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Immune Enteral Nutrition Can Improve Outcomes in Medical-Surgical Patients with ARDS: A Prospective Randomized Controlled Trial

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Abstract

Objective—To determine if early continuous enteral feeding of a diet containing eicosapentaenoic acid (EPA), gamma-linolenic acid (GLA), docosahexaenoic acid, and antioxidants in surgical-medical patients with ARDS improves Lung Injury Score (LIS), gas exchange, Multiple Organ Dysfunction (MOD) Score, ICU length of stay, and days on mechanical ventilation.

Methods—Prospective randomized 2-center double-blind controlled trial of 17 ARDS patients whom continuously tube-fed the experimental diet (n=9) or an isonitrogenous, isocaloric standard diet (n=8) at a minimum caloric delivery of 90% of basal energy expenditure.

Results—In the experimental group, there was a decrease in lung injury score ($p < 0.003$) and lower ventilation variables ($p < 0.001$). Patients in the experimental group had a statistically

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significant decrease in 28-day MOD score ($p < 0.05$). The length of ICU stay was significantly decreased in the experimental group (12.8 vs. 17.5 days; $p = 0.01$). The study was underpowered to detect any survival benefits between the two groups.

Conclusion—An EPA and GLA supplemented diet contributes to improved gas exchange in addition to decrease LIS, MOD scores and length of ICU stay in patients with ARDS. An EPA +GLA-enriched enteral diet may be an effective tool in the medical management of ARDS.

Keywords

Acute respiratory distress syndrome (ARDS); Eicosapentaenoic acid; Gamma-linolenic acid; Enteral nutrition; Antioxidant; Multiple organ dysfunction

Background

Acute respiratory distress syndrome (ARDS) is a complex multifactorial illness manifest clinically as refractory hypoxemia and pulmonary edema. Nearly 200,000 new cases of ARDS occur annually in US with a mortality rate of 32–45% [1]. It may be precipitated by acute inflammatory disorders secondary to trauma, burns, sepsis, pneumonia, pneumonitis or inhalation lung injury. Alterations in lung function are evident during early systemic responses to inflammatory effectors including complement activation and release and modulation of eicosanoids [2]. These events promote macrophage and neutrophil (PMN) accumulation and migration into the alveolar space, resulting in endothelial and alveolar damage by lysosomal contents and toxic oxygen species [3–8].

Dietary fatty acids may be oxidized for energy, stored in adipose tissue, or further metabolized to various long-chain polyunsaturated fatty acids (PUFA). Membrane derived PUFA's serve as substrates for formation of eicosanoid effectors (omega-3 and omega-6) of the inflammatory response to infection or injury, Figure 1 [9]. Arachadonic acid (AA) derived Thromboxane A₂ promotes vasoconstriction and platelet aggregation [10]. While, Leukotrienes C₄, D₄ and E₄ generated by 5-lipoxygenase induce arteriolar constriction and increase post-capillary permeability, leading to edema [10–13]. In the clinical setting, the over production and release of these cell effectors appear to be involved in the etiology of ARDS [14].

Experimental and clinical studies have shown that the host's immune response to a given stimulus can be down regulated by nutritional pretreatment with diets enriched with fish oil fatty acids including eicosapentaenoic (EPA) and docosahexaenoic (DHA) acids [15]. Similarly, attention has focused on borage seed oils containing relatively large amounts of gamma-linoleic acid (GLA) (18:3n6) which may be converted to dihomogammalinoleic acid (DGLA) (20:3n6) [16,17]. In a Brazilian study, amongst 165 patients with ARDS, those treated with a standard diet supplemented with EPA + GLA had significantly lower 28-day mortality, improved oxygenation status, more ventilator-free days, and lesser development of new organ dysfunction [18]. This was partially supported by an Israeli study which noted that amongst 100 ARDS patients, those treated with a standard diet supplemented with EPA + GLA showed significant improvement in oxygenation, lung compliance, and shorter ICU stays, however no statistical difference in survival was observed [19].

Methods

The protocol was a prospective, randomized controlled double-blind study in mixed surgical-medical intensive care unit (ICU) at two tertiary-care academic teaching hospitals in the United States over 18-months period (Table 1). Critically ill patients with a diagnosis of a condition resulting in ARDS and meeting inclusion criteria were considered for enrollment in the study and randomized into one of two the treatment groups. Inclusion criteria were age 18–80 years old; condition resulting in ARDS as defined by the American-European Consensus Conference [20]; endotracheal and enteral feeding tubes in place; Modified Lung Injury Scores (LIS) > 2.5; and multiple organ dysfunctions (MOD) score < 9. The lung injury score (LIS) was calculated by averaging the chest roentgenogram score, PaO₂/FIO₂ score, PEEP score, and respiratory system compliance score as proposed by Murray et al. (Table 2) [21]. The multiple organ dysfunction score (MOD), constructed using simple physiologic measures of dysfunction in six organ systems (Table 4) [22], mirrors organ dysfunction as the intensivist sees it and correlates strongly with the ultimate risk of ICU mortality and hospital mortality, Table 3. Patients were excluded if any of the following were present: left ventricular heart failure defined as pulmonary capillary wedge pressure > 18 mmHg or left ventricular ejection fraction < 40%.; lung cancer (primary or metastatic); acute lymphoblastic leukemia; active gastrointestinal bleed; immune suppression such as recent chemotherapy, prednisone more than 0.25 mg.Kg⁻¹.day⁻¹ or HIV disease; Glasgow Coma Scale < 5 secondary to head trauma; pregnancy; and admission MOD score > 9.

Patients with a clinical diagnosis of ARDS were enrolled by the study nurses who determined their APACHE III [23] score and randomly assigned each patient a number dedicating them to either the experimental (Oxepa[®] Ross Labs, Chicago, Illinois, USA) or control (Pulmocare[®] Ross Labs, Chicago, Illinois, USA) enteral formula on an alternating basis, Table 2. She retained the only copies of the full randomization schedule that were kept in a locked safe. A pharmacist blinded to the patient's clinical condition prepared the diet according to the number assigned to the patient. The treating physicians, caregivers, patients and families remained blinded to the diet selection at all times. Resting Energy Expenditure (REE) was calculated by the Weir formula [24] on enrollment day after measuring oxygen consumption and carbon dioxide production by the Deltatrac II calorimeter (Datex Instrumentation, Helsinki, Finland; VCO₂-Deltatrac)[25].

