TGF-β cascade regulation by PPP1 and its interactors – impact on prostate cancer development and therapy

Luís Korrodi-Gregório ^a, Joana Vieira Silva ^a, Luís Santos-Sousa ^a, Maria João Freitas ^a, Juliana Felgueiras ^a, Margarida Fardilha ^{a, *}

^a Signal Transduction Laboratory, Centre for Cell Biology, Biology Department, Health Sciences Department, University of Aveiro, Aveiro, Portugal

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Abstract

Protein phosphorylation is a key mechanism by which normal and cancer cells regulate their main transduction pathways. Protein kinases and phosphatases are precisely orchestrated to achieve the (de)phosphorylation of candidate proteins. Indeed, cellular health is dependent on the fine-tune of phosphorylation systems, which when deregulated lead to cancer. Transforming growth factor beta (TGF- β) pathway involvement in the genesis of prostate cancer has long been established. Many of its members were shown to be hypo- or hyperphosphorylated during the process of malignancy. A major phosphatase that is responsible for the vast majority of the serine/threonine dephosphorylation is the phosphorpotein phosphatase 1 (PPP1). PPP1 has been associated with the dephosphorylation of several proteins involved in the TGF- β cascade. This review will discuss the role of PPP1 in the regulation of several TGF- β signalling members and how the subversion of this pathway is related to prostate cancer development. Furthermore, current challenges on the protein phosphatases field as new targets to cancer therapy will be addressed.

Keywords: TGF-β • PPP1 • PIP • prostate cancer • phosphatase • signal transduction therapy

Introduction

TGF-β signalling pathway—an overview

The transforming growth factor beta (TGF- β) superfamily comprises over 42 members, all of which are generated from a single pre-propeptide precursor. Besides, TGF- β 1, 2 and 3, this superfamily includes the bone morphogenetic proteins (BMPs), the activins, the growth differentiation factors (GDFs) and the anti-muellerian hormone, among others [1]. Virtually, all types of cells produce and are sensitive to TGF- β superfamily members. These play fundamental roles in several cellular processes such as cell proliferation, adhesion, differentiation, apoptosis and migration, which may vary according to the ligand, the tissue and the conditions [2].

Transforming growth factor beta is a cytokine, with pleiotropic effects, that is produced mainly by fibroblasts and epithelial cells [3]. In the epithelium, $TGF-\beta$ inhibits cellular proliferation [4], whereas in

the mesenchyme, it promotes cellular proliferation [5, 6]. Other functions attributed to TGF- β are as follows: synthesis of extracellular matrix (ECM) [7], expression of integrins [8], modulation of immune response [9], angiogenesis [10] and wound healing [11]. Bone morphogenetic proteins, on the other hand, display a broad range of effects, even though sharing similar structure and signal transduction mechanisms. Among these, bone and cartilage formation and embryogenesis are the most relevant [12, 13]. Activins play crucial roles in the activation of follicle-stimulating hormone [14], erythropoiesis [15] and survival of neurons [16].

Transforming growth factor beta family ligands dimerize, most commonly forming homodimers, and propagate the signal by interacting with membrane surface receptors presented in the target cell [17]. A total of 12 transmembrane serine/threonine kinase receptors have been identified that are usually divided into two types: five

Tel.: 00351 91 8143947 Fax: 00351 234 370 200 E-mail: mfardilha@ua.pt

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^{*}Correspondence to: Margarida FARDILHA, Centro de Biologia Celular, Universidade de Aveiro, 3810-193 Aveiro. Portugal.

constitutively active type II receptors (TGF-BRII) and seven non-constitutively active type I receptors (TGF-BRI). Co-receptors, which lack catalytic activity, have also been identified, namely endoglin (CD105) and betaglycan (TGF-βRIII) [17]. Ligands display more affinity to the type II receptors, and the binding of the TGF-β to the type II receptor enables it to phosphorylate the GS domain of the type I receptor. activating its catalytic activity [2, 18, 19]. The type I receptors are denominated activin receptors-like kinases (ALKs) and once activated exert their catalytic activity by phosphorylating the C-terminal SxS domain of the main intracellular signal transducers of the pathway, the Smads [20]. Eight Smads have been identified in the human and mouse genomes: five regulatory Smads (R-Smads 1/2/3/5/8), one common Smad (Smad4, also known as Co-Smad) and two inhibitory Smads (I-Smads 6/7). The R-Smads, after being phosphorylated by the type I receptors, form trimers with the Co-Smad [20]. In general, ALKs 1/2/3/6 propagate the signals via Smads 1/5/8, whereas ALKs 4/5/7 propagate it through Smads 2/3 [2]. The fine dynamic equilibrium between these two opposing pathways often determines the ultimate outcome of the signal. After the formation of the complex, it is then translocated to the nucleus via microtubules and dyneins [21, 22]. The nuclear import is mainly done by importins, although direct interaction with nucleoporins is also described [23]. Once in the nucleus, the trimers act as trans-regulatory elements to activate or repress the expression of genes such as *Sp1*, *Id1* and *Myc*. R-Smads/ Co-Smads complex can also recruit transcription co-activators or co-repressors to modulate the amplitude of the activation/repression of the transcription [2].

Moreover, besides activating Smad-dependent signalling, $TGF-\beta$ can also activate other signalling pathways in a Smad-independent manner, such as mitogen-activating protein kinases (MAPKs), phosphatidylinositol 3-kinase (PI3K)/Akt and small GTPases [24, 25].

TGF-β in cancer

Transforming growth factor beta plays a pivotal role in a wide variety of diseases, as expected by its pleiotropic effects and ubiquitous expression, namely cardiovascular, connective tissue and neurological diseases, reproductive, developmental, skeletal and muscle disorders [26] and also in several cancers [27–32].

During cell malignant transformation, a number of alterations occur at molecular and cellular levels (genetic, epigenetic and somatic) and in the surrounding microenvironment, contributing to an increased survivability and proliferative advantage [27, 33–35]. The traditional hallmarks of cancer include: (i) insensitivity to antigrowth signals; (ii) evasion of apoptosis; (iii) self-sufficiency in growth signals; (iv) sustained angiogenesis; (v) limitless replicative potential; and (vi) tissue invasion and metastasis, all of which cooperate in providing malignant cells a selective advantage [36, 37]. Moreover, two new emerging hallmarks namely deregulation of the cellular energetics and avoidance of immune destruction have arisen [38].

In normal cells, a complex web of interconnected signalling pathways heavily regulates each of these functions. As a potent pleiotropic cytokine, $TGF-\beta$ acts in normal tissues as a formidable barrier to the development of cancer hallmarks, [31] inhibiting cellular proliferation

[39], migration and invasion [40], and promoting apoptosis [41], cell adhesion [40] and cellular differentiation [42]. However, TGF- β plays a dual role in cancer, since in late-stage tumours, the cellular machinery subverts the signalling pathway to promote the cancer progression [29].

Insensitivity to anti-growth signals

Transforming growth factor beta was initially named for its ability to promote the proliferation and transformation of mesenchymal cells in soft agar [43]. However, it is now established that TGF-B inhibits epithelial, endothelial and hematopoietic cell proliferation [44]. Transforming growth factor beta ability to mediate cytostasis occurs in the late G1 phase by preventing the progression to S phase through two synchronized events: (i) repression of Myc, Id1 and Id2; and (ii) induction of cyclin-dependent kinase (CDKs) inhibitors p15INK4b p21^{CIP} and p27^{KIP} [27, 31, 44, 45]. Nevertheless, virtually all epithelial-derived tumours (>85% of all human cancers) display partial or total resistance to TGF-β growth inhibition effects, being this a major cancer hallmark [27, 31]. Several mechanisms to override the cytostatic activities of TGF-B have been detected in cancer cells either due to inactivating mutations or repression of one or more genes of the TGF-β signalling pathway [32] (Fig. 1). As a result of this insensitiveness, cancer cells usually secrete larger amounts of TGF-β, being this production parallel to the cancer progression [28]. Of possible relevance, it has been shown that TGF-BRI is sufficient to mediate some cellular responses to TGF- β , such as expression of JunB and PAI-1, indicating that maybe the receptor levels are sufficient to mediate the response via TGF-βRI, but not the ones via TGF-βRII [46, 47].

Evasion of apoptosis

In general, TGF- β is pro-apoptotic in a Smad-dependent pathway [27, 48], although Smad-independent pathways can also play a role [49]. Apoptosis triggered by TGF- β can be both p53-dependent and p53-independent [27, 50, 51]. In fact, p53 has been shown to interact with Smad2 in a TGF- β -dependent fashion, thus linking two of the most important apoptotic mediators in human cells [52]. However, during malignant transformation, TGF- β signalling is altered to promote cell survival [31] (Fig. 1).

Self-sufficiency in growth signals

Cancer cells often acquire the ability to undergo proliferation when stimulated by TGF- β through the ability of inducing the expression of cytokines, growth factors and/or their receptors (Fig. 1). It has been also shown to activate the Ras/Raf/MAPK pathway [27]. Therefore, by overexpression of growth signals or constitutive activation of downstream pathways, independence of exogenous growth factors is achieved [27].

Sustained angiogenesis

Angiogenesis is a normal physiological process in which new blood vessels are formed from pre-existing vessels, being essential in wound healing and embryogenesis. Solid tumours usually cannot exceed 1–2 mm because of limited access to oxygen and nutrients.

