

REVIEW

Defining the nature of human $\gamma\delta$ T cells: a biographical sketch of the highly empathetic

Shirin Kalyan and Dieter Kabelitz

The elusive task of defining the character of $\gamma\delta$ T cells has been an evolving process for immunologists since stumbling upon their existence during the molecular characterization of the α and β T cell receptor genes of their better understood brethren. Defying the categorical rules used to distinctly characterize lymphocytes as either innate or adaptive in nature, $\gamma\delta$ T cells inhabit a hybrid world of their own. At opposing ends of the simplified spectrum of modes of antigen recognition used by lymphocytes, natural killer and $\alpha\beta$ T cells are particularly well equipped to respond to the 'missing self' and the 'dangerous non-self', respectively. However, between these two reductive extremes, we are chronically faced with the challenge of making peace with the 'safe non-self' and dealing with the inevitable 'distressed self', and it is within this more complex realm $\gamma\delta$ T cells excel thanks to their highly empathetic nature. This review gives an overview of the latest insights revealing the unfolding story of human $\gamma\delta$ T cells, providing a biographical sketch of these unique lymphocytes in an attempt to capture the essence of their fundamental nature and events that influence their life trajectory. What hangs in their balance is their nuanced ability to differentiate the friends from the foe and the pathological from the benign to help us adapt swiftly and efficiently to life's many stresses.

Cellular & Molecular Immunology (2013) 10, 21–29; doi:10.1038/cmi.2012.44; published online 22 October 2012

Keywords: antigen recognition; danger-associated molecular patterns; gamma delta T cell subsets; immune homeostasis; innate immunity; stress response

PROLOGUE

Since the time of their accidental discovery during the molecular characterization of the α and β T cell receptor (TCR) genes belonging to their relatively unambiguous brethren,¹ 'enigmatic' is still one of the most commonly used adjectives to describe $\gamma\delta$ T cells. This can be attributed to the fact that these unconventional lymphocytes have thus far been remarkably successful in thwarting most attempts at definition. Much of what we know about T cell biology and function comes from what has been discerned through studies done on $\alpha\beta$ T cells; however, one thing that is certain is that the majority of the rules governing the lives of $\alpha\beta$ T cells are surprisingly irrelevant when it comes to understanding the elusive character of $\gamma\delta$ T cells. Table 1 highlights some of the major differences between these two T cell subsets in humans.

Recognition by $\gamma\delta$ T cells is not typically in the context of classical major histocompatibility complex (MHC) class I or II molecules and neither is antigen processing required.^{2,3} These observations are corroborated by the fact that the majority of $\gamma\delta$ T cells are CD8 and CD4 negative. Crystal structure analysis of the $\gamma\delta$ TCR determined that the length and conformation of $\gamma\delta$ TCR resemble immunoglobulins (Ig) more than the $\alpha\beta$ TCR, which was taken to suggest that antigen recognition by $\gamma\delta$ T cells may be more similar to the binding of antibody to antigen rather than the MHC/peptide complex recognized by $\alpha\beta$ T

cells.⁴ The very basis of foreign antigen recognition by lymphocytes of the adaptive immune system is a contentious point for these unique T cells. Although $\gamma\delta$ T cells appear to have antigen recall memory by their demonstrated rapid expansion and antimicrobial response upon reinfection with a particular pathogen, much like lymphocytes of adaptive immunity,^{5–7} clonally expanded $\gamma\delta$ T cells with very restricted receptor repertoires are able to respond readily to a wide range of both infectious and non-infectious stressors. Natural killer (NK) cells share some characteristics with $\gamma\delta$ T cells as both are usually considered constituents of innate immunity, recognize transformed cells, play a prominent role in antiviral protection and are cytolytic lymphocytes.^{8–10} Like NK cells, human $\gamma\delta$ T cells express activating receptors, such as NKG2D that recognizes stress-inducible MHC class I-related MICA/MICB molecules and the UL16-binding proteins that are upregulated on malignant or stressed cells and induce cytotoxicity,^{11,12} and they are inhibited by the expression of killer Ig-like receptors, which bind certain MHC class I alleles expressed by non-malignant cells and trophoblasts.^{13–15} However, the versatility of $\gamma\delta$ T cells is further extended by their demonstrated ability to assume the role and appearance of professional antigen presenting cells.^{16,17} This unusual functional plasticity has given the impression that these multitasked T cells are 'Jacks of all-trades, but masters of none', and they are sometimes erroneously delegated to a niche occupied by vestigial

Institute of Immunology, Christian-Albrechts University Kiel, Kiel, Germany

Correspondence: Dr S Kalyan, Institute of Immunology, Christian-Albrechts University Kiel, Arnold-Heller Strasse 3, Haus 17, D-24105 Kiel, Germany.

E-mail: kalyan@immunologie.uni-kiel.de

or Prof D Kabelitz

E-mail: kabelitz@immunologie.uni-kiel.de

Received 26 July 2012; accepted 28 August 2012

Table 1 Major differences between human $\alpha\beta$ and $\gamma\delta$ T cells

Feature	$\alpha\beta$ T cells	$\gamma\delta$ T cells
Frequency in blood	• 65%–75% of PBMC	• <10% of PBMC (25%–60% gut)
MHC restriction	• CD4 ⁺ : MHC class II • CD8 ⁺ : MHC class I	• No MHC restriction • Possible roles of CD1 and MICA/B
CD4/CD8 expression	• ~60% CD4 ⁺ ; ~30% CD8 ⁺ ; <1% double positive; • <1% double negative	• Majority (>70) double negative; <1% CD4 ⁺ ; ~30% CD8 ⁺ $\alpha\alpha$ (as IELs in gut)
Antigen recognition	• Processed peptide/MHC	• Unprocessed, not peptides
TCR V gene germ line repertoire	• Large	• Small
TCR diversity	• Very diverse	• Relatively restricted expression despite high potential for junctional diversity; expression variance dictated by tissue localization
Function	• Adaptive immunity	• Immune regulation, surveillance and homeostasis

Abbreviations: IEL, intraepithelial lymphocyte; MHC, major histocompatibility complex; PBMC, peripheral blood mononuclear cell; TCR, T cell receptor.

features of modern anatomy by some who are left confounded as to their real purpose.

This review provides a biographical sketch of human $\gamma\delta$ T cells in an attempt to convey the essence of their fundamental nature and what may influence their life trajectory. Figure 1 illustrates the highly simplified paradigm for NK cells, $\alpha\beta$ T cells and the emerging view of where in the continuum that encompasses immune homeostasis and protection $\gamma\delta$ T cells take the throne. At opposing ends, NK and $\alpha\beta$ T cells are particularly well equipped to respond to the ‘missing self’ and the ‘dangerous non-self’, respectively, or as Klas Kärre had so aptly put it—‘to recognize a foreign submarine’.¹⁸ Between these two extremes, we are chronically faced with the challenge of making peace with the ‘safe non-self’ and dealing with the inevitable ‘distressed self’, and it is within this more complex realm $\gamma\delta$ T cells excel thanks to their highly empathetic nature. What hangs in the balance is the ability to dif-

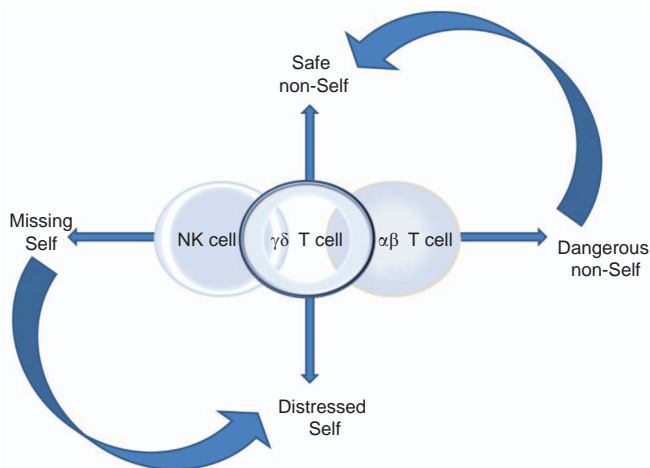


Figure 1 A simplified paradigm illustrating where in the continuum of immune protection and homeostasis $\gamma\delta$ T cells fall in relation to innate NK cells and the adaptive $\alpha\beta$ T cells. At the two extreme ends, NK and $\alpha\beta$ T cells are particularly well equipped to respond to the ‘missing self’ and the ‘dangerous non-self’, respectively. Between these two extremes, we are chronically faced with the challenge of making peace with the ‘safe non-self’ and dealing with the inevitable ‘distressed self’, and it is within this more complex realm $\gamma\delta$ T cells excel. It should be recognized that these different ‘selves’ and the immune response(s) that they trigger exist in a continuum and are modulated by the context in which they are presented. Both NK and $\alpha\beta$ T cells work with $\gamma\delta$ T cells to fill in the gaps of this spectrum—with NK cells contributing to responding to the ‘distressed self’ and $\alpha\beta$ T cells having some regulatory training to temper the response to the ‘safe non-self’. NK, natural killer.

ferentiate the friends from the foe and the pathological from the benign—and adapt swiftly and efficiently to life’s many stresses.

