

WJG 20th Anniversary Special Issues (8): Gastric cancer**Ethnic differences in gastric cancer genetic susceptibility:
Allele flips of interleukin gene**

Juwon Kim, Yoonjung Kim, Kyung-A Lee

Juwon Kim, Department of Laboratory Medicine, Yonsei University Wonju College of Medicine, Gangwon-do 220-701, South Korea

Yoonjung Kim, Kyung-A Lee, Department of Laboratory Medicine, Yonsei University College of Medicine, Seoul 135-720, South Korea

Author contributions: Kim J, Kim Y and Lee KA wrote and edited the manuscript.

Correspondence to: Kyung-A Lee, MD, PhD, Department of Laboratory Medicine, Yonsei University College of Medicine, 211 Eonju-ro, Gangnam-gu, Seoul 135-720, South Korea. kal1119@yuhs.ac

Telephone: +82-2-20193531 Fax: +82-2-20194822

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Abstract

Polymorphisms in promoter regions of inflammatory cytokines have been widely studied, and potentially functional polymorphisms have been discovered. Conflicting results from meta-analyses of interleukin (*IL*)-1 β and *IL*-10 polymorphisms show differences in gastric cancer susceptibilities between Caucasian and Asian populations. In particular, we note the suggestion of an allele flip in *IL*-1 β and *IL*-10 gene polymorphisms. In Asian populations, the *IL*-1 β -1464G/-511C/-31T haplotype indicates risk for gastric cancer, while the opposite haplotype, *IL*-1 β -1464C/-511T/-31C is the risk-related allele in Caucasians. Furthermore, while *IL*-10-1082G/-819C/-592C is associated with gastric cancer in Asians, *IL*-10-1082A/-819T/-592T is linked to gastric cancer risk in Caucasians. These seemingly contradictory results may be attributed to distinct carcinogenic mechanisms underlying the different gastric cancer subtypes. The allele flip observed in *IL*-10 and gastric cancer appears to reflect allelic heterogeneity, similar to that observed in *IL*-1 β . In this review, we focus on the allele flip phenomenon observed between different ethnic groups in

an effort to resolve certain controversial results from recent studies on interleukin polymorphism. In addition, we re-emphasize the importance of stratifying gastric cancer subtypes based on anatomical site and Lauren classification to prevent false associations arising through dilution of true ones.

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Key words: Allele flip; Gastric cancer; Interleukin gene

Core tip: In Asian populations, the highly expressed interleukin (*IL*)-1 β haplotype may increase risk for gastric cancer. Abundant *IL*-1 β expression determined by this haplotype may suppress gastric acid production in response to chronic *Helicobacter pylori* (*H. pylori*) infection, resulting in atrophic gastritis, the precursor of non-cardia gastric cancer. Conversely, the less expressive *IL*-1 β haplotype associates with gastric cardia cancer in Caucasians. Only low levels of *IL*-1 β are produced in response to *H. pylori* infection and gastric acid secretion is increased. Induction of gastroesophageal reflux disease may then promote cardia cancers.

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INTRODUCTION

Gastric cancer is the fourth most common cancer diagnosis in men worldwide, and the mortality rate is one of the highest among cancers. The incidence rate is especially high in East Asian countries including Japan, China and Korea^[1,2]. Of the various factors that contribute to

gastric cancer, including infectious, dietary, environmental and genetic factors, the chronic inflammatory state induced by *Helicobacter pylori* (*H. pylori*) infection is currently regarded as the most prevalent. Of note, however, only a small proportion of *H. pylori*-infected individuals develop gastric cancer, which implies that individual susceptibility, possibly genetic, is also involved^[1,3,4].

Associations of chronic inflammation with carcinogenic processes have prompted researchers to investigate the role that *H. pylori*-related inflammation may play in gastric cancer development. Thus, it was found that inflammatory cytokines, such as interleukin (IL)-1 β , also encoded by *IL-1B*, IL-1 receptor antagonist (*IL-1RA*), and tumor necrosis factor (TNF)- α are upregulated during *H. pylori* infection^[5-8]. Interest now converges on *IL-1B*, *IL-1RN*, *IL-8* and *IL-10*, encoding IL-1 β , IL-1RA, IL-8, and IL-10. The potent proinflammatory cytokine IL-1 β participates in a variety of cellular activities, including cell proliferation, differentiation and apoptosis^[9], in the amplification of immune response to infection, and as a potent inhibitor of gastric acid secretion^[10]. Studies on single nucleotide polymorphisms (SNPs) in the *IL-1B* promoter region reveal significant associations with gastric cancer that some meta-analyses support^[10,11]. Polymorphisms of *IL-10* and *IL-8* are also associated with gastric cancer risk^[10].

Other meta-analyses, however, present conflicting results with respect to *IL-1B* and *IL-10* polymorphisms and gastric cancer susceptibilities between Caucasian and Asian populations. On review of multiple meta-analyses, an “allele flip” between Asian and non-Asian groups is observed; most prominently in polymorphisms of *IL-1B* and *IL-10*. The allele flip refers to an inverse risk relationship of an allele in different groups or settings, for example, an allele found to be protective in one situation, but risk-related in another^[12]. A genuine allele flipping results from variations in allele frequencies and linkage disequilibrium (LD) that produce different patterns of risk association of a marker allele or haplotype across different ethnic groups^[13]. Alternatively, multiple loci may interact to create a disease phenotype^[14]. Finally, the phenomenon may be caused by allelic heterogeneity and locus heterogeneity, wherein different populations exhibit associations with alleles at different loci, through differences in genetic background or environment^[12]. Despite extensive review using meta-analysis, no clear explanation of allele flipping among these interleukin genes between different ethnic groups has emerged.

Here we concentrate on allele flips of *IL-1B* and *IL-10* polymorphisms in association with gastric cancer development in Asian and Caucasian groups. Of particular interest are the etiological significance of flipping in relation to genetic susceptibility and the incidence of gastric cancer at different anatomical sites.

GASTRIC CANCER EPIDEMIOLOGY AND CLASSIFICATION

Gastric cancer may be classified by histopathological

criteria as proposed by Lauren *et al*^[15] into two principal types, intestinal and diffuse, which differ in histology, pathogenesis, epidemiology, genetic profile, and prognosis^[16]. Based on anatomical site, stomach cancer may be classified as cardia or non-cardia (fundus, antrum, pylorus lesser curvature, and greater curvature)^[17]. Non-cardia intestinal gastric cancer is strongly associated with chronic inflammation related to *H. pylori* infection^[3,4]. For tumors arising in the proximal region of the stomach, namely the gastric cardia/gastroesophageal junction, inflammation due to chronic gastric acid secretion may be the driving force in carcinogenesis^[18,19]. Gastric cardia cancers are usually of the intestinal type^[20]. As proposed by Hansen *et al*^[21], the cardia cancers may comprise two distinct etiological subtypes, one non-cardia-like gastric cancer and the other resembling esophageal adenocarcinoma. The types of gastric cancer are summarized in Table 1. Diffuse-type gastric cancer is thought to arise through genetic changes in gastric cancer stem cells or epithelial precursor cells and usually lacks defined premalignant lesions^[22]. Recently, Shah *et al*^[20] proposed a classification of gastric cancer based on clinical and epidemiological data into three principal types: proximal nondiffuse gastric cancer, diffuse gastric cancer, and distal nondiffuse gastric cancer. These distinctions are supported by gene expression analysis.