The enteral diet was then delivered to all enrolled patients via a nasogastric, nasoduodenal, nasojejunal, or jejunostomy tubes. The enteral nutrition was delivered at a constant rate to achieve at least 65% REE within the first 24 hours, and then advanced to at least 90% REE within 72 hours. The first day that the patient received enteral nutrition at 90% REE or more was considered as study Day 1.

Patients in both EPA+GLA and control groups were ventilated by Puritan-Bennett 7200 series ventilator (Puritan Bennett, Carlsbad, CA). All the study measurements were obtained every morning between 7:00 AM and 12:00 PM before any changes were made in the patient's mechanical ventilation or oxygen delivery settings.

Enteral feeding with the EPA+GLA diet was continued for 7 days at that point enteral nutrition was converted to the standard feeding formula selected by the treating physician. Patients received enteral feeding until able to tolerate oral intake. However, daily evaluation of various clinical parameters continued until discharge from the ICU, then weekly until hospital discharge.

Outcome Measures

Primary outcome measures included oxygenation and modified Lung Injury Scores (LIS) assessed at days 1, 4, and 7. While secondary outcome measures included, Multiple Organ Dysfunction (MOD) scores, days on mechanical ventilation and total length of ICU stay.

Statistics

Patients were analyzed on an intention-to-treat basis. Statistical variables are expressed as mean values, standard deviations and medians as appropriate. Survival analysis was assessed using SAS Life Test (© SAS Institute Inc., Cary, North Carolina). When appropriate, mixed modeling was used to assess repeated measures.

Since repeated measures (respective variable) were found for each patient over a 7-day course, with respect to group, the multivariate mixed model analysis was applied to determine whether the measurements suggest significance within and between the experimental and control groups. In addition, Student's T-test was utilized for comparing the baseline values between the two groups. All *p* values were two-sided and significance was assigned at a threshold *alpha* of 0.05.

Ethics

The institutional review board at Southern Illinois University School of Medicine in addition to the review boards at Memorial Medical Center and Saint John's Hospital in Springfield, Illinois approved the protocol. A written informed consent was obtained from each patient or health care proxy before the enrollment of every study subject. Both principal investigators and the clinical coordinator were available 24-hours-per-day throughout the study to answer questions regarding safety, patient eligibility, and for reporting of any adverse events.

Results

The analysis was conducted in 17 out of 22 patients who were enrolled in the study. Reasons for exclusion from analysis was diarrhea (n=1) and withdrawal of care (n=3), see Figure 2. There were no significant differences between the two groups regarding age, gender; ICU admission diagnosis and APACHE III score at enrollment, Table 4. There was no significant difference in time from hospital admission to study entry between the study group (6.7 ± 2.3 days) and the control group (8.2 ± 3.5) ($P=0.3$). In addition, time to achievement of 90% of REE and duration of tube feeding was similar between the two groups. All enrolled patients were fed successfully for two weeks through naso-gastric, naso-duodenal or naso-jejunal route.

Primary outcome measures

Oxygenation and modified lung injury scores (LIS) oxygenation

There were no statistically significant baseline differences between the two groups regarding PaO₂, FiO₂, or PaO₂/FiO₂ ratio. Applying a multivariate mixed model analysis, results for PaO₂/FiO₂ ratio in the EPA+GLA group were statistically significant through Day 2 ($p < 0.01$), and then stabilized by day 7 (Table 5).

In addition, the LIS was calculated on entry to the study, and then on study days 1, 4 and 7. Applying a multivariate mixed model analysis; the results demonstrated a statistically significant difference in the EPA+GLA group from Day 1 through Day 4 with ($p < 0.003$). In addition, there was a statistically significant decline in LIS in the EPA+GLA group on Day 7 comparing to pretreatment value, Table 6.

Furthermore, a comparison of ventilation variables on days 1, 4, and 7 are listed in Table 7. Although ventilation variables (FIO₂, positive end-expiratory pressure, and minute ventilation) were similar between the two groups by day 7, lower ventilation variables were recorded in the EPA+GLA group patients with APACHE III scores >25 on admission to the ICU compared to the controls ($p < 0.01$).

Secondary outcome measures

Multiple organ dysfunctions—We investigated the incidence of development of new organ dysfunction in both groups. By applying a multivariate mixed model analysis, the overall results demonstrated a trend toward lower MOD scores for the EPA+GLA group from Day 1 through 7 ($p < 0.06$) which was statistically remains significant by day 28 ($p < 0.05$), Figure 3.

Length of mechanical ventilation and ICU stay—Patients fed the EPA+GLA diet supplemented with EPA, GLA, DHA, and antioxidants had a decreased length of stay in the ICU, 12.8 vs. 17.5 days for controls ($p = .01$), Figure 4. In addition, there was a trend towards a decreased number of ventilator dependant days in the EPA+GLA vs. control group (6.7 vs. 8.2), however these results failed to reach a statistical significance difference ($p = 0.3$).

Finally, although mortality was not an outcome measure, at the end of 28 days 0 of 9 patients died in the EPA+GLA group compared to 1 of 8 in the control group. However, such difference was not statistically significant, ($p=0.3$).

Discussion

Early animal studies illustrated that diets supplemented with EPA and GLA may have beneficial effects in models of acute lung injury. A swine study suggested that EPA and GLA supplemented diets attenuated acute lung injury following endotoxin challenge [26,27].

Later human studies examined the immunologic effects of feeding ICU patients EPA, DHA, GLA and antioxidants supplemented diets [28,29]. The aim effects were changes on pulmonary inflammation, eicosanoid mediators, endogenous antioxidants, and pulmonary function in patients with acute pulmonary inflammation.

Other multi-center prospective randomized clinical trials examining the effects of early enteral administration of a formula supplemented with arginine, nucleotides, and fish oil in ICU patients found that such a strategy was safe, well tolerated and contributed to a significant reduction in hospital length of stay and the frequency of nosocomial infection [30].

Several studies investigated the role for EPA and GLA enhanced diets in a variety of patient populations [31–36]. In one such study, patients with ARDS or ALI fed an enteral diet supplemented with Oxepa enteral nutrition. The experimental group patients had reduced pulmonary neutrophil recruitment with improved oxygenation, significantly less number of ventilator-dependant days, shorter length of ICU stay, and decreased mortality compared to patients fed the control diet (Pulmocare) [31].

