

## Original Article

# $\beta_3$ -Adrenoceptor activation attenuates atherosclerotic plaque formation in ApoE<sup>-/-</sup> mice through lowering blood lipids and glucose

Zhao-hong WANG<sup>1,2</sup>, Yan-fang LI<sup>1,2,\*</sup>, Yan-qing GUO<sup>1,2</sup><sup>1</sup>Department of Cardiology, Anzhen Hospital, Capital University of Medical Sciences, Beijing 100029, China; <sup>2</sup>Beijing Institute of Heart, Lung and Blood Vessel Diseases, Beijing 100029, China**Aim:** To examine the effects of  $\beta_3$ -adrenoceptor ( $\beta_3$ -AR) activation on atherosclerotic plaque development in ApoE<sup>-/-</sup> mice.**Methods:** Thirty six week-old male ApoE<sup>-/-</sup> mice on a high-fat diet were treated with atorvastatin (10 mg/kg<sup>-1</sup>d<sup>-1</sup>, po), BRL37344 ( $\beta_3$ -AR agonist, 1.65 or 3.30  $\mu$ g/kg, ip, twice a week) or SR52390A ( $\beta_3$ -AR antagonist, 50  $\mu$ g/kg, ip, twice a week) for 12 weeks. Wild-type C57BL/6J mice receiving a normal diet were taken as healthy controls. At the end of the treatments, serum levels of triglycerides (TG), total cholesterol (TC), high-density lipoprotein cholesterol (HDL-C), non-high density lipoprotein cholesterol (nHDL-C), glucose and insulin were measured. The thoracic aortas were dissected out, the area of atherosclerotic plaques and extent of fibrosis in the plaques were examined using HE and Masson's trichome staining, respectively.**Results:** Compared to wild-type mice, ApoE<sup>-/-</sup> mice fed on a high-fat diet exhibited prominent hyperlipidemia and insulin resistance, associated with large area of atherosclerotic plaques and great extent of fibrosis in aortas. Atorvastatin significantly decreased the serum levels of TC and nHDL-C, and reduced the plaque area and collagen content in aortas. BRL37344 significantly decreased the serum levels of TG, TC, nHDL-C, glucose and insulin, and increased HDL-C and the insulin sensitivity, and dose-dependently reduced the plaque area and collagen content in aortas. SR52390A treatment did not affect any parameters studied.**Conclusion:** The  $\beta_3$ -AR agonist impedes the progression of atherosclerosis in ApoE<sup>-/-</sup> mice, through improvement of the lipid and glucose profiles.**Keywords:** atherosclerosis; apolipoprotein E; ApoE<sup>-/-</sup> mice;  $\beta_3$ -adrenoceptor; BRL37344; SR52390A; atorvastatin; hyperlipidemia; glucose; insulin

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## Introduction

$\beta$  receptors were classified as  $\beta_1$ - and  $\beta_2$ -adrenoceptors ( $\beta$ -ARs) until the early 1980s, when a third member of the  $\beta$ -ARs was cloned from adipose tissue<sup>[1]</sup>.  $\beta_3$ -AR activation leads to lipolysis in white adipose tissue and thermogenesis in brown adipose tissue<sup>[2]</sup>.  $\beta_3$ -AR is a G protein-coupled receptor that activates adenylyl cyclase, cAMP/protein kinase A (PKA) and the PI3-kinase pathway<sup>[3]</sup>.

In the *ob/ob* mouse model for obesity,  $\beta_3$ -AR expression and function in white and brown adipose tissues are markedly reduced<sup>[4]</sup>.  $\beta_3$ -AR agonists could improve glycemic control and insulin sensitivity in addition to reducing plasma triglycerides (TG), free fatty acids and body weight in obese diabetic animals<sup>[5–7]</sup>.

Abnormal lipid and glucose metabolism plays a major role in the pathogenesis of atherosclerosis<sup>[8]</sup>. Because activation of  $\beta_3$ -AR improves lipid and glucose metabolism in obese and diabetic rodent models<sup>[5–7]</sup>, we propose that chronic  $\beta_3$ -AR activation could attenuate atherosclerosis. The current study examined the potential effects of a representative  $\beta_3$ -AR agonist and antagonist on the formation and fibrosis of atherosclerotic plaques in ApoE-deficient (ApoE<sup>-/-</sup>) mice. The concentrations of plasma lipids and glucose were also monitored in addition to insulin sensitivity.

## Materials and methods

### Reagents

BRL37344 (sodium-4-[-2[-2-hydroxy-2-(3-chloro-phenyl) ethylamine]propyl] phenoxyacetate) and SR52390A (3-(2-ethylphenoxy)-1-[(1S)-1,2,3,4-tetrahydronaph-1-ylamino]-2S-2-propanol oxalate) were purchased from Sigma (St Louis, MO, USA). Atorvastatin was obtained from Pfizer (Dalian, China).

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An ultra-sensitive rat insulin enzyme-linked immunosorbent assay (ELISA) kit was purchased from Crystal Chem (Downers Grove, PA, USA). TRIzol reagent was purchased from Invitrogen (Carlsbad, CA, USA). The reverse transcription-polymerase chain reaction (RT-PCR) kit (A3500) was obtained from Promega (Madison, WI, USA). All primers were synthesized by Invitrogen (Shanghai, China). The SYBR Green real-time PCR kit was purchased from Katara Bio (Shiga, Nagano, Japan).

