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REVIEW

Diversity of $\gamma\delta$ T-cell antigens

Willi K Born^{1,2}, M Kemal Aydintug¹ and Rebecca L O'Brien^{1,2}

In the last two decades, it has become clear that $\gamma\delta$ T cells recognize a diverse array of antigens including self and foreign, large and small, and peptidic and non-peptidic molecules. In this respect, $\gamma\delta$ antigens as a whole resemble more the antigens recognized by antibodies than those recognized by $\alpha\beta$ T cells. Because of this antigenic diversity, no single mechanism—such as the major histocompatibility complex (MHC) restriction of $\alpha\beta$ T cells—is likely to provide a basis for all observed T-cell antigen receptor (TCR)-dependent $\gamma\delta$ T-cell responses. Furthermore, available evidence suggests that many individual $\gamma\delta$ T cells are poly-specific, probably using different modes of ligand recognition in their responses to unrelated antigens. While posing a unique challenge in the maintenance of self-tolerance, this broad reactivity pattern might enable multiple overlapping uses of $\gamma\delta$ T-cell populations, and thus generate a more efficient immune response.

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INTRODUCTION

Determining which antigens (Ags) $\gamma\delta$ T cells recognize with their Tcell antigen receptors (TCRs) arguably remains the biggest challenge in the field. Significant progress has been made during the last several years in collecting individual examples of Ag responses of $\gamma\delta$ T cells and, in some cases, in delineating the mechanism of Ag recognition. The upshot of these studies is that $\gamma\delta$ T cells recognize diverse Ags, quite different from the uniform, major histocompatibility complex (MHC)-packaged protein fragments recognized by $\alpha\beta$ T cells, and more like the highly variable and diverse Ags recognized by B cells (Figure 1). This observation, along with a structural comparison of the $\gamma\delta$ TCRs with immunoglobulins, encouraged the hypothesis that ligand recognition by $\gamma\delta$ T cells is B cell-like.¹ However, despite obvious similarities, the $\gamma\delta$ TCRs are different from the B-cell receptors (BCRs) in structure and composition, and chances are that Ag recognition by $\gamma\delta$ T cells is a unique feature of this lymphocyte type, even if it resembles in some aspects Ag recognition by other lymphocytes. In this review, we will examine the diversity of Ags recognized by $\gamma \delta$ T cells, as well as cellular distributions of $\gamma\delta$ specificities, and possible consequences for self-tolerance in comparison with $\alpha\beta$ T cells and B cells.

THE $\gamma\delta$ T-CELL RECEPTORS

The $\gamma\delta$ TCRs are encoded by two distinct sets of genes, γ and δ , which rearrange like the Ig genes to form templates for diverse TCR protein molecules.² Although there are fewer $V\gamma$ and $V\delta$ genes than IgV and TCR $\alpha\beta$ V genes, the recombinatorial possibilities for generating the $\gamma\delta$ TCRs are almost infinite,³ largely due to the unique ability of the δ genes to rearrange D segments in tandem and to utilize all three reading frames. This creates multiple joints in the same single gene, each with the opportunity for N region additions and trimming. However,

this unique mechanism focuses potential $\gamma\delta$ TCR diversity on a relatively small region of the TCR surface, CDR3 δ . The significance of this narrow focus is not yet clear as will be further discussed below. Despite the potential for diversity, some commonly occurring $\gamma\delta$ TCRs are invariant,⁴ or nearly invariant, which has led to the concept of $\gamma \delta$ TCRs as innate pattern recognition receptors. However, other $\gamma\delta$ TCRs are diverse initially, and later selected during immune responses.^{5,6} At the level of lymphocyte populations, the $\gamma\delta$ TCRs appear to be less evenly distributed than the other adaptive receptors. In mice, where this has been studied in detail, subsets of $\gamma\delta$ T cells expressing different V γs become segregated already during ontogeny due to their sequential developmental patterns in the thymus.⁷ This temporal segregation may also dictate their spacial partition, as cells expressing different V γ s colonize different peripheral tissues.² γ δ T cells expressing different V γ s have different effects on the immune responses.⁸ Recent studies revealed a correlation between $TCR-V\gamma$ expression and functional differentiation, $5^{9,10}$ thus emphasizing the biological significance of the $\gamma\delta$ TCR segregation in the mouse. Nevertheless, specific evidence concerned with IL-17 producing $\gamma \delta$ T cells suggests that, at least in this case, the correlation between $\gamma\delta$ TCR expression and function is merely coincidental—waves of $\gamma\delta$ T cells expressing certain TCRs coincide with a thymic environment that temporarily favors TH17 differentiation.¹¹ Although there is still less experimental evidence, segregation of TCR-defined $\gamma\delta$ T-cell subsets (both spacial and functional) has also been found in other species and might well be a universal feature of these cells. With regard to Ag specificity, one immediate consequence of this is a partial repertoire restriction, and the association of certain repertoires with certain functions. This might be necessary in maintaining self-tolerance, or to channel the responses of $\gamma\delta$ T cells into patterns with predetermined outcomes. In any case, it suggests a degree of rigidity of the repertoire that seems to

¹Integrated Department of Immunology, National Jewish Health, Denver, CO, USA and ²University of Colorado Denver, Aurora, CO, USA Correspondence: Dr WK Born, National Jewish Health, 1400 Jackson Street, GB K409, Denver, CO 80206, USA. E-mail: bornw@njhealth.org

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Figure 1 γ δ T cells recognize a wide range of structurally different ligands. Unlike most $\alpha\beta$ T cells, $\gamma\delta$ T cells recognize ligands that vary much in size, composition and molecular structure, including MHC and non-MHC cell surface molecules, soluble proteins and smaller peptides, phospholipids, prenyl pyrophosphates and sulfatide. Some of these molecules appear to be recognized in complex with others (e.g. certain phospholipids in complex with CD1d or apo H), and most if not all are recognized while bound to or expressed on the cell surface. apo H, apolipoprotein H; MHC, major histocompatibility complex.