In our study, we expanded on the previously reported benefits of enteral diets enriched with Ω -3 and Ω -6 fatty acids and examined the effect of such a diet on the MOD score. Unlike number of the previous studies, we utilized the APACHE III score for objective comparison of the disease severity on entry to the study [18,19,31]. Furthermore, to maximize nutrient intake, we established enteral nutrition at 90% of REE within 72 hours of initiation of enteral feeding as compared to the 75% noted in another study [18].

In the present study, there was an early statistically significant difference in the $\text{PaO}_2/\text{FiO}_2$ ratio from Day 1 to Day 2 but then stabilized and became not significant for the remainder of the study. However, the LIS scores significantly improved in the EPA+GLA group from Day 1 to Day 4 with significant decline on Day 7 comparing to pretreatment value. Such finding again supports the better utility of the LIS comparing to the PaO_2 or $\text{PaO}_2/\text{FiO}_2$ ratio in assessment of pulmonary response to different therapeutic interventions [37]. Another interesting findings in this study was that patients with higher enrollment APACHE III scores displayed the greatest benefits when fed EPA+GLA. The latter had lower FiO_2 requirements, positive endexpiratory pressure, and minute ventilation requirements compared to controls ($p < 0.01$). This is somewhat similar to the findings from the PROWESS study, which showed that severe septic patients with the higher APACHE II scores, rather than APACHE III in our study had a lower mortality after treatment with recombinant activated protein C [38].

Another unique design feature of our study was the use of MOD for objective assessment of organs failure rather than relying on the percent of patients who developed a new organ failure in each group [31]. Overall, we found that patients who were fed EPA+GLA enriched diets had a moderately significant decrease in MOD score from days 1 through 7, reaching statistical significance by study day 28 ($p < 0.05$), Figure 3. These improvements in patient status translated into decreased length of ICU stay in the EPA+GLA fed group (12.8 vs. 17.5 days; $p = 0.01$) compared to the control group, Figure 4. Considering the daily ICU

expenditure in caring for ARDS patients, such decrease in length of ICU stay is expected to offset the cost of the EPA+ GLA enhanced enteral diet many fold. However, this study was not designed to assess that question and a larger pharmacoeconomics study with larger patient numbers is needed to investigate such issue

Finally, although mortality in both groups was recorded it was not one of the study outcome measures. In addition, the small number of patients enrolled in the current study precludes any solid conclusions to the survival benefits of the EPA+GLA enteral diet.

Limitations

The small sample size is a limiting factor in this study. In the areas where we did not find statistical significance, it is possible that significance exists but the study was underpowered to detect it. However, the magnitude of the clinical advantage detected even with the small sample size may suggest efficacy of the intervention. Nevertheless, our data will still need further validation in a prospective independent sample of patients in a much larger multi-center study. The primary author of the present study is currently conducting a larger clinical trial to investigate the survival benefits of the EPA+GLA enteral diet in an independent sample of ICU patients.

Conclusion

The findings of this non-pharmaceutically sponsored study support the previously reported benefits of an enteral diet supplemented with EPA, GLA, DHA and antioxidants on gas exchange and of length of ICU stay of ALI & ARDS patients. In addition, our study demonstrated that patients with higher APACHE III scores had additional benefit of decreasing 28-day MOD scores by receiving the EPA+GLA enteral diet. When one takes into account the low cost of this enteral therapy with its safety and minimal side effect profile, EPA+GLA-enriched enteral therapy may prove to be an integral component of our management strategy for ALI and ARDS. The latter is in agreement with the various scientific organizations clinical practice guidelines for nutrition support of mechanically, ventilated critically ill adult patients [39,40].

Abbreviations

ARDS	Acute Respiratory Distress Syndrome
ALI	Acute Lung Injury
AA	Arachadonic Acid
DHA	Docosahexaenoic Acid
DGLA	Dihomo-gammalinoleic Acid
EPA	Eicosapentaenoic Acid
GLA	Gamma-linolenic Acid
Ω-3	Omega-3

ICU	Intensive Care Unit
LT	Leukotriene
LA	Linoleic Acid
LOX	Lipoxygenase-derived hydroxyeicosatetraenoic acids (HETEs)
MOD	Multiple Organ Dysfunction
PMN	Neutrophil
PUFA	Polyunsaturated Fatty Acids
PG	Prostaglandin
TNF	Tumor Necrosis Factor
IL	Interleukin
REE	Resting Energy Expenditure
AM	Alveolar Macrophage
APACHE Score	Acute physiology, Age, Chronic Health Evaluation Score
RNA	Ribonucleic Acid
FO	Fish Oil
CO	Corn Oil
BO	Borage Oil
TX	Thromboxane
LIS	Lung Injury Scores (LIS)

References

1. Rubenfeld GD, Caldwell E, Peabody E, Weaver J, Martin DP, et al. Incidence and outcomes of acute lung injury. *N Engl J Med.* 2005; 353:1685–1693. [PubMed: 16236739]
2. Pingleton SK. Complications of acute respiratory failure. *Am Rev Respir Dis.* 1988; 137:1463–1493. [PubMed: 3059862]
3. Tate RM, Repine JE. Neutrophils and the adult respiratory distress syndrome. *Am Rev Respir Dis.* 1983; 128:552–559. [PubMed: 6351681]
4. Weiland JE, Davis WB, Holter JF, Mohammed JR, Dorinsky PM, et al. Lung neutrophils in the adult respiratory distress syndrome. Clinical and pathophysiologic significance. *Am Rev Respir Dis.* 1986; 133:218–225. [PubMed: 3004270]
5. Weiss SJ. Tissue destruction by neutrophils. *N Engl J Med.* 1989; 320:365–376. [PubMed: 2536474]
6. Brain JD. Lung macrophages: how many kinds are there? What do they do? *Am Rev Respir Dis.* 1988; 137:507–509. [PubMed: 3345032]
7. Nogare ARD, Toews GB. Characteristics of alveolar macrophages in an animal model of resolving pulmonary inflammation. *Am Rev Respir Dis.* 1990; 142:660–667. [PubMed: 2167624]
8. Sibille Y, Reynolds HY. Macrophages and polymorphonuclear neutrophils in lung defense and injury. *Am Rev Respir Dis.* 1990; 141:471–501. [PubMed: 2405761]
9. Simopoulos AP. Omega-3 fatty acids in health and disease and in growth and development. *Am J Clin Nutr.* 1991; 54:438–463. [PubMed: 1908631]