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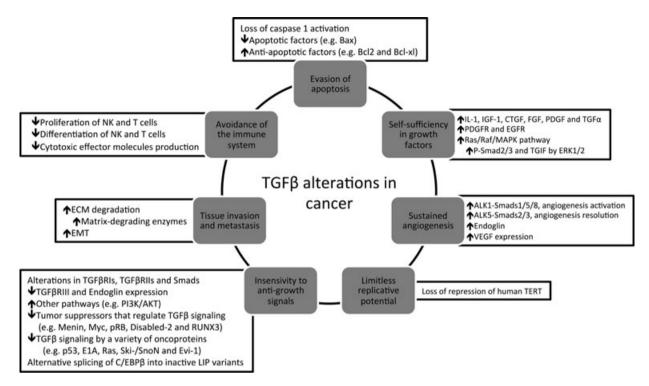


Fig. 1 Alterations in TGF- β signalling cascade associated with major cancer hallmarks. The traditional cancer hallmarks are disrupted at variable extent, with insensitivity to anti-growth signals comprising the most well-described alterations. White boxes: TGF- β superfamily ligands, receptors, downstream effectors or in the responses exerted by TGF- β signalling pathway; Grey boxes: major cancer hallmark. TGF- β , transforming growth factor beta; IL-1, interleucin-1; IGF-1, insulin-like growth factor-1; CTGF, connective tissue growth factor; FGF, fibroblast growth factor; PDGF, platelet-derived growth factor; TGF- α , transforming growth factor alpha; PDGFR, PDGF receptor; EGFR, epidermal growth factor receptor; MAPK, mitogen-activated protein kinase; TGIF, transforming growth-interacting factor; VEGF, vascular endothelial growth factor; TERT, telomerase reverse transcriptase; pRb, protein retinoblastoma; LIP, liver-enriched inhibitory protein; ECM, extracellular matrix; EMT, epithelial—mesenchymal transition.

being neoangiogenesis a way to circumvent this limitation. Ultimately, angiogenesis provides also a route for metastatic spread. *In vitro*, TGF- β can display either pro- or anti-angiogenic features. However, *in vivo* TGF- β is considered to have a pro-angiogenic role, being this action rather complex, as it appears to contribute to both phases of the angiogenic process: activation and resolution. Activation of ALK1 stimulates Smads1/5/8, regulating angiogenesis activation, while ALK5 regulates angiogenesis resolution *via* Smads2/3 [27, 31]. Another relevant aspect is the fact that TGF- β induces the expression of VEGF in a Smad2/3 and Src-dependent mechanism, which directly contributes to angiogenesis [53]. In cancer, pro-angiogenic effects of TGF- β seem to be up-regulated (Fig. 1).

Tissue invasion and metastasis

Transforming growth factor beta is a potent regulator of cell adhesion, ECM and motility. In normal cells, TGF- β stimulates the production of ECM by increasing the synthesis of collagen, fibronectin and other ECM proteins, decreasing the production of enzymes that degrade the ECM (*e.g.* heparinase, collagenase and stromelysin) and stimulating the production of proteins that inhibit ECM degradation (*e.g.* PAI-1 and TIM) (Fig. 1). However, cancer cells often respond to

TGF- β by stimulating the expression of matrix-degrading enzymes, thus contributing to ECM degradation and invasiveness through epithelial-to-mesenchymal transition (EMT) [27]. Epithelial-to-mesenchymal transition is a process that occurs during embryogenesis and a pathological feature in neoplasia, rheumatoid arthritis, chronic inflammation and fibrosis [29, 31], which consists in the transdifferentiation of immotile, adherent, polarized epithelial cells into highly motile, apolar mesenchymal cells. Transforming growth factor beta has been found to promote EMT through a combination of Smaddependent transcriptional events and Smad-independent effects on cell junction complexes [29] (Fig. 1).

Avoidance of the immune system

It has already been established that cancer initiation, promotion and progression are linked to aberrant and/or persistent inflammation within tumour microenvironment. In general, high levels of $TGF-\beta$ are seen in advanced cancers and are found to inactivate host antitumour immunosurveillance systems, which confer immune privilege to developing neoplasms and ensures for their continued progression (Fig. 1). Transforming growth factor beta signalling crucial role has been demonstrated by the fact that Smad3-defficient mice exhibited

defects in the responsiveness and chemotaxis of B and T cells and neutrophils [28, 31]. Therefore, it is now clear that TGF- β signalling affects all the hallmarks of cancer at some extent (Fig. 1). Cancer cells appear to selectively use the TGF- β responses that are advantageous.

Phosphatases in cancer

A common mechanism used by cells to either propagate or terminate intracellular signal transduction pathways is the reversible protein phosphorylation [54]. Many cellular processes are controlled by phosphorylation/dephosphorylation of structural or regulatory proteins that work as a molecular 'switch' [55, 56]. The reversible phosphorylation consists in the addition or removal of a negatively charged phosphate group mainly to serine, threonine or tyrosine residues calalyzed by protein kinases and protein phosphatases, respectively.

In humans, there are around 500 kinases, all sharing a related catalytic domain [57]. Uncontrolled kinase activity is associated with cell proliferation and is a common finding in human cancers [58]. Curiously, there are three to five times fewer phosphatases than kinases suggesting that the specificity of substrates is not only because of the catalytic subunits but also because of the regulatory subunits diversity [59].

Phosphoprotein phosphatase 1 (PPP1) is a major serine/threonine phosphatase and is expressed in all eukarvotic cells. This holoenzyme consists of a catalytic and at least one regulatory subunit. The catalytic subunit is encoded by three different genes (PPP1CA, PPP1CB and PPP1CC) that share near 90% of the amino acid sequence [55, 60]. The regulatory subunits account for PPP1 substrate diversity and consequently function [61, 62]. The regulatory subunits are known as PPP1 interacting proteins (PIPs) and, to date, more than 200 PIPs were identified with many more expected to be found [55, 63]. PIPs can be activity modulating proteins when their function is to inhibit or enhance PPP1 catalytic activity; targeting proteins, which are responsible for PPP1 subcellular localization or/and bring together PPP1 and specific substrates; and PPP1 substrates that associate with PPP1 and are dephosphorylated. Despite the function, every PIP interacts with PPP1. Although several PPP1 binding motifs have been already identified, such as RVxF and SILK, binding specificity between PIPs and PPP1 is obtained by differences in the number and the type of docking sites [64-67].

The evidence that reversible protein phosphorylation is essential for cellular function arises from the significant number of human diseases in which control of protein phosphorylation is impaired, like cancer and diabetes [60]. In cancer, imbalances of protein phosphorylation appear to be an important pathophysiological mechanism [60]. Constitutive activation of oncogenic kinases is one of the hall-marks observed in cancer cells, driving uncontrolled cell proliferation, invasion and metastasis [68]. Transmembrane kinases, such as epidermal growth factor receptor and platelet-derived growth factor receptor, and cytoplasmic kinases, such as Raf and Akt, are mutated or hyperactivated in several types of human cancer. For example, an increased Akt signalling, a serine/threonine kinase involved in the

control of cell size/growth, proliferation and survival [69], has been associated with poor clinical outcome in a variety of tumours, such as melanoma, breast and prostate [70].

Given that phosphorylation represents an essential element of cancer pathophysiology, it is not surprising that tumour suppressive functions have been linked to protein phosphatases [68, 71, 72]. PPP2 and PPP1 have been associated with cancer suppressive processes, such as inhibition of cell survival, proliferation and migration [68, 71]. Moreover, PPP1 tumour suppressive functions have been connected with some of its PIPs [73].

TGF- β signalling: role in prostate cancer pathogenesis

The different constituents of the prostate work as a functional unit, with the interactions between stroma and epithelium playing a pivotal role in normal prostate growth, development and function [74]. It has been established that androgens, testosterone and dihydrotestosterone (DHT) are the most potent and relevant mitogens of the normal prostate [74]. However, androgens actions are mainly indirect, through the stimulation of the production of diverse growth factors (e.g. EGF, TGF-α, KGF, IGF and bFGF) by stromal cells, which act mainly in a paracrine mode [74]. To counteract the effects of these growth factors, both in stroma (e.g. FGF) and epithelium (e.g. TGF- α and EGF), TGF-β has been identified as a key growth modulator in normal prostate, by inducing growth inhibition and differentiation [27, 47, 74]. Most prostate cancers arise as androgen-dependent, meaning that androgens drive cell proliferation. Prostate cancer progression usually involves the shifting to an androgen-independent state, sometimes with mutation or loss of the androgen receptor (AR) and an increasing impact of growth factors signalling pathways [75, 76]. This androgen resistance leads to an increase in the TGF-β production, which in turn promotes prostate cancer growth, viability and aggressiveness [47, 77]. Also, in vitro studies have shown that normal TGF-β-induced growth inhibition must be disrupted by the neoplasic surrounding environment because normal epithelial prostate cells are not able to protect themselves under any experimental condition [47].