SPECIES SPECIFICITY: THE BEST LAID PLANS OF MICE AND MEN OFT GO AWRY

Among the many fascinating and important conundrums that have yet to be solved for $\gamma\delta$ T cells is the large phenotypic difference between species when it comes to their relative numbers, location and antigen recognition. In both mice and primates, $\gamma\delta$ T cells comprise a small population of peripheral T cells (averaging 5% of the total peripheral T cell pool in Caucasians living in Western Europe and North America), and they are poorly represented within conventional T cell zones found in lymphoid tissue.¹⁹ In contrast, sheep, cattle, rabbits and chickens have much higher levels of circulating $\gamma\delta$ T cells.²⁰ The reason behind this disparity in the constitution of $\gamma\delta$ T cells between and within species is currently unknown, but the discrepancy seen between ‘low’ and ‘high’ $\gamma\delta$ T cell species has been correlated with the total level of TCR variable genes, which is represented largely by the variable potential of the $\alpha\beta$ TCR, in conjunction with the diversity present in the Ig variable genes. Thus, species having a high number of variable TCR and Ig genes tend to have low numbers of peripheral $\gamma\delta$ T cells (Table 2).²⁰ It can be speculated that the role of and exposure to microbes in the species’ lifecycle may be associated with these immune parameters. For example, omnivorous rodents and primates may have less of a need to maintain peace with a diverse population of microbiota in comparison to cows, sheep, rabbits and chickens for which microbes play a more significant role in digestion and metabolism.^{21–24} Therefore, the immune cell repertoire that recognize more variable ‘non-self’ as dangerous (i.e., the features that we depend on from the adaptive immune system) may be less advantageous when balancing the need to protect beneficial or non-threatening microbes that are needed for sustenance and/or to prevent chronic inflammation.

As shown in Table 3, the numbers of V γ and V δ gene segments in both mice and men are greatly limited in comparison to $\alpha\beta$ T cells, and despite their relative low levels in peripheral blood, $\gamma\delta$ T cells are found to be enriched at mucosal interfaces, such as the intestinal tract, that are heavily populated by largely non-pathogenic and symbiotic organisms.^{19,25,26} In the murine system, distinct subsets of $\gamma\delta$ T cells bearing specific pairs of the γ and δ TCR chains are targeted to particular epithelial locations during development such as the skin, lungs, uterus, vagina and tongue where they make up a significant portion of the local intraepithelial lymphocyte population and appear to respond to endogenous stress-induced antigens that have yet to be unambiguously identified.^{27–31} For some of these special niche localized $\gamma\delta$ T

Table 2 Comparative diversity and percent composition of peripheral $\gamma\delta$ T cells between species (adapted from Su *et al.*, 1999²⁰)

	Percent peripheral $\gamma\delta$ T cells	Diversity of $\alpha\beta$ T cell receptor variable genes	Diversity of immunoglobulin variable genes
Human	~5% (low)	High	High
Mouse	~5% (low)	High	High
Chicken	~20% (high)	Low	Low
Rabbit	~20% (high)	Low	Low
Sheep	~30% (high)	Low	Low
Cattle	~30% (high)	Low	Low

cells, such as the invariant dendritic epithelial V γ 3V δ 1 T cells found in murine skin,²⁸ no human equivalent has been found. However, it is interesting to note that the phenomenon observed at the specie level is extended to niches found within an organism and that an abundance of certain subsets of $\gamma\delta$ T cells are found where there is a vital need to guard against inappropriate immune responses to ‘safe non-selves’ at the same time as keeping in check the inevitable ‘distressed self’—a combinatorial function that neither $\alpha\beta$ T cells nor NK cells are especially designed to deal with alone.

‘Non-V δ 2 T cells’: the high art of non-self acceptance

In adult humans, two subsets of $\gamma\delta$ T cells, defined by the usage of either the V δ 2 or V δ 1 TCR, predominate. The majority of the tissue-associated $\gamma\delta$ T cells bear the V δ 1 TCR with V δ 3 and V δ 5 making up minor populations.^{32,33} In the literature, these are often collectively called the ‘non-V δ 2’ $\gamma\delta$ T cells. In addition to their role in maintaining immune homeostasis in the local microenvironment,²⁵ tissue-associated $\gamma\delta$ T cells, in both mice and men, play an important function in wound healing, removing distressed or transformed epithelial cells and subduing excessive inflammation.^{19,26,34–39} For example, in the gastrointestinal tract $\gamma\delta$ T cells play a non-redundant role in maintaining tolerance to food antigens as well as the intestinal flora.^{34,40} The V γ chain pairing of tissue-associated V δ 1 T cells in humans is less stringent in comparison to the highly restricted V γ 9V δ 2 T cells found in peripheral blood—though they prefer pairing with V γ 4 and V γ 5 over V γ 9.^{32,41} This greater diversity in pairing and structure is taken to imply that there exists a broader range of ligands that are recognized—but the recognition is thought to be largely introspective as it is usually in response to the changing states of self. The actual antigen specificity and mechanism of recognition of these non-V δ 2 T cells is still in the unraveling. The stress-induced molecules, MICA and MICB, have been found to be tumor antigens recognized by cancer infiltrating V δ 1 T cells.^{42,43} The redundancy of this recognition is a point in the pondering since both V δ 1 and V δ 2 T cells express NKG2D, an activating C-type lectin that is the known receptor for MICA, MICB and related UL16-binding protein.^{44,45} Nevertheless, the crystal structure analysis showed that the binding of V δ 1 T cells to MICA was mutually exclusive to NKG2D, although the complementary determining region (CDR) of the TCR appeared to have an uncharacteristically flat potential binding surface.⁴⁶ Extending this apparent tendency to respond to non-polymorphic MHC class I family-related proteins, CD1c, a molecule that is able to present both foreign and self lipid

antigens,⁴⁷ is another candidate ligand for V δ 1 T cells.⁴⁸ It remains to be seen exactly how broadly based these recognition patterns are within the V δ 1 T cell family since many of these studies have relied on a limited number of $\gamma\delta$ T cell clones or lines. Adding a new twist to this storyline, it was recently revealed by the working groups of Willcox and Dechanet-Merville that a ‘non-V δ 2’ T cell clone expressing V γ 4V δ 5 TCR was activated by the recognition of endothelial protein C receptor.⁴⁹ This interaction involved the CDR3 of V γ 4 and was similar in nature to the binding of antibody to antigen. Of particular note, recognition was highly dependent on the multimolecular stress signature that was induced by either infection by cytomegalovirus or malignant transformation.⁴⁹ Endothelial protein C receptor is another member of the CD1/MHC class I superfamily that is expressed on endothelium and trophoblasts⁵⁰ and can bind lipids similar to CD1 family members; however, binding of the V γ 4V δ 5 T cell clone was determined to be independent of the presence of any lipid antigen.⁴⁹

In continuation of the theme of sometimes needing to accept the ‘safe non-self’, the immunological state of pregnancy is an example of a time when this requirement is essential. It is still an enigma how the maternal immune system in mammals is not only able to prevent rejection of the fetal semiallograft, but also successfully support its growth for 9 months. Human V δ 1 T cells have been observed to be recruited to the maternal/fetal interface during pregnancy where their activation is believed to play a role in the induction of tolerance to paternal antigens.^{15,51–54} Furthermore, there is evidence suggesting that the early pregnancy human decidua is actually a site of extrathymic maturation of V δ 1 T cells.^{55,56} This enrichment of $\gamma\delta$ T cells at the maternal/fetal interface and their mysterious role in pregnancy is not unique to humans as the same has been observed in sheep⁵⁷ and mice⁵⁸, suggesting a central conserved role of $\gamma\delta$ T cells involved in pregnancy-related immune changes. In women with healthy pregnancies, the changes are not only found at the maternal/fetal interface since the number of V δ 1 T cells in circulation have also been reported to be increased in relation to the V δ 2 subset.^{59–61} Of note, some immune-related pregnancy complications where maternal immune tolerance is compromised and there is risk of premature pregnancy termination, a significant drop in the number of circulating V δ 1 T cells is observed.⁶² The cognate pregnancy-associated $\gamma\delta$ T cells in the murine system were found to directly recognize an unknown antigen on trophoblasts *via* their TCR, and this recognition was MHC-unrestricted and, intriguingly, not species-specific.⁶³ Although expression

Table 3 Comparative diversity of the α , β , δ and γ TCR chains in mice and men³²

	Human	Mice
V γ	~6 (5 from same family and 1 distantly related)	~6 (2 from same family and 4 broadly diverged)
V δ	~8 functional genes (only 3 commonly used—VD1, VD2 and VD3)	~16 (6 homologous and 10 distinct)
V α	~42	~75
V β	~47	~23

Abbreviation: TCR, T cell receptor.

of inhibitory non-classical MHC class I molecules, such as HLA-E and HLA-G, on trophoblasts appear to play a role in suppressing V δ 2 T cells during pregnancy, they are unlikely to be the candidates recognized by the V δ 1 T cells actively on guard.¹⁵ Mammals require a reliable mechanism to maintain immune tolerance to 'safe non-selves', as a failure to do so would be extinction. In this regard, V δ 1 T cells seem to have a conserved acumen for brokering peace for the benefit of generations to come.