10. Kinsella JE, Broughton KS, Whelan JW. Dietary unsaturated fatty acids: interactions and possible needs in relation to eicosanoid synthesis. *J Nutr Biochem.* 1990; 1:123–141. [PubMed: 15539195]
11. Chauncey JB, Simon RH, Peters-Golden M. Rat alveolar macrophages synthesize leukotriene B₄ and 12-hydroxyeicosatetraenoic acid from alveolar epithelial cell-derived arachidonic acid. *Am Rev Respir Dis.* 1988; 138:928–935. [PubMed: 2849344]
12. Lewis RA, Austen KF, Soberman RJ. Leukotrienes and other products of the 5-lipoxygenase pathway. Biochemistry and relation to pathobiology in human diseases. *N Engl J Med.* 1990; 323:645–655. [PubMed: 2166915]
13. Lindgren, JA.; Edenius, C.; Samuelsson, B. Eicosanoid metabolism and function nutritional modulation. In: Kinney, JM.; Borum, PR., editors. *Perspectives in Clinical Nutrition.* Vol. Chapter 26. Baltimore: Urban and Schwanenberg; 1989.
14. Zimmerman GA, Renzetti AD, Hill HR. Granulocyte adherence in pulmonary and systemic arterial blood samples from patients with adult respiratory distress syndrome. *Am Rev Respir Dis.* 1984; 129:798–804. [PubMed: 6721278]
15. Pomposelli JJ, Flores EA, Blackburn GL, Zeisel SH, Bistran BR. Diets enriched with n-3 fatty acids ameliorate lactic acidosis by improving endotoxin-induced tissue hypoperfusion in guinea pigs. *Ann Surg.* 1991; 213:166–176. [PubMed: 1992944]
16. Tate GA, Mandell BF, Karmali RA, Laposata M, Baker DG, et al. Suppression of monosodium urate crystal-induced acute inflammation by diets enriched with gamma-linolenic acid and eicosapentaenoic acid. *Arthritis Rheum.* 1988; 31:1543–1551. [PubMed: 2848532]
17. Tate G, Mandell BF, Laposata M, Ohliger D, Baker DG, et al. Suppression of acute and chronic inflammation by dietary gamma linolenic acid. *J Rheumatol.* 1989; 16:729–734. [PubMed: 2550629]
18. Pontes-Arruda A, Aragão AM, Albuquerque JD. Effects of enteral feeding with eicosapentaenoic acid, γ -linolenic acid, and antioxidants in mechanically ventilated patients with severe sepsis and septic shock. *Crit Care Med.* 2006; 34:2325–2333. [PubMed: 16850002]
19. Singer P, Theilla M, Fisher H, Gibstein L, Grozovski E, et al. Benefit of an enteral diet enriched with eicosapentaenoic acid and gamma-linolenic acid in ventilated patients with acute lung injury. *Crit Care Med.* 2006; 34:1033–1038. [PubMed: 16484911]
20. Bernard GR, Artigas A, Brigham KL, Carlet J, Falke K, et al. Report of the American-European Consensus conference on acute respiratory distress syndrome: definitions, mechanisms, relevant outcomes, and clinical trial coordination. Consensus Committee. *J Crit Care.* 1994; 9:72–81. [PubMed: 8199655]
21. Murray JF, Matthay MA, Luce JM, Flick MR. An expanded definition of the adult respiratory distress syndrome. *Am Rev Respir Dis.* 1988; 138:720–723. [PubMed: 3202424]
22. Marshall JC, Cook DJ, Christou NV, Bernard GR, Sprung CL, et al. Control Diet Experimental Diet Multiple organ dysfunction score: a reliable descriptor of a complex clinical outcome. *Crit Care Med.* 1995; 23:1638–1652. [PubMed: 7587228]
23. Knaus WA, Wagner DP, Draper EA, Zimmerman JE, Bergner M, et al. The APACHE III prognostic system. Risk prediction of hospital mortality for critically ill hospitalized adults. *Chest.* 1991; 100:1619–1636. [PubMed: 1959406]
24. Weir JB. New methods for calculating metabolic rate with special reference to protein metabolism. *J Physiol.* 1949; 109:1–9. [PubMed: 15394301]
25. Epstein CD, Peerless JR, Martin JE, Malangoni MA. Comparison of methods of measurements of oxygen consumption in mechanically ventilated patients with multiple trauma: the Fick method versus indirect calorimetry. *Crit Care Med.* 2000; 28:1363–1369. [PubMed: 10834679]
26. Palombo JD, Lydon EE, Chen PL, Bistran BR, Forse RA. Fatty acid composition of lung, macrophage and surfactant phospholipids after short-term enteral feeding with n-3 lipids. *Lipids.* 1994; 29:643–649. [PubMed: 7815899]
27. Palombo JD, DeMichele SJ, Lydon EE, Gregory TJ, Banks PL, et al. Rapid modulation of lung and liver macrophage phospholipid fatty acids in endotoxemic rats by continuous enteral feeding with n-3 and gamma-linolenic fatty acids. *Am J Clin Nutr.* 1996; 63:208–219. [PubMed: 8561062]
28. Bower RH, Cerra FB, Bershady B, Licari JJ, Hoyt DB, et al. Early enteral administration of a formula (impact) supplemented with arginine, nucleotides, and fish oil in intensive care unit

