influence of sample type on the relationship of methylation with the asthma phenotypes. In essence, this demonstrates the effect of methylation independent of the sample source (the residual effect after sample type is accounted for) on the outcomes of interest.

## Results

### Study population

One hundred and seventy seven children enrolled in the School Inner City Asthma Study had blood or saliva-derived DNA analyzed for methylation of the *ADRB2* gene. The mean age of the study participants was 7.9 years old and gender was evenly represented. Table I illustrates the demographic and baseline characteristics of the cohort and distribution of outcome measures. The group was racially and ethnically diverse with over one third of participants reporting to be Black and another one third Hispanic. Nearly 20% were of mixed race and/or ethnicity. Based on annual household income, greater than one third of participants were impoverished. Lung function was within the normal predicted range for all subjects.

## ADRB2 methylation patterns

Eight of eleven typed *ADRB2* methylation sites met quality control criteria. The average variability from the mean for duplicate samples was 0.34, and between blood and saliva was 0.30. Figure 1 illustrates the location of the CpG sites in relation to the *ADRB2* gene as well as representative histograms for several of the methylation sites, grouped by clade. The proportion of methylation generally ranged from 0 – 6% with the average % methylation of 2.4% (Table 2). CpG sites were clustered based on high intra-site correlation of methylation (table E1). For two of the three groups, there was a high correlation of percent methylation for sites within the group ( $r^2 = 0.73$  for positions 4 and 5 [clade 2];  $r^2 = 0.75$ – $0.79$  between positions 6, 7 and 8 [clade 3] (Supplementary Table E1)). However, despite their physical proximity to one another and shared primer sets, correlation between sites 1, 2 and 3 (clade 1) were not high ( $r^2 = 0.54$  for all comparisons).

## Associations of ADRB2 methylation status with asthma severity phenotypes

We next assessed the relationship of *ADRB2* methylation to measures of asthma severity. In univariate analysis (Table 3), average % methylation demonstrated a strong association with dyspnea and notable trends with other markers of improved asthma control, including decreased asthma symptoms, less frequent rescue medication use, fewer school absentee days, and higher measures of lung function. After adjusting for age, race, gender, preterm birth, family history of asthma, diagnosis of eczema and sample source % methylation of the *ADRB2* promoter continued to exhibit a strong inverse association with dyspnea (OR = 0.2; 95% CI 0.1 – 0.6,  $p = 0.002$ ; table 4). Furthermore, sensitivity analysis by DNA sample type and, separately, the residual effect after accounting for sample type was consistent with the findings of full cohort. Each clade showed strong, but not significant, associations with decreased dyspnea (table 4).

## Methylation quantitative trait locus (mQTL) analysis

Recent data suggests that DNA sequence variation (for example, single nucleotide polymorphisms, SNPs) can confer allele-specific methylation changes at adjacent CpG sites. It is therefore possible that our observed associations of *ADRB2* CpG methylation with asthma severity phenotypes is an indirect consequence of neighboring functional DNA sequence variants. Given that a common coding variant – a substitution of Arginine for Glycine at amino acid 16 – has been inconsistently associated with asthma severity and pharmacogenetic phenotypes, we genotyped the Arg16Gly variant and tested for association with the average % methylation at the *ADRB2* promoter. We found no evidence of allele-specific differences in methylation, suggesting that our observed methylation-phenotype relationships are not due to indirect genetic effects for this SNP.

## Discussion

We found that increased DNA methylation at the promoter region of the *ADRB2* gene was associated with decreased asthma symptom severity, as measured by report of dyspnea, and trended toward an inverse association with airway obstruction measured by spirometry. These results provide evidence that epigenetic modification of the *ADRB2* gene may directly influence the clinical asthma phenotype observed in school-aged children at high risk for



asthma morbidity. This is the first evidence that increased methylation at the *ADRB2* gene may be associated with improved asthma phenotypes.

While the association demonstrated here is compelling, the interpretation of these findings in the absence of gene expression and functional assays related to *ADRB2* methylation is limited. DNA CpG hypermethylation typically leads to gene silencing and overall decreased expression of the gene product. These effects are frequently tissue specific, and understanding the potential mechanisms by which methylation of *ADRB2* may promote milder asthma requires consideration of the main asthma-relevant tissues in which *ADRB2* is expressed: airway epithelium and smooth muscle, and peripheral blood mononuclear cells (PBMCs).