be absent in B cells. Along the same vein, there is no evidence of somatic mutation in the $\gamma\delta$ TCRs. Therefore, any affinity maturation of $\gamma\delta$ T cells has to be driven by CDR3 selection, although $\gamma\delta$ TCR– ligand interactions described so far are low affinity by comparison with class-switched BCRs. On the other hand, ligand recognition by $\gamma\delta$ T cells is not limited to MHC-presented peptides. Affinities of TCR–ligand interactions seem to vary considerably, and in some reported cases were of higher affinity than those of MHC-restricted $\alpha\beta$ TCRs. In this regard and by comparison with $\alpha\beta$ T cells at least, the $\gamma\delta$ responses show more variation and plasticity.¹²

The density of cell surface-expressed $\gamma\delta$ TCRs is generally similar to that of $\alpha\beta$ TCRs, but there are differences between the two types of TCR in terms of their association with proteins of the CD3 complex. Of note, murine $\gamma\delta$ TCR complexes lack CD3 δ , and upon activation instead express FceR1 γ , in contrast to the $\alpha\beta$ TCRs and to human $\gamma\delta$ TCRs.¹³ At least some of the $\gamma\delta$ TCRs appear to signal more efficiently than the $\alpha\beta$ TCRs, which is described in detail elsewhere.^{13,14} This might lower the activation threshold for certain $\gamma \delta$ T cells. Nevertheless, the $\gamma\delta$ TCRs overall resemble the other cell surfaceexpressed adaptive lymphocyte receptors, the BCRs and the $\alpha\beta$ TCRs, suggesting that they provide similar sensitivities to the lymphocytes that carry them.

MECHANISM OF LIGAND RECOGNITION

We already mentioned the hypothesis, proposed by Chien and collaborators,¹ that ligand recognition by $\gamma\delta T$ cells is B cell-like. This idea is based on structural and functional observations. Thus, CDR3 regions of the $\gamma\delta$ TCRs resemble Ig CDRs in terms of length and variability.¹⁵ As is the case with Ig heavy and light chains, respectively, $TCR-\delta$ has extensive CDR3s whereas TCR- γ has short CDR3s. Moreover, TCR- δ CDR3s are far more diverse, similar to IgH CDR3s. In contrast, TCR-a and TCR- β CDR3s are intermediate in length, and similar to each other both in length and diversity, which may be a requirement for the docking on the surface of MHC molecules and the recognition of MHC-bound small peptides. However, with $\gamma\delta$ TCRs, most of the potential for diversity is concentrated in CDR3d, while BCRs are

diverse in CDR1 and 2 as well, and have, in addition, the option of affinity maturation through somatic mutation, which implies large differences in ligand recognition. Not only will B cells recognize a wider array of ligands, but also will they interact with them through higher affinity.

Other observations support the idea of B cell-like ligand recognition by $\gamma\delta$ T cells, but they also illustrate differences. Thus, there is a conspicuous absence of reports of MHC-restricted Ag recognition by $\gamma\delta$ T cells,¹⁶ which is the main mode of ligand recognition by $\alpha\beta$ T cells. Possible exceptions to this observation will be discussed below. However, in some cases it has been shown that Ags must be presented to be stimulatory for $\gamma\delta$ T cells.¹⁷ These presented Ags tend to be small, and in soluble form might not be capable of TCR cross-linking, a prerequisite for activation via immunoreceptors. Whether any Ags in solution can be recognized and can trigger responses of $\gamma\delta$ T cells is not yet known, although this seems likely when such Ags are multivalent.¹⁸ An example might be the response to an insulin peptide, which can be elicited from isolated single hybridoma cells (in the absence of antigen-presenting cells (APCs) or other hybridoma cells) expressing an insulin peptide-reactive $\gamma \delta$ TCR.¹⁹ Whether responses to cell surface-expressed molecules such as CD1c, CD1d, MICA/B and T10/22 have a special significance in $\gamma\delta$ TCR-mediated ligand recognition remains unclear. Unlike the $\alpha\beta$ TCRs, which have an inherent bias for MHC recognition associated with certain dedicated amino acids,^{20,21} no such bias has been reported for the $\gamma\delta$ TCRs. In fact, judging from the interaction of T10/22-reactive $\gamma\delta$ TCRs with their ligand, where specificity is largely determined by a single D segment within TCR- δ ,²² there is no reason to expect a similar bias for the $\gamma\delta$ TCRs. Similarly, no inherent MHC bias seems to exist with the BCRs. However, it remains possible that $\gamma\delta$ TCRs have inherent biases for the recognition of cell surface molecules other than MHC,²³ and given the limitation of the repertoire outside of CDR3 δ , this even seems likely.²⁴ No such bias or restricting element has been firmly established, however. Perhaps the biggest difference to Ag recognition via BCRs is that so many conventional Ags seem to be incapable of eliciting responses by $\gamma\delta$ T cells. To our knowledge, specific TCR-mediated responses of $\gamma\delta$ T cells have not been elicited to Ags such as ovalbumin, hen egg lysozyme, cytochrome C and many others, all of which are recognized by antibodies. This is clearly not due to an inability of $\gamma \delta$ T cells to recognize proteins—in fact, there may be more proteinaceous than non-proteinaceous ligands for the $\gamma\delta$ TCRs. Nor is it due to an inability of $\gamma\delta$ T cells to undergo clonal selection following immunization—there are well-documented examples of such selection among peripheral $\gamma\delta$ T cells. It may have to do, however, with the fact that large portions of the $\gamma\delta$ TCR are comparatively invariant, and the highly variable area is limited to CDR3 δ , i.e. one segment of the $\gamma\delta$ TCR combining site. It seems likely that this particular restriction of variability holds a clue that might eventually help to explain the Ag preferences of $\gamma\delta$ T cells.²⁴

SPECIFIC EXAMPLES OF LIGANDS

The number of bona fide ligands for $\gamma\delta$ TCRs is still relatively small. Nevertheless, our aim was not to provide a complete list but rather to highlight the differences and diversity of ligands recognized.