V δ 2 T cells: to know the complex burden of being human

In the peripheral blood of most healthy adult humans and some other primates such as the rhesus monkey, the vast majority of $\gamma\delta$ T cells bear the V δ 2 TCR—usually paired with the V γ 9 chain.⁶⁴ These special canonical V γ 9V δ 2 T cells recognize non-peptidic phosphorylated antigens, such as isopentyl pyrophosphate (IPP), that are metabolites in the essential isoprenoid biosynthesis pathway present in virtually all living organisms.⁶ Endogenously, IPP and its stereoisomer dimethylallyl diphosphate are substrates produced in the mevalonate pathway for cholesterol metabolism; however, they can also be produced by the deoxyxylulose pathway commonly used by organisms, such as *Escherichia coli* and certain plant cells, that lack the critical HMG-CoA reductase enzyme of the mevalonate pathway.^{65,66} It was later determined that the stimulatory capacity of IPP and dimethylallyl diphosphate is actually fairly weak in comparison to some of the upstream intermediates of the alternative pathway of isoprenoid biosynthesis, such as (*E*)-4-hydroxy-3-methyl-but-2-enyl pyrophosphate,⁶⁷ and it is usually in the context of malignancy or distress that cellular endogenous levels are able to awaken the effector functions of V γ 9V δ 2 T cells.⁶⁸ A class of drugs called aminobisphosphonates used for the prevention of bone fragility fractures and certain alkylamine compounds that are ubiquitously dispersed in everything from plants to amniotic fluid are also able to stimulate V γ 9V δ 2 T cells^{69,70} by virtue of their ability to induce cellular accumulation of IPP. Both these compounds modulate IPP levels by blocking the metabolic activity of farnesyl pyrophosphate synthase, a key enzyme in the mevalonate pathway.^{71,72}

It is unclear whether these 'phospho-antigens' (pAgs) require presentation *via* some cell-surface accessory molecule; however, the fact that contact with cells of human origin is necessary for their ability to fully activate V γ 9V δ 2 T cells seems to suggest that some kind of assistance is necessary.⁷³ Recent developments in regards to alternative potential endogenous stress-induced ligands for V γ 9V δ 2 T cells provide new venues that may reveal how these cellular short-lived substrates can be potentially stabilized and presented extracellularly to the stress surveillance capabilities inherent in these lymphocytes. Scotet and colleagues⁷⁴ first reported that tumor recognition by V γ 9V δ 2 T cells could be mediated by the ectopic expression of ATP synthase/F1-ATPase, which is normally expressed on the internal membrane of mitochondria, and this interaction was enhanced by the cobinding of apolipoprotein A-I that is usually present in serum. Literally along the same vein, it was later observed that shear stress experienced by endothelial cells also led to the translocation of the ATP synthase β chain to the cell surface which resulted in the binding and activation of V γ 9V δ 2 T cells.⁷⁵ Reminiscent of the previous findings with F1-ATPase, the response of $\gamma\delta$ T cells to endothelial cells expressing ATP synthase β was significantly potentiated by the coincident accumulation of cholesterol in the cell membrane, and this interaction led to the secretion of inflammatory cytokines by $\gamma\delta$ T cells and upregulation of vascular cell adhesion molecules on endothelial cells.⁷⁵ This phenomenon was taken to suggest that endothelial dysfunction, char-

acterized by the disturbed flow created by shear stress, and hypercholesterolemia work synergistically to activate $\gamma\delta$ T cells and the endothelium⁷⁵—providing a novel mechanism contributing to cardiovascular pathology. In an attempt to reconcile the established reactivity of V γ 9V δ 2 T cells to non-peptidic pAgs and these new observations of alternative stress-induced ligands, experiments were carried out to demonstrate that ATP synthase can also potentially bind to pAgs, and it was concluded that it seemingly has the appropriate features to function as a possible pAg presenting molecule for V γ 9V δ 2 T cells.⁷⁶ More recently, another putative V γ 9V δ 2 T cell tumor antigen, human MutS homologue 2 (MSH2), was identified using a peptide-based affinity screening system for the CDR3 of the δ chain.⁷⁷ MSH2 is normally located in the nucleus where it functions as a DNA mismatch repair gene, but it is often mutated in a number of different types of epithelial cancers and can be ectopically expressed.⁷⁸ Transformation of normal human B cells by Epstein–Barr virus or subjecting renal carcinoma cell lines to oxidative stress also led to an increased surface expression of MSH2 and rendered them susceptible to V γ 9V δ 2 T cell-mediated cell lysis that could be blocked by the use of anti-MSH2 antibodies or downregulation of its gene expression.^{78,79} These observations were collectively taken to suggest that MSH2 may be yet another damage-associated molecular pattern recognized by V γ 9V δ 2 T cells. An overlooked point worth considering is the fact that MSH2 also has an intrinsic ability to bind and hydrolyze ATP,⁸⁰ which may be the property providing the common mechanism leading to the mobilization of human peripheral blood $\gamma\delta$ T cells. Both ATP synthase/F1-ATPase and MSH2 are evolutionarily conserved molecules that serve essential stress-sensitive homeostatic functions in prokaryotes and eukaryotes and can bind nucleotide derivatives, and this may confer the potential to present low molecular weight pAgs to V γ 9V δ 2 T cells' clever immunosurveillance strategy. This is still in large part conjecture, and much of the story is still in the telling given that neither F1-ATPase nor MSH2 are absolutely indispensable for pAg recognition. Exogenously added pyrophosphate antigens readily stimulate V γ 9V δ 2 T cells in the presence of human accessory cells, yet both these molecules require some stress-induced event for their ectopic expression—which argues for there existing some other more widely expressed candidate(s) that would be more exclusive to the primate lineage. Harly and colleagues,⁸¹ from the working groups of Scotet and Bonneville, may have uncovered at least one of the missing pieces of the story in the form of a molecule known generically as CD277—or BTN3A. This unassuming molecule is a member of the butyrophilin/B7-like group of proteins belonging to the Ig superfamily that are found clustered at the extended end of the classical MHC I genes on human chromosome 6.⁸² In a series of elegant experiments involving specific activating and inhibitory antibodies to CD277 as well as domain-swapping and knockdown experiments, the Scotet–Bonneville group shows convincing evidence that this widely expressed molecule provides the necessary support that pAgs need to awaken V γ 9V δ 2 T cell effector functions.⁸¹ The fact that no functional orthologue of CD277 exists in rodents adds further credence to its purported role as the long-awaited missing link—or what may very well turn out to be a series of interconnected links.

Given the fact that these unique pAg-reactive V γ 9V δ 2 T cells have no known equivalent in either rodents or ruminants, it is perplexing why their effective stress surveillance tactic appears to be solely a primate advantage. It is of considerable interest to determine where along our common evolutionary lineage they first made their appearance. With the emergence of the peripheral V γ 9V δ 2 T cell subset, primates seem to have done away with the functional niches of a

number of other various $\gamma\delta$ T cell subsets and chiseled it down to mostly two highly specialized arsenals—the one in the periphery often found fanning the flames at the first site of trouble and the one entrenched in the tissues often putting out the fire before it starts.

rites of passage

First there were $\gamma\delta$ T cells

Befitting their deeper evolutionary roots, the ontogeny of $\gamma\delta$ T cells precedes $\alpha\beta$ T cells in expression. $\gamma\delta$ TCR gene rearrangement can be detected by embryonic day 14 in the murine thymus⁸³ and week 8 in humans,³² and canonical subsets can also be detected extrathymically during fetal development in both species.^{83,84} The most striking feature is the change from having fairly diverse pairs of $\gamma\delta$ T cells (of which V δ 1⁺ serve as the majority in cord blood at birth) to increasingly restricted pairings, with V γ 9V δ 2 T cells becoming the major subset with very limited receptor diversity by adulthood (Figure 2).³² By 1 year of life, the majority of peripheral blood V γ 9V δ 2 T cells are non-naïve, readily produce IFN γ upon stimulation and have cytolytic granules.⁸⁵ It was concluded that this phenomenon was evidence of ‘the earliest immunological maturation of any lymphocyte compartment in humans.’⁸⁵ In contrast, the V δ 1 T cells remaining in the periphery in early life were found to be still naïve—similar to $\alpha\beta$ T cells.⁸⁵ This could be because the antigens that ‘non-V δ 2’ T cells recognize are more abundantly expressed in the tissue microenvironment where they are targeted, and an analysis of V δ 1 T cells in their true niches, such as the intestine, in this early period may very well find that they too have a mature phenotype. The absolute number of $\gamma\delta$ T cells in the periphery increases from birth to about 10 years of age, which is congruent with the expansion of the V γ 9V δ 2 T cell subset going from a minor population at birth to usually >75% of circulating $\gamma\delta$ T cells.³² Although this clonal expansion has been seen as evidence of the vital role that these T cells play in responding to environmental challenges in early life, it has not been demonstrated that this phenomenon is in fact primarily in response to environmental challenges and not, at least in part, in response to endogenous stimuli as an