Ligands

In prostate cancer, there is a dramatic increase in TGF- β 1 mRNA and protein levels, which are correlated with high Gleason score, bone metastasis, angiogenesis and poor clinical outcome [27, 78] (Fig. 2). Exogenous TGF- β causes auto-induction in both normal and malignant cell lines at low concentrations. However, such effect is only seen at high concentrations in malignant cells [79]. In prostate cancer, TGF- β -mediated apoptosis involves: (*i*) caspase 1 activation [80]; (*ii*) up-regulation of pro-apoptotic factors (*e.g.* Bax, p27^{KIP1}); and/or (*iii*) down-regulation of antiapoptotic factors (*e.g.* Bcl-2 and Bcl-xl) [51, 81] (Fig. 2). Bcl-2 overexpression plays a role in the development of androgen-independent prostate cancer cells, conferring

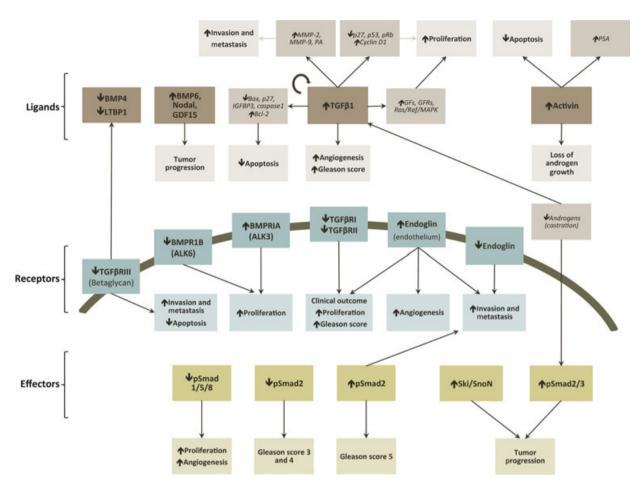


Fig. 2 TGF- β signalling cascade alterations that lead to prostate cancer. Main effects or alterations related to the TGF- β signalling pathway that drive to prostate cancer hallmarks. Brown boxes: ligands; Blue boxes: receptors; Green boxes: downstream effectors of the TGF- β signalling pathway; Light brown boxes: alteration in other targets (italics). Grey, light blue and light green boxes: alteration in cancer cell hallmarks or effects related to the ligands/other targets, receptors or downstream effectors, respectively; Black arrows inside boxes: increase/decrease or activation/inactivation; Black arrows: effect or alteration. PSA, prostate-specific androgen; MMP-2, matrix metalloproteinase 2; MMP-9, matrix metalloproteinase 9; PA, plasminogen activator; pRb, protein retinoblastoma; BMP, bone morphogenetic protein; LTBP1, latent TGF- β binding protein 1; GDF15, growth differentiation factor 15; IGFBP3, insulin-like growth factor binding protein 3; GF, growth factors; GFR, growth factor receptor; MAPK, mitogen-activated protein kinase; BMPR, bone morphogenetic protein receptor; TGF- β R, TGF- β receptor; ALK, activin receptor-like kinase; EGFR, epidermal growth factor receptor.

resistance to apoptosis by an antagonistic effect in caspase 1 activation, providing resistance to radio and chemotherapies [82]. Moreover, loss of p27 $^{\text{KIP1}}$ in prostate cancer has been firmly established and regarded as a prognostic marker of increased recurrence and reduced survival [83]. Even though most studies are centred in TGF- β , other ligands of the TGF- β superfamily may also play pivotal roles in prostate cancer (e.g. Activins, BMP6, GDF15 and Nodal/BMP16) [52, 84].

Receptors

Up to 30% of prostate cancer cases have down-regulation or absence of a TGF- β receptor, while no alterations in Smads are usually found

[27]. Mutations of TGF- β RII are common in lung and laryngeal cancers, but not in prostate cancer [52]. Nevertheless, some prostate cancer cells express a truncated TGF- β RI mRNA transcript [85], lack a TGF- β RII gene [86], have TGF- β RI epigenetically regulated [87, 88], or carry some sort of TGF- β RII mutation [89]. The fact that TGF- β RI and TGF- β RII are decreased in metastasis *versus* primary tumours may indicate an active role of this alteration in cancer progression [90] (Fig. 2). Therefore, the absence of TGF- β Rs in prostate tumour cells leads to growth inhibition resistance, thus resulting in clonal expansion of these cells [74, 91–94]. Acquired resistance can also be developed as a result of alterations in downstream genes such as p27 (repression), Cyclin D1 (induction), p53 and protein retinoblastoma (mutations) or *via* alterations in other pathways, like Akt/mTOR [83, 95–97].

The BMPRs appear to elicit different responses, with a shift from BMPR1B towards BMPRIA being associated with cancer [98, 99].

Several studies have also reported the loss of TGF- β RIII as the most common alteration during prostate cancer progression, being this alteration even more evident in metastasis [100] (Fig. 2). This loss occurs at both the mRNA and the protein levels and is subject of either direct or indirect epigenetic regulation. Knockdown of TGF- β RIII in prostate epithelial cells led to alterations in 101 genes associated with prostate cancer and its restoration decreased tumour growth, angiogenesis and increased apoptosis [100]. Moreover, loss of TGF- β RIII correlates with disease state, metastatic disease and prostate-specific antigen (PSA) recurrence [100].

In a similar manner, endoglin levels are lower in prostate cancer cells *versus* normal prostate cells, and even lower in metastasis (Fig. 2). Endoglin has been found to inhibit invasiveness, metastasis formation and motility while increasing cell adhesion, neovascularization and growth [101–104]. In contrast, levels are higher in endothelial cells, being this associated with ongoing angiogenesis [98, 102].

Effectors

Smad alterations in prostate cancer are also found, although not as extensively described as alterations in TGF- β ligand and its receptors. In the initial stages of prostate tumour development, ALK2-Smad1/5/8 signalling is promoted to increase the growth and neovascularization, whereas in late-stage tumours, there is a shift towards ALK5-Smad2/3 signalling that leads to the acquisition of malignant capabilities, namely enhanced invasiveness, migration and metastasis formation [101, 103]. Also, endoglin may play a pivotal role in modulating this equilibrium [102]. Smad4 promoter methylation has also been reported [105]. High levels of Ski, a co-repressor of Smad2/3 (preferentially Smad3), were detected only in prostate cancer cells [84] (Fig. 2).

Interconnections between TGF- β signalling and other signalling pathways

The relationship between TGF- β and androgens is also relevant. For instance, TGF- β has been linked to castration-induced apoptosis. After androgen withdrawal, TGF- β and TGF- β Rs mRNA levels were up-regulated, at least transiently [74, 78, 106] (Fig. 2). Androgens have also been found to down-regulate Smads expression and activation. Dihydrotestosterone bounds to AR and this complex was able to bind to active Smad3, inhibiting the association of Smad3 with Smad binding element and, therefore, blocking the signal. Also, DHT leads to the inactivation of Sp1 suppressing its binding to TGF- β RII promoter, decreasing TGF- β RII levels [52, 107, 108]. It has been recognized that the AR status determines the sensitivity of prostate cancer cells to TGF- β -mediated apoptosis [109] and the ability to evade is of paramount importance in the development of cancer, especially in prostate [27].

TGF- β pathway is regulated by protein phosphatases

Transforming growth factor beta signalling regulates numerous cellular responses. Thus, its activity is tightly controlled by reversible phosphorylation as well as other post-translational modifications, such as ubiquitination, SUMOylation and acetylation [110]. The TGF- β pathway is activated by phosphorylation at the membrane receptors level and at the cytosolic Smad proteins, its primary effectors. In contrast, dephosphorylation of TGF- β signalling players provides a counterbalance mechanism that limits the duration and intensity of the signal, contributing to its termination. Despite the long-standing suspected influence of protein phosphatases in TGF- β signalling, concrete data only started to emerge recently [111].

Protein phosphatases and TGF-B pathway

An increasing number of protein phosphatases, particularly of the serine/threonine phosphatases family, have been reported to regulate the TGF- β pathway through interactions with both receptors and Smad proteins [112] (Fig. 3).

Phosphoprotein phosphatase 2 is a serine/threonine phosphatase in which the core structure consists of a catalytic and structural subunit. Although dimeric PPP2 can exist as such, it is normally associated with a third subunit that is regulatory. This subunit acts as a targeting and substrate-specifying unit, hence regulating this phosphatase and defining its physiological function. Phosphoprotein phosphatase 2 holoenzymes are important physiological regulators of proper checkpoint functioning during the cell cycle, which contributes significantly to their tumour suppressive abilities. It is well known that PPP2 inhibits both TGF-βRI and R-Smads [113] (Fig. 3). The regulatory B subunit family of PPP2 is composed by four homologous genes, two of which have opposite effects on TGF- β pathway. B α (PPP2R2A) enhances TGF-β/Activin/Nodal signal by regulating the basal levels of ALK5, whereas Bδ (PPP2R2D) negatively modulates these signals by limiting the receptor activity [114]. It has also been shown that TGF-βRI activation enables PPP2R2D binding to its cytoplasmic domain resulting in the phosphatase activation. Henceforth, PPP2R2D recruits PPP2C, PPP2A-β (PPP2R1B) and other regulatory subunits to bind and dephosphorylate p70s6k, a serine/threonine kinase crucial to progression of G₁/S phases of cell cycle [115]. Phosphoprotein phosphatase 2 also dephosphorylates the C-terminal SxS motif of Smad3 under hypoxic conditions without interfering with Smad2 phosphorylation state [113]. In BMP signalling pathway, the regulatory subunit Bß of PPP2 (PPP2R2B) has been recognized as an interactor of both BMPRII and Smad1 (Fig. 3). More specifically, it dephosphorylates BMPRII contributing to the inactive state of the receptor complex and reverses the phosphorylation by MAPK in the Smad1 linker region, enabling the Smad complex translocation into the nucleus [116].