MHC-LIKE LIGANDS

Despite the fact that there may be no inherent MHC bias in the $\gamma\delta$ TCRs—none has been reported as of this writing—MHC molecules were investigated as ligands for the $\gamma\delta$ TCR even prior to the landmark studies by Matis and Bluestone.25,26 The pair of related T-locus Ags,

T10/22, may be considered prototypic, because crystal structures of these Ags, as well as of a $\gamma\delta$ TCR engaged with T22, have been available for some time now.27,28 These structures show that the T Ags do not present peptides, and that the $\gamma\delta$ TCR (KN6) binds to T22 at an angle, mainly using CDR3 δ amino-acid side chains for the interaction. This is much unlike the binding of $\alpha\beta$ TCRs to MHC molecules, where CDR1 and 2 of both TCR- α and β , mainly interact with the MHC surface, and the CDR3s with the peptide in the groove. The repertoire of T10/22 recognizing $\gamma\delta$ TCRs is diverse, including several V γs and V δs , with a shared motif in CDR3 δ (W-(S)EGYEL).²⁹ Although expressing the motif is sufficient for ligand recognition, these TCRs can have widely different affinities for T22, suggesting that non-motif amino-acid side chains are involved in the interaction as well. Approximately 0.4% of lymphoid $\gamma\delta$ T cells in mice recognize T22. The biological significance of this specificity remains to be determined. However, because T10/22 appear to be induced by cell stress, it is possible that T10/22-specific $\gamma \delta$ T cells play a role as monitors of tissue health.

Similarly to T10/22, the classical MHC molecules I- $E^{k,b,s}$ have been identified early as potential ligands for $\gamma\delta$ T cells. This has been confirmed later in binding studies, which again did not reveal any role for presented peptides.³⁰ Post-translational modification of these classical MHC molecules appears to be critical for $\gamma\delta$ T-cell recognition. However, binding affinities are low, the population of murine $\gamma\delta$ T cells capable of recognizing these ligands remains to be investigated, and the biological role of I-E recognition by $\gamma \delta$ T cells is still unclear.

Although a number of human $\gamma\delta$ T-cell lines and clones were characterized early on as MHC-specific, it was not formally shown that their responses were TCR-mediated. This was rectified more recently in the case of a human $\gamma\delta$ response to HLA B58 (Kaiser A, Fisch P, 5th International $\gamma\delta$ T cell Conference, 31 May–12 June 2012, Freiburg, GER). Here, it was shown by TCR transfer into a mouse hybridoma cell line that the specific reactivity to the alloantigen HLA B58 is mediated by the $\gamma\delta$ TCRs, and that this type of recognition supports cytolysis. Interestingly, because the human $\gamma\delta$ TCR in question differentiates between HLA B*5802 and HLA B*5801, which differ only in three amino acids in the floor of the peptide binding groove, a bound peptide might play a role in this particular case.

Human $\gamma \delta$ T cells (as well as $\alpha \beta$ T cells) also recognize group 1 CD1 molecules (CD1a, b, c). 31 These molecules are primarily expressed on professional Ag presenting cells where they present lipid Ags (glycolipids and certain microbial lipids) to the T cells. The $V\delta1^+$ subset of human $\gamma\delta$ T cells, which is mainly found in the tissues, shows prominent reactivity to CD1c, and produces IFN- γ and granulysin in the course of such responses. It has been suggested that the CD1c-reactive cells, dependent in part also on inflammatory cytokines and co-stimulation via NKG2D, provide protection against microbial infections prior to the more slowly developing responses of Ag-specific $\alpha\beta$ T cells.31,32 Finally, it has been reported that group 1 CD1 molecules can present lipid A to human $\gamma \delta$ T cells.³³ The related group 2 CD1 molecule, CD1d, appears to be recognized by both human and murine $\gamma\delta$ T cells.^{34–37} The same molecule has been studied in detail as a ligand for classical natural killer T cells,³⁸ also both in mice and humans. With natural killer T cells, CD1d serves as presenter of certain lipids, and there is some evidence that $\gamma\delta$ T cells in humans and mice also recognize a CD1d/lipid complex. Thus, cloned human $\gamma\delta$ T cells responded to phosphatidyl ethanol amine (PE) in a manner dependent on CD1d, which suggested CD1d-restricted recognition of this phospholipid.34,35,39 The phospholipid cardiolipin (CL) binds to CD1d and murine $\gamma \delta$ T cells responded both in vivo and in vitro to

CL dependent on the presence of CD1d, suggesting that this phospholipid is also presented by CD1d, and is thus recognized by the $\gamma\delta$ T cells.³⁶ Finally, CD1d appears to present sulfatide, an abundant myelin glycosphingolipid, to human $\gamma\delta$ T cells expressing V δ 1.⁴⁰ Thus, the CD1d-restricted recognition by $\gamma\delta$ T cells of small molecules might be the format closest to MHC-restricted Ag recognition by $\alpha\beta$ T cells.