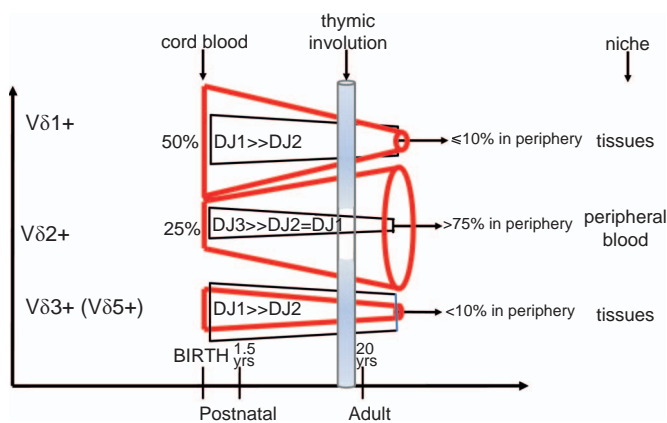


Figure 2 Schematic diagram of the changes in the diversity of $\gamma\delta$ T cell repertoire through development in humans. At birth, $\gamma\delta$ T cells show high junctional diversity, which is generated through the addition of ‘P’ (palindromic) and ‘N’ (non-templated) nucleotides while combining the V-D-J segments of their TCR. Despite the limited variable genes available for $\gamma\delta$ T cells, their potential for junctional diversity rivals that of $\alpha\beta$ T cells. However, with age, the diversity that is observed at birth gets noticeably restricted, especially for the main pool of circulating $\gamma\delta$ T cells, the V γ 9V δ 2 T cell subset, which are a minor population in cord blood but become the predominant subset in circulation by 1 year of life. TCR, T cell receptor.³²

extension of the adaptive changes taking place within the newborn as the immune system is being initiated.

Defined by experiences tainted by innate tendencies

It should be noted that what has been described for the early life and developmental trajectories of $\gamma\delta$ T cells is based largely on studies conducted in North America and Western Europe, and it is now appreciated that there are great interindividual and regional differences in the make-up of one’s $\gamma\delta$ T cell repertoire. A small study looking at ethnic differences in peripheral $\gamma\delta$ T cell numbers found Turkish and ‘non-Japanese Asians’ (which comprised of a handful of people from Iran, India, Thailand and China) had significantly higher levels of $\gamma\delta$ T cells (a median of 9.3% and 9.2%, respectively, of the total T cell pool) in comparison to Swedes (4.2%) and Japanese Asians (4.5%).⁸⁶ A childhood history of tuberculosis in one Japanese man was associated with an unusually high percentage of $\gamma\delta$ T cells (23.5%), which was similar to some of those sampled from Turkey and non-Japanese Asian countries. Thus, it was concluded that environmental factors, rather than genetic, are likely to be the driving force behind these ethnic and regional disparities in the proportion of $\gamma\delta$ T cells found in peripheral blood. It is interesting to note the regional and interindividual differences in the distribution and number of $\gamma\delta$ T cells is in line with the ‘hygiene hypothesis’ which purports that the highly sterilized environment of industrialized or ‘Westernized’ nations impacts the developmental trajectory of the immune system that is partly shaped by its reciprocal interaction with the microbial world at an early age.⁸⁷ This is speculated to be a major contributive factor for the higher prevalence of immune and inflammatory disorders present in industrialized countries.^{87,88}

Although a thorough study of potential genetic determinants is still lacking, its contribution to the observed phenotype of $\gamma\delta$ T cells should not be ignored. An analysis of the repertoire of peripheral blood $\gamma\delta$ T cells in the West African region of Ghana found that healthy people there had a five-fold higher level of circulating V δ 1 T cells in comparison to their European counterparts.⁸⁹ This reversed ratio of V δ 1/V δ 2 T cells showed no age dependency nor did it appear to be an antigen-driven event.⁸⁹ This region is highly endemic for *Plasmodium falciparum*, the causative agent of malaria, for which the V δ 1 T cell subset seems to have a special immunoregulatory function.^{90,91} It is therefore feasible that this change in $\gamma\delta$ T cell distribution observed in never-infected individuals from this West African population stems from evolutionary selection based on an immune advantage by those having a high proportion of V δ 1 T cells—which would again align with the association of there being some adaptive influence of microbial exposure on the make-up of $\gamma\delta$ T cells in an organism. Other infectious diseases that are associated with the recruitment and expansion of specifically the V δ 1 T cell subset in the periphery are cytomegalovirus,^{33,92} Epstein–Barr virus^{93,94} and HIV.^{95,96} In most of these cases, the specific activation and expansion appears to be driven by the presentation of endogenous molecules rather than virally derived antigens—keeping with the notion that these T cells are particularly well adapted to sensing the turmoil within rather than relying exclusively on external validation.

Tales told by battle scars and tree rings

A number of chronic inflammatory diseases are also associated with perturbed $\gamma\delta$ T cell distributions.^{97–101} In some cases, such as systemic sclerosis, rheumatoid arthritis, Takayasu’s arteritis and Wegner’s granulomatosis, an inverse ratio of peripheral blood $\gamma\delta$ T cells is seen again such that the V δ 1 T cell subset outnumber the V δ 2 T

cells.^{100,101} (and unpublished data) This occurrence is often associated with the concomitant loss of peripheral V γ 9V δ 2 T cells—possibly due to their tendency to undergo activation-induced cell death *via* FAS-mediated apoptosis which occurs following repeated stimulation of their TCR.^{102–104} Death by exhaustion appears to be a consequence for the highly empathetic in an environment subjected to chronic distress. An example of this is seen in people with Crohn's disease who were found to have a global deficit in the number of $\gamma\delta$ T cells present.⁹⁸ Evidence that this may be a result of activation-induced cell death as opposed to an inherent immune anomaly of the disease comes from the observation that treatment with infliximab, an anti-TNF α agent, resulted in clonal expansion of $\gamma\delta$ T cells in Crohn's patients on treatment¹⁰⁵, highlighting the potential *in vivo* role of TNF α signalling in $\gamma\delta$ T cell self-modulation. However, *in vitro* studies looking at the functional effects of infliximab on $\gamma\delta$ T cells derived from patients with Behçet's disease or rheumatoid arthritis, in contrast, found that the expansion and reactivity of these excitable T cells was notably impaired,^{106,107} underscoring again the often discrepant outcomes when attempting to extrapolate findings from manipulated artificial settings *in vitro* to what occurs *in vivo*. This has been a particularly challenging hurdle when trying to implement the remarkable anti-cancer arsenal of $\gamma\delta$ T cells for cancer immunotherapy.^{10,108–110} Despite the ready expansion and cytolytic effector functions of $\gamma\delta$ T cells demonstrated at the bench, attempts to harness this potential in the clinical setting have been disappointing due to the inevitable loss of these cells that is observed in many cancer patients.^{111,112} It is therefore of great interest to find ways to keep these cells from collapsing from the often overwhelming task of meeting the many needs of the distressed self.

Of course the cumulative stressors experienced over a lifetime also take their toll, and a gradual loss of peripheral $\gamma\delta$ T cell pool is found to occur with age in the industrialized nations studied.^{113–115} The life events that affect $\gamma\delta$ T cell function are not limited to infectious and physiological insults, but also include psychological stress that especially mobilize terminally differentiated 'effector memory' cells characterized as being negative for cell surface marker CD27 but expressing CD45RA.¹¹⁶ This phenotype represents $\gamma\delta$ T cells that have already tackled some hurdles in their life and have reduced proliferative capacity, but are still able to pull the trigger given their highly cytolytic potential.¹¹⁷ Thus, differences in $\gamma\delta$ T cell numbers in relation to parameters such as gender and race that are inconsistent across the various populations studied^{89,113–115} may partly reflect the discrepant burdens that contribute to overall life stress experienced by members of these groups. For example, African Americans in Baltimore were found to have significantly lower peripheral blood $\gamma\delta$ T cells in comparison to their Caucasian counterparts;¹¹³ however, Africans from Ghana have significantly more $\gamma\delta$ T cells (of both the V δ 1 and V δ 2 T cell variety) in comparison to Europeans.⁸⁹ This observation makes it highly suspect that the racial difference observed in the North American cohort is due to an intrinsic property of African Americans to have fewer $\gamma\delta$ T cells, and it may reflect the different cumulative life stresses they face that take a toll on immune function.

'While there is tea, there is hope.'