Protein phosphatase Mg^{2+}/Mn^{2+} dependent 1A (PPM1A) is induced by activated TGF- β RI (Fig. 3). This protein dephosphorylates the C-terminal SxS motif of activated Smads2/3 and Smads1/5/8 in

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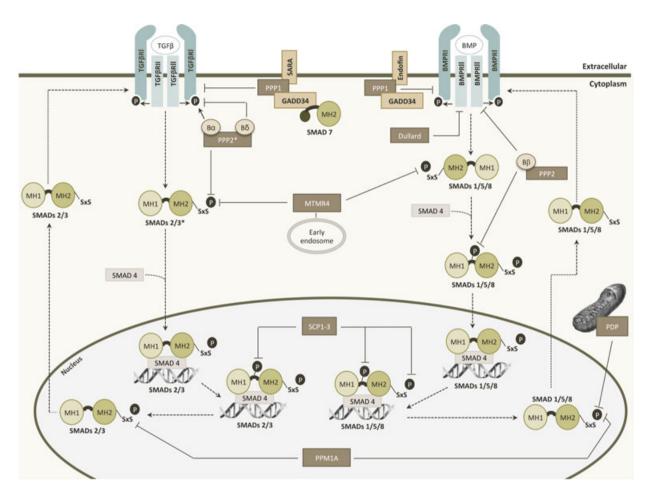


Fig. 3 Regulation of TGF- β signalling by phosphatases. With exception of Dullard and PDP, which only act on BMP signalling, all phosphatases contribute to both pathways. PPP1 and Dullard act exclusively on receptors, whereas MTMR4, SCPs, PDP and PPM1A act exclusively on R-Smads. PPP2 can act both on receptors and R-Smads. *, PPP2 dephosphorylates the C-terminal SxS motif of Smad3 without interfering with Smad2 phosphorylation state; Dashed arrows: sequential events on TGF- β and BMP signalling pathways; Black arrows and black bars: phosphorylation or dephosphorylation events, respectively; Brown boxes: phosphatases involved; Beige boxes: PPP1 interacting proteins. PPP1, phosphoprotein phosphatase 1; PPP2, phosphoprotein phosphatase 2; PPM1A, protein phosphatase Mg²⁺/Mn²⁺ dependent 1A; PDP, pyruvate dehydrogenase phosphatase; MTMR4, myotubularin-related protein 4; SCP, small C-terminal domain phosphatase; TGF- β R, TGF- β R receptor; BMPR, bone morphogenetic protein receptor; SARA, Smad anchor for receptor activation; GADD34, growth arrest and DNA damage-inducible protein 34.

the nucleus, thus promoting the dissociation of Smad heteromeric complexes. Subsequently, R-Smads undergo nuclear export, and the TGF- β -mediated anti-proliferative and translational effects are attenuated [117]. Moreover, PPM1A dephosphorylates RanBP3, enhancing its aptitude to export nuclear Smad2/3 and promotes Smad1 proteasomal degradation [113, 118]. Pyruvate dehydrogenase phosphatase is a mitochondrial phosphatase that also acts on Smad1 C-terminal SxS motif (Fig. 3), being the molecular mechanism underlying this interaction still unclear [119]. A member of dual specificity phosphatase family named myotubularin-related protein 4 (MTMR4) down-regulates the TGF- β signalling through interaction and dephosphorylation of activated Smads2/3 on early endosomes (Fig. 3). This dephosphorylation consequently blocks their nuclear translocation [120]. Recently, it has also been described the MTMR4 involvement

in the dephosphorylation of the SxS motif in BMP signalling R-Smads [121].

Also, a role for the aspartate-based phosphatases (FCPs and SCPs) in TGF- β signalling has been described (Fig. 3). In humans, there are three forms of small C-terminal domain phosphatases (SCP1-3) that bind with high affinity to Smad1, dephosphorylating it [118]. Furthermore, they dephosphorylate linker region residues of R-Smads. More precisely, SCPs1-3 dephosphorylate the residues Ser245, Ser250 and Ser255 of Smad2 and analogues sites on Smad3, Thr8 and Thr179 of Smad3, and Ser187, Ser195, Ser206 and Ser214 of Smad1 [122]. Another aspartate phosphatase from the FCP family named Dullard interacts with the BMPRII resulting in its degradation and consequent inhibition of BMPRI signalling [123, 124].

PPP1 and TGF-β receptor activity

Past studies in *Drosophila melanogaster* Dpp (Decapentaplegic) signalling found that PPP1 catalytic subunit (PPP1C) acts as a negative regulator of TGF-β signalling through its binding to Smad anchor for receptor activation protein (SARA; Fig. 3). Also, disruption of this binding leads to the hyperphosphorylation of the Dpp type I receptor [125]. Further studies in mammalian cells showed that SARA presents PPP1 to ALK5 receptor promoting its dephosphorylation and consequent signal attenuation. This targeting involves the inhibitory Smad7 and another PIP, GADD34 (PPP1R15B). It has also been shown that Smad7 recruits PPP1C to ALK1, inhibiting Smad 1/5/8 dependent pathway [126, 127]. The PIP that recruits PPP1 to ALK1 still needs to be elucidated; however, a Smad anchor for BMP signalling called Endofin was recently discovered. In a similar way of what happens to SARA, PPP1 also binds to Endofin and GADD34 to dephosphorylate the ALK3 and ALK6 receptors but without any intervention of the inhibitory Smad7 [128].

This TGF- β RI dephosphorylation is a critical reversible mechanism in the regulation of TGF- β signalling in several cellular contexts such as cellular stress, DNA damage and cellular growth [112].

Interestingly, PPP1 (and also PPP2) activity is required to maintain endothelial cells in a resting state [129]. Inhibition of PPP1 activity promotes endothelial cells migration consistent with its negative role in ALK1-induced activation of these cells [126].

PPP1 and PIPs in prostate cancer

Protein dephosphorylation at the serine and threonine residues has important roles in regulating both cell survival and cell differentiation [130]. Several evidence shows that PPP1 regulates multiple signalling pathways modulating cell apoptosis and differentiation [131]. PPP1 regulates the two major human tumour suppressors, which disruption has also been associated with prostate cancer, p53 and pRb [132, 133]. Phosphoprotein phosphatase 1 dephosphorylates p53 to attenuate its transcriptional and pro-apoptotic activity [7]. Also, pRb dephosphorylation by PPP1 negatively regulates apoptosis [6]. Phosphoprotein phosphatase 1 dephosphorylates Akt, regulating its activity and also its downstream targets to suppress differentiation and promote apoptosis [134].

Alteration of PPP1 gene expression and activity is associated with multiple human cancers, including prostate cancer. Over the past two decades, it has become evident that PPP1 versatility is achieved by its ability to interact with multiple PIPs, many of which have been associated with crucial processes implicated in carcinogenesis, such as cell cycle, apoptosis and cell migration.

NIPP1

Phosphoprotein phosphatase 1/Nuclear inhibitor of protein phosphatase 1 (NIPP1) complex has been recently described as a regulator of cell migration in prostate cancer cells [135]. Nuclear inhibitor of protein phosphatase 1, is ubiquitously expressed and was initially char-

acterized as a PPP1 inhibitor [136]. Phosphoprotein phosphatase 1 has already been described as a regulator of cell polarity and migration, namely in controlling enteric nerve cell migration [137, 138]. Recently, Martin-Granados has described a role for PPP1/NIPP1 in directing migration of human cancer cells [135] (Fig. 4). Genetic disruption of PPP1 and NIPP1 decreases directional migration in PC3 cells. It has been shown that the PPP1/NIPP1 complex controls directed cell migration *via* up-regulation of Cdc42, and it has been suggested that the complex may contribute to the migratory properties of cancer cells [135].

Androgen receptor

The AR is a ligand activated transcription factor, which plays a central role in prostate cancer development and progression, with androgen deprivation therapy being the standard treatment for prostate cancer [139, 140]. Chen *et al.* have shown that AR and PPP1 interact, and that PPP1 inhibition enhanced proteasome-mediated AR degradation. On the other hand, PPP1 overexpression increased AR expression and markedly enhanced AR transcriptional activity in prostate cancer cells [141] (Fig. 4). In addition, both AR and PPP1 undergo nuclear translocation when prostate cancer cells are stimulated by androgen. Phosphoprotein phosphatase 1 regulates AR protein stability and nuclear localization through its dephosphorylation at Ser650. Moreover, AR may function as a PPP1 regulatory subunit and mediate PPP1 recruitment to chromatin, where it can also modulate transcription and splicing events [141].