In the airways, increased  $\beta_2$ AR density has been associated with fatal and severe asthma in children and adults[32][33], however these findings are inconsistent[34]. In untreated adults with mild asthma there is an inverse relationship between FEV1 percent predicted and pulmonary receptor density measured by positron emission tomography (PET)[35]. Additionally, gene expression assays have demonstrated increased *ADRB2* expression in bronchial mucosa in adults with severe asthma compared to those with mild to moderate disease[36] and maximal expression in those with persistent airflow limitation[37]. Therefore, it is plausible that suppressing beta 2 adrenergic receptor expression by methylation of *ADRB2* in the bronchial mucosa and smooth muscle may lead to a milder asthma phenotype.

In peripheral mononuclear cells, the  $\beta_2$ AR is known to mediate a proinflammatory pathway favoring the expression of Th2 cytokines[38, 39]. DNA methylation, among other epigenetic modulations, was recently shown decrease *ADRB2* gene expression in Th2 driven cells in a mouse model[40]. In regards to the current study in which genetic analysis from blood and spit saliva, both samples representative of mononuclear cells, the effect of increased methylation on improved asthma symptoms and suggestion of better airway caliber may be mediated through decreased expression of Th2 cytokines. Mechanistically, this would favorably affect airways inflammation, thereby mitigating airway obstruction and asthma symptoms.

Despite these associations, results from pharmacologic studies have demonstrated expression of *ADRB2* in airway epithelial cells and reduced airway hyperresponsiveness[41] and similarly, drug induced downregulation of leukocyte  $\beta_2$ AR has been associated with increased symptoms and worsened lung function[4, 6, 42]. These studies suggest that decreased *ADRB2* expression leads to greater airway hyperresponsiveness and a  $T_H2$  predominant phenotype, contrary to the paradigm we propose. Fu, et al.[22] recently demonstrated findings consistent with this alternative mechanistic hypothesis. In their cohort of school-aged children they found a positive relationship between whole blood *ADRB2* methylation and asthma severity. Both our study and that by Fu et al. demonstrate similar distribution of methylation, age of participants and definition of asthma, though the methylated region of *ADRB2*, the population, and the timing of assessment differed substantially. The current study enrolled a racially and ethnically diverse population and measured outcomes at the time of genetic sample acquisition, and focused on methylation

upstream of the transcriptional start site of *ADRB2*, while Fu et al. studied methylation of the 5' UTR downstream of the transcriptional start site in Caucasian children and assessed outcomes over the subsequent year as annualized severity[22].

This study has several potential limitations. We studied DNA isolated from either whole blood or saliva (which contains a mix of leukocytes and monocytes[44] as well as buccal epithelial cells), raising the possibility that the heterogeneity of these cell types may influence our results due to tissue specific differences in methylation patterns. Several lines of investigation reassure us that this is not the case. First, saliva and blood methylation demonstrated similar distributions at each site. Second, the relationship of methylation to the outcomes was not altered by the inclusion of sample type as a covariate. Third, completely removing the effect of sample source by analyzing the residuals from the main analysis with the outcomes did not substantially change the findings of this analysis. Finally, we performed a sensitivity analysis stratified by sample type which confirmed the effect size and directionality of the main study results. These analyses demonstrate that despite the lack of adequate sample size to find significant effect, the findings in the selected tissue are consistent with the findings of the overall cohort. Together, these results are reassuring that our findings are not due to the use of two tissue types. We are further limited in the narrow portion of the *ADRB2* gene in which we focused for assessment of methylation. This region was chosen for its high probability for affecting gene transcription and thereby, expression, however, we concede that other regions may be important in this regard. Furthermore, expression and function studies are lacking for the *ADRB2* gene and the influence of methylation and were beyond the scope of this study.

The findings presented here are an important step in elucidating the relationship between methylation at the *ADRB2* gene clinical asthma phenotypes. Mechanistic studies to determine the effect of methylation on gene transcription and B2AR function are needed to fully understand the consequences of methylation of *ADRB2*. Vigorous investigation of epigenetic DNA modification of *ADRB2* may offer the opportunity to discover new pathways in the disease and new pharmacologic targets[45]. Clarifying the role of epigenetic effects on the *ADRB2* gene, as well as other candidate genes, allows for further understanding of complex asthma phenotypes by determining functional modifications to genomic expression. Extensive allele specific and haplotype specific methylation has been described across the genome[46–49]. While there was no genotype-specific methylation pattern at the ARG16GLY position in our data, it will be important to investigate the relationship between methylation of the *ADRB2* gene and a more exhaustive array of polymorphisms in future studies before concluding that methylation is independent of genotype. We anticipate that integrative studies that explore the joint contributions of both genetic and epigenetic variation on phenotype will advance our understanding of complex diseases, particularly those with both significant genetic and environmental contributions, such as asthma.