- patients: results of a multicenter, prospective, randomized, clinical trial. *Crit Care Med.* 1995; 23:436–449. [PubMed: 7874893]
29. Braga M, Gianotti L, Cestari A, Vignali A, Pellegatta F, et al. Gut function and immune and inflammatory responses in patients perioperatively fed with supplemented enteral formulas. *Arch Surg.* 1996; 131:1257–1264. [PubMed: 8956766]
 30. Kemen M, Senkal M, Mumme A. Early postoperativimmunonutrition: clinical outcome and cost-benefit analysis. *JPEN.* 1996; 20(Suppl):S24.
 31. Gadek JE, DeMichele SJ, Karlstad MD, Pacht ER, Donahoe M, et al. Effect of enteral feeding with eicosapentaenoic acid, gamma-linolenic acid, and antioxidants in patients with acute respiratory distress syndrome. Enteral Nutrition in ARDS Study Group. *Crit Care Med.* 1999; 27:1409–1420. [PubMed: 10470743]
 32. Heller AR, Rossler S, Litz RJ, Stehr SN, Heller SC, et al. Omega-3 fatty acids improve the diagnosis-related clinical outcome. *Crit Care Med.* 2006; 34:972–979. [PubMed: 16484909]
 33. Pacht ER, DeMichele SJ, Nelson JL, Hart J, Wennberg AK, et al. Enteral nutrition with eicosapentaenoic acid, gamma-linolenic acid, and antioxidants reduces alveolar inflammatory mediators and protein influx in patients with acute respiratory distress syndrome. *Crit Care Med.* 2003; 31:491–500. [PubMed: 12576957]
 34. Mayes T, Gottschlich MM, Kagan RJ. An evaluation of the safety and efficacy of an anti-inflammatory, pulmonary enteral formula in the treatment of pediatric burn patients with respiratory failure. *J Burn Care Res.* 2008; 29:82–88. [PubMed: 18182902]
 35. Nadkarni V, DeMichele S, Goldstein B, Checchia P, Ayad O, et al. Safety of an enteral formula enriched with eicosapentaenoic and g-linolenic acids in critically ill children with acute lung injury. *Crit Care Med.* 2005; 33:A99.
 36. Elamin EM, Hughes LF, Drew D. Effect of enteral nutrition with eicosapentaenoic acid (EPA), gamma-linolenic acid (GLA), and antioxidants reduces alveolar inflammatory mediators and protein influx in patients with acute respiratory distress syndrome (ARDS). *Chest.* 2005; 128:225S.
 37. Miller AC, Rivero A, Ziad S, Smith DJ, Elamin EM. Influence of nebulized unfractionated heparin and N-acetylcysteine in acute lung injury after smoke inhalation injury. *J Burn Care Res.* 2009; 30:249–256. [PubMed: 19165116]
 38. Bernard GR, Vincent JLV, LaTerre PF, LaRosa SP, Dhainaut JF, et al. Efficacy and safety of recombinant human activated protein C for severe sepsis. *NEJM.* 2001; 344:699–709. [PubMed: 11236773]
 39. Kreymann KG, Berger MM, Deutz NE, Hiesmayr M, Jolliet P, et al. ESPEN Guidelines on Enteral Nutrition: Intensive Care. *Clin Nutr.* 2006; 25:210–223. [PubMed: 16697087]
 40. McClave SA, Martindale RG, Vanek VW, McCarthy M, Roberts P, et al. Guidelines for the Provision and Assessment of Nutrition Support Therapy in the Adult Critically Ill Patient: Society of Critical Care Medicine and American Society for Parenteral and Enteral Nutrition(A.S.P.E.N.). *JPEN J Parenter Enteral Nutr.* 2009; 33:277–316. [PubMed: 19398613]

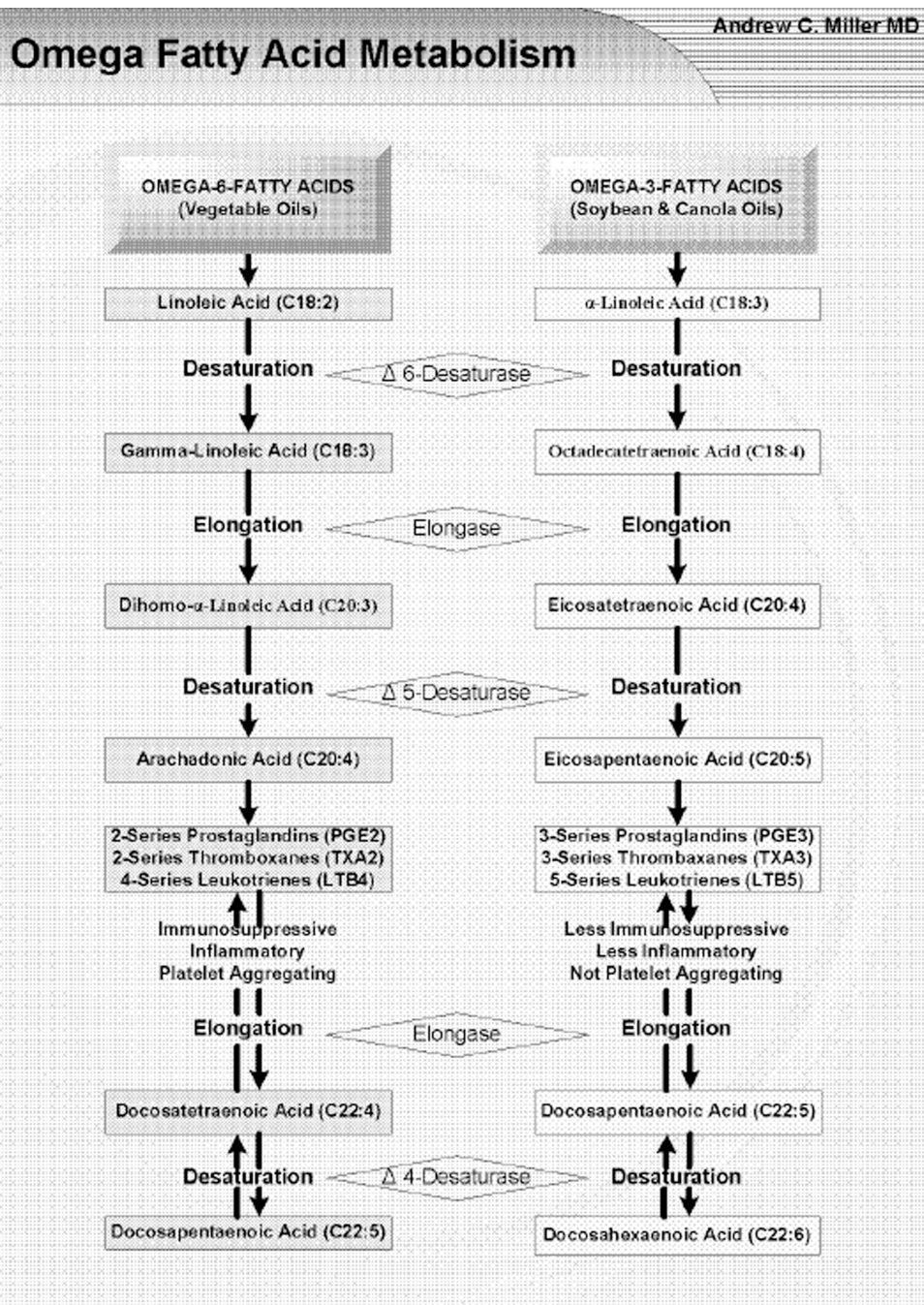


Figure 1.
The metabolism of Omega-3- and Omega-6-Fatty Acids.