#### Mice

Homozygous ApoE<sup>-/-</sup><sup>[9]</sup> and C57BL/6J mice were provided by Beijing Vital River Laboratory Animal Technology (Certificate No: SCXK 2006-0008). The mice were housed in a specific pathogen-free facility under a 12/12 h light-dark cycle at 23 °C and were used after at least one week of adaptation. ApoE<sup>-/-</sup> mice were given a high-fat diet containing 0.15% (*w/w*) cholesterol and 21% (*w/w*) fat starting from 10 weeks of age. C57BL/6J mice received normal chow throughout the experiment. All mice had unrestricted access to tap water. Animal care and use were carried out in accordance with the Chinese standards (GB14925-2001).

#### Experimental protocols

Starting from 36 weeks of age, ApoE<sup>-/-</sup> mice randomly received atorvastatin (10 mg/kg by daily gavage), the β<sub>3</sub>-AR agonist BRL37344 [1.65 or 3.30 μg/kg by intraperitoneal (ip) injection, twice a week], the β<sub>3</sub>-AR antagonist SR52390A (50 μg/kg by ip injection, twice a week), or a saline vehicle of the same volume (ip injection, twice a week) for 12 weeks (*n*=10 per treatment condition). Wild-type (WT) C57BL/6J mice that received a normal diet and were treated with vehicle were included as healthy controls (*n*=10). Body weight was monitored at 10, 20, 30, 36, 42 and 48 weeks of age.

#### Serum lipids, glucose and insulin

Blood samples were collected 6 h after fasting using a retro-orbital approach. Serum glucose, TG, total cholesterol (TC) and high-density lipoprotein cholesterol (HDL-C) were examined with a CX7 analyzer (Beckman Coulter, Fullerton, CA, USA). Non-high density lipoprotein cholesterol (nHDL-C) was determined using Friedewald's formula<sup>[10, 11]</sup>. The serum insulin concentration was determined by ELISA.

#### Morphometric analysis of atherosclerotic plaques

A section of the thoracic aorta between the distal end of the aortic arch and the diaphragm was dissected after anesthetization with an intraperitoneal injection of sodium pentobarbital (50 mg/kg). Tissue samples were immersed in 10% formaldehyde, dehydrated and embedded in optimum cutting temperature compound (OCT). Cross-sections (6 μm) were prepared every 100 μm. Eight sections were prepared from each animal subject for hematoxylin and eosin (H&E) or Masson's trichrome staining<sup>[12]</sup>. Images were captured using a light microscope (DP 70, Olympus, Tokyo, Japan) and analyzed using an image analysis program (Image-Pro Plus 3.0) by an

observer who was blinded to the treatment condition. The assessment of the lumen and lesion areas was performed using a method that was described by Paigen and colleagues<sup>[13]</sup>. The extent of the fibrosis is presented as the percent area stained divided by the total plaque area.

#### Transmission electron microscopy

The midsection of the thoracic aorta was fixed in 2.5% glutaraldehyde followed by osmium tetroxide treatment, dehydrated in a graded ethanol series and embedded for cross-sectioning to identify monocytes that adhered to the endothelial wall. Smooth muscle cells (SMCs) and elastic fiber layers were observed with a transmission electron microscope. Two animals from each group were examined (12 tissue samples×5 sections for a total of 60 sections).

#### Quantitative real-time PCR

RNA extraction, reverse transcription-PCR and quantitative real-time PCR of β<sub>3</sub>-AR mRNA in epididymal white adipose tissue were performed as previously described<sup>[14]</sup> using a SYBR Green real-time PCR kit. The PCR primer sequences were as follows: forward 5'-TCGACATGTTCTCCACAAA-3' and reverse 5'-GATGGTCCAAGATGGTGCTT-3' (NCBI reference sequence: NM\_013462.3). The reaction was performed as follows: 40 cycles of denaturation at 94 °C for 30 s, annealing at 60 °C for 30 s and extension at 72 °C for 30 s. GAPDH mRNA was used as a control (NCBI reference sequence: NM\_008084.2) and amplified with the following primers: forward 5'-ACCCAGAAGACTGTGGATGG-3' and reverse 5'-ACATTGGGGGTAGGAACAC-3'. The results were normalized against GAPDH and expressed as fold changes using the 2<sup>-ΔΔCT</sup> method<sup>[15]</sup>. The experiment was performed in triplicate.

#### Body fat

Whole body fat was measured in conscious mice using a quantitative magnetic resonance method (Echo MRI 3-in-1; Echo Medical Systems, Houston, TX, USA)<sup>[16]</sup>.