Fortunately, there is a good amount of evidence suggesting that the adaptogenic properties of $\gamma\delta$ T cells can be protected and enhanced in those who need it most, namely those burdened with chronic stress. $\gamma\delta$ T cells seem particularly responsive to certain nutrient properties found in various food and supplement products, especially those with

Table 4 Bioactive compounds that have been studied to enhance $\gamma\delta$ T cell function

Compound	Reference
Tea (L-theonine)	69,123,127
Alkylamines	69,72,118
Heat-treated mistletoe	125
Plant tannins; procyanidins (e.g. from unripe apple peels)	121,122,129
Polysaccharides from Acai berry and Yamao	119,120
Concord grape juice and fruit/vegetable concentrate	126,128
Aged garlic	124

high antioxidant values (Table 4).^{69,118–127} In a double-blind placebo-controlled randomized control trial, capsules containing fruit and vegetable concentrate (containing, among other compounds, standardized levels of β -carotene, vitamin C, vitamin E, folate and calcium) were given to law school students, taken as a representative of young adults living with modern day stresses, over a period of 11 weeks.¹²⁸ $\gamma\delta$ T cells were found to be specifically increased (by about 30%; no change in circulating $\alpha\beta$ T cells) in parallel to an increase in the antioxidant capacity of the study subjects, and *in vitro* analyses showed that this effect was associated with a more tempered inflammatory response upon phorbol myristate acetate stimulation.¹²⁸ For some of these compounds with high antioxidant properties, their impact on $\gamma\delta$ T cells may be partly derived from their direct effect on the distress levels of cells that $\gamma\delta$ T cells are sensitive to. For example, treating renal carcinoma cells subjected to oxidative stress with the strong antioxidant, N-acetylcysteine, blunted the inflammatory response and activation of V γ 9V δ 2 T cells by downregulating the ectopic expression of the stress-induced molecule, MSH2.⁷⁹ Therefore, some of these immunomodulatory bioactive compounds, which include diverse items like procyanidins from unripe apple peels¹²⁹ and polysaccharide components of the Acai berry,¹¹⁹ their noted influence on $\gamma\delta$ T cells may be through the modulation of the type of stress signals displayed by cells that mobilize their activity. In contrast, other compounds, such as the aforementioned alkylamines (e.g. ethylamine, propylamine, butylamine and amylamine),⁶⁹ L-theonine from tea⁶⁹ and heat-treated mistletoe extracts,¹²⁵ the effect may be more direct and specific to human peripheral blood V γ 9V δ 2 T cells through the modulation of isoprenoid synthesis and the generation of pAgs. Understanding the mechanism by which $\gamma\delta$ T cell function is affected by various immunomodulatory compounds is central to formulating specific effective protocols for the targeted management of the various ailments these unique cells have the capacity to help treat and/or influence, including cancer, immunodeficiency and autoimmunity. Importantly, antioxidants could play an essential role in providing a buffer to prevent the untimely burn-out of these often over-worked but highly empathetic unconventional lymphocytes.

EPILOGUE

In light of their story, it can be appreciated that the difficulty in accurately defining $\gamma\delta$ T cells within existing prototypes stems from the fact that these T cells operate according to their own exclusive paradigm, as illustrated in Figure 1. In many ways they are the true 'adaptive' T cells as they are able to sense changes within the self that have the potential to disturb equilibrium, and they help the 'self' adapt as efficiently as possible to life's many challenges. However, unlike $\alpha\beta$ T cells, their own multifaceted but sensitive character remains relatively invariant in the face of the diverse stressors to which they help the self acclimatize. One strategy that $\gamma\delta$ T cells use to fulfill the complex nature of their task is to creatively merge the instincts of

the innate with the diversity of the adaptive immune systems. Thus, some of the antigens that they recognize are evolutionarily shared by the self and non-self and their subsequent response is contextually guided by the level of distress that they sense from within—often in the context of the contingent presence of other damage-associated molecular patterns. Given that different organisms are subjected to different types of stressors that impact their livelihood, it is not surprising that the phenotypic repertoire and distribution of $\gamma\delta$ T cells between species is quite diverse and tailored for the specific needs that they help meet. In fact, it was noted that substantial differences can be observed in the $\gamma\delta$ T cell fingerprint born between individuals even of the same species depending on the nature and extent of their life stress exposures—somewhat like the story held within the rings of a tree trunk. It still remains to be seen how these interindividual differences in $\gamma\delta$ T cell profiles are associated with health outcomes. For diseases such as cancer, a relationship between higher levels of circulating responsive $\gamma\delta$ T cells and a more favorable outcome has been observed;¹⁰⁸ however, there is still much to be learned about the potential contribution of $\gamma\delta$ T cells in providing general resiliency to malaise. An important challenge is to determine how to best support these intuitive T cells with their burden—some food for thought while destressing over a cup of afternoon tea.

COMPETING FINANCIAL INTERESTS

The authors have no conflict of interest to disclose.

ACKNOWLEDGEMENTS

SK is supported by a fellowship from the Alexander von Humboldt Foundation of Germany. DK acknowledges grant support from the Deutsche Forschungsgemeinschaft (DFG WE 3559/2-1) and is also supported by the 'Inflammation-at-Interfaces' Cluster of Excellence funded by the Deutsche Forschungsgemeinschaft (DFG).

