

Draft Genome Sequence of Quorum-Sensing and Quorum-Quenching Pseudomonas aeruginosa Strain MW3a

Kok-Gan Chan, Cheng Siang Wong, Wai-Fong Yin, Xin Yue Chan

Division of Genetics and Molecular Biology, Institute of Biological Sciences, Faculty of Science, University of Malaya, Kuala Lumpur, Malaysia

Pseudomonas aeruginosa has a broad range of habitation, from aquatic environments to human lungs. The coexistence of quorum-sensing and quorum-quenching activities occurs in *P. aeruginosa* strain MW3a. In this work, we present the draft genome sequence of *P. aeruginosa* MW3a, an interesting bacterium isolated from a marine environment.

Received 5 March 2014 Accepted 26 March 2014 Published 17 April 2014

Citation Chan K-G, Wong CS, Yin W-F, Chan XY. 2014. Draft genome sequence of quorum-sensing and quorum-quenching *Pseudomonas aeruginosa* strain MW3a. Genome Announc. 2(2):e00258-14. doi:10.1128/genomeA.00258-14.

Copyright © 2014 Chan et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 3.0 Unported license.

Address correspondence to Kok-Gan Chan, kokgan@um.edu.my.

Pseudomonas aeruginosa is classified under the Gammaproteo-bacteria and lives in a broad range of environments, including natural environments, eukaryotic hosts, and man-made products (1–3). P. aeruginosa utilizes its quorum-sensing ability to communicate within its community in order to coordinate community behaviors that enhance adaptivity to various environments (4–6). Aside from quorum sensing, P. aeruginosa also possesses quorum-quenching mechanisms that enzymatically degrade its quorum-sensing signaling molecules (7). The degradation of its quorum-sensing signaling compound enables the self-regulation of P. aeruginosa quorum sensing, while the degraded compound acts as the bacterial growth nutrient (7, 8).

P. aeruginosa strain MW3a was isolated from the subsurface level (5 cm beneath sea level) in the Strait of Malacca using KGm medium (9, 10). Preliminary studies have shown that this isolate possesses both quorum-sensing and quorum-quenching abilities. N-Dodecanoyl-L-homoserine lactone and N-3-oxotetradecanoyl-L-homoserine lactone were detected from the spent supernatant of this isolate by high-resolution mass spectrometry. In addition, N-acyl-homoserine lactone (AHL) degradation was observed by rapid-resolution liquid chromatography, with a preference on AHL with a 3-oxo group substitution.

Total genomic DNA of *P. aeruginosa* strain MW3a was extracted and purified with the QIAamp DNA minikit (Qiagen, Germany). Subsequently, the purified genomic DNA was subjected to whole-genome shotgun sequencing on an Illumina HiSeq (Illumina, Inc., USA) platform. Quality reads were *de novo* assembled with the CLC Genomics Workbench 6.0.5 (CLC bio, Denmark). Gene prediction was performed with the prokaryotic gene prediction algorithm Prodigal (version 2.60) (11), while rRNAs were predicted with RNAmmer (12). Subsequently, the genome sequence was annotated with BLASTx against the NCBInt/nr and UniProt databases (13, 14).

The whole-genome sequencing generated 25,826,420 pairedend reads, with an average length of 101 bp. The filtered reads were de novo assembled into 240 contigs with a length of \geq 200 bp, and an N_{50} of 81.6 kb was generated. The draft genome of *P. aeruginosa* MW3a contains 6,665,300 bases, with an average coverage of 366fold and a G+C content of 66.29%. The gene prediction resulted in 6,288 open reading frames (ORFs), and a copy each of 5S rRNA, 16S rRNA, and 23S rRNA was identified.

Based on the BLAST result, two AHL-based quorum-sensing homologs were detected from the draft genome of *P. aeruginosa* MW3a. The *rhl* quorum-sensing system, which is responsible for short-chain AHL synthesis, was found in contig 7, while the *las* system, which synthesizes long-chain AHL, was carried in contig 158. On the other hand, the *quiP* and *pvdQ* genes that encode AHL acylase were found in contigs 98 and 107, respectively.

Nucleotide sequence accession numbers. This whole-genome shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession no. JAQK00000000. The version described in this paper is the first version, JAQK01000000.

ACKNOWLEDGMENTS

Kok-Gan Chan thanks the University of Malaya for the financial support given under the High Impact Research Grant (UM-MOHE HIR Nature Microbiome grants UM.C/625/1/HIR/MOHE/CHAN/14/1 and H-50001-A000027).