The stress-induced MHC class I-related molecules MICA and MICB were also found to be recognized by human $\gamma\delta$ T cells derived from the intestinal epithelia. These cells expressed diverse V δ 1⁺ TCRs. Both α 1 and α 2 domains of MICA/B were involved in the recognition, but Ag processing was not required.⁴¹ Interestingly, the human $\gamma \delta$ T cells also recognized MIC proteins derived from other primate species despite extensive amino-acid changes in the α 1 and α 2 domains, perhaps due to a single conserved site. 42 Further examination revealed that MICA/ B are broadly expressed on carcinomas of the lung, breast, kindney, ovary, prostate and colon where they are recognized by tumor-infiltrating V δ 1⁺ $\gamma\delta$ T cells, which may affect tumor survival.⁴³ However, MICA is also a ligand for NKG2D, which is expressed on natural killer cells, CD8⁺ $\alpha\beta$ T cells and $\gamma\delta$ T cells. All of these cells can be activated via NKG2D, which complicates analysis of the $\gamma\delta$ responses.⁴⁴ MICA engagement by NKG2D also enhances responses of $V\gamma9 + V\delta2 + \gamma\delta T$ cells to non-peptide Ags.45 Nevertheless, a study with soluble MICA tetramers confirmed binding to $V\delta1^+$ $\gamma\delta$ TCRs and suggested that MIC delivers both TCR-dependent signal 1 and NKG2D-dependent signal 2 in the appropriate $\gamma \delta$ T cells.⁴⁶ A crystal structure of a MICreactive V δ 1⁺ γ δ TCR is now available, revealing a surprisingly flat potential binding surface. Furthermore, it appears that MIC binding by the TCR and by NKG2D is mutually exclusive, perhaps forcing sequential recognition.⁴⁷

Finally, a study presented by C. R. Willcox at the 5th International γδ T Cell Conference in Freiburg, Germany (31 May–2 June 2012) indicates that the endothelial protein C receptor has joined the ranks of MHC-like $\gamma\delta$ TCR ligands too. Endothelial protein C receptor is a TCR ligand that is expressed on cytomegalovirus-infected cells and on tumor cells. It is recognized by a human $V\gamma 4V\delta 5^+$ clone sensitive to the conformation of the ligand. Recognition—via CDR3—also depends on endothelial protein C receptor expression levels and costimulation, both of which were found to be stress-regulated. This specificity was deemed Ig-like as well.

OTHER PROTEINS EXPRESSED ON THE CELL SURFACE

A study of anti tumor responses by human $\gamma\delta$ T cells revealed interactions of the V γ 9V δ 2 $\gamma\delta$ TCR with an F1-ATPase-related structure expressed on the surface of the Burkitt's lymphoma cell line Daudi and certain other tumor lines but not another Burkitt's lymphoma, Raji.⁴⁸ This observation showed that cell surface-expressed proteins other than MHC molecules can be recognized by $\gamma\delta$ T cells. Moreover, response-inhibitory effects of antibodies directed against a serum protein, apolipoprotein A1, suggested involvement of this protein as well, and this was confirmed in molecular binding experiments using surface plasmon resonance, in which a soluble V γ 9V δ 2 TCR construct bound both purified F1-ATPase and a delipidated form of apolipoprotein A1. Finally, consistent with the idea that a trimolecular complex of these molecules provides the basis for $\gamma\delta$ T-cell ligand recognition, apolipoprotein A1 was found to be required for optimal responses of V γ 9V δ 2 T cells by tumor target cell lines expressing F1-ATPase. Involvement of an apolipoprotein has been observed in another $\gamma\delta$ response as well ⁴⁹ and will be further discussed below.

Based on other studies, also with Daudi cells,^{50–52} it seems clear that F1-ATPase is not the only non-MHC-related protein on the cell surface that might be recognized by $\gamma\delta$ T cells. In fact, screening experiments of murine cell lines and macrophages with soluble $\gamma\delta$ TCR constructs have implied the presence of multiple additional ligands,53,54 although their precise molecular nature remains to be determined.

SOLUBLE PROTEINS

Probably the first defined Ag reported to stimulate specific responses of human $\gamma\delta$ T cells was tetanus toxoid, a potent immunogen derived from the protein tetanospasmin of *Clostridium tetani.*^{55,56} Responses included IFN- γ production, induction of IL-2R expression and proliferation, and were limited to clones and T-cell somatic hybrids expressing certain $\gamma\delta$ TCRs. Despite these auspicious beginnings, no further studies on the tetanus toxoid response of $\gamma\delta$ T cells have been reported and the underlying mechanism remains uncertain.

Further bacterial proteins reported to elicit specific responses of human $\gamma\delta$ T cells include the unrelated staphylococcal enterotoxin A (SEA) 57 and the toxin listeriolysin O (LLO).⁵⁸ Several studies examined the responses of human $\gamma\delta$ T cells to bacterial super-Ags, such as SEA, SEB, SEE and TSST.⁵⁹ Interestingly, $V\gamma9^+$ clones killed SEApulsed targets but did not proliferate in response to such stimulation, whereas V γ 9-negative γ δ clones proliferated. The cytotoxic reactivity of the $\gamma\delta$ T cells was found to be more restricted such that a given clone might respond to SEA but not SEE and vice versa, in contrast to $\alpha\beta$ T cells, which often respond to multiple SEs. Although limited to certain $\gamma\delta$ TCRs, binding interactions between SEA or other super-Ags and the $\gamma\delta$ TCR have not yet been demonstrated, and the analysis of the responses is complicated by observations indicating that some of the cytotoxic activity of the V γ 9⁺ clones is actually mediated by staphylococcal enterotoxin-specific antibodies that bind to Fc receptors of the $\gamma\delta$ T cells.

Both rodent and primate $\gamma\delta$ T cells respond to Listeria monocytogenes.^{58,60–63} In a study with human $\gamma\delta$ T cells, live bacteria were required to elicit responses of $\gamma \delta$ T cells in vitro. A $\gamma \delta$ T cell line derived from such culture responded specifically to the large LLO-derived peptide LLO 470–508. Since this portion of LLO shows a high degree of homology with other Gram-positive bacterial toxins, it was deemed probable that such toxins might also be recognized.⁵⁸ In this system, involvement of the $\gamma\delta$ TCR remains to be demonstrated. The large size of the stimulatory peptide, which also contains a cysteine, raises the possibility that the LLO-reactive $\gamma\delta$ T cells might recognize a conformational epitope, but this aspect has not been explored.