In the United States, rates of gastric cancer at all sites decreased from 1978 to 2005, while cardia cancer rates increased in the early years and plateaued^[17]. With respect to histological type, intestinal-type cancer decreased at all sites except at the gastric cardia^[17]. The declining prevalence of *H. pylori* infection has most likely contributed to downward trends in intestinal-type gastric cancer at non-cardia sites, particularly in Caucasians^[17]. Among Asians living in the *H. pylori* endemic region, there is ample evidence that non-cardia types outnumber cardia types, although the incidence of cardia-type gastric cancers has increased in recent years^[23].

IL-1 GENE POLYMORPHISM

Non-cardia, intestinal type gastric cancer

The IL-1 family includes the cytokines IL-1 α , IL-1 β and IL-1RA, encoded by three genes, *IL-1A*, *IL-1B* and *IL-1RN* on chromosome 2q14^[24,25]. Three polymorphisms in the promoter region of *IL-1B*, including *IL-1B*-1464 (G/C; rs1143623; previously known as -1476), *IL-1B*-511 (C/T; rs16944), and *IL-1B*-31 (T/C; rs1143627), are widely studied in association with gastric cancer risk. Studies of the *IL-1B* polymorphism have revealed an increased risk of gastric cancer with proinflammatory phenotype in Caucasian carriers of *IL-1B*-31C and *IL-1B*-511T^[11]. In meta-analyses, however, the *IL-1B* and *IL-1RN* polymorphisms imply different levels of risk for Asians and Caucasians. A landmark meta-analysis by Persson *et al*^[26] revealed a consistent negative association of *IL-1B*-31C with gastric cancer in Asians. Other data show an even more significant increase in risk for non-cardia gastric cancer related to *IL-1B*-511T and *IL-1RN**2 alleles, but

Table 1 Differing risk associations for cytokine polymorphisms according to race and tumor type

Tumor type	Non-cardia or cardia with atrophy (non-cardia-like), intestinal		Cardia (esophageal), intestinal		Diffuse	
	Asian	Non-asian	Non-asian	Asian	Asian	Non-asian
Race						
Direction of association						
<i>H. pylori</i> infection	↑	↑	↓	↓	↑	↑
Gastric acid secretion	↓	↓	↑	↑	-	-
IL-1β production	↑	↑	↓	↓	-	-
IL-10 production	-	↓	-	↑	-	-
Allele flips of IL-1B and IL-10 (risk haplotype)						
IL-1B-1464/-511/-31	GCT	CTC	CTC	-	-	-
IL-10-1082/-819/-592	-	ATA	-	GCC	-	-
Genetic factors other than IL-1B and IL-10 ¹	IL-8, ZBTB20, PRKAA1	IL-1RN			MUC1, PSCA	IL-1RN, TNFA
Molecular classification by gene expression analysis ^[20]	Distal non-diffuse		Proximal non-diffuse		Diffuse	

¹Information based on meta-analyses^[10,26] and GWAS^[69-71,81]. ↑: Increase; ↓: Decrease. *H. pylori*: *Helicobacter pylori*; IL: Interleukin; PSCA: Prostate stem cell antigen.

only among Caucasians, while the *IL-1B*-511T allele may be protective against gastric cancer among Asians^[27,28]. In addition, complete LD between *IL-1B*-31 and -511 has been found^[11]. In East Asian populations, *IL-1B*-31TT homozygosity may be associated with increased risk for intestinal-type gastric cancer^[29].

Another polymorphism in the *IL-1B* promoter region at -1464 may be associated with gastric cancer, and -1464G is a putative risk allele in Asians^[30]. Moreover, -1464G is closely linked to *IL-1B*-511C/-31T alleles previously designated as risk alleles in Asians. In contrast, -1464C, in LD with *IL-1B*-511T/-31C and a risk allele among Caucasians, is associated with decreased risk of gastric cancer in the Chinese population^[31]. In atrophic gastritis, a precursor lesion in gastric cancer, -1464CC, may be associated with atrophic gastritis in the antrum among Caucasians^[32]. The haplotype associated with gastric cancer risk in Asians (*IL-1B*-1464G/-511C/-31T) may imply the opposite level of risk in Caucasians, among whom the *IL-1B*-1464C/-511T/-31C is the putative risk allele.

However, in a country such as China, comprising multiple ethnic groups with diverse geographical and historical roots, allelic heterogeneity with respect to gastric cancer prevalence and *H. pylori* infection status is apparent^[33]. Of note, -511TT is associated with an increased risk of gastric cancer in low-risk regions of China, an association that might be less obvious in high-risk regions^[33]. This is similar to the situation wherein -511TT is associated with increased gastric cancer risk in a Caucasian population with lower gastric cancer risk and *H. pylori* infection, whereas -511CC is the risk allele among Asian population, namely China, Korea and Japan, where gastric cancer risk is high. Therefore, it is plausible that *IL-1B*-1464G/-511C/-31T are the risk alleles for gastric cancer among Asians, while the exact opposite is true for Caucasians, indicating the existence of a genuine allele flip in the *IL-1B* gene polymorphisms with respect to gastric cancer risk.

To test the influence of haplotype on IL-1β expres-

sion, Chen *et al.*^[34] investigated the effect of SNPs in the *IL-1B* promoter region in terms of haplotype context. Of note, the SNPs at -1464, -511 and -31 in the promoter region expressed functional activities that were influenced by haplotype context^[34]. This observation was confirmed in a subsequent study *in vivo*, by the finding of a positive association between haplotype pairs containing *IL-1B*-1464G/-511C/-31T and levels of IL-1β expression in Caucasian subjects, despite previous understanding of *IL-1B*-511T/-31C as the proinflammatory allele^[35,36]. In an *in vitro* study, -31T expressed stronger promoter activity than -31C by virtue of retaining the TATA sequence, and showed greater binding affinity for transcription factors as well^[36]. The haplotype consisting of *IL-1B*-1464G/-511C/-31T shows a positive association with lung cancer risk and higher *IL-1B* gene expression in Caucasians^[37]. In evaluating transcriptional activities of individual SNPs, the -1464C SNP had higher transcriptional activity by itself^[38]. Placing the SNPs in haplotype context, however, the G allele at -1464 in the -1464G/-511C/-31T haplotype combination expressed higher transcriptional and translational activities, underscoring the influence of other SNPs in the genetic environment on individual SNPs^[38].