Fer/pRb

Several lines of evidence implicate Fer, a tyrosine kinase, in the progression and growth of malignant tumours [142, 143]. Fer levels in malignant prostate tumours are significantly higher than those detected in benign tumours [17]. Furthermore, down-regulation of Fer impaired the proliferation of prostate carcinoma cells and led to their arrest at the G0/G1 phase [17]. Fer amino acid sequence analysis revealed two PPP1 binding motifs in the kinase domain, which most probably mediate the interaction of this kinase with PPP1 (Fig. 4). Another important protein, pRb,

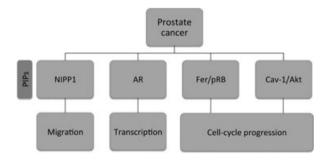


Fig. 4 PPP1/PIPs complexes and their potential roles in prostate cancer. NIPP1, nuclear inhibitor of protein phosphatase 1; AR, androgen receptor; pRB, protein retinoblastoma; Cav-1, Caveolin-1.

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when hypophosphorylated sequesters E2F, a transcription factor required for G1-S transition, thereby preventing cell cycle progression in stress conditions such as ultraviolet radiation, ionizing radiation or hypoxia stress [73, 144]. Down-regulation of Fer potentiates the activation of PPP1 that has been shown to dephosphorylate and activate pRb leading to its growth suppressive state and cell cycle arrest in malignant cells [17, 143, 145]. Therefore, the up-regulation of Fer in prostate cancer cells leads to the inactivation of PPP1 that culminates in pRb hyperphosphorylation resulting in a poor G1-S transition control and tumourigenesis [145].

Cav-1/Akt

Caveolin-1 (Cav-1) is a ubiquitously expressed integral membrane protein, with antiapoptotic activity in prostate cancer cells, functioning downstream of androgenic stimulation [146, 147]. Caveolin-1 has been reported to be overexpressed in prostate cancer cells and is associated with disease progression [148, 149]. Li et al. found that Cav-1 interacts with and inhibits PPP1 and PPP2 [150]. Analysis of putative substrates for PPP1 and PPP2 revealed that Cav-1-mediated inhibition of PPP1 and PPP2 leads to an increase in PDK1, Akt and ERK1/2 activities. This unravels a novel mechanism of Akt activation in prostate cancer (Fig. 4). Through its binding and inhibition of PPP1 and PPP2, Cav-1 is able to maintain Akt activated, leading to sustained activation of downstream oncogenic Akt targets and increased cell survival [150]. This mechanism could explain TGF-\u03b3-mediated apoptosis disruption by AKT pathway [151]. An increase in Akt signalling has been correlated with poor clinical outcome in prostate cancer [110]. These findings support the concept of PPP1 and/or PPP2 as tumour suppressor proteins and further support the notion that Cav-1 is an important metastasis-related gene.

Conclusions

Prostate cancer is a common disease. In developed countries, it is the most incident cancer and the third in mortality. Up to 50% of elderly men have small clinically insignificant tumours and the lifetime risk of developing prostate cancer is, in the more developed areas, circa 7.8% [74, 152]. Most human prostatic carcinomas are initially responsive to androgen ablation therapy and radiotherapy. However, when prostatic carcinomas become castration-resistant, radical prostatectomy is the only option for treatment [153]. Nonetheless, it has several side effects, including stricture of the vesico-urethral anastomosis, urinary incontinence and impotence [154]. After metastasis formation no curative treatment is currently available being surgical or medical castration the current standard palliative treatment [74]. Importantly, metastasis spread has been associated with most prostate cancer deaths [74, 155]. Recently, the U.S. Food and Drug Administration (FDA) approved a new therapeutic option to treat men with metastatic castration-resistant prostate cancer. However, this AR antagonist named Xtandi (MDV3100, enzalutamide, Astellas Pharma US, Inc., Northbrook, IL, USA) also has some side effects as seizures and weakness [156]. Therefore, it is very important to identify nonandrogenic and less aggressive therapeutic agents capable of reducing the proliferation of prostatic carcinoma [157]. Currently, PSA levels, digital rectal examination and histological grading are the most commonly used detection techniques, although the utility of these methods (especially PSA) are currently in discussion, thus providing a need for new molecular biomarkers [21]. A better knowledge of prostate cancer biology can provide the basis for such identification.

Among the different growth factors associated with normal prostate functioning, it has not been completely established which, if any, are associated with the acquisition of tumour cell autonomy. Transforming growth factor beta signalling pathway components have been regarded as possible targets for prostate cancer therapy for several years [158]. In fact, genistein acts through activation of Smad1, thus suppressing prostate cancer cell invasion, in an ALK2-dependent way [102]. Also, the delivery of oncolytic adenoviruses targeting TGF-B signalling resulted in less tumour burden, osteoclasts and trabecular and cortical destruction, thus representing a possible treatment for prostate cancer bone metastasis [159]. The targeting of more general TGF- β effectors, like p53, has also been defended [160]. The possibility of using TGF- β as a biomarker for prostate cancer has also been discussed for long [161]. Serum levels correlate with tumour burden, metastasis and serum PSA. Also, TGF-β presence in semen is probably related with tumour stage [74].

Because of the critical effect of phosphatases and kinases on TGF- β pathway and cancer metabolic control, inhibitors of these molecules may represent good alternative treatment options. Although kinases continue to retain the primary focus as drug targets for cancer therapy, phosphatases are receiving increasing attention [162].

Despite all the efforts made by researchers and pharmaceutical companies in this area, the only FDA-approved drugs targeting a protein phosphatase are cyclosporine A and FK506. These drugs are used as immunosuppressors and inhibit PPP3 (calcineurin). Unfortunately, because of the numerous functions of PPP1 and PPP2 catalytic subunits, the long-term usage of non-selective or marginal selective enzyme inhibitors is associated with nephrotoxicity and hepatotoxicity because of the inhibition of a number of critical cellular processes. For these reasons, it will be interesting to target PIPs instead of protein phosphatases directly as they are more event, tissue and subcellular compartment specific [60]. Nowadays, two targeted PPP1-PIP complexes have been described. The level of PPP1-GADD34 complex is diminished in cells treated with salubrinal, a small molecule that protect the cell from ER-stress-induced apoptosis [54]. Studies have suggested that this compound constitutes a potential treatment for the herpes simplex virus infection as it inhibits the virus replication [54]. The other complex involves PPP1 and histone deacetylases (HDACs) and is an attractive target to anti-tumour drugs. Trichostatin A, for example, disrupts the interaction between PPP1 and HDAC6 in glioblastoma and prostate cancer cells [54]. Here, we presented increasing evidence that makes PPP1-PIP complexes attractive targets to pursue in the near future for prostate cancer therapy. However, PPP1-PIPs complexes previously described may not be suitable for targeting because of the several functions and the ubiquitous expression of those PIPs. Thereof, a deeper knowledge of the PPP1 interactome in both normal and malignant prostate is required. This information could lead to a new understanding of PPP1-PIPs complexes functions and which PIPs are better targets for more focused therapeutical approaches in prostate cancer.

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Conflicts of interest

The authors confirm that there are no conflicts of interest.

References

- Kingsley DM. The TGF-beta superfamily: new members, new receptors, and new genetic tests of function in different organisms. Genes Dev. 1994; 8: 133–46.
- Dennler S, Goumans MJ, ten Dijke P. Transforming growth factor beta signal transduction. J Leukoc Biol. 2002; 71: 731–40.
- Lee C, Sintich SM, Mathews EP, et al.
 Transforming growth factor-beta in benign and malignant prostate. Prostate. 1999; 39: 285–90
- Moses HL. TGF-beta regulation of epithelial cell proliferation. Mol Reprod Dev. 1992; 32: 179–84.
- Imamura T, Hikita A, Inoue Y. The roles of TGF-beta signaling in carcinogenesis and breast cancer metastasis. *Breast Cancer*. 2012; 19: 118–24.
- Fuxe J, Karlsson MC. TGF-beta-induced epithelial-mesenchymal transition: a link between cancer and inflammation. Semin Cancer Biol. 2012; 22: 455–61.
- Nakamura T, Miller D, Ruoslahti E, et al.
 Production of extracellular matrix by glomerular epithelial cells is regulated by transforming growth factor-beta 1. Kidney Int. 1992; 41: 1213–21.
- Mamuya FA, Duncan MK. aV integrins and TGF-beta-induced EMT: a circle of regulation. J Cell Mol Med. 2012; 16: 445–55.
- de Visser KE, Kast WM. Effects of TGF-beta on the immune system: implications for cancer immunotherapy. *Leukemia*. 1999; 13: 1188–99
- Mahmoud M, Upton PD, Arthur HM. Angiogenesis regulation by TGFbeta signalling: clues from an inherited vascular disease. Biochem Soc Trans. 2011; 39: 1659–66.
- Huang C, Akaishi S, Ogawa R. Mechanosignaling pathways in cutaneous scarring. *Arch Dermatol Res.* 2012; 304: 589–97.
- 12. **Cao X, Chen D.** The BMP signaling and *in vivo* bone formation. *Gene.* 2005; 357: 1–8.
- Guo J, Wu G. The signaling and functions of heterodimeric bone morphogenetic pro-