REFERENCES

- Chong TM, Koh CL, Sam CK, Choo YM, Yin WF, Chan KG. 2012. Characterization of quorum sensing and quorum quenching soil bacteria isolated from Malaysian tropical montane forest. Sensors (Basel) 12: 4846–4859. http://dx.doi.org/10.3390/s120404846.
- Erickson DL, Endersby R, Kirkham A, Stuber K, Vollman DD, Rabin HR, Mitchell I, Storey DG. 2002. Pseudomonas aeruginosa quorumsensing systems may control virulence factor expression in the lungs of patients with cystic fibrosis. Infect. Immun. 70:1783–1790. http://dx.doi .org/10.1128/IAI.70.4.1783-1790.2002.
- Lanini S, D'Arezzo S, Puro V, Martini L, Imperi F, Piselli P, Montanaro M, Paoletti S, Visca P, Ippolito G. 2011. Molecular epidemiology of a *Pseudomonas aeruginosa* hospital outbreak driven by a contaminated disinfectant-soap dispenser. PLoS One 6:e17064. http://dx.doi.org/10.1371/journal.pone.0017064.
- 4. Lee J, Wu J, Deng Y, Wang J, Wang C, Wang J, Chang C, Dong Y, Williams P, Zhang LH. 2013. A cell-cell communication signal integrates quorum sensing and stress response. Nat. Chem. Biol. 9:339–343. http://dx.doi.org/10.1038/nchembio.1225.
- 5. Waters CM, Bassler BL. 2005. Quorum sensing: cell-to-cell communi-

- cation in bacteria. Annu. Rev. Cell Dev. Biol. 21:319–346. http://dx.doi.org/10.1146/annurev.cellbio.21.012704.131001.
- Cámara M. 2006. Quorum sensing: a cell-cell signalling mechanism used to coordinate behavioral changes in bacterial populations. Membr. Comput. 4361:42–48. http://dx.doi.org/10.1007/11963516_3.
- Sio CF, Otten LG, Cool RH, Diggle SP, Braun PG, Bos R, Daykin M, Cámara M, Williams P, Quax WJ. 2006. Quorum quenching by an N-acyl-homoserine lactone acylase from *Pseudomonas aeruginosa* PAO1. Infect. Immun. 74:1673–1682.
- Huang JJ, Han JI, Zhang LH, Leadbetter JR. 2003. Utilization of acylhomoserine lactone quorum signals for growth by a soil pseudomonad and *Pseudomonas aeruginosa* PAO1. Appl. Environ. Microbiol. 69: 5941–5949. http://dx.doi.org/10.1128/AEM.69.10.5941-5949.2003.
- 9. Wong CS, Yin WF, Choo YM, Sam CK, Koh CL, Chan KG. 2012. Coexistence of quorum-quenching and quorum-sensing in tropical marine *Pseudomonas aeruginosa* strain MW3A. World J. Microbiol. Biotechnol. 28:453–461. http://dx.doi.org/10.1007/s11274-011-0836-x.
- 10. Chan KG, Yin WF, Sam CK, Koh CL. 2009. A novel medium for the

- isolation of N-acylhomoserine lactone-degrading bacteria. J. Ind. Microbiol. Biotechnol. 36:247-251. http://dx.doi.org/10.1007/s10295-008-0491-x.
- Doug H, Gwo-Liang C, Philip LC, Miriam L, Frank L, Loren H. 2010. Prodigal: prokaryotic gene recognition and translation initiation site identification. BMC Bioinformatics 11:119. http://dx.doi.org/10.1186/1471-2 105-11-119.
- Lagesen K, Hallin P, Rødland EA, Staerfeldt HH, Rognes T, Ussery DW. 2007. RNAmmer: consistent and rapid annotation of ribosomal RNA genes. Nucleic Acids Res. 35:3100-3108. http://dx.doi.org/10.1093/nar/gkm160.
- 13. Apweiler R, Bairoch A, Wu CH, Barker WC, Boeckmann B, Ferro S, Gasteiger E, Huang H, Lopez R, Magrane M, Martin MJ, Natale DA, O'Donovan C, Redaschi N, Yeh LS. 2004. UniProt: the universal protein knowledgebase. Nucleic Acids Res. 32:D115–D119.
- 14. Chan XY, Chua KH, Puthucheary SD, Yin WF, Chan KG. 2012. Draft genome sequence of an *Aeromonas* sp. strain 159 clinical isolate that shows quorum-sensing activity. J. Bacteriol. 194:6350. http://dx.doi.org/10.1128/JB.01642-12.