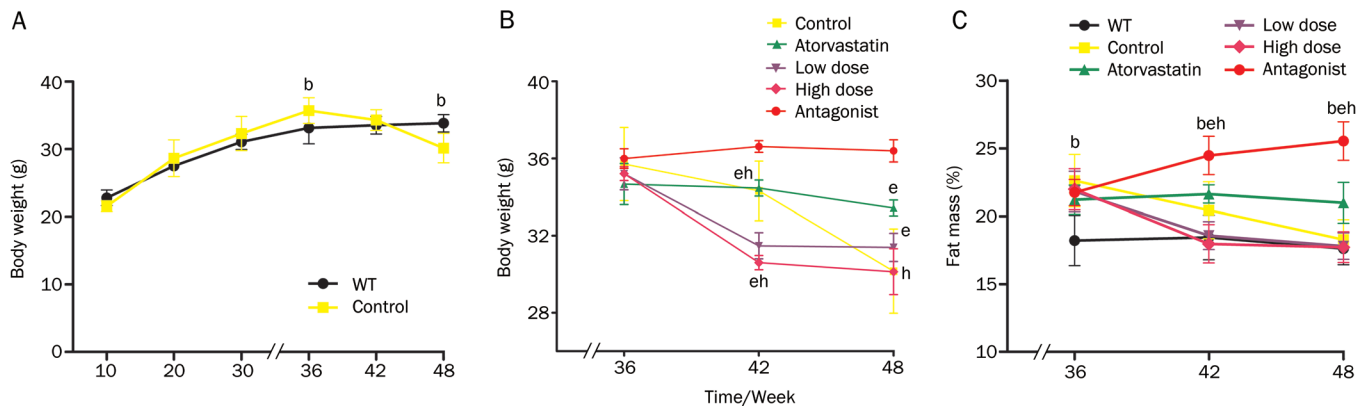
#### Statistical analysis

The results are expressed as the mean±SD. The data were evaluated by one-way analysis of variance (ANOVA) followed by the Bonferroni *t*-test (equal variances) or Dunn's multiple comparison test (unequal variances) for pair-wise comparisons. The statistical significance was set at *P*<0.05.

## Results

#### Body weight and fat

Across the treatment conditions, the mice had similar body weights prior to the experiment at 10 weeks of age. Immediately prior to the drug treatment (36 weeks of age), ApoE<sup>-/-</sup> mice receiving vehicle were significantly heavier than the WT control mice (*P*=0.038). At the completion of the experiment (48 weeks of age), ApoE<sup>-/-</sup> mice receiving vehicle were significantly lighter than the WT controls (*P*=0.01) (Figure 1A). Atorvastatin treatment increased the body weight



**Figure 1.** Body weight and fat mass at different ages. (A) Body weights in the ApoE<sup>-/-</sup> control and WT groups were measured from 10 to 48 weeks of age. (B) The change in body weight with different interventions from 36 to 48 weeks of age (*n*=10 per group). (C) The fat mass was measured in ApoE<sup>-/-</sup> and WT mice at 36, 42, and 48 weeks of age (*n*=6 per group). Mean±SD. <sup>b</sup>*P*<0.05 compared with WT mice. <sup>e</sup>*P*<0.05 compared with ApoE<sup>-/-</sup> control mice. <sup>h</sup>*P*<0.05 compared with the atorvastatin treatment group.

(*P*=0.032 vs ApoE<sup>-/-</sup> vehicle control at 48 weeks of age) (Figure 1B), whereas the β<sub>3</sub>-AR agonist decreased the body weight (*P*<0.001 vs ApoE<sup>-/-</sup> vehicle control at 42 weeks of age). However, at a later time point (48 weeks), the weights of ApoE<sup>-/-</sup> mice receiving BRL37344 were similar to those of the ApoE<sup>-/-</sup> control mice. The β<sub>3</sub>-AR antagonist increased the body weight of mice at 42 and 48 weeks of age (*P*<0.01 vs ApoE<sup>-/-</sup> vehicle control).

ApoE<sup>-/-</sup> mice initially had a higher body fat content (*P*<0.001 vs WT control at 36 weeks of age). During the next 12 weeks, however, the fat mass of ApoE<sup>-/-</sup> mice gradually declined (Figure 1C). Atorvastatin did not affect the fat mass of ApoE<sup>-/-</sup> mice (*P*=0.019). The β<sub>3</sub>-AR agonist decreased the fat mass (*P*<0.01 vs ApoE<sup>-/-</sup> vehicle control), and the β<sub>3</sub>-AR antagonist increased the fat mass (*P*<0.01 vs the ApoE<sup>-/-</sup> vehicle control).

### Lipid analysis

ApoE<sup>-/-</sup> mice developed severe hypercholesterolemia and hypertriglyceridemia, showing a 27.7-fold increase in the very low-density lipoprotein/low-density lipoprotein cholesterol (VLDL/LDL-C) ratio and a 62% reduction in the HDL-C/TC ratio compared with WT controls of the same age (Table 1). Atorvastatin reduced serum TC by 45.5% and the VLDL/LDL-C ratio by 46.9% but did not alter HDL cholesterol or TG. At 1.65 and 3.30 μg/kg, the β<sub>3</sub>-AR agonist decreased plasma TG by 41% and 51% (vs ApoE<sup>-/-</sup> mice receiving vehicle), TC

by 18% and 23%, and VLDL/LDL-C by 26% and 34%, respectively; additionally, the plasma HDL-C was increased by 30% and 52%, and the HDL-C/TC ratio was increased by 9.7% and 115.7%, respectively. A posthoc analysis indicated that the higher dose of 3.30 μg/kg had a stronger effect (*P*<0.05 vs 1.65 μg/kg). The β<sub>3</sub>-AR antagonist did not affect serum lipids (*P*>0.05 for all) (Table 1).