- Saito H, Kranz DM, Takagaki Y, Hayday AC, Eisen HN, Tonegawa S. A third rearranged and expressed gene in a clone of cytotoxic T lymphocytes. *Nature* 1984; **312**: 36–40.
- Shin S, El-Diwany R, Schaffert S, Adams EJ, Garcia KC, Pereira P *et al*. Antigen recognition determinants of gammadelta T cell receptors. *Science* 2005; **308**: 252–255.
- Yamashita S, Tanaka Y, Harazaki M, Mikami B, Minato N. Recognition mechanism of non-peptide antigens by human gammadelta T cells. *Int Immunol* 2003; **15**: 1301–1307.
- Li H, Lebedeva MI, Llera AS, Fields BA, Brenner MB, Mariuzza RA. Structure of the Vdelta domain of a human gammadelta T-cell antigen receptor. *Nature* 1998; **391**: 502–506.
- Shao L, Huang D, Wei H, Wang RC, Chen CY, Shen L *et al*. Expansion, reexpansion, and recall-like expansion of Vgamma2Vdelta2 T cells in smallpox vaccination and monkeypox virus infection. *J Virol* 2009; **83**: 11959–11965.
- Morita CT, Jin C, Sarikonda G, Wang H. Nonpeptide antigens, presentation mechanisms, and immunological memory of human Vgamma2Vdelta2 T cells: discriminating friend from foe through the recognition of prenyl pyrophosphate antigens. *Immunol Rev* 2007; **215**: 59–76.
- Chen ZW, Letvin NL. Adaptive immune response of Vgamma2Vdelta2 T cells: a new paradigm. *Trends Immunol* 2003; **24**: 213–219.
- Rey J, Veuillen C, Vey N, Bouabdallah R, Olive D. Natural killer and gammadelta T cells in haematological malignancies: enhancing the immune effectors. *Trends Mol Med* 2009; **15**: 275–284.
- Shafi S, Vantourout P, Wallace G, Antoun A, Vaughan R, Stanford M *et al*. An NKG2D-mediated human lymphoid stress surveillance response with high interindividual variation. *Sci Transl Med* 2011; **3**: 113–124.
- Beetz S, Marischen L, Kabelitz D, Wesch D. Human gamma delta T cells: candidates for the development of immunotherapeutic strategies. *Immunol Res* 2007; **37**: 97–111.
- Wrobel P, Shojaei H, Schitteck B, Gieseler F, Wollenberg B, Kalthoff H *et al*. Lysis of a broad range of epithelial tumour cells by human gamma delta T cells: involvement of NKG2D ligands and T-cell receptor- versus NKG2D-dependent recognition. *Scand J Immunol* 2007; **66**: 320–328.
- Fisch P, Meuer E, Pende D, Rothenfusser S, Viale O, Kock S *et al*. Control of B cell lymphoma recognition via natural killer inhibitory receptors implies a role for human Vgamma9/Vdelta2 T cells in tumor immunity. *Eur J Immunol* 1997; **27**: 3368–3379.
- Halary F, Peyrat MA, Champagne E, Lopez-Botet M, Moretta A, Moretta L *et al*. Control of self-reactive cytotoxic T lymphocytes expressing gamma delta T cell receptors by natural killer inhibitory receptors. *Eur J Immunol* 1997; **27**: 2812–2821.
- Moris A, Rothenfusser S, Meuer E, Hangretinger R, Fisch P. Role of gammadelta T cells in tumor immunity and their control by NK receptors. *Microbes Infect* 1999; **1**: 227–234.
- Barakonyi A, Kovacs KT, Miko E, Szereday L, Varga P, Szekeres-Bartho J. Recognition of nonclassical HLA class I antigens by gamma delta T cells during pregnancy. *J Immunol* 2002; **168**: 2683–2688.
- Brandes M, Willmann K, Moser B. Professional antigen-presentation function by human gammadelta T Cells. *Science* 2005; **309**: 264–268.
- Brandes M, Willmann K, Bioley G, Levy N, Eberl M, Luo M *et al*. Cross-presenting human gammadelta T cells induce robust CD8⁺ alpha beta T cell responses. *Proc Natl Acad Sci USA* 2009; **106**: 2307–2312.
- Karre K. How to recognize a foreign submarine. *Immunol Rev* 1997; **155**: 5–9.
- Kabelitz D, Marischen L, Oberg HH, Holtmeier W, Wesch D. Epithelial defence by gamma delta T cells. *Int Arch Allergy Immunol* 2005; **137**: 73–81.
- Su C, Jakobsen I, Gu X, Nei M. Diversity and evolution of T-cell receptor variable region genes in mammals and birds. *Immunogenetics* 1999; **50**: 301–308.
- Qu A, Brulc JM, Wilson MK, Law BF, Theoret JR, Joens LA *et al*. Comparative metagenomics reveals host specific metavirulomes and horizontal gene transfer elements in the chicken cecum microbiome. *PLoS One* 2008; **3**: e2945.
- Hoover WH, Miller TK. Rumen digestive physiology and microbial ecology. *Vet Clin North Am Food Anim Pract* 1991; **7**: 311–325.
- Ross JA, Scott A, Gardner IC. Bacterial localization in the caecum of the rabbit. *Microbios* 1987; **52**: 51–63.
- Stevens CE, Hume ID. Contributions of microbes in vertebrate gastrointestinal tract to production and conservation of nutrients. *Physiol Rev* 1998; **78**: 393–427.
- Ismail AS, Severson KM, Vaishnava S, Behrendt CL, Yu X, Benjamin JL *et al*. Gammadelta intraepithelial lymphocytes are essential mediators of host-microbial homeostasis at the intestinal mucosal surface. *Proc Natl Acad Sci USA* 2011; **108**: 8743–8748.
- Ismail AS, Behrendt CL, Hooper LV. Reciprocal interactions between commensal bacteria and gamma delta intraepithelial lymphocytes during mucosal injury. *J Immunol* 2009; **182**: 3047–3054.
- Chodaczek G, Papanna V, Zal MA, Zal T. Body-barrier surveillance by epidermal gammadelta TCRs. *Nat Immunol* 2012; **13**: 272–282.
- Jameson J, Havran WL. Skin gammadelta T-cell functions in homeostasis and wound healing. *Immunol Rev* 2007; **215**: 114–122.
- Jameson JM, Cauvi G, Witherden DA, Havran WL. A keratinocyte-responsive gamma delta TCR is necessary for dendritic epidermal T cell activation by damaged keratinocytes and maintenance in the epidermis. *J Immunol* 2004; **172**: 3573–3579.
- Boismenu R, Havran WL. Gammadelta T cells in host defense and epithelial cell biology. *Clin Immunol Immunopathol* 1998; **86**: 121–133.
- Havran WL, Chen Y, Boismenu R. Innate functions of epithelial gamma delta T cells. *Adv Exp Med Biol* 1998; **452**: 29–35.
- McVay LD, Carding SR. Generation of human gammadelta T-cell repertoires. *Crit Rev Immunol* 1999; **19**: 431–460.
- Halary F, Pitard V, Dlubek D, Krzysiek R, de la Salle H, Merville P *et al*. Shared reactivity of V(delta)2(neg) (gamma)(delta) T cells against cytomegalovirus-infected cells and tumor intestinal epithelial cells. *J Exp Med* 2005; **201**: 1567–1578.
- Locke NR, Stankovic S, Funda DP, Harrison LC. TCR gamma delta intraepithelial lymphocytes are required for self-tolerance. *J Immunol* 2006; **176**: 6553–6559.
- Holtmeier W. Compartmentalization gamma/delta T cells and their putative role in mucosal immunity. *Crit Rev Immunol* 2003; **23**: 473–488.
- Hayday A, Gibbons D. Brokering the peace: the origin of intestinal T cells. *Mucosal Immunol* 2008; **1**: 172–174.
- Trejdosiewicz LK. Intestinal intraepithelial lymphocytes and lymphoepithelial interactions in the human gastrointestinal mucosa. *Immunol Lett* 1992; **32**: 13–19.
- Zocchi MR, Poggi A. Role of gammadelta T lymphocytes in tumor defense. *Front Biosci* 2004; **9**: 2588–2604.
- Maeurer MJ, Martin D, Walter W, Liu K, Zitvogel L, Halusczyk K *et al*. Human intestinal Vdelta1⁺ lymphocytes recognize tumor cells of epithelial origin. *J Exp Med* 1996; **183**: 1681–1696.
- Bhagat G, Naiyer AJ, Shah JG, Harper J, Jabri B, Wang TC *et al*. Small intestinal CD8⁺ TCRgammadelta⁺ NKG2A⁺ intraepithelial lymphocytes have attributes of regulatory cells in patients with celiac disease. *J Clin Invest* 2008; **118**: 281–293.
- Holtmeier W, Kabelitz D. gammadelta T cells link innate and adaptive immune responses. *Chem Immunol Allergy* 2005; **86**: 151–183.
- Groh V, Steinle A, Bauer S, Spies T. Recognition of stress-induced MHC molecules by intestinal epithelial gammadelta T cells. *Science* 1998; **279**: 1737–1740.
- Groh V, Rhinehart R, Secrist H, Bauer S, Grabstein KH, Spies T. Broad tumor-associated expression and recognition by tumor-derived gamma delta T cells of MICA and MICB. *Proc Natl Acad Sci USA* 1999; **96**: 6879–6884.
- Rincon-Orozco B, Kunzmann V, Wrobel P, Kabelitz D, Steinle A, Herrmann T. Activation of V gamma 9V delta 2 T cells by NKG2D. *J Immunol* 2005; **175**: 2144–2151.
- Vivier E, Tomasello E, Paul P. Lymphocyte activation via NKG2D: towards a new paradigm in immune recognition? *Curr Opin Immunol* 2002; **14**: 306–311.
- Xu B, Pizarro JC, Holmes MA, McBeth C, Groh V, Spies T *et al*. Crystal structure of a gammadelta T-cell receptor specific for the human MHC class I homolog MICA. *Proc Natl Acad Sci USA* 2011; **108**: 2414–2419.