It is also possible that the allele flip seen in *IL-1B* polymorphisms results from allelic heterogeneity reflecting differences in clinical backgrounds between Asian and Caucasian populations, such as the prevalence of *H. pylori*-related premalignant gastric lesions and cancer arising at different anatomical sites. The presence of *H. pylori* infection is strongly associated with risk of non-cardia intestinal gastric cancer. The *IL-1B*-1464G/511C/-31T haplotype, with high mucosal IL-1β expression, is believed to be a proinflammatory allele that produces IL-1β in excess in response to *H. pylori* infection, and suppresses gastric acid secretion. Prolonged hypochlorhydria provides an environment favorable to *H. pylori* survival, leading to atrophic gastritis or intestinal metaplasia, and subsequently to non-cardia intestinal-type gastric cancer; the predominant subtype among East Asians. The direct

suppression of gastrin secretion by excess IL-1 β expression in association with the -31T allele may be directly observed^[39].

In non-cardia intestinal-type gastric cancer in Caucasians, *IL-1RN**2 may be an essential factor. IL-1RA, encoded by the *IL-1RN* gene, is a naturally occurring anti-inflammatory cytokine that competes with the binding of IL-1 to its receptor^[40]. The second intron of *IL-1RN* includes a penta-allelic 86-bp variable number tandem repeat producing two repeats (*IL-1RN**2) or three to six repeats (*IL-1RN**L)^[40]. As El-Omar *et al*^[11] have suggested, the presumably proinflammatory *IL-1RN**2 and *IL-1B*-31C haplotypes, presenting risk factors for *H. pylori*-related gastric cancer, may be in strong LD in Caucasian populations. Accordingly, low acid secretion shows a significant positive association with *IL-1RN**2, and homozygosity for this allele increases risk of hypochlorhydria. In an *in vitro* study, *IL-1RN**2 evidently increased production of IL-1B, regardless of the allele type of *IL-1B*, indicating that *IL-1RN**2 has a decisive role, not the IL-1B polymorphisms^[41]. In the Human Genome Epidemiology Network (HuGE) meta-analysis, the association of *IL-1RN**2 with gastric cancer detected appears to be confined to non-Asian populations, because overall frequency of the *IL-1RN**2 allele among Asians is low, if measurable^[26,32].

Cardia, intestinal-type gastric cancer

Studies of gene associations in cardia cancer are conducted mostly with Caucasian subjects because non-cardia, intestinal-type cancer predominates among Asians. In an unusual investigation, Kamangar *et al*^[42] first divided cancer into cardia and non-cardia gastric adenocarcinoma and then tested associations with *H. pylori*. *H. pylori* showed a strong positive risk association with non-cardia gastric cancer but an inverse association with cardia gastric cancer risk^[42]. Some studies have found that *H. pylori* infection is associated with decreased risk of adenocarcinoma arising near the esophagogastric junction^[42-47]. This may be explained by the tendency of *H. pylori* colonization to induce gastric atrophy, with reduced acid secretion, thereby reducing acid reflux into the esophagus, and reducing risk of esophageal or junction cancer^[42,48]. The cardia cancers were positively associated with gastroesophageal reflux disease (GERD)^[49]. The *IL-1B*-1464C/-511T/-31C allele among Caucasians is associated with low levels of IL-1 β expression in response to *H. pylori* infection or other inflammatory stimuli, and could not efficiently suppress gastric acid. Increased acid production following a subsequent inflammatory response would then produce GERD-like symptoms and promote cardia cancers.

IL-10 GENE POLYMORPHISM

IL-10, encoded by the *IL-10* gene at chromosome 1q31.1, is an anti-inflammatory cytokine. Three polymorphisms in the *IL-10* promoter, namely *IL-10*-1082

(G/A; rs1800896), -819 (C/T; rs1800871), and -592 (C/A; rs1800872), are shown to influence inflammation in response to infection at the transcriptional level^[50-52]. An allele flip in *IL-10* polymorphisms with respect to gastric cancer risk is also observed. In Caucasian populations, risk for non-cardia gastric cancers in association with the -1082AA genotype may be increased twofold, while the -1082G allele is the risk allele in cardia gastric cancer in studies of Asians, independent of *H. pylori* infection^[53-55]. In a meta-analysis, *IL-10*-1082G carriers showed a significant increase in risk of developing gastric cancer, especially for cardia-type gastric cancer among Asians^[56,57]. A recent meta-analysis by Yu *et al*^[58] showed a significantly negative association of *IL-10*-819TT with gastric cancer risk in Asians, in accordance with the previous finding that *IL-10*-819CC is a risk allele^[59]. Furthermore, identification of *IL-10*-592AA as a protective allele for total gastric cancer incidence in Asians supports *IL-10*-1082G/-819C/-592C as the risk haplotype^[60,61].

Evidence indicates that selection mechanisms operating on the *IL-10* region differ among ethnic groups. In Asian populations, with relatively high prevalence of chronic *H. pylori* infection, *IL-10*-1082A, is found more frequently than in Caucasian populations. Relatively low IL-10 expression by *IL-10*-1082A promotes elimination of *H. pylori* infection, and this may exert positive selective pressure on the haplotype. In Caucasian populations *H. pylori* infection is less prevalent, and greater IL-10 production would be advantageous in defense against infectious and inflammatory diseases^[55]. This may explain the relatively high frequency of the *IL-10*-1082G allele in Caucasian populations^[55]. Evidence for balancing selection within the IL-10 promoter region is consistently reported in studies of European populations^[62,63].

In Caucasian populations with low rates of *H. pylori* infection and premalignant lesions, the -1082A allele imposes risk for gastric cancer through low IL-10 production and consequent excess of proinflammatory cytokines. This promotes inflammation of the gastric mucosa, which may increase frequency of the mutation^[55,64]. These findings are consistent with observations of carcinogenesis in non-cardia cancer. In Asian populations, wherein *H. pylori* infection and premalignant lesions such as atrophic gastritis and intestinal metaplasia are more common, high-expression *IL-10*-1082G may suppress cytotoxic anti-tumor T-cell activity and thereby promote tumor progression^[55,65]. In high-risk populations, IL-10 may play an essential role in advanced stages of gastric cancer^[55]. In the Taiwanese population, *IL-10*-1082G may be linked to gastric cancer risk and advanced cancer, and cardia location of gastric cancer may be associated with a high-producing *IL-10* genotype^[66]. Carriers of the *IL-10*-1082G/-819C/-592C haplotype may be susceptible to virulent *H. pylori* strains with a high capability to colonize and adapt, and also to gastric cancer development^[67]. As compared to carriers of the *IL-10*-1082A/-819T/-592A haplotype, those with the *IL-10*-1082G/-819C/-592C carriers show higher mucosal levels of IL-10 mRNA,

which may result in a diminished proinflammatory response and capacity to control *H. pylori* infection^[67]. Actually, it appears that the *IL-10* genotype at -1082 is sufficient to establish a risk relationship with gastric cancer, because the *IL-10*-1082 genotype correlates well with mucosal *IL-10* mRNA levels, and *IL-10*-1082G fully represents the high-expression *IL-10*-1082G/-819C/-592C haplotype^[55,67]. Consequently, the allele flip of *IL-10* observed in gastric cancer represents allelic heterogeneity, similar to that observed in *IL-1B*.