### Serum glucose and insulin

ApoE<sup>-/-</sup> mice given a high-fat diet displayed a significant increase in serum glucose and insulin. BRL37344 decreased serum glucose and insulin (*P*<0.01 vs ApoE<sup>-/-</sup> vehicle control) (Table 2). BRL37344 also increased the insulin sensitivity index (ISI=Ln[1/(Ins×Glu)]); however, the effects were not dose-dependent. At both doses, BRL37344 reduced glucose, insulin and the ISI to levels that were even lower than those achieved with atorvastatin (*P*<0.01). The β<sub>3</sub>-AR antagonist SR52390A did not affect serum glucose or insulin levels (*P*>0.05 vs ApoE<sup>-/-</sup> mice receiving vehicle).

### Effects on atherosclerotic lesions

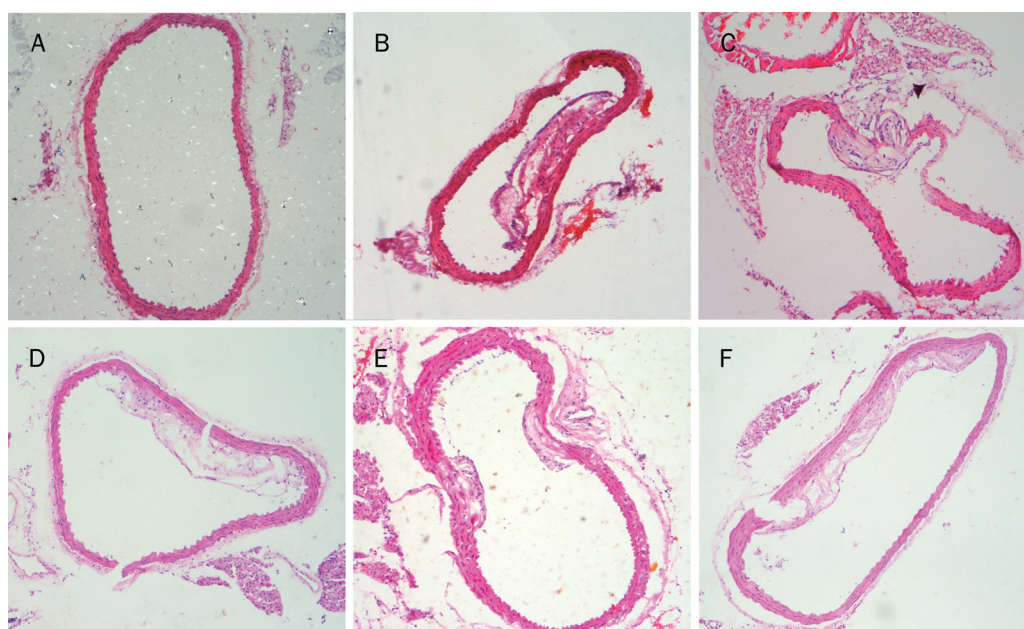
At 48 weeks of age, apparent atherosclerotic lesions were observed in the ApoE<sup>-/-</sup> mice but not in the WT controls (Figure 2A and 2B). Atorvastatin markedly decreased the atherosclerotic plaque area and increased the lumen area (*P*<0.01 vs ApoE<sup>-/-</sup> vehicle control) (Figure 2C). BRL37344 dose-

**Table 1.** Lipid profile of WT and ApoE<sup>-/-</sup> mice. Results are expressed as the mean±SD. <sup>c</sup>*P*<0.01 vs WT mice. <sup>f</sup>*P*<0.01 vs ApoE<sup>-/-</sup> control mice. <sup>l</sup>*P*<0.01 vs atorvastatin treatment group. <sup>h</sup>*P*<0.01 vs low-dose β<sub>3</sub>-AR agonist treatment group.

|                     | <i>n</i> | WT        | Control                 | Atorvastatin             | Low dose agonist          | High dose agonist         | Antagonist               |
|---------------------|----------|-----------|-------------------------|--------------------------|---------------------------|---------------------------|--------------------------|
| TC (mmol/L)         | 10       | 2.38±0.34 | 22.25±2.43 <sup>c</sup> | 12.09±1.26 <sup>cf</sup> | 17.80±0.73 <sup>fl</sup>  | 15.57±0.67 <sup>cfi</sup> | 20.94±1.89 <sup>ci</sup> |
| TG (mmol/L)         | 10       | 0.62±0.12 | 3.13±0.21 <sup>c</sup>  | 2.91±0.25 <sup>c</sup>   | 1.81±0.24 <sup>cfi</sup>  | 1.55±0.15 <sup>cfi</sup>  | 2.95±0.24 <sup>c</sup>   |
| HDL-C (mmol/L)      | 10       | 1.68±0.29 | 2.67±0.36 <sup>c</sup>  | 2.08±0.26 <sup>f</sup>   | 3.46±0.30 <sup>cfi</sup>  | 4.05±0.47 <sup>cfi</sup>  | 2.60±0.47 <sup>ci</sup>  |
| VLDL/LDL-C (mmol/L) | 10       | 0.67±0.16 | 19.02±2.37 <sup>c</sup> | 9.76±1.25 <sup>cf</sup>  | 14.11±0.57 <sup>cfi</sup> | 11.50±0.84 <sup>cfi</sup> | 18.28±1.54 <sup>ci</sup> |
| HDL-C/TC (%)        | 10       | 70.8±6.7  | 12.1±2.1 <sup>c</sup>   | 17.5±3.9 <sup>cf</sup>   | 19.4±1.6 <sup>cf</sup>    | 26.1±3.2 <sup>cfi</sup>   | 12.4±2.2 <sup>c</sup>    |