- 47 Haig NA, Guan Z, Li D, McMichael A, Raetz CR, Xu XN. Identification of self-lipids presented by CD1c and CD1d proteins. *J Biol Chem* 2011; **286**: 37692–37701.
- 48 Spada FM, Grant EP, Peters PJ, Sugita M, Melian A, Leslie DS *et al*. Self-recognition of CD1 by gamma/delta T cells: implications for innate immunity. *J Exp Med* 2000; **191**: 937–948.
- 49 Willcox CR, Pitard V, Netzer S, Couzi L, Salim M, Silberzahn T, *et al*. Cytomegalovirus and tumor stress surveillance by binding of a human gammadelta T cell antigen receptor to endothelial protein C receptor. *Nat Immunol* 2012; **13**: 872–879.
- 50 Montes R, Puy C, Molina E, Hermida J. Is EPCR a multi-ligand receptor? Pros and cons. *Thromb Haemost* 2012; **107**: 815–826.
- 51 Mincheva-Nilsson L, Hammarstrom S, Hammarstrom ML. Human decidua leukocytes from early pregnancy contain high numbers of gamma delta+ cells and show selective down-regulation of alloreactivity. *J Immunol* 1992; **149**: 2203–2211.
- 52 Bonney EA, Pudney J, Anderson DJ, Hill JA. Gamma-delta T cells in midgestation human placental villi. *Gynecol Obstet Invest* 2000; **50**: 153–157.
- 53 Nagaeva O, Jonsson L, Mincheva-Nilsson L. Dominant IL-10 and TGF-beta mRNA expression in gamma delta T cells of human early pregnancy decidua suggests immunoregulatory potential. *Am J Reprod Immunol* 2002; **48**: 9–17.
- 54 Fan DX, Duan J, Li MQ, Xu B, Li DJ, Jin LP. The decidua gamma-delta T cells up-regulate the biological functions of trophoblasts via IL-10 secretion in early human pregnancy. *Clin Immunol* 2011; **141**: 284–292.
- 55 Hayakawa S, Shiraishi H, Saitoh S, Satoh K. Decidua as a site of extrathymic V gamma I T-cell differentiation. *Am J Reprod Immunol* 1996; **35**: 233–238.
- 56 Mincheva-Nilsson L, Kling M, Hammarstrom S, Nagaeva O, Sundqvist KG, Hammarstrom ML *et al*. Gamma delta T cells of human early pregnancy decidua: evidence for local proliferation, phenotypic heterogeneity, and extrathymic differentiation. *J Immunol* 1997; **159**: 3266–3277.
- 57 Meeusen E, Fox A, Brandon M, Lee CS. Activation of uterine intraepithelial gamma delta T cell receptor-positive lymphocytes during pregnancy. *Eur J Immunol* 1993; **23**: 1112–1117.
- 58 Heyborne KD, Cranfill RL, Carding SR, Born WK, O'Brien RL. Characterization of gamma delta T lymphocytes at the maternal-fetal interface. *J Immunol* 1992; **149**: 2872–2878.
- 59 Barakonyi A, Miko E, Varga P, Szekeres-Bartho J. V-chain preference of gamma/delta T-cell receptors in peripheral blood during term labor. *Am J Reprod Immunol* 2008; **59**: 201–205.
- 60 Polgar B, Barakonyi A, Xynos I, Szekeres-Bartho J. The role of gamma/delta T cell receptor positive cells in pregnancy. *Am J Reprod Immunol* 1999; **41**: 239–244.
- 61 Barakonyi A, Polgar B, Szekeres-Bartho J. The role of gamma/delta T-cell receptor-positive cells in pregnancy: part II. *Am J Reprod Immunol* 1999; **42**: 83–87.
- 62 Szereday L, Barakonyi A, Miko E, Varga P, Szekeres-Bartho J. gamma/delta T-cell subsets, NKG2A expression and apoptosis of V delta 2+ T cells in pregnant women with or without risk of premature pregnancy termination. *Am J Reprod Immunol* 2003; **50**: 490–496.
- 63 Heyborne K, Fu YX, Nelson A, Farr A, O'Brien R, Born W. Recognition of trophoblasts by gamma delta T cells. *J Immunol* 1994; **153**: 2918–2926.
- 64 Wang H, Lee HK, Bukowski JF, Li H, Mariuzza RA, Chen ZW *et al*. Conservation of nonpeptide antigen recognition by rhesus monkey V gamma 2V delta 2 T cells. *J Immunol* 2003; **170**: 3696–3706.
- 65 Tanaka Y, Morita CT, Tanaka Y, Nieves E, Brenner MB, Bloom BR. Natural and synthetic non-peptide antigens recognized by human gamma delta T cells. *Nature* 1995; **375**: 155–158.
- 66 Jomaa H, Feurle J, Luhs K, Kunzmann V, Tony HP, Herderich M *et al*. Vgamma9/Vdelta2 T cell activation induced by bacterial low molecular mass compounds depends on the 1-deoxy-D-xylulose 5-phosphate pathway of isoprenoid biosynthesis. *FEMS Immunol Med Microbiol* 1999; **25**: 371–378.
- 67 Hintz M, Reichenberg A, Altincicek B, Bahr U, Gschwind RM, Kollas AK *et al*. Identification of (E)-4-hydroxy-3-methyl-but-2-enyl pyrophosphate as a major activator for human gammadelta T cells in *Escherichia coli*. *FEBS Lett* 2001; **509**: 317–322.
- 68 Gober HJ, Kistowska M, Angman L, Jenó P, Mori L, de Libero G. Human T cell receptor gammadelta cells recognize endogenous mevalonate metabolites in tumor cells. *J Exp Med* 2003; **197**: 163–168.
- 69 Bukowski JF, Morita CT, Brenner MB. Human gamma delta T cells recognize alkylamines derived from microbes, edible plants, and tea: implications for innate immunity. *Immunity* 1999; **11**: 57–65.
- 70 Kunzmann V, Bauer E, Feurle J, Weissinger F, Tony HP, Wilhelm M. Stimulation of gammadelta T cells by aminobisphosphonates and induction of antiplasma cell activity in multiple myeloma. *Blood* 2000; **96**: 384–392.
- 71 Thompson K, Rogers MJ. Statins prevent bisphosphonate-induced gamma,delta-T-cell proliferation and activation *in vitro*. *J Bone Miner Res* 2004; **19**: 278–288.
- 72 Thompson K, Rojas-Navea J, Rogers MJ. Alkylamines cause Vgamma9Vdelta2 T-cell activation and proliferation by inhibiting the mevalonate pathway. *Blood* 2006; **107**: 651–654.
- 73 Green AE, Lissina A, Hutchinson SL, Hewitt RE, Temple B, James D *et al*. Recognition of nonpeptide antigens by human V gamma 9V delta 2 T cells requires contact with cells of human origin. *Clin Exp Immunol* 2004; **136**: 472–482.
- 74 Scotet E, Martinez LO, Grant E, Barbaras R, Jenó P, Guiraud M *et al*. Tumor recognition following Vgamma9Vdelta2 T cell receptor interactions with a surface F1-ATPase-related structure and apolipoprotein A-I. *Immunity* 2005; **22**: 71–80.
- 75 Fu Y, Hou Y, Fu C, Gu M, Li C, Kong W *et al*. A novel mechanism of gamma/delta T-lymphocyte and endothelial activation by shear stress: the role of ecto-ATP synthase beta chain. *Circ Res* 2011; **108**: 410–417.
- 76 Mookerjee-Basu J, Vantourout P, Martinez LO, Perret B, Collet X, Perigaud C *et al*. F1-adenosine triphosphatase displays properties characteristic of an antigen presentation molecule for Vgamma9Vdelta2 T cells. *J Immunol* 2010; **184**: 6920–6928.
- 77 Chen H, He X, Wang Z, Wu D, Zhang H, Xu C *et al*. Identification of human T cell receptor gammadelta-recognized epitopes/proteins via CD3delta peptide-based immunobiochemical strategy. *J Biol Chem* 2008; **283**: 12528–12537.
- 78 Dai Y, Chen H, Mo C, Cui L, He W. Ectopically expressed human tumor biomarker MutS homologue 2 is a novel endogenous ligand that is recognized by human gammadelta t cells to induce innate anti-tumor/virus immunity. *J Biol Chem* 2012; **287**: 16812–16819.
- 79 Mo C, Dai Y, Kang N, Cui L, He W. Ectopic expression of human MutS homologue 2 on renal carcinoma cells is induced by oxidative stress with interleukin-18 promotion via p38 mitogen-activated protein kinase (MAPK) and c-Jun N-terminal kinase (JNK) signaling pathways. *J Biol Chem* 2012; **287**: 19242–19254.
- 80 Heinen CD, Cyr JL, Cook C, Punja N, Sakato M, Forties RA *et al*. Human MSH2 (hMSH2) protein controls ATP processing by hMSH2-hMSH6. *J Biol Chem* 2011; **286**: 40287–40295.
- 81 Harly C, Guillaume Y, Nedellec S, Peigne CM, Monkonen H, Monkonen J *et al*. Key implication of CD277/Butyrophilin-3 (BTN3A) in cellular stress sensing by a major human gammadelta T cell subset. *Blood* 2012; **120**: 2269–2279.
- 82 Rhodes DA, Stammers M, Malcherek G, Beck S, Trowsdale J. The cluster of BTN genes in the extended major histocompatibility complex. *Genomics* 2001; **71**: 351–362.
- 83 Carding SR, Kyles S, Jenkinson EJ, Kingston R, Bottomly K, Owen JJ *et al*. Developmentally regulated fetal thymic and extrathymic T-cell receptor gamma delta gene expression. *Genes Dev* 1990; **4**: 1304–1315.
- 84 McVay LD, Carding SR. Extrathymic origin of human gamma delta T cells during fetal development. *J Immunol* 1996; **157**: 2873–2882.
- 85 de Rosa SC, Andrus JP, Perfetto SP, Mantovani JJ, Herzenberg LA, Herzenberg LA *et al*. Ontogeny of gamma delta T cells in humans. *J Immunol* 2004; **172**: 1637–1645.
- 86 Esin S, Shigematsu M, Nagai S, Eklund A, Wigzell H, Grunewald J. Different percentages of peripheral blood gamma delta+ T cells in healthy individuals from different areas of the world. *Scand J Immunol* 1996; **43**: 593–596.
- 87 Romagnani S. The increased prevalence of allergy and the hygiene hypothesis: missing immune deviation, reduced immune suppression, or both? *Immunology* 2004; **112**: 352–363.
- 88 Garn H, Renz H. Epidemiological and immunological evidence for the hygiene hypothesis. *Immunobiology* 2007; **212**: 441–452.
- 89 Hviid L, Akanmori BD, Loizon S, Kurtzhals JA, Ricke CH, Lim A *et al*. High frequency of circulating gamma delta T cells with dominance of the v(delta)1 subset in a healthy population. *Int Immunol* 2000; **12**: 797–805.
- 90 Hviid L, Kurtzhals JA, Adabayeri V, Loizon S, Kemp K, Goka BQ *et al*. Perturbation and proinflammatory type activation of V delta 1+ gamma delta T cells in African children with Plasmodium falciparum malaria. *Infect Immunol* 2001; **69**: 3190–3196.
- 91 Ho M, Tongtawe P, Kriangkum J, Wimonwattawate T, Pattanapanyasat K, Bryant L *et al*. Polyclonal expansion of peripheral gamma delta T cells in human Plasmodium falciparum malaria. *Infect Immunol* 1994; **62**: 855–862.
- 92 Vermijlen D, Brouwer M, Donner C, Liesnard C, Tackoen M, van Rysselberge M *et al*. Human cytomegalovirus elicits fetal gammadelta T cell responses in utero. *J Exp Med* 2010; **207**: 807–821.
- 93 Orsini DL, Res PC, van Laar JM, Muller LM, Soprano AE, Kooy YM *et al*. A subset of V delta 1+ T cells proliferates in response to Epstein-Barr virus-transformed B cell lines *in vitro*. *Scand J Immunol* 1993; **38**: 335–340.
- 94 Orsini DL, van Gils M, Kooy YM, Struyk L, Klein G, van den Elsen P *et al*. Functional and molecular characterization of B cell-responsive V delta 1+ gamma delta T cells. *Eur J Immunol* 1994; **24**: 3199–3204.
- 95 Zheng NN, McElrath MJ, Sow PS, Meshner A, Hawes SE, Stern J *et al*. Association between peripheral gammadelta T-cell profile and disease progression in individuals infected with HIV-1 or HIV-2 in West Africa. *J Acquir Immune Defic Syndr* 2011; **57**: 92–100.
- 96 Rossol R, Dobmeyer JM, Dobmeyer TS, Klein SA, Rossol S, Wesch D *et al*. Increase in Vdelta1+ gammadelta T cells in the peripheral blood and bone marrow as a selective feature of HIV-1 but not other virus infections. *Br J Haematol* 1998; **100**: 728–734.
- 97 Oswald E, Fisch P, Jakob T, Bruckner-Tuderman L, Martin SF, Rensing-Ehl A. Reduced numbers of circulating gammadelta T cells in patients with bullous pemphigoid. *Exp Dermatol* 2009; **18**: 991–993.
- 98 Andreu-Ballester JC, Amigo-Garcia V, Catalan-Serra I, Gil-Borras R, Ballester F, Almela-Quilis A *et al*. Deficit of gammadelta T lymphocytes in the peripheral blood of patients with Crohn's disease. *Dig Dis Sci* 2011; **56**: 2613–2622.
- 99 Kiladjian JJ, Visentin G, Viey E, Chevret S, Eclache V, Stirnemann J *et al*. Activation of cytotoxic T-cell receptor gammadelta T lymphocytes in response to specific stimulation in myelodysplastic syndromes. *Haematologica* 2008; **93**: 381–389.
- 100 Chauhan SK, Tripathy NK, Sinha N, Nityanand S. T-cell receptor repertoire of circulating gamma delta T-cells in Takayasu's arteritis. *Clin Immunol* 2006; **118**: 243–249.
- 101 Giacomelli R, Matucci-Cerinic M, Cipriani P, Ghersetich I, Lattanzio R, Pavan A *et al*. Circulating Vdelta1+ T cells are activated and accumulate in the skin of systemic sclerosis patients. *Arthritis Rheum* 1998; **41**: 327–334.
- 102 Gan YH, Lui SS, Malkovsky M. Differential susceptibility of naive and activated human gammadelta T cells to activation-induced cell death by T-cell receptor cross-linking. *Mol Med* 2001; **7**: 636–643.
- 103 Gan YH, Wallace M, Malkovsky M. Fas-dependent, activation-induced cell death of gammadelta cells. *J Biol Regul Homeost Agents* 2001; **15**: 277–285.