GENETIC FACTORS OTHER THAN IL-1B AND IL-10

Diffuse-type gastric cancer

Intestinal-type gastric cancer follows a multistep progression that usually initiates in chronic gastritis, whereas diffuse-type gastric cancer lacks defined premalignant lesions; diffuse-type gastric cancer is therefore suspected to be more influenced by genetics factors^[22,68]. Genome-wide association studies (GWASs) reveal some additional gastric cancer susceptibility loci^[69]. Detected in a Japanese GWAS, an SNP (rs2976392) in the prostate stem cell antigen (*PSCA*) gene, which encodes a glycosylphosphatidylinositol-anchored cell surface antigen, shows a significant association with diffuse-type gastric cancer^[70]. Two SNPs (rs2070803 and rs4072037) in mucin 1 (*MUC1*) also show positive risk associations with diffuse-type gastric cancer in Asian populations^[69-71]. We found that normal T cell expressed and secreted (*RANTES*)-403A presents a significant increase in risk for diffuse-type gastric cancer in an Asian male population, when stratified by Lauren classification and sex^[72].

Non-cardia, intestinal-type gastric cancer

The chemokine *IL-8* participates in the initiation and amplification of acute inflammatory reactions as well as in the maintenance of chronic inflammatory processes^[73]. Evidence now links *IL-8* to tumorigenesis, angiogenesis and metastasis^[74-76]. A meta-analysis has shown an increased risk of *IL-8*-251A (T/A; rs4073) allele in several cancers, including gastric cancer, among Asians, but no such correlations among Europeans, suggesting racial differences in disease susceptibility with respect to *IL-8* polymorphisms^[77]. A case-control study has also shown no significant association between *IL-8*-251 polymorphism and increased risk of gastric cancer, whereas the association remains in Asians^[78]. In gastric cardia adenocarcinoma (non-cardia like), but not esophageal squamous cell carcinoma, the AGT/AGC haplotypes of *IL-8* polymorphisms showed a fourfold increase in relative risk in a high-risk Chinese population^[79]. Unfortunately, most association studies on *IL-8* polymorphisms have focused on a single polymorphism at -251, without considering its haplotype structure. The *IL-8*-251A allele resides on two different haplotypes, and only one of these is associated with disease^[80]. In other words, information regarding the *IL-8* haplotype structure is essential in determining the

true relationship between *IL-8* polymorphisms and gastric cancer development.

Finally, the rs13361707 SNP in the first intron of protein kinase, AMP-activated, $\alpha 1$ catalytic subunit (PPKAA1) and rs9841504 in the intron of zinc finger and BTB domain containing protein 20 (ZBTB20) emerge as susceptibility loci from GWAS analysis^[81].

CONCLUSION

Here, we aimed to summarize our rapidly evolving understanding of polymorphic structure in the interleukin promoter region and the involvement of *IL-1B* and *IL-10* polymorphisms in gastric cancer development. Analysis of these polymorphisms offers possible explanations for the allele flip observed in associations of *IL-1B* and *IL-10* with gastric cancer risk. The epidemiology of gastric cancer subtypes suggests a difference in genetic background between Asian and Caucasian groups. Among Asians, the *IL-1B*-1464G/-511C/-31T haplotype presents a risk allele for gastric cancer. This corresponds physiologically to increased *IL-1 β* expression in response to chronic *H. pylori* infection, which may inhibit gastric acid production and promote atrophic gastritis and non-cardia gastric cancer. Then, what about the gastric cardia cancer, which affects only a minority of Asians? In Asians, the highly expressive *IL-10* allele may serve to augment the inflammatory response to colonization by virulent strains of *H. pylori*, and, following malignant transformation high *IL-10* production, may tend to suppress anti-tumor cytotoxic T-cell response, thereby contributing to tumor progression. *IL-1 β* expression influences the initiation of cancer in response to *H. pylori* infection, whereas *IL-10* influences tumor progression after malignant transformation. Conversely, the less-expressive haplotype of *IL-1B* is associated with gastric cancer risk in Caucasians, specifically cancer of the gastric cardia. In this setting, low levels of *IL-1 β* produced in response to *H. pylori* infection may increase gastric acid secretion, promoting gastric cardia cancers through induction of GERD. Concerning non-cardia gastric cancers in Caucasians, the less expressive *IL-10* haplotype may promote metaplasia in the distal portion of the stomach by augmenting inflammatory response, while the *IL-1RN**2 polymorphism contributes by activating *IL-1 β* production. In conclusion, stratifying gastric cancer subtypes according to both anatomical site and the Lauren histological classification is essential in establishing genetic risk factors, because different subtypes follow different pathways of development and failure to consider this may produce false associations.

REFERENCES

- 1 Guggenheim DE, Shah MA. Gastric cancer epidemiology and risk factors. *J Surg Oncol* 2013; **107**: 230-236 [PMID: 23129495 DOI: 10.1002/jso.23262]
- 2 Jemal A, Bray F, Center MM, Ferlay J, Ward E, Forman D. Global cancer statistics. *CA Cancer J Clin* 2011; **61**: 69-90 [PMID: 21296855 DOI: 10.3322/caac.20107]
- 3 Peek RM, Blaser MJ. Helicobacter pylori and gastrointestinal