**Table 2.** Glucose and insulin profiles of WT and ApoE<sup>-/-</sup> mice. Results are expressed as the mean±SD. ISI: insulin sensitive index (ISI)=Ln[1/(Ins×Glu)].  
°P<0.01 vs WT mice. <sup>f</sup>P<0.01 vs ApoE<sup>-/-</sup> control mice. <sup>l</sup>P<0.01 vs atorvastatin treatment group.

|             | n  | WT         | Control                 | Atorvastatin             | Low dose agonist          | High dose agonist         | Antagonist              |
|-------------|----|------------|-------------------------|--------------------------|---------------------------|---------------------------|-------------------------|
| Glu (mg/dL) | 10 | 136.9±10.1 | 269.0±10.4 <sup>c</sup> | 251.4±12.2 <sup>cf</sup> | 198.9±6.45 <sup>cfi</sup> | 200.4±10.4 <sup>cfi</sup> | 270.9±8.53 <sup>c</sup> |
| Ins (ng/mL) | 10 | 1.23±0.09  | 6.99±0.53 <sup>c</sup>  | 6.98±0.32 <sup>c</sup>   | 4.55±0.29 <sup>cfi</sup>  | 4.60±0.25 <sup>cfi</sup>  | 7.11±0.35 <sup>c</sup>  |
| ISI         | 10 | -5.12±0.05 | -7.53±0.11 <sup>c</sup> | -7.47±0.07 <sup>c</sup>  | -6.80±0.09 <sup>cfi</sup> | -6.82±0.10 <sup>cfi</sup> | -7.56±0.06 <sup>c</sup> |



**Figure 2.** Morphology of the atherosclerotic lesions in WT and ApoE<sup>-/-</sup> mice. Cryosections of the thoracic aorta area were obtained from WT mice (A), ApoE<sup>-/-</sup> control mice (B), atorvastatin-treated mice (C), low-dose BRL37344-treated mice (D), high-dose BRL37344-treated mice (E) and SR52390A-treated mice (F) prior to staining with hematoxylin and eosin (H&E). Scale bars=200 μm.

independently decreased the atherosclerotic area to a level comparable to that achieved with atorvastatin (Figure 2D and 2E). SR52390A treatment did not produce any effects (Figure 2F) (Table 3).

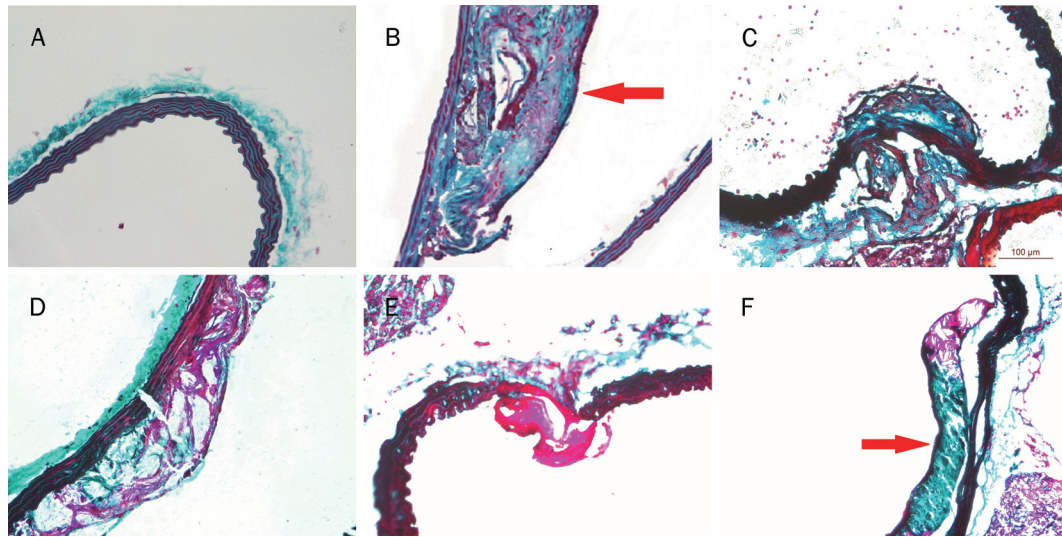
The ApoE<sup>-/-</sup> control mice displayed prominent fibrous caps on their atherosclerotic lesions (Figure 3B). Fibrous caps were not apparent in mice receiving atorvastatin or BRL37344 at either dose (Figure 3C–3E; *P*<0.01 for all). SR52390A had no effect (Figure 3F) (Table 3).

Transmission electron microscopy revealed normal SMCs

in the WT control mice (Figure 4A). The ApoE<sup>-/-</sup> control mice had fewer swollen SMCs with rarefaction of the nuclear chromosome; obscured, swollen mitochondria; lack of cristae; and fiber hyperplasia (Figure 4B). Atorvastatin decreased these pathological changes (Figure 4C). In mice receiving BRL37344, there were fewer normal SMCs and a large number of swollen SMCs with mild fiber hyperplasia (regardless of dose) (Figure 4D and 4E). In mice receiving SR52390A, there were also fewer swollen SMCs (Figure 4F).