- 104 Kabelitz D. Effector functions and control of human gammadelta T-cell activation. *Microbes Infect* 1999; **1**: 255–261.
- 105 Kelsen J, Dige A, Schwindt H, D'Amore F, Pedersen FS, Agnholt J *et al*. Infliximab induces clonal expansion of gammadelta-T cells in Crohn's disease: a predictor of lymphoma risk? *PLoS One* 2011; **6**: e17890.
- 106 Giardina AR, Accardo-Palumbo A, Ciccia F, Ferrante A, Principato A, Impastato R *et al*. Blocking TNF *in vitro* with infliximab determines the inhibition of expansion and interferon gamma production of Vgamma9/Vdelta2 T lymphocytes from patients with active rheumatoid arthritis. A role in the susceptibility to tuberculosis? *Reumatismo* 2009; **61**: 21–26.
- 107 Accardo-Palumbo A, Giardina AR, Ciccia F, Ferrante A, Principato A, Impastato R *et al*. Phenotype and functional changes of Vgamma9/Vdelta2 T lymphocytes in Behcet's disease and the effect of infliximab on Vgamma9/Vdelta2 T cell expansion, activation and cytotoxicity. *Arthritis Res Ther* 2010; **12**: R109.
- 108 Kalyan S, Wesch D, Kabelitz D. Aminobisphosphonates and Toll-like receptor ligands: recruiting Vgamma9/Vdelta2 T cells for the treatment of hematologic malignancy. *Curr Med Chem* 2011; **18**: 5206–5216.
- 109 Kabelitz D. Human gammadelta T lymphocytes for immunotherapeutic strategies against cancer. *F1000 Med Rep* 2010; **2**: 45.
- 110 Kabelitz D, Wesch D, He W. Perspectives of gammadelta T cells in tumor immunology. *Cancer Res* 2007; **67**: 5–8.
- 111 Martinet L, Poupot R, Fournie JJ. Pitfalls on the roadmap to gammadelta T cell-based cancer immunotherapies. *Immunol Lett* 2009; **124**: 1–8.
- 112 Lopez RD. Human gammadelta-T cells in adoptive immunotherapy of malignant and infectious diseases. *Immunol Res* 2002; **26**: 207–221.
- 113 Cairo C, Armstrong CL, Cummings JS, Deetz CO, Tan M, Lu C *et al*. Impact of age, gender, and race on circulating gammadelta T cells. *Hum Immunol* 2010; **71**: 968–975.
- 114 Caccamo N, Dieli F, Wesch D, Jomaa H, Eberl M. Sex-specific phenotypical and functional differences in peripheral human Vgamma9/Vdelta2 T cells. *J Leukoc Biol* 2006; **79**: 663–666.
- 115 Michishita Y, Hirokawa M, Guo YM, Abe Y, Liu J, Ubukawa K *et al*. Age-associated alteration of gammadelta T-cell repertoire and different profiles of activation-induced death of Vdelta1 and Vdelta2 T cells. *Int J Hematol* 2011; **94**: 230–240.
- 116 Anane LH, Edwards KM, Burns VE, Zanten JJ, Drayson MT, Bosch JA. Phenotypic characterization of gammadelta T cells mobilized in response to acute psychological stress. *Brain Behav Immun* 2010; **24**: 608–614.
- 117 Dieli F, Poccia F, Lipp M, Sireci G, Caccamo N, di Sano C *et al*. Differentiation of effector/memory Vdelta2 T cells and migratory routes in lymph nodes or inflammatory sites. *J Exp Med* 2003; **198**: 391–397.
- 118 Percival SS, Bukowski JF, Milner J. Bioactive food components that enhance gammadelta T cell function may play a role in cancer prevention. *J Nutr* 2008; **138**: 1–4.
- 119 Holderness J, Schepetkin IA, Freedman B, Kirpotina LN, Quinn MT, Hedges JF *et al*. Polysaccharides isolated from Acai fruit induce innate immune responses. *PLoS One* 2011; **6**: e17301.
- 120 Graff JC, Kimmel EM, Freedman B, Schepetkin IA, Holderness J, Quinn MT *et al*. Polysaccharides derived from Yamao (*Funtumia elastica*) prime gammadelta T cells *in vitro* and enhance innate immune responses *in vivo*. *Int Immunopharmacol* 2009; **9**: 1313–1322.
- 121 Holderness J, Jackiw L, Kimmel E, Kerns H, Radke M, Hedges JF *et al*. Select plant tannins induce IL-2Ralpha up-regulation and augment cell division in gammadelta T cells. *J Immunol* 2007; **179**: 6468–6478.
- 122 Daughenbaugh KF, Holderness J, Graff JC, Hedges JF, Freedman B, Graff JW *et al*. Contribution of transcript stability to a conserved procyanidin-induced cytokine response in gammadelta T cells. *Genes Immun* 2011; **12**: 378–389.
- 123 Rowe CA, Nantz MP, Bukowski JF, Percival SS. Specific formulation of *Camellia sinensis* prevents cold and flu symptoms and enhances gamma,delta T cell function: a randomized, double-blind, placebo-controlled study. *J Am Coll Nutr* 2007; **26**: 445–452.
- 124 Nantz MP, Rowe CA, Muller CE, Creasy RA, Stanilka JM, Percival SS. Supplementation with aged garlic extract improves both NK and gammadelta-T cell function and reduces the severity of cold and flu symptoms: A randomized, double-blind, placebo-controlled nutrition intervention. *Clin Nutr* 2012; **31**: 337–344.
- 125 Fischer S, Scheffler A, Kabelitz D. Activation of human gamma delta T-cells by heat-treated mistletoe plant extracts. *Immunol Lett* 1996; **52**: 69–72.
- 126 Rowe CA, Nantz MP, Nieves C Jr, West RL, Percival SS. Regular consumption of concord grape juice benefits human immunity. *J Med Food* 2011; **14**: 69–78.
- 127 Kamath AB, Wang L, Das H, Li L, Reinhold VN, Bukowski JF. Antigens in tea-beverage prime human Vgamma 2Vdelta 2 T cells *in vitro* and *in vivo* for memory and nonmemory antibacterial cytokine responses. *Proc Natl Acad Sci USA* 2003; **100**: 6009–6014.
- 128 Nantz MP, Rowe CA, Nieves C Jr, Percival SS. Immunity and antioxidant capacity in humans is enhanced by consumption of a dried, encapsulated fruit and vegetable juice concentrate. *J Nutr* 2006; **136**: 2606–2610.
- 129 Holderness J, Hedges JF, Daughenbaugh K, Kimmel E, Graff J, Freedman B *et al*. Response of gamma delta T cells to plant-derived tannins. *Crit Rev Immunol* 2008; **28**: 377–402.