Early studies with mycobacterial extracts and purified protein derivative (PPD) revealed responses of human and murine $\gamma \delta$ T cells to mycobacterial Ags.64–66 The murine response to PPD was found to be limited to cells expressing V γ 1,⁶⁷ suggesting γ δ TCR involvement in the response to a mycobacterial protein molecule. Nevertheless, subsequent studies with human $\gamma\delta$ T cells focused on non-peptidic Ags contained within but not specific to mycobacteria (see below). More recently, defined mycobacterial proteins were also found to elicit $\gamma \delta$ Tcell responses. One of these is ESAT-6, a small, highly immunogenic protein, which is part of a transmembrane secretion pathway in M. tuberculosis,⁶⁸ and a critical virulence factor.⁶⁹ A response to ESAT-6 was initially observed in $\gamma\delta$ T cells of cattle experimentally infected with M. bovis.⁷⁰ WC1⁺ bovine $\gamma\delta$ T cells responded to this protein with proliferation and IFN- γ secretion and changes in CD45 isoforms.^{71,72} A $\gamma\delta$ response to ESAT-6 was also observed in patients with active pulmonary tuberculosis.⁷³ Others reported that ESAT-6 directly induces purified $\gamma\delta$ T effector memory cells from tuberculin skin testpositive patients to produce IFN- γ , and that CD4⁺ $\alpha\beta$ T cells regulate this response.⁷⁴ However, this observation has been challenged and it requires further investigation.^{75,76} Since $\alpha\beta$ T cells also respond to ESAT-6, and for possible application in a tuberculosis vaccine,⁷⁷ it will be important to show that an $\alpha\beta$ T cell-independent $\gamma\delta$ response exists.

 $\gamma\delta$ T cells protect mice from herpes simplex virus (HSV) type 1induced lethal encephalitis,⁷⁸ and $\gamma \delta$ T cells were isolated that recognize and respond to HSV glycoprotein 1.⁷⁹ Further characterization of a clone expressing $V\gamma 2V\delta 8$ revealed that this response does not require classical Ag-processing pathways and in fact can occur in the absence of any APCs,⁸⁰ similar to the findings with small peptide Ags described below.^{19,81} The HSV glycoprotein 1-reactive $\gamma\delta$ clone responded in a B cell-like manner to a conformational epitope, and it did not require any glycosylation of the Ag. The epitope was localized to the solventexposed amino terminus of the protein, and it was sensitive to sulfhydryl reduction.⁸² The same clone failed to respond to glycoprotein 1 derived from HSV-2, suggesting that amino acid differences near the N-terminus and specific to these orthologues are critical for recognition.⁷⁹ These data provided convincing evidence that soluble non-MHC proteins can be recognized by $\gamma\delta$ T cells and suggest that such reactivity might play an important role in host protection against viral infections.⁸³

In a study of anti-tumor responses by human T cells, TCR- δ^+ cytotoxic cells with specificity for an immunoglobulin idiotype of autologous B-cell tumors were detected.⁸⁴ Further investigation showed that the $\gamma\delta$ CTLs specifically recognized the λ light-chain protein.⁸⁵ However, the anti-tumor response was not inhibited by λ -specific antibodies, and cells transfected with a λ -chain construct that could not be expressed on the cell surface were still lysed suggesting that the cytolytic $\gamma \delta$ T cells recognized the λ -protein in a processed form. There was no indication of MHC-mediated Ag presentation, but because antibodies against a member of the HSP-70 family, which is expressed on the surface of the target cells, inhibited the anti tumor response, the authors suggested that λ might be recognized by the cytolytic $\gamma \delta$ T cells in the form of an HSP-70-presented peptide Ag. Despite such intriguing findings, no further reports on this response have appeared. The postulated antigenic λ peptide has not been confirmed, and it remains unclear if the λ protein needs to retain some secondary structure to be recognized. Ig λ contains several cysteines, and in view of the studies with HSV $gI_s⁸²$ as well as our own recent observations with an insulin peptide (Kemal Aydintug M, unpublished, see below), it might be worthwhile to determine whether the redox state of the λ -chain influences its antigenicity.

Most recently, the target specificity of a human $\gamma\delta$ TCR derived from a presumed autoreactive and pathogenic $\gamma \delta$ T-cell clone has been determined. This amazing story goes back to a rare case of human autoimmune myositis where infiltration of skeletal muscle with $\gamma\delta$ T cells instead of the more common autoreactive $\alpha\beta$ T cells was observed.⁵ It was found that the muscle-infiltrating $\gamma \delta$ T cells were clonally expanded and express an uncommon $V\gamma$ 1.3V δ 2⁺ $\gamma\delta$ TCR.⁸⁶ Based on responses of transfectomas, as well as binding studies with soluble TCR constructs, ^{87,88} it appears now that this $\gamma \delta$ TCR (named M88) recognizes a common conformational motif on the surface of several proteins from different species, including bacterial and human aminoacyl-tRNA synthetases.⁸⁹ For example, M88 recognizes histidyltRNA synthetase, which is also targeted by autoantibodies in patients with myositis. One of the soluble proteins recognized, the E. coli translation initition factor 1, has been mutagenized extensively to show that a short α -helical stretch including amino acids 39–42 of the E. coli translation initition factor 1 constitutes a critical part of the epitope for

M88. Several aspects of this response seem noteworthy including the focus of a $\gamma\delta$ TCR on a conformational epitope, which is reminiscent of the findings with HSV glycoprotein 1 ,⁸² the multispecificity of the $\gamma\delta$ TCR, which recognizes an epitope shared between various unrelated proteins, and the convergence of $\gamma\delta$ and antibody responses, which focus on the same Ags.