## Supplementary Material

Refer to Web version on PubMed Central for supplementary material.



## Acknowledgments

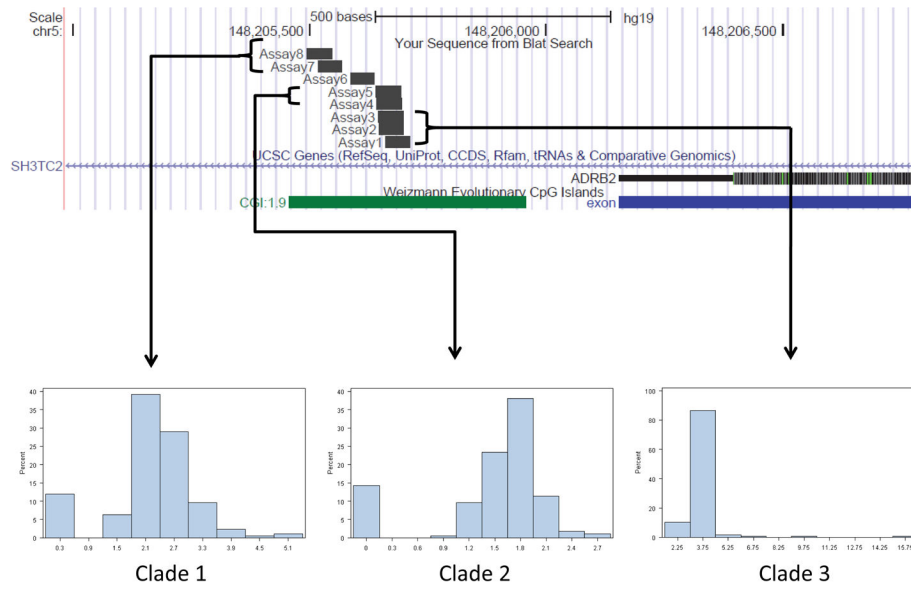
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**Figure 1.** Upper panel: UCSC genome browser map of methylation sites investigated. Lower Panel: selected histograms for methylation Clades. UCSC Genome Browser on Human Feb. 2009 (GRCh37/hg19) Assembly.

**Table 1**

Descriptive characteristics of the School Inner City Asthma Study participants

Characteristic	Number of Subjects (%), except where noted
Female	80 (45.2)
<b>Race</b>	
White	8 (4.5)
Black	67 (37.9)
Hispanic	61 (34.5)
Mixed	33 (18.6)
Other	8 (4.5)
Age, years, mean (SD)	7.9 (2.0)
<b>Poverty</b>	
Household income <\$25,000	67(37.9)
Household income \$25,000	83 (46.9)
Family history of asthma <sup>d</sup>	137 (77.4)
Diagnosis of eczema	88 (49.7)
Diagnosis of allergic rhinitis	29 (16.4)
<b>Environmental tobacco smoke exposure</b>	
Never	48 (27.1)
Occasional or more frequent	129 (72.9)
Preterm birth <sup>b</sup>	25 (14.1)
<b>Spirometry<sup>c</sup></b>	
FEV1 percent predicted, mean (SD)	107.2 (20.1)
FEV1/FVC ratio × 100, mean (SD)	88.2 (6.3)
Bronchodilator response, mean % change (SD)	12.4 (14.1)
School absences due to asthma in previous 12 months, Days, median (IQR)	4 (1, 8)
Number of unscheduled healthcare visits for asthma in previous 12 months, median (range)	0 (0,1)
Dyspnea <sup>d</sup>	26 (14.7)
Frequent rescue medication use <sup>d</sup>	40 (22.6)
Frequent nighttime awakening due to asthma <sup>d</sup>	82 (46.3)
Perceived poor asthma control <sup>d</sup>	48 (27.1)
Wheeze/dyspnea with exercise <sup>e</sup>	78 (44.1)
Wheeze/dyspnea without exercise <sup>e</sup>	37 (20.9)
Awaken with wheeze/dyspnea <sup>e</sup>	61 (34.5)
School absence due to asthma <sup>e</sup>	39 (22.0)
Missed any daily activities due to asthma <sup>e</sup>	40 (22.6)

SD, standard deviation; FEV1, forced expiratory volume in 1 second; FEV1/FVC, FEV1 per forced vital capacity; Bronchodilator response = ((post-beta agonist FEV1 – pre- beta agonist FEV1)/ post-beta agonist FEV1) × 100.