- teins. *Cytokine Growth Factor Rev.* 2012; 23: 61–7.
- Gregory SJ, Kaiser UB. Regulation of gonadotropins by inhibin and activin. Semin Reprod Med. 2004; 22: 253–67.
- Yu J, Dolter KE. Production of activin A and its roles in inflammation and hematopoiesis. Cytokines Cell Mol Ther. 1997; 3: 169–77
- Ageta H, Tsuchida K. Multifunctional roles of activins in the brain. Vitam Horm. 2011; 85: 185–206.
- Lutz M, Knaus P. Integration of the TGFbeta pathway into the cellular signalling network. Cell Signal. 2002; 14: 977–88.
- Boyd FT, Cheifetz S, Andres J, et al. Transforming growth factor-beta receptors and binding proteoglycans. J Cell Sci Suppl. 1990: 13: 131–8.
- de Caestecker M. The transforming growth factor-beta superfamily of receptors. Cytokine Growth Factor Rev. 2004; 15: 1–11.
- Moustakas A, Heldin CH. The regulation of TGFbeta signal transduction. *Development*. 2009; 136: 3699–714.
- Dong C, Li Z, Alvarez R Jr, et al. Microtubule binding to Smads may regulate TGF beta activity. Mol Cell. 2000; 5: 27–34.
- Jin Q, Gao G, Mulder KM. Requirement of a dynein light chain in TGFbeta/Smad3 signaling. J Cell Physiol. 2009; 221: 707–15.
- Chen X, Xu L. Mechanism and regulation of nucleocytoplasmic trafficking of smad. *Cell Biosci.* 2011; 1: 40.
- 24. **Zhang YE.** Non-Smad pathways in TGF-beta signaling. *Cell Res.* 2009; 19: 128–39.
- Massague J. How cells read TGF-beta signals. Nat Rev Mol Cell Biol. 2000; 1: 169– 78
- Gordon KJ, Blobe GC. Role of transforming growth factor-beta superfamily signaling pathways in human disease. *Biochim Bio*phys Acta. 2008; 1782: 197–228.
- Elliott RL, Blobe GC. Role of transforming growth factor Beta in human cancer. J Clin Oncol. 2005; 23: 2078–93.

- Pasche B. Role of transforming growth factor beta in cancer. J Cell Physiol. 2001; 186: 153–68.
- Massague J. TGFbeta in Cancer. Cell. 2008; 134: 215–30.
- Padua D, Massague J. Roles of TGFbeta in metastasis. Cell Res. 2009: 19: 89–102.
- Tian M, Neil JR, Schiemann WP. Transforming growth factor-beta and the hallmarks of cancer. *Cell Signal*. 2011; 23: 951–62
- Jakowlew SB. Transforming growth factorbeta in cancer and metastasis. Cancer Metastasis Rev. 2006; 25: 435–57.
- 33. **Blagosklonny MV.** Molecular theory of cancer. *Cancer Biol Ther.* 2005; 4: 621–7.
- Harris CC. Molecular basis of multistage carcinogenesis. *Princess Takamatsu Symp*. 1991: 22: 3–19.
- 35. **Martin GS.** Cell signaling and cancer. *Cancer Cell*. 2003: 4: 167–74.
- 36. **Hanahan D, Weinberg RA.** The hallmarks of cancer. *Cell.* 2000; 100: 57–70.
- Blagosklonny MV. Cell immortality and hallmarks of cancer. Cell Cycle. 2003; 2:
- Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. *Cell.* 2011; 144: 646–74.
- Moses HL, Yang EY, Pietenpol JA. TGFbeta stimulation and inhibition of cell proliferation: new mechanistic insights. *Cell*. 1990; 63: 245–7.
- Holting T. Transforming growth factor-beta
 1 is a negative regulator for differentiated thyroid cancer: studies of growth, migration, invasion, and adhesion of cultured follicular and papillary thyroid cancer cell lines.
 J Clin Endocrinol Metab. 1994; 79: 806–13.
- Lomo J, Blomhoff HK, Beiske K, et al. TGF-beta 1 and cyclic AMP promote apoptosis in resting human B lymphocytes. J Immunol. 1995; 154: 1634–43.
- Vaughan MB, Howard EW, Tomasek JJ.
 Transforming growth factor-beta1 promotes the morphological and functional

- differentiation of the myofibroblast. *Exp Cell Res.* 2000: 257: 180–9.
- Frolik CA, Dart LL, Meyers CA, et al. Purification and initial characterization of a type beta transforming growth factor from human placenta. Proc Natl Acad Sci USA. 1983; 80: 3676–80.
- Siegel PM, Massague J. Cytostatic and apoptotic actions of TGF-beta in homeostasis and cancer. Nat Rev Cancer. 2003; 3: 807–21
- Polyak K, Kato JY, Solomon MJ, et al. p27Kip1, a cyclin-Cdk inhibitor, links transforming growth factor-beta and contact inhibition to cell cycle arrest. Genes Dev. 1994: 8: 9–22.
- Chen RH, Ebner R, Derynck R. Inactivation of the type II receptor reveals two receptor pathways for the diverse TGF-beta activities. Science. 1993; 260: 1335–8.
- Barrack ER. TGF-β in prostate cancer: a growth inhibitor that can enhance tumorigenicity. *Prostate*. 1997; 31: 61–70.
- Yamamura Y, Hua X, Bergelson S, et al. Critical role of Smads and AP-1 complex in transforming growth factor-beta -dependent apoptosis. J Biol Chem. 2000; 275: 36295–302.
- Perlman R, Schiemann WP, Brooks MW, et al. TGF-beta-induced apoptosis is mediated by the adapter protein Daxx that facilitates JNK activation. Nat Cell Biol. 2001; 3: 708–14.
- Francis JM, Heyworth CM, Spooncer E, et al. Transforming growth factor-beta 1 induces apoptosis independently of p53 and selectively reduces expression of Bcl-2 in multipotent hematopoietic cells. J Biol Chem. 2000; 275: 39137–45.
- Motyl T, Grzelkowska K, Zimowska W, et al. Expression of bcl-2 and bax in TGFbeta 1-induced apoptosis of L1210 leukemic cells. Eur J Cell Biol. 1998; 75: 367–74.
- Zhu B, Kyprianou N. Transforming growth factor beta and prostate cancer. Cancer Treat Res. 2005; 126: 157–73.
- Mirzoeva S, Franzen CA, Pelling JC. Apigenin inhibits TGF-beta-induced VEGF expression in human prostate carcinoma cells via a Smad2/3- and Src-dependent mechanism. Mol Carcinog. 2013. Doi:10. 1002/mc.22005
- McConnell JL, Wadzinski BE. Targeting protein serine/threonine phosphatases for drug development. *Mol Pharmacol.* 2009; 75: 1249–61.
- Fardilha M, Esteves SL, Korrodi-Gregorio L, et al. Identification of the human testis protein phosphatase 1 interactome. Biochem Pharmacol. 2011: 82: 1403–15.

- Cohen P. The origins of protein phosphorylation. Nat Cell Biol. 2002: 4: E127–30.
- Hanks SK, Hunter T. Protein kinases 6. The eukaryotic protein kinase superfamily: kinase (catalytic) domain structure and classification. FASEB J. 1995; 9: 576–96.
- 58. Kondapalli L, Soltani K, Lacouture ME. The promise of molecular targeted therapies: protein kinase inhibitors in the treatment of cutaneous malignancies. J Am Acad Dermatol. 2005; 53: 291–302.
- Fardilha M, Esteves SL, Korrodi-Gregorio L, et al. Protein phosphatase 1 complexes modulate sperm motility and present novel targets for male infertility. Mol Hum Reprod. 2011: 17: 466–77.
- Fardilha M, Esteves SL, Korrodi-Gregorio L, et al. The physiological relevance of protein phosphatase 1 and its interacting proteins to health and disease. Curr Med Chem. 2010; 17: 3996–4017.
- Cohen PT. Protein phosphatase 1–targeted in many directions. *J Cell Sci.* 2002; 115: 241–56.
- Virshup DM, Shenolikar S. From promiscuity to precision: protein phosphatases get a makeover. Mol Cell. 2009; 33: 537–45
- Bollen M. Combinatorial control of protein phosphatase-1. *Trends Biochem Sci.* 2001; 26: 426–31.
- 64. Barford D, Das AK, Egloff MP. The structure and mechanism of protein phosphatases: insights into catalysis and regulation. Annu Rev Biophys Biomol Struct. 1998; 27: 133–64
- Egloff MP, Johnson DF, Moorhead G, et al. Structural basis for the recognition of regulatory subunits by the catalytic subunit of protein phosphatase 1. EMBO J. 1997; 16: 1876–87.
- 66. Bollen M, Peti W, Ragusa MJ, et al. The extended PP1 toolkit: designed to create specificity. Trends Biochem Sci. 2010; 35: 450–8.
- Heroes E, Lesage B, Gornemann J, et al. The PP1 binding code: a molecular-lego strategy that governs specificity. FEBS J. 2013; 280: 584–95.
- Haesen D, Sents W, Ivanova E, et al. Cellular inhibitors of Protein Phosphatase PP2A in cancer. Biomed Res. 2012; 23: 197–211.
- Fayard E, Tintignac LA, Baudry A, et al. Protein kinase B/Akt at a glance. J Cell Sci. 2005; 118: 5675–8.
- Cheng JQ, Lindsley CW, Cheng GZ, et al.
 The Akt/PKB pathway: molecular target for cancer drug discovery. Oncogene. 2005; 24: 7482–92.