- tract adenocarcinomas. *Nat Rev Cancer* 2002; **2**: 28-37 [PMID: 11902583 DOI: 10.1038/nrc703]
- 4 **Uemura N**, Okamoto S, Yamamoto S, Matsumura N, Yamaguchi S, Yamakido M, Taniyama K, Sasaki N, Schlemper RJ. Helicobacter pylori infection and the development of gastric cancer. *N Engl J Med* 2001; **345**: 784-789 [PMID: 11556297 DOI: 10.1056/NEJMoa001999]
 - 5 **El-Omar EM**. The importance of interleukin 1beta in Helicobacter pylori associated disease. *Gut* 2001; **48**: 743-747 [PMID: 11358884]
 - 6 **El-Omar EM**, Carrington M, Chow WH, McColl KE, Bream JH, Young HA, Herrera J, Lissowska J, Yuan CC, Rothman N, Lanyon G, Martin M, Fraumeni JF, Rabkin CS. The role of interleukin-1 polymorphisms in the pathogenesis of gastric cancer. *Nature* 2001; **412**: 99 [PMID: 11808612 DOI: 10.1038/35083631]
 - 7 **El-Omar EM**, Chow WH, Rabkin CS. Gastric cancer and H. pylori: Host genetics open the way. *Gastroenterology* 2001; **121**: 1002-1004 [PMID: 11606513]
 - 8 **Furuta T**, El-Omar EM, Xiao F, Shirai N, Takashima M, Sugimura H. Interleukin 1beta polymorphisms increase risk of hypochlorhydria and atrophic gastritis and reduce risk of duodenal ulcer recurrence in Japan. *Gastroenterology* 2002; **123**: 92-105 [PMID: 12105837]
 - 9 **Dinarello CA**. Biologic basis for interleukin-1 in disease. *Blood* 1996; **87**: 2095-2147 [PMID: 8630372]
 - 10 **Schneider BG**, Camargo MC, Ryckman KK, Sicinski LA, Piazuelo MB, Zabaleta J, Correa P, Williams SM. Cytokine polymorphisms and gastric cancer risk: an evolving view. *Cancer Biol Ther* 2008; **7**: 157-162 [PMID: 18059184]
 - 11 **El-Omar EM**, Carrington M, Chow WH, McColl KE, Bream JH, Young HA, Herrera J, Lissowska J, Yuan CC, Rothman N, Lanyon G, Martin M, Fraumeni JF, Rabkin CS. Interleukin-1 polymorphisms associated with increased risk of gastric cancer. *Nature* 2000; **404**: 398-402 [PMID: 10746728 DOI: 10.1038/35006081]
 - 12 **Clarke GM**, Cardon LR. Aspects of observing and claiming allele flips in association studies. *Genet Epidemiol* 2010; **34**: 266-274 [PMID: 20013941 DOI: 10.1002/gepi.20458]
 - 13 **Evans DM**, Cardon LR. A comparison of linkage disequilibrium patterns and estimated population recombination rates across multiple populations. *Am J Hum Genet* 2005; **76**: 681-687 [PMID: 15719321 DOI: 10.1086/429274]
 - 14 **Lin PI**, Vance JM, Pericak-Vance MA, Martin ER. No gene is an island: the flip-flop phenomenon. *Am J Hum Genet* 2007; **80**: 531-538 [PMID: 17273975 DOI: 10.1086/512133]
 - 15 **Lauren P**. The two histological main types of gastric carcinoma: diffuse and so-called intestinal-type carcinoma. an attempt at a histo-clinical classification. *Acta Pathol Microbiol Scand* 1965; **64**: 31-49 [PMID: 14320675]
 - 16 **Nardone G**. Review article: molecular basis of gastric carcinogenesis. *Aliment Pharmacol Ther* 2003; **17** Suppl 2: 75-81 [PMID: 12786617]
 - 17 **Wu H**, Rusiecki JA, Zhu K, Potter J, Devesa SS. Stomach carcinoma incidence patterns in the United States by histologic type and anatomic site. *Cancer Epidemiol Biomarkers Prev* 2009; **18**: 1945-1952 [PMID: 19531677 DOI: 10.1158/1055-9965.epi-09-0250]
 - 18 **Blot WJ**, Devesa SS, Kneller RW, Fraumeni JF. Rising incidence of adenocarcinoma of the esophagus and gastric cardia. *JAMA* 1991; **265**: 1287-1289 [PMID: 1995976]
 - 19 **Crew KD**, Neugut AI. Epidemiology of gastric cancer. *World J Gastroenterol* 2006; **12**: 354-362 [PMID: 16489633]
 - 20 **Shah MA**, Khanin R, Tang L, Janjigian YY, Klimstra DS, Gerdes H, Kelsen DP. Molecular classification of gastric cancer: a new paradigm. *Clin Cancer Res* 2011; **17**: 2693-2701 [PMID: 21430069 DOI: 10.1158/1078-0432.ccr-10-2203]
 - 21 **Hansen S**, Vollset SE, Derakhshan MH, Fyfe V, Melby KK, Aase S, Jellum E, McColl KE. Two distinct aetiologies of cardia cancer; evidence from premorbid serological markers of gastric atrophy and Helicobacter pylori status. *Gut* 2007; **56**: 918-925 [PMID: 17317788 DOI: 10.1136/gut.2006.114504]