<sup>a</sup>Family history includes immediate family plus grandparents

<sup>b</sup>less than 37 weeks gestational age

<sup>c</sup> FEV1 percent predicted and FEV1/FVC n=145, Bronchodilator response n=93

<sup>d</sup> Dyspnea: shortness of breath greater than twice weekly over the previous 4 weeks; Frequent rescue medication use: determined as twice weekly over previous 4 weeks; Frequent nighttime awakening due to asthma: Greater than twice in the previous 4 weeks; perceived poor asthma control: caregiver report regarding control over previous 4 week period.

<sup>e</sup> Reported presence of symptom over previous 4 week period.



**Table 2**  
Distribution of percent methylation of CpG sites in  $\beta$ -2 Adrenergic Receptor gene

	CpG Sites	Location, Chromosome 5 $\alpha$	Mean, %	Standard Deviation	Interquartile range
Clade 1	Position 1	148205686-7	3.1	0.5	2.8, 3.2
	Position 2	148205673-4	3.5	1.1	3.1, 3.6
	Position 3	148205671-2	2.7	0.5	2.5, 3.0
Clade 2	Position 4	148205669-70	1.3	0.7	1.2, 1.6
	Position 5	148205667-8	1.5	0.7	1.4, 1.8
	Position 6	148205612-3	2.1	1.0	1.9, 2.7
Clade 3	Position 7	148205542-3	2.2	0.8	1.9, 2.6
	Position 8	148205520-1	2.8	0.6	2.4, 3.0
	Average % Methylation	148205520 - 148205687	2.4	0.5	2.2, 2.7

<sup>a</sup>UCSC Genome Browser on Human Feb. 2009 (GRCh37/hg19) Assembly.

**Table 3**

Univariate analysis of average %methylation of ADRB2 and asthma outcomes

Outcome Severity <sup>a</sup>	Mean ADRB2 Methylation		
	Odds Ratio	95% confidence interval	P-value
Dyspnea	0.2	0.1 – 0.5	0.0006
Frequent nighttime awakening due to asthma	0.7	0.4 – 1.3	0.22
Frequent rescue medication use	0.4	0.2 – 0.8	0.01
Perceived poor asthma control	0.6	0.3 – 1.2	0.14
Wheeze/dyspnea with exercise	0.6	0.3 – 1.1	0.08
Wheeze/dyspnea without exercise	0.6	0.3 – 1.3	0.21
Awaken with wheeze/dyspnea	0.7	0.4 – 1.3	0.22
School absence due to asthma	0.7	0.3 – 1.4	0.26
Missed any activities due to asthma	0.6	0.3 – 1.2	0.14
<b>Spirometry<sup>b</sup></b>			
FEV1%	1.4	-5.0 – 7.8	0.67
Post bronchodilator change in FEV1	-2.9	-8.2 – 2.4	0.28
FEV1/FVC Ratio	1.0	-1.0 – 2.9	0.30
No. of days of school absence due to asthma/12 months	-0.2	-0.7 – 0.3	0.47
No. of unscheduled healthcare visits due to asthma/12 months	-0.1	-0.6 – 0.4	0.76

<sup>a</sup>Severity parameters tested using logistic regression adjusting for sample type

<sup>b</sup>Quantitative outcomes tested by linear regression analysis for spirometry outcomes and Wald test for Poisson distribution for number of school absences and number of unscheduled healthcare visits, beta coefficients are reported. All analyses adjusted for sample type.

**Table 4**

The association between percent methylation of ADRB2 and Dyspnea

Predictor	Odds Ratio <sup>a</sup>	95% Confidence Interval	Dyspnea		p-value
			% Methylation Positive Response mean (SD) <sup>b</sup>	% Methylation Negative Response mean (SD) <sup>b</sup>	
Average % methylation of ADRB2	0.2	0.1 – 0.6	2.1 (0.5)	2.4 (0.5)	0.002
Clade 1	0.2	0.04 – 0.7	2.9 (0.5)	3.1 (0.6)	0.01
Clade 2	0.4	0.2 – 0.8	1.1 (0.7)	1.5 (0.6)	0.01
Clade 3	0.4	0.2 – 0.7	2.0 (0.6)	2.4 (0.7)	0.004

<sup>a</sup> adjusted for age, race, gender, preterm birth, family history of asthma, diagnosis of eczema and sample type<sup>b</sup> means derived from univariate comparisons