SMALLER PEPTIDES

In addition to the $\gamma\delta$ responses to intact native proteins, a number of responses to smaller peptide Ags have been reported as well. These have been listed elsewhere in greater detail.⁹⁰ Briefly, using hybridomas with murine $\gamma \delta$ T cells, responses to peptides as short as seven amino acids were observed.⁹¹ The first report of a response to a small, defined peptide Ag involved a peptide derived from a mycobacterial heat shock protein, HSP-65.⁸¹ The peptide (p180–196) was recognized by a large number of clones expressing murine $V\gamma1$,⁹² and it was shown by TCR transfer that the hybridoma response was TCRdependent.⁹¹ Unlike $\alpha\beta$ T cells recognizing small peptides in an MHC-restricted fashion, $\gamma\delta$ hybridomas responding to these peptides did not require APCs,⁹² in this regard resembling B cells.

Because the shortest HSP-derived stimulatory peptide (FGLQLEL) resembled a motif shared by unfolded proteins which bind to the molecular chaperone HSP-70 BiP,⁹³ we also examined unrelated peptides having this motif for their ability to elicit $\gamma\delta$ responses. Several were stimulatory, albeit not as strongly as the HSP peptide (data unpublished). Although we have previously suggested that this might reflect an ability of $\gamma\delta$ T cells to recognize unfolded proteins, it might instead indicate that $\gamma\delta$ T cells recognize, rather than singular aminoacid sequences, secondary structure of peptides such as, for example, a helices.

More recently, we have studied a different peptide response of murine $\gamma \delta$ T cells involving a major insulin epitope in the non-obese diabetic mouse model of type 1 autoimmune diabetes. The Ins2 B:9-23 peptide is a naturally occurring Ag contained in certain β cells of the pancreatic islets, ⁹⁴ which is recognized by diabetogenic I-Ag⁷restricted CD4⁺ $\alpha\beta$ T cells, and by B lymphocytes.⁹⁵ In a hybridoma collection generated to screen the TCR-V β repertoire of B:9-23 peptide-reactive $\alpha\beta$ T cells, a peptide-reactive clone was found expressing a $\gamma\delta$ TCR.¹⁹ This hybridoma (SP9D11) expressed V γ 4 paired with $V\delta$ 10. In addition to the peptide, the hybridoma responded to a preparation of pancreatic islet cells, but not to intact insulin. The peptide response was TCR-dependent and, like the $\gamma\delta$ hybridomas recognizing HSP-65-derived peptides, SP9D11 did not require APCs, unlike all B:9-23-reactive $\alpha\beta$ T cells. In fact, the $\gamma\delta$ response did not require any accessory cells, because isolated individual SP9D11 hybridoma cells still responded to the peptide when tested in a single-cell assay.¹⁹ Unlike $\gamma\delta$ cells recognizing HSP-derived peptides, the repertoire of B:9-23- reactive $\gamma \delta$ T cells includes different V γs in addition to several V δ s. Thus, different portions of the $\gamma\delta$ TCR might be critical in recognizing the insulin peptide Ag. Whereas two C-terminal amino acids of the insulin peptide appear to be dispensable for the $\gamma\delta$ response, the Nterminal one is required. Surprisingly, the $\gamma\delta$ response also required the tyrosine in position 16, in this regard resembling I-Ag⁷-restricted $\alpha\beta$ T cells. However, in contrast to the $\alpha\beta$ T cells, $\gamma\delta$ cells also require the cysteine in position $19¹⁹$, suggesting that the peptide might be stimulatory as a dimer. This was recently confirmed (Kemal Aydintug M, manuscript in preparation). It is also noteworthy that B:9-23 represents an α -helical segment of the insulin B chain. Some of this secondary structure might be retained by the peptide and could be important for $\gamma\delta$ recognition.

Several other defined peptides have been either implied as Ag for $\gamma\delta$ T cells or actually found to be stimulatory, including the already mentioned processed Ig λ light-chain protein,⁸⁵ a peptide derived from tetanus toxin (C. tetani, $1235-1246$)⁹⁶ and a LLO-derived peptide $(470-508).$ ⁵⁸ However, the molecular details of these responses remain to be determined.

Finally, some studies involving random heterocopolymers of two or more amino acids have been reported as well. Most prominently, a response of a $\gamma\delta$ hybridoma to poly GT, a polymer of approximately 100 amino acids containing glutamic acid and tyrosine at a 1 : 1 ratio, was published as early as 1989.⁹⁷ We later showed that essentially all $V\gamma1^{\hat{+}}$ $\gamma\delta$ T cells respond to poly GT,¹⁸ as well as cells expressing other $\gamma\delta$ TCRs, but we were unable to confirm the original claim that poly GT is recognized in the context of $Qa-1^b$. With its random sequence, poly GT is not a defined Ag and does not have a defined structure. However, it seems probable that segments of individual pGT molecules assume secondary structures, some of which might meet stimulatory requirements. pGT is also anionic, and we have pointed out before that several polyanionic structures are capable of stimulating V γ 1 responses, perhaps due to particular properties of V γ 1 itself.¹⁶

NON-PEPTIDIC AGS

Original observations that large numbers of $\gamma\delta$ T cells were responsive to mycobacteria and mycobacterial Ags included both human and murine cells.64–66,98 However, while many murine cells were found to react with mycobacterial PPD and HSP-65,⁶⁶ human mycobacteriareactive $\gamma\delta$ T cells failed to react with PPD and HSP-65.⁹⁹ Searching for alternative Ags, Pfeffer and colleagues¹⁰⁰ found that most human mycobacteria-reactive $\gamma\delta$ T cells responded to Ags contained in fractions of mycobacterial lysates with a molecular mass of \leq 3 kDa. Moreover, these Ags were protease-resistant. In contrast, only few human $\gamma\delta$ T cells responded to PPD and HSP-65. These surprising observations were rapidly followed by studies identifying several mycobacterial and synthetic non-peptidic Ags for human $\gamma\delta$ T cells, first the mycobacteria-derived TUBag4, a 5' triphosphorylated thymidine-containing compound with additional structurally related stimulatory molecules¹⁰¹ and then synthetic alkyl phosphates. In particular monoethyl phosphate,¹⁰² and subsequently mycobacteriaderived isopentenyl pyrophosphate and related prenyl pyrophosphate derivatives were found to be stimulatory.¹⁰³ Since these early reports, many additional Ags of essentially the same type, now often somewhat imprecisely referred to as 'phosphoantigens', have been described. Although some are only weak stimulators, others were found that are extremely potent, most prominently perhaps HMBPP, ¹⁰⁴ a metabolite in the 2-C-methyl-D-erythritol-4 pathway for isoprenoid synthesis.