 - 22 **Hamilton JP**, Meltzer SJ. A review of the genomics of gastric cancer. *Clin Gastroenterol Hepatol* 2006; **4**: 416-425 [PMID: 16616344 DOI: 10.1016/j.cgh.2006.01.019]
 - 23 **Song HR**, Kim HN, Kweon SS, Choi JS, Shim HJ, Cho SH, Chung IJ, Park YK, Kim SH, Choi YD, Joo KW, Shin MH. Common genetic variants at 1q22 and 10q23 and gastric cancer susceptibility in a Korean population. *Tumour Biol* 2014; **35**: 3133-3137 [PMID: 24254309 DOI: 10.1007/s13277-013-1409-4]
 - 24 **Patterson D**, Jones C, Hart I, Bleskan J, Berger R, Geyer D, Eisenberg SP, Smith MF, Arend WP. The human interleukin-1 receptor antagonist (IL1RN) gene is located in the chromosome 2q14 region. *Genomics* 1993; **15**: 173-176 [PMID: 8432529 DOI: 10.1006/geno.1993.1025]
 - 25 **Smith DE**, Renshaw BR, Ketchem RR, Kubin M, Garka KE, Sims JE. Four new members expand the interleukin-1 superfamily. *J Biol Chem* 2000; **275**: 1169-1175 [PMID: 10625660]
 - 26 **Persson C**, Canedo P, Machado JC, El-Omar EM, Forman D. Polymorphisms in inflammatory response genes and their association with gastric cancer: A HuGE systematic review and meta-analyses. *Am J Epidemiol* 2011; **173**: 259-270 [PMID: 21178102 DOI: 10.1093/aje/kwq370]
 - 27 **Loh M**, Koh KX, Yeo BH, Song CM, Chia KS, Zhu F, Yeoh KG, Hill J, Iacopetta B, Soong R. Meta-analysis of genetic polymorphisms and gastric cancer risk: variability in associations according to race. *Eur J Cancer* 2009; **45**: 2562-2568 [PMID: 19375306 DOI: 10.1016/j.ejca.2009.03.017]
 - 28 **Xue H**, Lin B, Ni P, Xu H, Huang G. Interleukin-1B and interleukin-1 RN polymorphisms and gastric carcinoma risk: a meta-analysis. *J Gastroenterol Hepatol* 2010; **25**: 1604-1617 [PMID: 20880168 DOI: 10.1111/j.1440-1746.2010.06428.x]
 - 29 **Chang YW**, Jang JY, Kim NH, Lee JW, Lee HJ, Jung WW, Dong SH, Kim HJ, Kim BH, Lee JL, Chang R. Interleukin-1B (IL-1B) polymorphisms and gastric mucosal levels of IL-1beta cytokine in Korean patients with gastric cancer. *Int J Cancer* 2005; **114**: 465-471 [PMID: 15551344 DOI: 10.1002/ijc.20724]
 - 30 **Lee KA**, Ki CS, Kim HJ, Sohn KM, Kim JW, Kang WK, Rhee JC, Song SY, Sohn TS. Novel interleukin 1beta polymorphism increased the risk of gastric cancer in a Korean population. *J Gastroenterol* 2004; **39**: 429-433 [PMID: 15175940 DOI: 10.1007/s00535-003-1315-4]
 - 31 **Schmidt HM**, Ha DM, Taylor EF, Kovach Z, Goh KL, Fock KM, Barrett JH, Forman D, Mitchell H. Variation in human genetic polymorphisms, their association with Helicobacter pylori acquisition and gastric cancer in a multi-ethnic country. *J Gastroenterol Hepatol* 2011; **26**: 1725-1732 [PMID: 21649724 DOI: 10.1111/j.1440-1746.2011.06799.x]
 - 32 **Kupcinskas L**, Wex T, Kupcinskas J, Leja M, Ivanauskas A, Jonaitis LV, Janciauskas D, Kiudelis G, Funka K, Sudraba A, Chiu HM, Lin JT, Malfertheiner P. Interleukin-1B and interleukin-1 receptor antagonist gene polymorphisms are not associated with premalignant gastric conditions: a combined haplotype analysis. *Eur J Gastroenterol Hepatol* 2010; **22**: 1189-1195 [PMID: 20631624 DOI: 10.1097/MEG.0b013e32833cf3d5]
 - 33 **Zeng ZR**, Hu PJ, Hu S, Pang RP, Chen MH, Ng M, Sung JJ. Association of interleukin 1B gene polymorphism and gastric cancers in high and low prevalence regions in China. *Gut* 2003; **52**: 1684-1689 [PMID: 14633943]
 - 34 **Chen H**, Wilkins LM, Aziz N, Cannings C, Wyllie DH, Bingle C, Rogus J, Beck JD, Offenbacher S, Cork MJ, Rafie-Kolpin M, Hsieh CM, Kornman KS, Duff GW. Single nucleotide polymorphisms in the human interleukin-1B gene affect transcription according to haplotype context. *Hum Mol Genet* 2006; **15**: 519-529 [PMID: 16399797 DOI: 10.1093/hmg/ddi469]
 - 35 **Rogus J**, Beck JD, Offenbacher S, Huttner K, Iacoviello L,