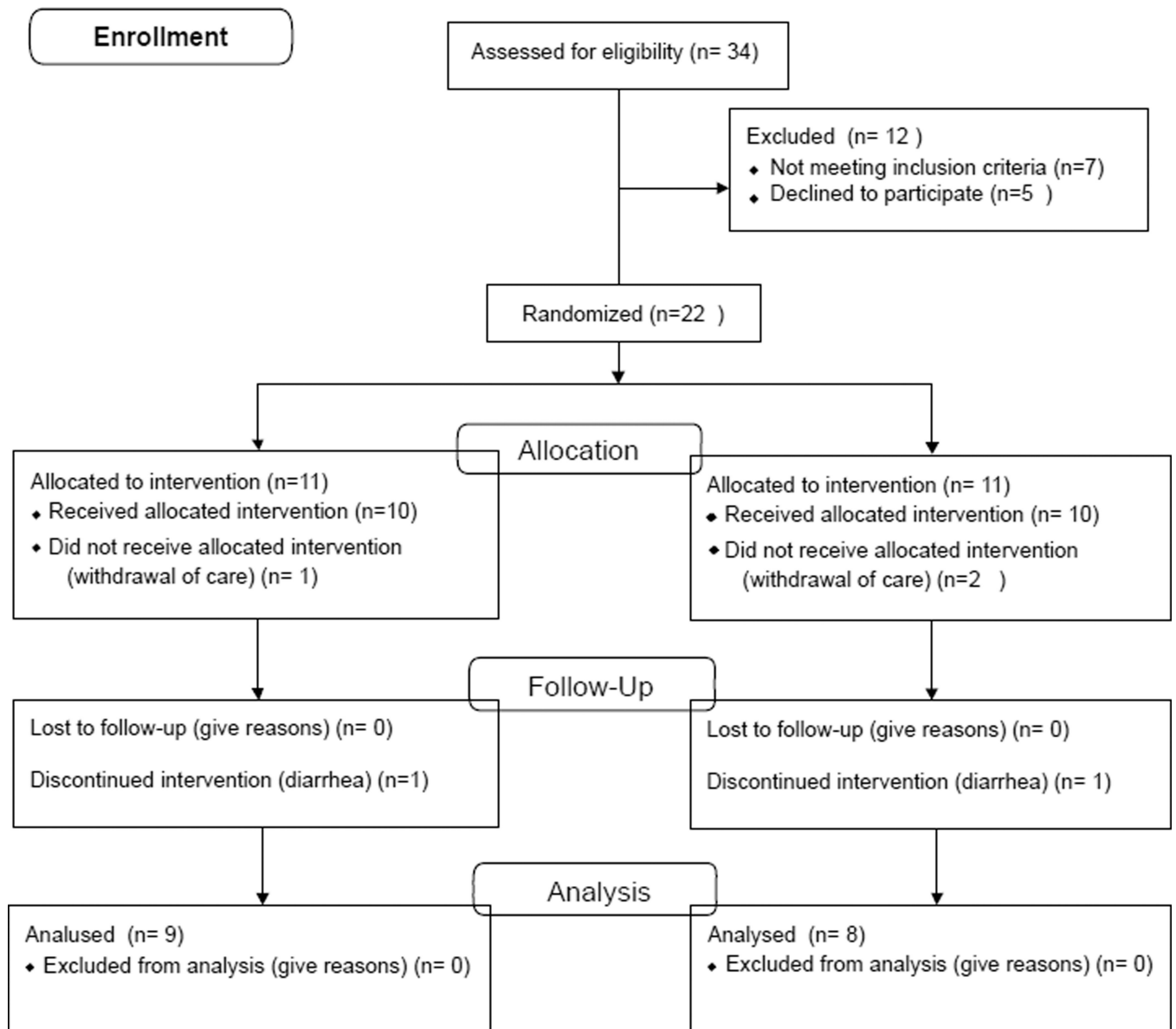


Figure 2.
Study flow diagram.