- Peng A, Maller JL. Serine/threonine phosphatases in the DNA damage response and cancer. *Oncogene*. 2010; 29: 5977–88.
- Shiloh Y. ATM and related protein kinases: safeguarding genome integrity. Nat Rev Cancer. 2003; 3: 155–68.
- Giacinti C, Giordano A. RB and cell cycle progression. *Oncogene*. 2006; 25: 5220–7.
- Wikstrom P, Damber J, Bergh A. Role of transforming growth factor-beta1 in prostate cancer. *Microsc Res Tech.* 2001; 52: 411–9.
- Gioeli D. Signal transduction in prostate cancer progression. Clin Sci. 2005; 108: 293–308.
- Saraon P, Jarvi K, Diamandis EP. Molecular alterations during progression of prostate cancer to androgen independence. Clin Chem. 2011; 57: 1366–75.
- Steiner MS, Barrack ER. Transforming growth factor-beta 1 overproduction in prostate cancer: effects on growth in vivo and in vitro. Mol Endocrinol. 1992; 6: 15–25.
- Wikstrom P, Stattin P, Franck-Lissbrant I, et al. Transforming growth factor beta1 is associated with angiogenesis, metastasis, and poor clinical outcome in prostate cancer. Prostate. 1998; 37: 19–29.
- Yu N, Kozlowski JM, Park II, et al. Overexpression of transforming growth factor beta1 in malignant prostate cells is partly caused by a runaway of TGF-beta1 autoinduction mediated through a defective recruitment of protein phosphatase 2A by TGF-beta type I receptor. Urology. 2010; 76: 1519 e8–13.
- Inman GJ, Allday MJ. Apoptosis induced by TGF-beta 1 in Burkitt's lymphoma cells is caspase 8 dependent but is death receptor independent. J Immunol. 2000; 165: 2500–10.
- Reynolds AR, Kyprianou N. Growth factor signalling in prostatic growth: significance in tumour development and therapeutic targeting. Br J Pharmacol. 2006; 147: S144– 52
- Bruckheimer EM, Kyprianou N. Bcl-2 antagonizes the combined apoptotic effect of transforming growth factor-beta and dihydrotestosterone in prostate cancer cells. *Prostate*. 2002; 53: 133–42.
- Zeng L, Rowland RG, Lele SM, et al. Apoptosis incidence and protein expression of p53, TGF-beta receptor II, p27Kip1, and Smad4 in benign, premalignant, and malignant human prostate. Hum Pathol. 2004; 35: 290–7.
- 84. **Vo BT, Cody B, Cao Y**, *et al.* Differential role of Sloan-Kettering Institute (Ski)

- protein in Nodal and transforming growth factor-beta (TGF-beta)-induced Smad signaling in prostate cancer cells. *Carcinogenesis*. 2012; 33: 2054–64.
- Yang HK, Kang SH, Kim YS, et al. Truncation of the TGF-beta type II receptor gene results in insensitivity to TGF-beta in human gastric cancer cells. Oncogene. 1999; 18: 2213–9.
- Kehrl JH. Transforming growth factor-beta: an important mediator of immunoregulation. Int J Cell Cloning. 1991; 9: 438–50.
- Zhang Q, Rubenstein JN, Jang TL, et al. Insensitivity to transforming growth factorbeta results from promoter methylation of cognate receptors in human prostate cancer cells (LNCaP). Mol Endocrinol. 2005; 19: 2390–9.
- 88. **Zhang Q, Chen L, Helfand BT, et al.** TGFbeta regulates DNA methyltransferase expression in prostate cancer, correlates with aggressive capabilities, and predicts disease recurrence. *PLoS ONE*. 2011; 6: e25168.
- Wang J, Sun L, Myeroff L, et al. Demonstration that mutation of the type II transforming growth factor beta receptor inactivates its tumor suppressor activity in replication error-positive colon carcinoma cells. J Biol Chem. 1995; 270: 22044–9.
- Guo Y, Jacobs SC, Kyprianou N. Downregulation of protein and mRNA expression for transforming growth factor-beta (TGFbeta1) type I and type II receptors in human prostate cancer. *Int J Cancer*. 1997; 71: 573–9.
- Blanchere M, Saunier E, Mestayer C, et al. Alterations of expression and regulation of transforming growth factor beta in human cancer prostate cell lines. J Steroid Biochem Mol Biol. 2002; 82: 297–304.
- Miles FL, Tung NS, Aguiar AA, et al. Increased TGF-beta1-mediated suppression of growth and motility in castrate-resistant prostate cancer cells is consistent with Smad2/3 signaling. Prostate. 2012; 72: 1339–50.
- Guo Y, Kyprianou N. Restoration of transforming growth factor beta signaling pathway in human prostate cancer cells suppresses tumorigenicity via induction of caspase-1-mediated apoptosis. Cancer Res. 1999; 59: 1366–71.
- 94. Guo Y, Kyprianou N. Overexpression of transforming growth factor (TGF) beta1 type II receptor restores TGF-beta1 sensitivity and signaling in human prostate cancer cells. Cell Growth Differ. 1998; 9: 185–93.
- 95. **Slingerland J, Pagano M.** Regulation of the cdk inhibitor p27 and its deregulation in cancer. *J Cell Physiol*. 2000; 183: 10–7.

- Hu XT. TGFbeta-mediated formation of pRb-E2F complexes in human myeloid leukemia cells. Biochem Biophys Res Commun. 2008; 369: 277–80.
- Ananth S, Knebelmann B, Gruning W, et al. Transforming growth factor beta1 is a target for the von Hippel-Lindau tumor suppressor and a critical growth factor for clear cell renal carcinoma. Cancer Res. 1999; 59: 2210–6.
- Ide H, Yoshida T, Matsumoto N, et al. Growth regulation of human prostate cancer cells by bone morphogenetic protein-2. Cancer Res. 1997; 57: 5022–7.
- 99. Huang F, Newman E, Theodorescu D, et al. Transforming growth factor beta 1 (TGF beta 1) is an autocrine positive regulator of colon carcinoma U9 cells in vivo as shown by transfection of a TGF beta 1 antisense expression plasmid. Cell Growth Differ. 1995; 6: 1635–42.
- Gatza CE, Oh SY, Blobe GC. Roles for the type III TGF-beta receptor in human cancer. Cell Signal. 2010; 22: 1163–74.
- Romero D, O'Neill C, Terzic A, et al. Endoglin regulates cancer-stromal cell interactions in prostate tumors. Cancer Res. 2011; 71: 3482–93.
- 102. Lakshman M, Huang X, Ananthanarayanan V, et al. Endoglin suppresses human prostate cancer metastasis. Clin Exp Metastasis. 2011; 28: 39–53.
- Romero D, Terzic A, Conley BA, et al. Endoglin phosphorylation by ALK2 contributes to the regulation of prostate cancer cell migration. Carcinogenesis. 2010; 31: 359–66.
- 104. Liu Y, Jovanovic B, Pins M, et al. Over expression of endoglin in human prostate cancer suppresses cell detachment, migration and invasion. Oncogene. 2002; 21: 8272–81.
- 105. Aitchison AA, Veerakumarasivam A, Vias M, et al. Promoter methylation correlates with reduced Smad4 expression in advanced prostate cancer. Prostate. 2008; 68: 661–74
- 106. Fuzio P, Ditonno P, Rutigliano M, et al. Regulation of TGF-beta1 expression by androgen deprivation therapy of prostate cancer. Cancer Lett. 2012; 318: 135–44.
- 107. Stravodimos K, Constantinides C, Manousakas T, et al. Immunohistochemical expression of transforming growth factor beta 1 and nm-23 H1 antioncogene in prostate cancer: divergent correlation with clinicopathological parameters. Anticancer Res. 2000; 20: 3823–8.
- 108. Cardillo MR, Petrangeli E, Perracchio L, et al. Transforming growth factor-beta

- expression in prostate neoplasia. *Anal Quant Cytol Histol*. 2000: 22: 1–10.
- 109. Zhu ML, Partin JV, Bruckheimer EM, et al. TGF-beta signaling and androgen receptor status determine apoptotic cross-talk in human prostate cancer cells. Prostate. 2008; 68: 287–95.
- Inoue Y, Imamura T. Regulation of TGFbeta family signaling by E3 ubiquitin ligases. Cancer Sci. 2008; 99: 2107–12.
- Massague J, Seoane J, Wotton D. Smad transcription factors. *Genes Dev.* 2005; 19: 2783–810.
- Liu T, Feng XH. Regulation of TGF-beta signalling by protein phosphatases. Biochem J. 2010; 430: 191–8.
- Heikkinen PT, Nummela M, Leivonen SK, et al. Hypoxia-activated Smad3-specific dephosphorylation by PP2A. J Biol Chem. 2010: 285: 3740–9.
- 114. Batut J, Schmierer B, Cao J, et al. Two highly related regulatory subunits of PP2A exert opposite effects on TGF-beta/Activin/ Nodal signalling. Development. 2008; 135: 2927–37.
- 115. Petritsch C, Beug H, Balmain A, et al. TGF-beta inhibits p70 S6 kinase via protein phosphatase 2A to induce G(1) arrest. Genes Dev. 2000; 14: 3093–101.
- 116. Bengtsson L, Schwappacher R, Roth M, et al. PP2A regulates BMP signalling by interacting with BMP receptor complexes and by dephosphorylating both the C-terminus and the linker region of Smad1. J Cell Sci. 2009; 122: 1248–57.
- 117. Lin X, Duan X, Liang YY, et al. PPM1A functions as a Smad phosphatase to terminate TGFbeta signaling. Cell. 2006; 125: 915–28.
- 118. Knockaert M, Sapkota G, Alarcon C, et al. Unique players in the BMP pathway: small C-terminal domain phosphatases dephosphorylate Smad1 to attenuate BMP signaling. Proc Natl Acad Sci USA. 2006; 103: 11940–5.
- Chen HB, Shen J, Ip YT, et al. Identification of phosphatases for Smad in the BMP/DPP pathway. Genes Dev. 2006; 20: 648–53.
- 120. Yu J, Pan L, Qin X, et al. MTMR4 attenuates transforming growth factor beta (TGFbeta) signaling by dephosphorylating R-Smads in endosomes. J Biol Chem. 2010; 285: 8454–62.
- 121. Yu J, He X, Chen YG, et al. Myotubularinrelated protein 4 (MTMR4) attenuates BMP/Dpp signaling by dephosphorylation of Smad proteins. J Biol Chem. 2013; 288: 79–88
- 122. Sapkota G, Knockaert M, Alarcon C, et al. Dephosphorylation of the linker regions of