- Latella MC, de Gaetano M, Wang HY, Kornman KS, Duff GW. IL1B gene promoter haplotype pairs predict clinical levels of interleukin-1beta and C-reactive protein. *Hum Genet* 2008; **123**: 387-398 [PMID: 18369665 DOI: 10.1007/s00439-008-0488-6]
- 36 **Chakravorty M**, Ghosh A, Choudhury A, Santra A, Hembrum J, Roychoudhury S. Interaction between IL1B gene promoter polymorphisms in determining susceptibility to Helicobacter pylori associated duodenal ulcer. *Hum Mutat* 2006; **27**: 411-419 [PMID: 16550552 DOI: 10.1002/humu.20299]
- 37 **Landvik NE**, Hart K, Skaug V, Stangeland LB, Haugen A, Zienolddiny S. A specific interleukin-1B haplotype correlates with high levels of IL1B mRNA in the lung and increased risk of non-small cell lung cancer. *Carcinogenesis* 2009; **30**: 1186-1192 [PMID: 19461122 DOI: 10.1093/carcin/bgp122]
- 38 **Landvik NE**, Hart K, Haugen A, Zienolddiny S. Functional analysis of a lung cancer risk haplotype in the IL1B gene regulatory region. *J Hum Genet* 2012; **57**: 747-752 [PMID: 22951596 DOI: 10.1038/jhg.2012.106]
- 39 **Chakravorty M**, Datta De D, Choudhury A, Roychoudhury S. IL1B promoter polymorphism regulates the expression of gastric acid stimulating hormone gastrin. *Int J Biochem Cell Biol* 2009; **41**: 1502-1510 [PMID: 19166966 DOI: 10.1016/j.biocel.2008.12.017]
- 40 **Tarlow JK**, Blakemore AI, Lennard A, Solari R, Hughes HN, Steinkasserer A, Duff GW. Polymorphism in human IL-1 receptor antagonist gene intron 2 is caused by variable numbers of an 86-bp tandem repeat. *Hum Genet* 1993; **91**: 403-404 [PMID: 8500797]
- 41 **Santtila S**, Savinainen K, Hurme M. Presence of the IL-1RA allele 2 (IL1RN*2) is associated with enhanced IL-1beta production in vitro. *Scand J Immunol* 1998; **47**: 195-198 [PMID: 9519856]
- 42 **Kamangar F**, Dawsey SM, Blaser MJ, Perez-Perez GI, Pietinen P, Newschaffer CJ, Abnet CC, Albanes D, Virtamo J, Taylor PR. Opposing risks of gastric cardia and noncardia gastric adenocarcinomas associated with Helicobacter pylori seropositivity. *J Natl Cancer Inst* 2006; **98**: 1445-1452 [PMID: 17047193 DOI: 10.1093/jnci/djj393]
- 43 **Aromaa A**, Kosunen TU, Knekt P, Maatela J, Teppo L, Heironen OP, Härkönen M, Hakama MK. Circulating anti-Helicobacter pylori immunoglobulin A antibodies and low serum pepsinogen I level are associated with increased risk of gastric cancer. *Am J Epidemiol* 1996; **144**: 142-149 [PMID: 8678045]
- 44 **Ye W**, Held M, Lagergren J, Engstrand L, Blot WJ, McLaughlin JK, Nyrén O. Helicobacter pylori infection and gastric atrophy: risk of adenocarcinoma and squamous-cell carcinoma of the esophagus and adenocarcinoma of the gastric cardia. *J Natl Cancer Inst* 2004; **96**: 388-396 [PMID: 14996860]
- 45 **Hansen S**, Melby KK, Aase S, Jellum E, Vollset SE. Helicobacter pylori infection and risk of cardia cancer and non-cardia gastric cancer. A nested case-control study. *Scand J Gastroenterol* 1999; **34**: 353-360 [PMID: 10365894]
- 46 **de Martel C**, Llosa AE, Farr SM, Friedman GD, Vogelstein JH, Orentreich N, Corley DA, Parsonnet J. Helicobacter pylori infection and the risk of development of esophageal adenocarcinoma. *J Infect Dis* 2005; **191**: 761-767 [PMID: 15688293 DOI: 10.1086/427659]
- 47 **Chow WH**, Blaser MJ, Blot WJ, Gammon MD, Vaughan TL, Risch HA, Perez-Perez GI, Schoenberg JB, Stanford JL, Rotterdam H, West AB, Fraumeni JF. An inverse relation between cagA+ strains of Helicobacter pylori infection and risk of esophageal and gastric cardia adenocarcinoma. *Cancer Res* 1998; **58**: 588-590 [PMID: 9485003]
- 48 **Richter JE**, Falk GW, Vaezi MF. Helicobacter pylori and gastroesophageal reflux disease: the bug may not be all bad. *Am J Gastroenterol* 1998; **93**: 1800-1802 [PMID: 9772034 DOI: 10.1111/j.1572-0241.1998.00523.x]
- 49 **Derakhshan MH**, Malekzadeh R, Watabe H, Yazdanbod A, Fyfe V, Kazemi A, Rakhshani N, Didevar R, Sotoudeh M, Zolfeghari AA, McColl KE. Combination of gastric atrophy, reflux symptoms and histological subtype indicates two distinct aetiologies of gastric cardia cancer. *Gut* 2008; **57**: 298-305 [PMID: 17965056 DOI: 10.1136/gut.2007.137364]
- 50 **Bidwell J**, Keen L, Gallagher G, Kimberly R, Huizinga T, McDermott MF, Oksenberg J, McNicholl J, Pociot F, Hardt C, D'Alfonso S. Cytokine gene polymorphism in human disease: on-line databases. *Genes Immun* 1999; **1**: 3-19 [PMID: 11197303 DOI: 10.1038/sj.gene.6363645]
- 51 **Hurme M**, Lahdenpohja N, Santtila S. Gene polymorphisms of interleukins 1 and 10 in infectious and autoimmune diseases. *Ann Med* 1998; **30**: 469-473 [PMID: 9814833]
- 52 **Eskdale J**, Keijsers V, Huizinga T, Gallagher G. Microsatellite alleles and single nucleotide polymorphisms (SNP) combine to form four major haplotype families at the human interleukin-10 (IL-10) locus. *Genes Immun* 1999; **1**: 151-155 [PMID: 11196662 DOI: 10.1038/sj.gene.6363656]
- 53 **El-Omar EM**, Rabkin CS, Gammon MD, Vaughan TL, Risch HA, Schoenberg JB, Stanford JL, Mayne ST, Goedert J, Blot WJ, Fraumeni JF, Chow WH. Increased risk of noncardia gastric cancer associated with proinflammatory cytokine gene polymorphisms. *Gastroenterology* 2003; **124**: 1193-1201 [PMID: 12730860]
- 54 **Gammon MD**, Schoenberg JB, Ahsan H, Risch HA, Vaughan TL, Chow WH, Rotterdam H, West AB, Dubrow R, Stanford JL, Mayne ST, Farrow DC, Niwa S, Blot WJ, Fraumeni JF. Tobacco, alcohol, and socioeconomic status and adenocarcinomas of the esophagus and gastric cardia. *J Natl Cancer Inst* 1997; **89**: 1277-1284 [PMID: 9293918]
- 55 **Won HH**, Kim JW, Kim MJ, Kim S, Park JH, Lee KA. Interleukin 10 polymorphisms differentially influence the risk of gastric cancer in East Asians and Caucasians. *Cytokine* 2010; **51**: 73-77 [PMID: 20363151 DOI: 10.1016/j.cyto.2010.03.007]
- 56 **Ni P**, Xu H, Xue H, Lin B, Lu Y. A meta-analysis of interleukin-10-1082 promoter polymorphism associated with gastric cancer risk. *DNA Cell Biol* 2012; **31**: 582-591 [PMID: 22335769 DOI: 10.1089/dna.2011.1440]
- 57 **Zhou Y**, Li N, Zhuang W, Liu GJ, Wu TX, Yao X, Du L, Wei ML, Wu XT. Interleukin-10 -1082 promoter polymorphism associated with gastric cancer among Asians. *Eur J Cancer* 2008; **44**: 2648-2654 [PMID: 18707865 DOI: 10.1016/j.ejca.2008.07.017]
- 58 **Yu Z**, Liu Q, Huang C, Wu M, Li G. The interleukin 10 -819C/T polymorphism and cancer risk: a HuGE review and meta-analysis of 73 studies including 15,942 cases and 22,336 controls. *OMICS* 2013; **17**: 200-214 [PMID: 23574339 DOI: 10.1089/omi.2012.0089]
- 59 **Chen KF**, Li B, Wei YG, Peng CJ. Interleukin-10 -819 promoter polymorphism associated with gastric cancer among Asians. *J Int Med Res* 2010; **38**: 1-8 [PMID: 20233508]
- 60 **Xue H**, Lin B, An J, Zhu Y, Huang G. Interleukin-10-819 promoter polymorphism in association with gastric cancer risk. *BMC Cancer* 2012; **12**: 102 [PMID: 22436502 DOI: 10.1186/1471-2407-12-102]
- 61 **Zhu Y**, Wang J, He Q, Zhang JQ. The association between interleukin-10-592 polymorphism and gastric cancer risk: a meta-analysis. *Med Oncol* 2011; **28**: 133-136 [PMID: 20087693 DOI: 10.1007/s12032-010-9417-3]
- 62 **Ferrer-Admetlla A**, Bosch E, Sikora M, Marqués-Bonet T, Ramírez-Soriano A, Muntasell A, Navarro A, Lazarus R, Calafell F, Bertranpetit J, Casals F. Balancing selection is the main force shaping the evolution of innate immunity genes. *J Immunol* 2008; **181**: 1315-1322 [PMID: 18606686]
- 63 **Wilson JN**, Rockett K, Keating B, Jallow M, Pinder M, Sisay-Joof F, Newport M, Kwiatkowski D. A hallmark of balancing selection is present at the promoter region of interleukin 10. *Genes Immun* 2006; **7**: 680-683 [PMID: 16943796 DOI: 10.1038/sj.gene.6364336]
- 64 **Lundin BS**, Enarsson K, Kindlund B, Lundgren A, Johnsson