Despite their small size, recognition of the prenyl pyrophosphates by the V γ 9V δ 2 $\gamma\delta$ TCRs depends on all CDRs.¹⁰⁵ This may be more understandable considering that these small molecules appear to be presented on the surface of target cells^{17,106,107} and thus might be 'seen' in a certain context. However, the mechanism of presentation remains mostly unclear, and it might vary between individual Ags.

Finally, there are small molecular compounds that stimulate prenyl pyrophosphate-reactive $\gamma\delta$ T cells indirectly, by blocking farnesyl pyrophosphate synthase in the mevalonate pathway, which increases cellular IPP levels.^{108,109} These include bisphosphonates¹¹⁰ and alkylamines.¹¹¹

It should be noted that the response to prenyl pyrophosphates is limited to a single subset of $\gamma\delta$ T cells present in primates, both human and non-human. This response already received and clearly deserves much attention, because of its potential therapeutic significance and because so many human peripheral blood $\gamma\delta$ T cells show this specificity, but there is no indication that it is representative of $\gamma\delta$ specificities in general.

PHOSPHOLIPIDS

In the 1990s, our lab used a collection of hybridomas generated with $\gamma\delta$ T cells from normal untreated mice to screen for ligands recognized by the $\gamma\delta$ TCR. Because we had already found responses to other anionic molecules (oligonucleotides and peptides), we also tested anionic phospholipids. We promptly found TCR-dependent responses to CL and the related phospholipids phosphatidylglycerol and phosphatidic acid, but not to other phospholipids (phosphatidylinositol, phosphatidylserine, phosphatidylcholine and phosphorylcholine). Only cells expressing $V\gamma1$ responded, and the response was dependent on the presence of a serum factor, which we tentatively identified as apolipoprotein H (apo H, β_2 -glycoprotein 1). These findings suggested a connection with numerous observations made with so-called antiphospholipid antibodies, which develop in response to some infections but also in patients suffering from autoimmune diseases. Some such antibodies react with apo H, or complexes of apo H and CL.¹¹² Because the $\gamma\delta$ cells required CL and serum factors in stoichiometric ratios in order to respond optimally, we suggested that they might recognize CL and apo H in a complex as do antiphospholipid antibodies.⁴⁹ This remains to be confirmed, however. Although we observed anti CL responses only with hybridomas, recently another group reported that normal splenic and hepatic $\gamma\delta$ T cells from healthy mice proliferated in vitro in response to CL. Such cells were also activated in vivo following transfer of CL-pulsed dendritic cells and here, they noted a requirement for CD1d expression by the DC. They finally demonstrated that CL binds to CD1d and provided a crystal structure of this complex.³⁶ They suggested that $\gamma \delta$ T cells can recognize this complex but further analysis will be required to confirm this proposed mechanism.

Human $\gamma\delta$ T cells were reported to recognize PE, apparently also in the context of (human) CD1 $d³⁵$. The investigators found that cloned peripheral blood or nasal mucosa-derived $\gamma\delta$ T cells from cypress pollen-sensitive subjects responded to pollen-derived PE after incubation in vitro. The response was specific for PE and limited to molecules with partially saturated fatty acid side chains. In a subsequent study, the same group found that human $\gamma\delta$ T cells derived from the duodenal mucosa also contained a high percentage of CD1d-restricted PEreactive $\gamma\delta$ T cells.³⁹ Taken together, the studies of three independent groups involving mice and humans make a good case that phospholipids also belong into the category of ligands recognized by $\gamma\delta$ T cells.

Finally, a just published study already mentioned above indicates that the myelin-derived glycosphingolipid sulfatide is also recognized, by human V δ 1⁺ $\gamma\delta$ T cells.⁴⁰ This new observation hints at the possibility of other not yet identified categories of non-peptidic Ags for $\gamma\delta$ T cells.