**Table 3.** Atherosclerotic lesions of WT and ApoE<sup>-/-</sup> mice. Results are expressed as the mean±SD. LA, lumen area; PA, plaque area; CA, collagen area.  
°P<0.01 vs WT mice. <sup>f</sup>P<0.01 vs ApoE<sup>-/-</sup> control mice. <sup>l</sup>P<0.01 vs atorvastatin treatment group. <sup>i</sup>P<0.01 vs low-dose β<sub>3</sub>-AR agonist treatment group.

|  | n | WT         | Control                 | Atorvastatin             | Low dose agonist          | High dose agonist         | Antagonist              |
|--|---|------------|-------------------------|--------------------------|---------------------------|---------------------------|-------------------------|
| LA (×10 <sup>4</sup> μm <sup>2</sup> ) | 6 | 60.83±2.44 | 42.02±2.30 <sup>c</sup> | 50.36±2.42 <sup>cf</sup> | 46.67±2.39 <sup>cfi</sup> | 50.27±1.45 <sup>cfi</sup> | 41.53±2.30 <sup>c</sup> |
| PA (×10 <sup>4</sup> μm <sup>2</sup> ) | 6 | 0          | 21.26±2.47 <sup>c</sup> | 12.39±1.40 <sup>cf</sup> | 16.60±1.35 <sup>cfi</sup> | 11.94±0.92 <sup>cfi</sup> | 19.92±1.04 <sup>c</sup> |
| CA (×10 <sup>4</sup> μm <sup>2</sup> ) | 6 | 0          | 9.88±1.32 <sup>c</sup>  | 3.45±0.77 <sup>cf</sup>  | 5.20±1.02 <sup>cfi</sup>  | 3.46±0.83 <sup>cfi</sup>  | 9.74±1.12 <sup>c</sup>  |
| PA/LA (%)                              | 6 | 0          | 50.82±7.43 <sup>c</sup> | 24.56±1.97 <sup>cf</sup> | 35.57±2.51 <sup>cfi</sup> | 23.80±2.39 <sup>cfi</sup> | 48.11±4.20 <sup>c</sup> |
| CA/PA (%)                              | 6 | 0          | 47.19±9.35 <sup>c</sup> | 27.90±5.71 <sup>cf</sup> | 31.30±5.64 <sup>cfi</sup> | 29.11±7.54 <sup>cfi</sup> | 49.07±6.94 <sup>c</sup> |



**Figure 3.** Morphology of the fibrous plaques of WT mice and ApoE<sup>-/-</sup> mice. Cryosections of the thoracic aorta area were obtained from WT mice (A), ApoE<sup>-/-</sup> control mice (B), atorvastatin-treated mice (C), low-dose BRL37344-treated mice (E), high-dose BRL37344-treated mice (E) and SR52390A-treated mice (F) prior to staining with Masson's trichrome. The fibrous plaques with fibrous caps contained smooth muscle cells in B and F (red arrows). Scale bars=100 μm.

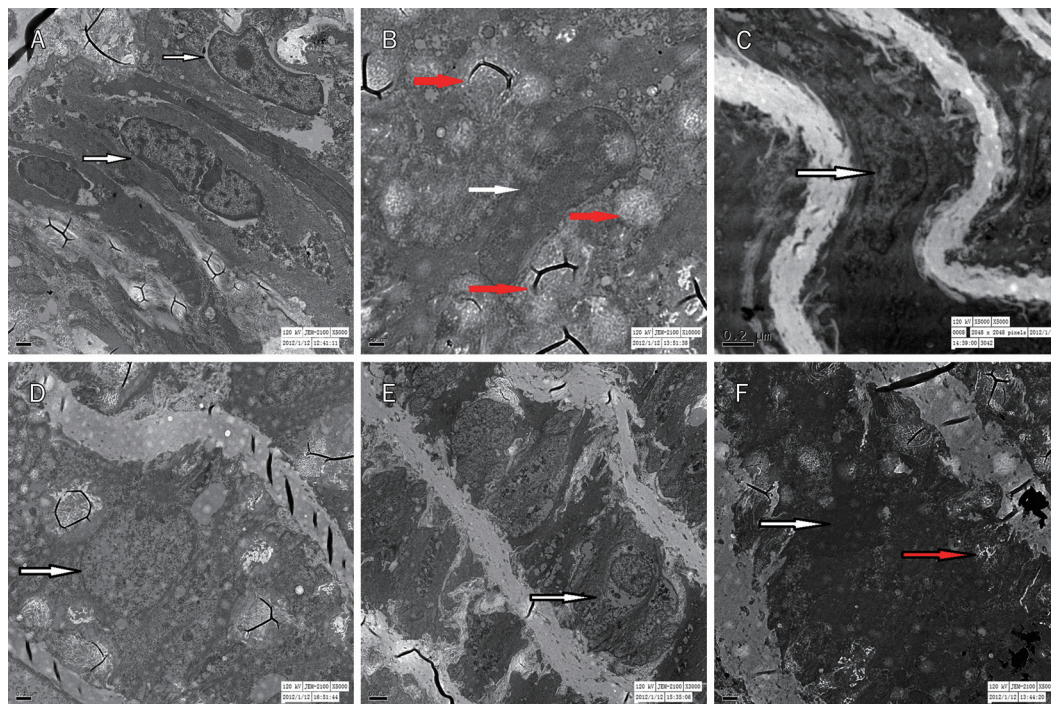
#### β<sub>3</sub>-adrenoceptor mRNA expression in white adipose tissue