- Smad1 and Smad2/3 by small C-terminal domain phosphatases has distinct outcomes for bone morphogenetic protein and transforming growth factor-beta pathways. *J Biol Chem.* 2006; 281: 40412–9.
- 123. Satow R, Kurisaki A, Chan TC, et al. Dullard promotes degradation and dephosphorylation of BMP receptors and is required for neural induction. Dev Cell. 2006; 11: 763–74.
- 124. Sakaguchi M, Sharmin S, Taguchi A, et al. The phosphatase Dullard negatively regulates BMP signalling and is essential for nephron maintenance after birth. Nat Commun. 2013; 4: 1398.
- Bennett D, Alphey L. PP1 binds Sara and negatively regulates Dpp signaling in Drosophila melanogaster. *Nat Genet*. 2002; 31: 419–23.
- 126. Shi W, Sun C, He B, et al. GADD34-PP1c recruited by Smad7 dephosphorylates TGFbeta type I receptor. J Cell Biol. 2004; 164: 291–300.
- 127. Valdimarsdottir G, Goumans MJ, Itoh F, et al. Smad7 and protein phosphatase 1alpha are critical determinants in the duration of TGF-beta/ALK1 signaling in endothelial cells. BMC Cell Biol. 2006; 7: 16.
- Shi W, Chang C, Nie S, et al. Endofin acts as a Smad anchor for receptor activation in BMP signaling. J Cell Sci. 2007; 120: 1216–24.
- 129. Gabel S, Benefield J, Meisinger J, et al. Protein phosphatases 1 and 2A maintain endothelial cells in a resting state, limiting the motility that is needed for the morphogenic process of angiogenesis. Otolaryngol Head Neck Surg. 1999; 121: 463–8.
- Mumby MC, Walter G. Protein serine/threonine phosphatases: structure, regulation, and functions in cell growth. *Physiol Rev.* 1993; 73: 673–99.
- Moorhead GB, Trinkle-Mulcahy L, Ulke-Lemee A. Emerging roles of nuclear protein phosphatases. Nat Rev Mol Cell Biol. 2007; 8: 234–44.
- 132. Amatangelo MD, Goodyear S, Varma D, et al. c-Myc expression and MEK1-induced Erk2 nuclear localization are required for TGF-beta induced epithelial-mesenchymal transition and invasion in prostate cancer. Carcinogenesis. 2012; 33: 1965–75.
- 133. Li DW, Liu JP, Schmid PC, et al. Protein serine/threonine phosphatase-1 dephosphorylates p53 at Ser-15 and Ser-37 to modulate its transcriptional and apoptotic activities. Oncogene. 2006; 25: 3006–22.
- 134. Xiao L, Gong LL, Yuan D, et al. Protein phosphatase-1 regulates Akt1 signal transduction pathway to control gene expres-

- sion, cell survival and differentiation. *Cell Death Differ*. 2010: 17: 1448–62.
- Martin-Granados C, Prescott AR, Van Dessel N, et al. A role for PP1/NIPP1 in steering migration of human cancer cells. PLoS ONE. 2012; 7: e40769.
- Van Eynde A, Wera S, Beullens M, et al.
 Molecular cloning of NIPP-1, a nuclear inhibitor of protein phosphatase-1, reveals homology with polypeptides involved in RNA processing. J Biol Chem. 1995; 270: 28068-74
- Hendrickx A, Beullens M, Ceulemans H, et al. Docking motif-guided mapping of the interactome of protein phosphatase-1. Chem Biol. 2009: 16: 365–71.
- Zhang Y, Kim TH, Niswander L. Phactr4 regulates directional migration of enteric neural crest through PP1, integrin signaling, and cofilin activity. *Genes Dev.* 2012; 26: 69–81.
- Massie CE, Lynch A, Ramos-Montoya A, et al. The androgen receptor fuels prostate cancer by regulating central metabolism and biosynthesis. EMBO J. 2011; 30: 2719–33.
- Heinlein CA, Chang C. Androgen receptor in prostate cancer. Endocr Rev. 2004; 25: 276–308.
- 141. Chen S, Kesler CT, Paschal BM, et al. Androgen receptor phosphorylation and activity are regulated by an association with protein phosphatase 1. J Biol Chem. 2009; 284: 25576–84.
- 142. Allard P, Zoubeidi A, Nguyen LT, et al. Links between Fer tyrosine kinase expression levels and prostate cell proliferation. Mol Cell Endocrinol. 2000; 159: 63–77.
- 143. Pasder O, Shpungin S, Salem Y, et al. Downregulation of Fer induces PP1 activation and cell-cycle arrest in malignant cells. Oncogene. 2006; 25: 4194–206.
- 144. Berndt N, Dohadwala M, Liu CW. Constitutively active protein phosphatase 1alpha causes Rb-dependent G1 arrest in human cancer cells. Curr Biol. 1997: 7: 375–86.
- 145. **Kaelin WG Jr.** Functions of the retinoblastoma protein. *BioEssays*. 1999; 21: 950–8.
- Freeman MR, Yang W, Di Vizio D. Caveolin-1 and prostate cancer progression. Adv Exp Med Biol. 2012; 729: 95–110.
- 147. Thompson TC, Tahir SA, Li L, et al. The role of caveolin-1 in prostate cancer: clinical implications. Prostate Cancer Prostatic Dis. 2010; 13: 6–11.
- Yang G, Truong LD, Wheeler TM, et al. Caveolin-1 expression in clinically confined human prostate cancer: a novel prognostic marker. Cancer Res. 1999; 59: 5719–23.
- 149. Karam JA, Lotan Y, Roehrborn CG, et al. Caveolin-1 overexpression is associated

- with aggressive prostate cancer recurrence. *Prostate*. 2007: 67: 614–22.
- 150. Li L, Ren CH, Tahir SA, et al. Caveolin-1 maintains activated Akt in prostate cancer cells through scaffolding domain binding site interactions with and inhibition of serine/threonine protein phosphatases PP1 and PP2A. Molecular and Cellular Biology. 2003; 23: 9389–404.
- 151. Chen RH, Su YH, Chuang RL, et al. Suppression of transforming growth factor-beta-induced apoptosis through a phosphatidylinositol 3-kinase/Akt-dependent pathway. Oncogene. 1998; 17: 1959– 68
- Jemal A, Bray F, Center MM, et al. Global cancer statistics. CA Cancer J Clin. 2011; 61: 69–90.
- 153. Feldman BJ, Feldman D. The development of androgen-independent prostate cancer. Nat Rev Cancer. 2001; 1: 34–45.
- Hugosson J, Stranne J, Carlsson SV. Radical retropubic prostatectomy: a review of outcomes and side-effects. *Acta Oncol.* 2011: 50: 92–7.
- Denmeade SR, Isaacs JT. A history of prostate cancer treatment. Nat Rev Cancer. 2002; 2: 389–96.
- Scher HI, Fizazi K, Saad F, et al. Increased survival with enzalutamide in prostate cancer after chemotherapy. N Engl J Med. 2012; 367: 1187–97.
- 157. Rokhlin OW, Cohen MB. Differential sensitivity of human prostatic cancer cell lines to the effects of protein kinase and phosphatase inhibitors. Cancer Lett. 1995; 98: 103–10
- 158. Kirkbride KC, Blobe GC. Inhibiting the TGFbeta signalling pathway as a means of cancer immunotherapy. Expert Opin Biol Ther. 2003; 3: 251–61.
- 159. Hu Z, Gupta J, Zhang Z, et al. Systemic delivery of oncolytic adenoviruses targeting transforming growth factor-beta inhibits established bone metastasis in a prostate cancer mouse model. Hum Gene Ther. 2012; 23: 871–82.
- 160. Komarova EA, Gudkov AV. Suppression of p53: a new approach to overcome side effects of antitumor therapy. *Biochemistry*. 2000: 65: 41–8
- 161. Thompson TC, Truong LD, Timme TL, et al. Transforming growth factor beta 1 as a biomarker for prostate cancer. J Cell Biochem Suppl. 1992; 16H: 54–61.
- 162. Jailkhani N, Chaudhri VK, Rao KV. Regulatory cascades of protein phosphatases: implications for cancer treatment. Anticancer Agents Med Chem. 2011; 11: 64–77.