- E, Quiding-Järbrink M, Svennerholm AM. The local and systemic T-cell response to *Helicobacter pylori* in gastric cancer patients is characterised by production of interleukin-10. *Clin Immunol* 2007; **125**: 205-213 [PMID: 17826353 DOI: 10.1016/j.clim.2007.07.011]
- 65 **Karnes WE**, Samloff IM, Siurala M, Kekki M, Sipponen P, Kim SW, Walsh JH. Positive serum antibody and negative tissue staining for *Helicobacter pylori* in subjects with atrophic body gastritis. *Gastroenterology* 1991; **101**: 167-174 [PMID: 2044906]
- 66 **Wu MS**, Wu CY, Chen CJ, Lin MT, Shun CT, Lin JT. Interleukin-10 genotypes associate with the risk of gastric carcinoma in Taiwanese Chinese. *Int J Cancer* 2003; **104**: 617-623 [PMID: 12594817 DOI: 10.1002/ijc.10987]
- 67 **Rad R**, Dossumbekova A, Neu B, Lang R, Bauer S, Saur D, Gerhard M, Prinz C. Cytokine gene polymorphisms influence mucosal cytokine expression, gastric inflammation, and host specific colonisation during *Helicobacter pylori* infection. *Gut* 2004; **53**: 1082-1089 [PMID: 15247172 DOI: 10.1136/gut.2003.029736]
- 68 **Tan IB**, Ng I, Tai WM, Tan P. Understanding the genetic basis of gastric cancer: recent advances. *Expert Rev Gastroenterol Hepatol* 2012; **6**: 335-341 [PMID: 22646255 DOI: 10.1586/egh.12.7]
- 69 **Saeki N**, Ono H, Sakamoto H, Yoshida T. Genetic factors related to gastric cancer susceptibility identified using a genome-wide association study. *Cancer Sci* 2013; **104**: 1-8 [PMID: 23057512 DOI: 10.1111/cas.12042]
- 70 **Sakamoto H**, Yoshimura K, Saeki N, Katai H, Shimoda T, Matsuno Y, Saito D, Sugimura H, Tanioka F, Kato S, Matsukura N, Matsuda N, Nakamura T, Hyodo I, Nishina T, Yasui W, Hirose H, Hayashi M, Toshiro E, Ohnami S, Sekine A, Sato Y, Totsuka H, Ando M, Takemura R, Takahashi Y, Ohdaira M, Aoki K, Honmyo I, Chiku S, Aoyagi K, Sasaki H, Ohnami S, Yanagihara K, Yoon KA, Kook MC, Lee YS, Park SR, Kim CG, Choi IJ, Yoshida T, Nakamura Y, Hirohashi S. Genetic variation in PSCA is associated with susceptibility to diffuse-type gastric cancer. *Nat Genet* 2008; **40**: 730-740 [PMID: 18488030 DOI: 10.1038/ng.152]
- 71 **Saeki N**, Saito A, Choi IJ, Matsuo K, Ohnami S, Totsuka H, Chiku S, Kuchiba A, Lee YS, Yoon KA, Kook MC, Park SR, Kim YW, Tanaka H, Tajima K, Hirose H, Tanioka F, Matsuno Y, Sugimura H, Kato S, Nakamura T, Nishina T, Yasui W, Aoyagi K, Sasaki H, Yanagihara K, Katai H, Shimoda T, Yoshida T, Nakamura Y, Hirohashi S, Sakamoto H. A functional single nucleotide polymorphism in mucin 1, at chromosome 1q22, determines susceptibility to diffuse-type gastric cancer. *Gastroenterology* 2011; **140**: 892-902 [PMID: 21070779 DOI: 10.1053/j.gastro.2010.10.058]
- 72 **Kim J**, Kim JW, Kim Y, Lee KA. Differential association of RANTES-403 and IL-1B-1464 polymorphisms on histological subtypes in male Korean patients with gastric cancer. *Tumour Biol* 2014; **35**: 3765-3770 [PMID: 24323564 DOI: 10.1007/s13277-013-1498-0]
- 73 **Du H**, Bay BH, Mahendran R, Olivo M. Endogenous expression of interleukin-8 and interleukin-10 in nasopharyngeal carcinoma cells and the effect of photodynamic therapy. *Int J Mol Med* 2002; **10**: 73-76 [PMID: 12060853]
- 74 **Bendre MS**, Gaddy-Kurten D, Mon-Foote T, Akel NS, Skinner RA, Nicholas RW, Suva LJ. Expression of interleukin 8 and not parathyroid hormone-related protein by human breast cancer cells correlates with bone metastasis in vivo. *Cancer Res* 2002; **62**: 5571-5579 [PMID: 12359770]
- 75 **Koch AE**, Polverini PJ, Kunkel SL, Harlow LA, DiPietro LA, Elner VM, Elner SG, Strieter RM. Interleukin-8 as a macrophage-derived mediator of angiogenesis. *Science* 1992; **258**: 1798-1801 [PMID: 1281554]
- 76 **Li A**, Dubey S, Varney ML, Dave BJ, Singh RK. IL-8 directly enhanced endothelial cell survival, proliferation, and matrix metalloproteinases production and regulated angiogenesis. *J Immunol* 2003; **170**: 3369-3376 [PMID: 12626597]
- 77 **Wang N**, Zhou R, Wang C, Guo X, Chen Z, Yang S, Li Y. -251 T/A polymorphism of the interleukin-8 gene and cancer risk: a HuGE review and meta-analysis based on 42 case-control studies. *Mol Biol Rep* 2012; **39**: 2831-2841 [PMID: 21681427 DOI: 10.1007/s11033-011-1042-5]
- 78 **Canedo P**, Castanheira-Vale AJ, Lunet N, Pereira F, Figueiredo C, Gioia-Patricola L, Canzian F, Moreira H, Suriano G, Barros H, Carneiro F, Seruca R, Machado JC. The interleukin-8-251T/*A polymorphism is not associated with risk for gastric carcinoma development in a Portuguese population. *Eur J Cancer Prev* 2008; **17**: 28-32 [PMID: 18090907 DOI: 10.1097/CEJ.0b013e32809b4d0f]
- 79 **Savage SA**, Abnet CC, Mark SD, Qiao YL, Dong ZW, Dawsey SM, Taylor PR, Chanock SJ. Variants of the IL8 and IL8RB genes and risk for gastric cardia adenocarcinoma and esophageal squamous cell carcinoma. *Cancer Epidemiol Biomarkers Prev* 2004; **13**: 2251-2257 [PMID: 15598788]
- 80 **Hull J**, Ackerman H, Isles K, Usen S, Pinder M, Thomson A, Kwiatkowski D. Unusual haplotypic structure of IL8, a susceptibility locus for a common respiratory virus. *Am J Hum Genet* 2001; **69**: 413-419 [PMID: 11431705 DOI: 10.1086/321291]
- 81 **Shi Y**, Hu Z, Wu C, Dai J, Li H, Dong J, Wang M, Miao X, Zhou Y, Lu F, Zhang H, Hu L, Jiang Y, Li Z, Chu M, Ma H, Chen J, Jin G, Tan W, Wu T, Zhang Z, Lin D, Shen H. A genome-wide association study identifies new susceptibility loci for non-cardia gastric cancer at 3q13.31 and 5p13.1. *Nat Genet* 2011; **43**: 1215-1218 [PMID: 22037551 DOI: 10.1038/ng.978]

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