β<sub>3</sub>-AR mRNA in epididymal white adipose tissue was decreased in ApoE<sup>-/-</sup> mice compared with WT controls ( $P < 0.01$ ) (Figure 5). BRL37344 increased β<sub>3</sub>-AR mRNA; however, the effects were not dose-dependent. Neither atorvasta-

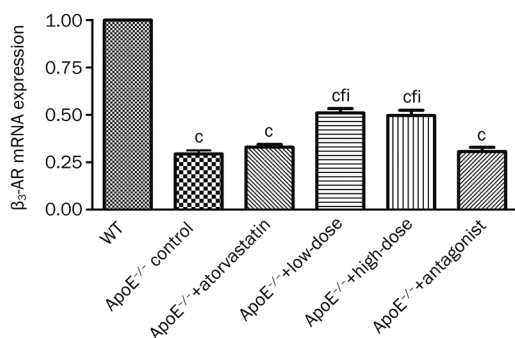
tin nor the β<sub>3</sub>-AR antagonist affected the β<sub>3</sub>-AR mRNA level.

#### Discussion

Atherosclerotic disease is the leading cause of morbidity and mortality in both developed and developing countries. It



**Figure 4.** Transmission electron microscopy images of thoracic aortas obtained from each group. The morphology of SMCs (white arrow) was normal (without fiber hyperplasia) in the WT group (A); Fewer swollen SMCs were noted with obvious fiber hyperplasia (red arrow) in the control and SR52390A-treated groups (B and F); the morphology of thoracic aortas was essentially normal in the atorvastatin group (C); there were many swollen SMCs with mild fiber hyperplasia in mice treated with low and high doses of BRL37344 (D and E).



**Figure 5.**  $\beta_3$ -AR mRNA expression in epididymal adipose tissue.  $\beta_3$ -AR mRNA expression was quantified by real-time PCR and normalized to GAPDH mRNA levels. The results are expressed as the mean  $\pm$  SD ( $n=10$  per group). <sup>c</sup> $P<0.01$  vs WT mice. <sup>cfi</sup> $P<0.01$  vs ApoE<sup>-/-</sup> control mice. <sup>c</sup> $P<0.01$  vs atorvastatin treatment group.

is associated with a metabolic and vascular cluster of diseases. Major risk factors for atherosclerosis include hyperlipidemia, glucose intolerance, hypertension, smoking and aging. Hyperlipidemia is defined as an increased flux of free fatty acids (FFA); increased TG, TC, and nHDL-C levels; and a reduced HDL-C level. Previous studies have shown that early events in atherogenesis include the accumulation of lipoprotein aggregates in the subendothelial space and vessel damage caused by apoB-containing low- and very-low-density lipoproteins<sup>[17,18]</sup>. Statins improve cholesterol metabolism by reducing nHDL-C and prevent the progression of atherosclerotic disease<sup>[19]</sup>. HDL removes excess cholesterol from cells, transporting it to the liver for excretion<sup>[20]</sup>. A low HDL-C level is commonly found in subjects with elevated serum TG and VLDL, a profile that is associated with a high risk of coronary heart disease (CHD)<sup>[21]</sup>.

Recent studies have shown that the  $\beta_3$ -AR agonists can alleviate disorders of glucose and lipid metabolism, increase insulin sensitivity and minimize inflammation<sup>[5,6]</sup>. It is known that the metabolism of lipids and glucose, insulin resistance and inflammation play major roles in atherosclerotic progression. However, the effects of  $\beta_3$ -AR on atherosclerotic disease are not known. In this study,  $\beta_3$ -AR agonists or antagonist was given to the old ApoE<sup>-/-</sup> mice on high-fat diet for 12 weeks and their effects on atherosclerotic development were observed<sup>[22,23]</sup>.

Compared with WT control mice, ApoE<sup>-/-</sup> mice were significantly heavier before BRL37344 treatment, and BRL37344 treatment decreased body weight. We speculated that ApoE deficiency prevented the development of obesity, with decreased fat accumulation in the liver and adipose tissues. These results are consistent with the reports by Kypreos *et al*<sup>[24]</sup>. Atorvastatin did not reduce body weight or fat mass in ApoE<sup>-/-</sup> mice, suggesting that atorvastatin indirectly affected food intake and fat synthesis. The  $\beta_3$ -AR agonist BRL37344 reduced body weight and fat mass in ApoE<sup>-/-</sup> mice. In contrast, the  $\beta_3$ -AR antagonist SR52390A increased body weight and fat mass. These findings are consistent with the hypothesis that  $\beta_3$ -AR

activation could reduce food intake and accelerate lipolysis in ApoE<sup>-/-</sup> mice on a high-fat diet<sup>[7,25]</sup>.