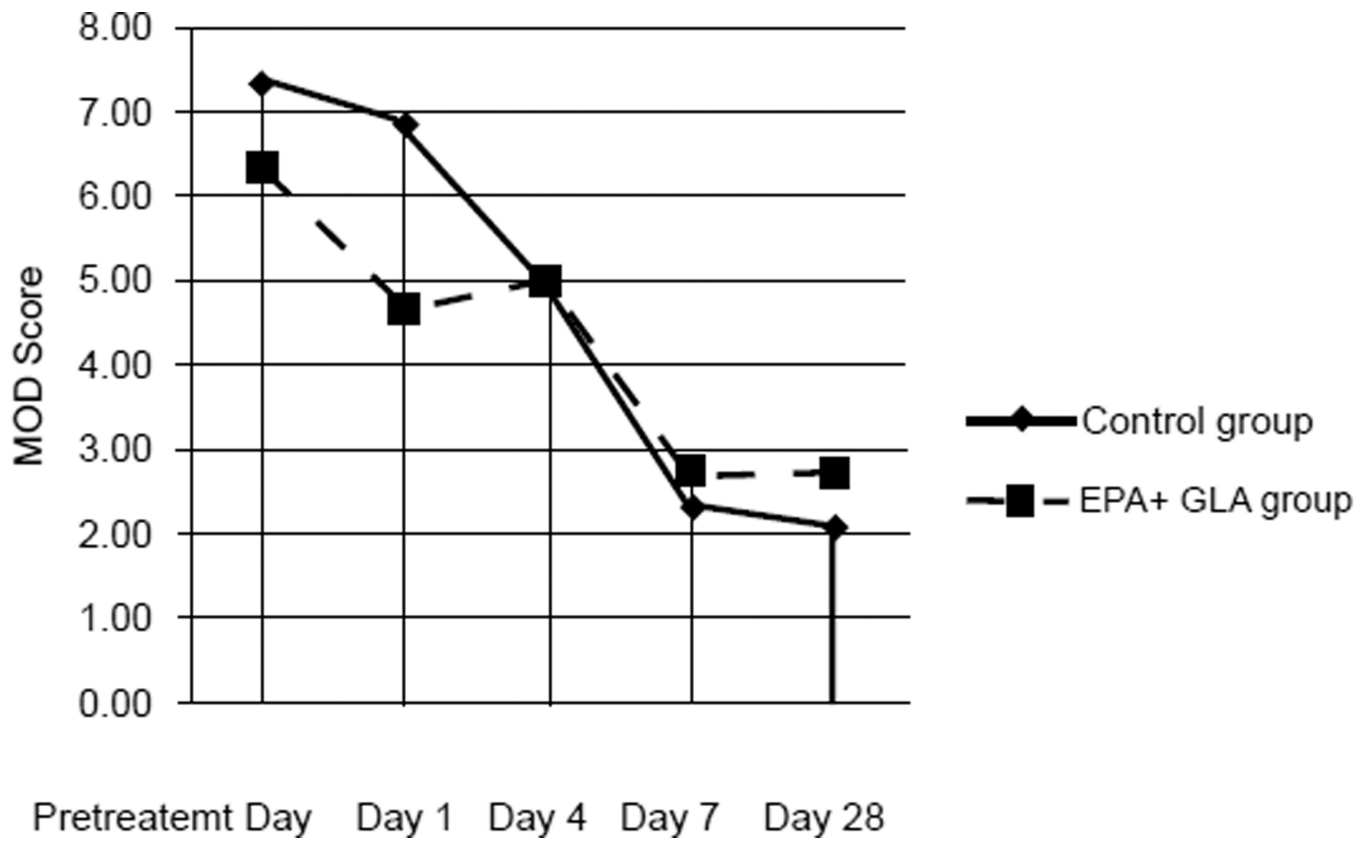
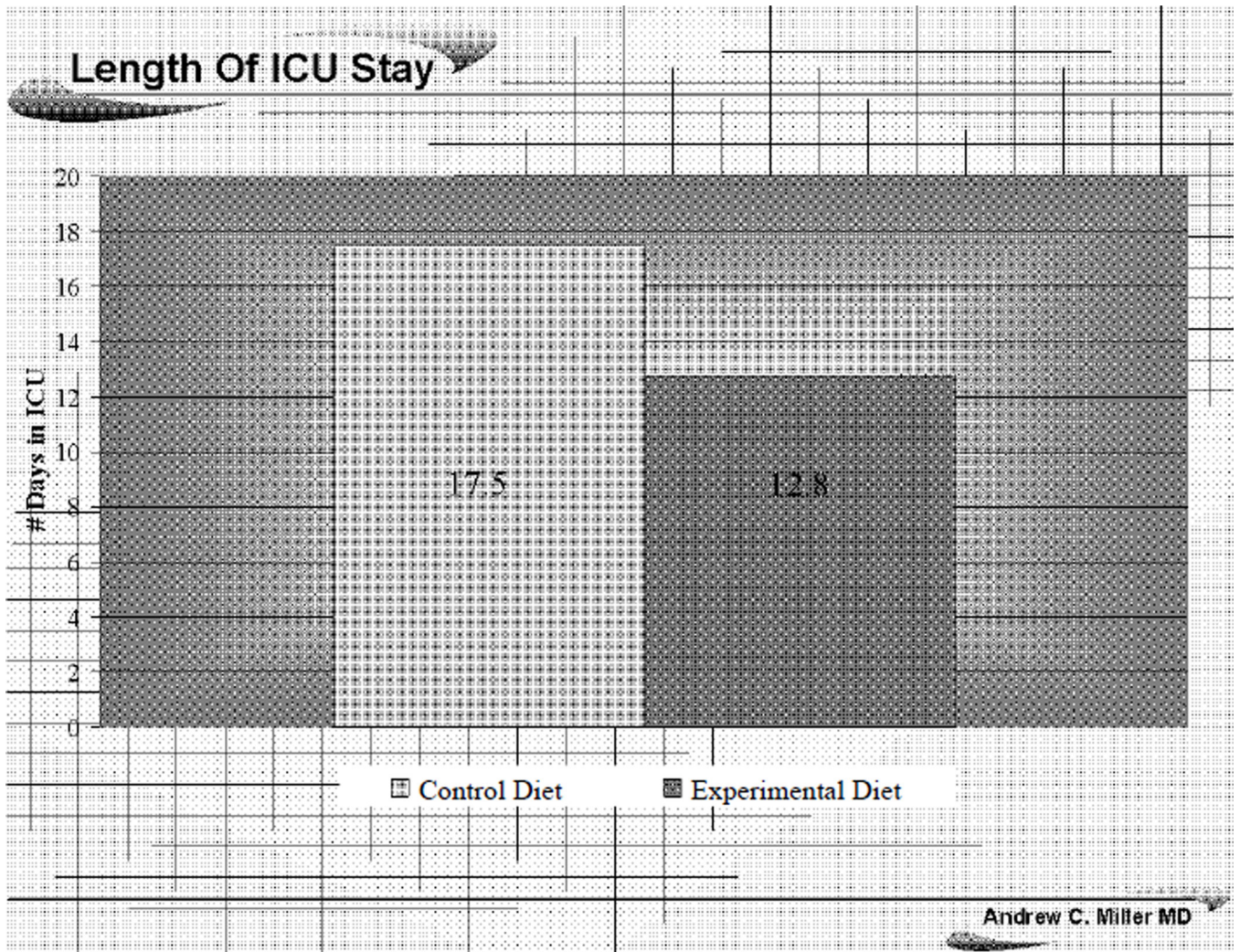


Figure 3.

Effects of EPA and GLA enteral feedings on Multiple Organ Dysfunction (MOD) scores comparing to the control group.



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Figure 4.
The effects of EPA+GPA diet on length of ICU stay comparing to the control group.

Table 1

Comparison of study dietary regimens.

Parameter	Control Diet	Experimental Diet
Caloric Distribution		
% Protein	16.7	16.7
% Fat	55.2	55.2
% Carbohydrate	28.1	28.1
Nutrient Sources		
Protein	Sodium & calcium caseinates	Sodium & calcium caseinates
Fat	Corn oil	Canola oil, MCT, borage oil, fish oil
Carbohydrate Sources		
	Hydrolyzed cornstarch, Sucrose	Hydrolyzed cornstarch, Sucrose
Other Specific Nutrients		
	Ultra-trace minerals, High vitamin E, High vitamin C and β -carotene, Carnitine, Taurine	Ultra-trace minerals, High vitamin E, High vitamin C and β -carotene, Carnitine, Taurine
Kilocalories/milliliter		
	1.5	1.5
Osmolality (mOSm/kg H₂O)		
	465	465
Calorie-to-Nitrogen ratio		
	150:1	150:1
mL/day to meet RDA *		
	947	947
Form		
	Ready to feed liquid	Ready to feed liquid

Both the control and experimental groups were matched with regard to percentage of protein, fat and carbohydrate in their respective diets as well as feed concentration (Kcal/ml), osmolality, and calorie-to-nitrogen ratio.

* RDA: Recommended Daily Allowance.

Table 2

Components and individual values of the lung injury score.

Value		
1	Chest roentgenogram Score	
	No alveolar consolidation	0
	Alveolar consolidation confined to 1 quadrant	1
	Alveolar consolidation confined to 2 quadrants	2
	Alveolar consolidation confined to 3 quadrants	3
	Alveolar consolidation in all 4 quadrants	4
2	Hypoxemia score	
	PaO ₂ /FIO ₂	300
	PaO ₂ /FIO ₂	225–299
	PaO ₂ /FIO ₂	175–224
	PaO ₂ /FIO ₂	100–174
	PaO ₂ /FIO ₂	<100
3	PEEP score (when ventilated)	
	PEEP	5 cm H ₂ O
	PEEP	6–8 cm H ₂ O
	PEEP	9–11 cm H ₂ O
	PEEP	12–14 cm H ₂ O
	PEEP	15 cm H ₂ O
4	Respiratory system compliance score	
	Compliance	80 ml/cm H ₂ O
	Compliance	60–79 ml/cm H ₂ O
	Compliance	40–59 ml/cm H ₂ O
	Compliance	20–39 ml/cm H ₂ O
	Compliance	19 ml/cm H ₂ O
The final value is obtained by dividing the aggregate sum by the number of components that were used.		
<u>Score</u>		
No lung injury- 0		
Mild-to-moderate lung injury –0.1–2.5		
Severe lung injury (ARDS) >2.5		

* Abbreviations: PaO₂/FIO₂ = Arterial oxygen tension to inspired oxygen concentration ratio; PEEP = Positive end-expiratory pressure.

Table 3

The Multiple Organ Dysfunction Score.

Organ System	Score				
	0	1	2	3	4
Respiratory ^a (PO ₂ /FIO ₂ ratio)	>300	226–300	151–225	76–150	75
Renal ^b (serum creatinine)	100	101–200	201–350	351–500	>500
Hepatic ^c (serum bilirubin)	20	21–60	61–120	121–240	>240
Cardiovascular ^d (PAR)	10.0	10.1–15.0	15.1–20.0	20.1–30.0	>30.0
Hematologic ^e (Platelet count)	>120	81–120	51–80	21–50	20
Neurologic ^f (Glasgow Coma Score)	15	13–14	10–12	7–9	6

^aThe PO₂/FIO₂ ratio is calculated without reference to the use or mode of mechanical ventilation, and without reference to the use or level of positive end-expiratory pressure;

^bThe serum creatinine concentration is measured in µmol/L without reference to the use of dialysis;

^cthe serum bilirubin concentration is measured in µmol/L

^dthe pressure-adjusted heart rate (PAR) is calculated as the product of the heart rate (HR) multiplied by the ratio of the right atrial (central venous) pressure (RAP) to the mean arterial pressure (MAP);
PAR= HR × RAP/ mean BP;

^ethe platelet count is measured in platelets/ml 10⁻³;

^fthe Glasgow coma Score is preferably calculated by the patient's nurse, and is scored conservatively (for the patient receiving sedation or muscle relaxants, normal function is assumed, unless there is evidence of intrinsically altered mentation)

Table 4

Patient characteristics at time of enrollment.

Variables	Study (n=9)/ SD	Control (n=8) / SD	<i>p</i>
Age (years)	50.0 / 22.2	55.2 / 16.5	0.6
Gender			
Male	5	3	0.4
Female	4	5	0.6
APACHE III score	60.33 / 14.6	52.9 / 9.5	0.2
Diagnosis			
Medical	7	6	0.4
*Surgical (none trauma)	2	2	0.8
REE Kcal/day (Indirect Calorimetry)	1700 / 236	1681 / 294	0.3
LIS	2.83 / 0.28	2.84 / 0.51	0.9
MOD Score	7.33 / 1.4	6.12 / 2.0	0.3

Acute Physiology, Age, Chronic Health Evaluation (APACHE) III score, resting energy expenditure (REE), Lung Injury Score (LIS), and Multiple Organ Dysfunction (MOD) Score.

* Surgery prior to ICU admission

Table 5

Comparison of mean PaO₂/FiO₂ ratios on days 1, 2, 4 and 7.

Group	Day 1 PaO₂/FiO₂	Day 2 PaO₂/FiO₂	Day 4 PaO₂/FiO₂	Day 7 PaO₂/FiO₂
EPA+GLA	157	149	162	178
Control	138	142	145	201
P-value	0.01	0.12	0.38	0.11

Table 6

Comparison of mean Lung Injury Score from pretreatment day through study Day 7.

	Pretreatment Day	Study Day 1	Study Day 4	Study Day 7
Study group (n=9)	2.83±0.2	2.77±0.4	2.5±0.5	2.1±0.6*
Control group (n=8)	2.84±0.3	2.91±0.4	2.4±0.6	2.3±0.4

* Statistically significant decline comparing to pretreatment mean (P=0.04)

Table 7

Comparison of ventilation variables on days 1, 4, and 7.

Variable	Study Day	No.	Control (mean/median)	No.	EPA+GLA (mean/median)
FIO ₂	1	8	0.54 / 0.5	9	0.52 / 0.6
	4	8	0.53 / 0.48	7	0.57 / 0.45
	7	7	0.43 / 0.43	6	0.48 / 0.4
PEEP (cm H ₂ O)	1	8	9.38 / 8	9	9.44 / 10
	4	8	8.63 / 8.5	7	8.29 / 8
	7	6	5.5 / 5	6	8.67 / 7.5
PIP (cm H ₂ O)	1	8	31.38 / 30.5	9	28.11 / 29
	4	7	28.14 / 30	5	32.8 / 35
	7	5	28.2 / 28	5	35 / 31
PaO ₂ (mm Hg)	1	8	72.75 / 68.5	9	81.56 / 76
	4	7	87 / 78	7	92.29 / 89
	7	5	86.4 / 85	7	92.71 / 90
Minute Ventilation (L/min) *	1	8	9.76 / 8.82	9	11.94 / 10.5
	4	8	9.09 / 9.35	5 ^{**}	16.03 / 17.6
	7	6	12.41 / 11.54	6	13.26 / 12.38

EPA: Eicosapentaenoic Acid; GLA: γ -linolenic acid; PEEP: Positive End-expiratory Pressure; PIP: Peak Inspiratory Pressure.

* Minute Ventilation (L/min) = (tidal volume in L) X (total ventilation rate in breaths/min)

** Data was missing for 1 patient.