The aged ApoE<sup>-/-</sup> mice developed prominent dyslipidemia (eg, high serum TG, TC and nHDL-C levels and a low HDL-C/TC ratio) as well as atherosclerotic lesions when given a high-fat diet. These results are consistent with reports by Kawashima *et al*<sup>[16]</sup> and Zadelaar *et al*<sup>[26]</sup>. The  $\beta_3$ -AR agonist reduced the levels of TG, TC, and nHDL-C and markedly raised the HDL-C level and the HDL-C/TC ratio. Such findings are consistent with previous studies showing that chronic  $\beta_3$ -AR activation could increase the oxidative capacity of adipocytes within white adipose tissue by upregulating the expression of uncoupling protein (UCP) 1<sup>[27-30]</sup>.  $\beta_3$ -AR activation could also reduce the levels of nHDL-C and chylomicron in addition to raising the HDL-C level via the secretion of hormone-sensitive lipase and adiponectin and the upregulation of adiponectin receptor expression<sup>[6,31-33]</sup>. In the present study, atorvastatin did not improve TG or HDL-C levels; however, it reduced TC and nHDL-C levels in ApoE<sup>-/-</sup> mice. Atorvastatin could inhibit the synthesis of cholesterol by inhibiting 3-hydroxy-3-methylglutaryl-coenzyme A (HMG-CoA) in the liver, and by doing so, it could regulate TG and HDL-C metabolism. The current study suggested that the  $\beta_3$ -AR agonist is superior relative to atorvastatin for increasing HDL-C and HDL-C/TC while decreasing TG.

In our experiments, ApoE<sup>-/-</sup> mice on a high-fat diet displayed hyperglycemia, hyperinsulinemia and insulin resistance. In agreement with previous studies in obesity/diabetes animal models<sup>[5,6,34]</sup>, the stimulation of  $\beta_3$ -AR improved glycemic control, as evidenced by reduced hyperinsulinemia and hyperglycemia and elevated ISI in ApoE<sup>-/-</sup> mice. Mechanistically, the  $\beta_3$ -AR agonist could upregulate insulin receptors and glucose transporters in the peripheral tissues, promoting glucose uptake and insulin-independent utilization in adipose tissue and skeletal muscle and reduced glycogen synthesis in the liver<sup>[35]</sup>.  $\beta_3$ -AR activation has been reported to enhance insulin action in the liver and adipose tissue by mediating the insulin-like growth factor I (IGF-I)/insulin receptor signaling pathways of skeletal muscle<sup>[7]</sup>. Although there was a slight decrease in the insulin level in our study, atorvastatin did not reduce the glucose level or the ISI. These results suggest that atorvastatin is not sufficient for glycemic control in ApoE<sup>-/-</sup> mice. Atorvastatin inhibits the synthesis of fats by inhibiting the expression of sterol regulatory element binding protein-1 (SREBP-1) in the liver; however, it has less influence on glucose metabolism<sup>[36]</sup>. Indeed, atorvastatin has been shown to decrease the expression of the glucose transporter GLUT-4 in adipocytes and impair glucose tolerance<sup>[37]</sup>.

Quantitative real-time PCR confirmed  $\beta_3$ -AR mRNA expression in white adipose tissue; this expression was downregulated in the ApoE<sup>-/-</sup> control mice and by SR52390A treatment. The  $\beta_3$ -AR mRNA level was increased by BRL37344 and atorvastatin. These results are consistent with previous studies on obese/diabetic mice<sup>[4,6,38]</sup>. The basis of these findings could be the inhibitory effects of insulin on  $\beta_3$ -AR expression<sup>[39]</sup>. Previous studies have shown that the  $\beta_3$ -AR agonist CL-316,234

increases  $\beta_3$ -AR mRNA levels and improves glucose metabolism. Similar results were obtained in this study with ApoE<sup>-/-</sup> mice. Upregulation of  $\beta$ -AR may further increase the effect of the  $\beta_3$ -AR agonist on plasma lipids, glucose and anti-insulin resistance<sup>[6]</sup>. Neither atorvastatin nor SR52390A increased the  $\beta_3$ -AR mRNA level in the white adipose tissue of ApoE<sup>-/-</sup> mice. These results could partially explain the failure of both drugs to improve glycemic control.

Our study demonstrated that the  $\beta_3$ -AR agonist could reduce the atherosclerotic plaque area and collagen content and increase the lumen area in the thoracic aorta of ApoE<sup>-/-</sup> mice. The observed changes in HDL-C could activate the liver orphan receptor (LXR), accelerate reverse cholesterol transport (RCT), and prevent the progression of atherosclerosis<sup>[40]</sup>. Additionally,  $\beta_3$ -AR agonists can increase the protein levels of adiponectin and adiponectin receptors, which contribute to increases in the expression of apolipoprotein A-1 (apoA-I) and ATP-binding cassette transporter A1 (ABCA1) in liver tissue<sup>[6, 41]</sup>. ApoA-1 and ABCA1 transport excess peripheral cholesterol to the liver, thus preventing cholesterol accumulation and atherosclerotic plaque formation<sup>[42, 43]</sup>.

In conclusion, this study demonstrated that  $\beta_3$ -AR activation could impede the progression of atherosclerosis, possibly through improving lipid and glucose metabolism. At a high dose, the  $\beta_3$ -AR agonist BRL37344 was similar to atorvastatin with respect to its ability to reduce atherosclerosis.

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### Author contribution

Yan-fang LI designed the research, Zhao-hong WANG and Yan-qing GUO performed the research, Yan-qing GUO analyzed the data, and Zhao-hong WANG wrote the paper.

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