DISTRIBUTION OF $\gamma\delta$ T-CELL SPECIFICITIES

Despite an enormous potential for receptor diversity in CDR3 δ ,³ clear evidence of peripheral selection of $\gamma\delta$ T cells δ and the occasional emergence of $\gamma\delta$ T-cell clones,⁵ most specificities identified so far are not clone-specific. It is not clear whether this is mainly a bias of the investigators—polyclonal responses are more easily spotted—or an intrinsic property of $\gamma\delta$ T-cell recognition. The response to poly GT in mice, for example, is shared by V γ 1⁺ cells, V γ 2⁺ cells and others. Similarly, the response to PPD is common to most V γ 1⁺ cells. It could be argued that these examples are inadequate because both stimuli are composed of a heterogeneous mix of molecules, but the response to a fully defined peptide (HSP-65 p180–196) was also shared by most V γ 1⁺ cells. Similarly, many V γ 1⁺ cells respond to CL, and all of these responses are demonstrably TCR-dependent. In contrast to these responses, the insulin peptide B:9-23 is recognized by some $\gamma \delta$ T cells expressing V γ 4, but also by some expressing V γ 1 and by others expressing at least one additional V γ . B:9-23 reactive $\gamma \delta$ T cells seem to be frequent in the non-obese diabetic mouse strain but not in C57BL/6 mice, but this needs to be confirmed. The much studied response to T10/22 is shared by many different cells expressing the TCR- δ CDR3 motif W(S)EGYEL, which is largely based on one reading frame of D δ 2, in murine $\gamma \delta$ T cells expressing several different V γ s and V δ s. In humans, the responses to prenyl pyrophosphates are shared by many V γ 9V δ 2⁺ cells, with only some bias for particular γ -chain CDR3 amino acids,¹⁰⁵ and many V δ 1⁺ cells respond to MICA/B. However, the recognition by a polymyositis associated $\gamma\delta$ TCR of a peptide motif on amino-acyl tRNA synthetases is remarkable because the TCR is derived from clonally expanded $\gamma\delta$ T cells, and both CDR3s seem to play a role. Here, the $\gamma\delta$ TCR seems to provide specificity for a conformational motif shared by a number of related and some unrelated proteins. Albeit still sketchy, these data taken together suggest that ligand recognition by $\gamma\delta$ T cells can occur in different modes, e.g. involving none, one or two CDR3s, and specificities might be distributed over larger and smaller subsets of $\gamma\delta$ T cells, depending on the Ag. A logical consequence of this is that many individual $\gamma \delta$ T cells are multispecific. Indeed, this is what we find. For example, a single murine $\gamma\delta$ TCR expressed by a hybridoma clone might support responses to poly GT, PPD, HSP-60 p180–196 and CL. Over the years, we observed several cells with such broad reactivity pattern. More interestingly, perhaps, we recently found that a cell that recognizes T22, because it expresses the appropriate CDR3 δ motif also responded to pGT and even to the insulin peptide B:9-23 (Kemal Aydintug M, unpubl. observ.). However, it seems that some $\gamma\delta$ T cells are more likely to be multispecific than others. In mice, $V\gamma1^+ \gamma \delta T$ cells seem to have a greater propensity for multispecificity than $V\gamma 4^+$ or $V\gamma 6^+$ cells. In this regard, $V\gamma1^+$ cells in mice are reminiscent of the polyspecific B1 B cells. In humans, multispecific $\gamma\delta$ T cell clones have been observed as well, mostly with $V\gamma9V\delta2^+$ cells. Here, mycobacteria-reactive cells also might respond to tetanus toxoid, to non-mycobacteria-derived phosphoantigens, or to phospholipids in the context of CD1d.

CONSEQUENCES OF MULTI-SPECIFICITY

With all multispecific lymphocytes, the question of self-tolerance looms heavily. Selection becomes more complex if there are several specificities of auto-Ags within one responder. Negative selection of high affinity clones also removes their low affinity specificities as well as possible specificities for non-self Ags. A requirement for intermediate to low affinities for several Ags might diminish a large cell population to a small one that fits all the selection criteria. On the other hand, surviving multispecific cells might be more useful than monospecific ones, simply because they can be employed in several different settings. Moreover, as has been argued before, specificities shared by many $\gamma\delta$ T cells would make for faster responses by eliminating the need for prior clonal expansion.

CONCLUSIONS

A review of the Ags recognized by $\gamma\delta$ T cells reveals a remarkable diversity (Figure 1). This likely precludes a single mechanism of ligand recognition akin to that of MHC-restricted $\alpha\beta$ T cells, but does not preclude Ig-like Ag recognition. Some subsets of $\gamma\delta$ T cells resemble B1 B cells in their broad ligand specificity, while others more resemble innate $\alpha\beta$ T cells. How $\gamma\delta$ T cells are kept in check to prevent autoaggressive reactivity remains unclear.

- 1 Chien Y-H, Jores R, Crowley MP. Recognition by γ/δ T cells. Annu Rev Immunol 1996; 14: 511–532.
- 2 Haas W, Pereira P, Tonegawa S. γ/δ T cells. Annu Rev Immunol 1993; 11: 637–685.
3 Davis MM. Biorkman P.J. T cell antigen receptor genes and T cell recognition. Nature Davis MM, Bjorkman PJ. T cell antigen receptor genes and T cell recognition. Nature
- 1988; 334: 395–402. 4 Asarnow DM, Kuziel WA, Bonyhadi M, Tigelaar RE, Tucker PW, Allison JP. Limited diversity of $\gamma\delta$ antigen receptor genes of Thy-1⁺ dendritic epidermal cells. Cell 1988; 55: 837–847.
- 5 Hohlfeld R, Engel AG, Ii K, Harper MC. Polymyositis mediated by T lymphocytes that express that $\sqrt{\delta}$ receptor. N Engl J Med 1991; 324: 877–881.
- 6 Roark CL, French JD, Taylor MA, Bendele AM, Born WK, O'Brien RL. Exacerbation of collagen-induced arthritis by oligoclonal, iL-17-producing $\gamma \delta$ T cells. J Immunol 2007; 179: 5576–5583.
- 7 Havran W, Allison JP. Developmentally ordered appearance of thymocytes expressing different T cell antigen receptors. Nature 1988; 335: 443-445
- 8 O'Brien RL, Roark CL, Jin N, Aydintug MK, French JD, Chain JL et al. $\gamma\delta$ T cell receptors: functional correlations. Immunol Rev 2007; 215: 77–88.
- 9 Jin N, Roark CL, Miyahara N, Taube C, Aydintug MK, Wands JM et al. Allergic airway hyperresponsiveness-enhancing $\gamma\delta$ T cells develop in normal untreated mice and fail to produce IL-4/13, unlike Th2 and NKT cells. J Immunol 2009; 182: 2002-2010.
- Narayan K, Sylvia KE, Malhotra N, Yin CC, Martens G, Vallerskog T et al. Intrathymic programming of effector fates in three molecularly distinct $\gamma\delta$ T cell subtypes. Nature Immunol 2012; 13: 511–518
- 11 Haas JD, Ravens S, Duber S, Sandrock I, Oberdorfer L, Kashani E et al. Development of interleukin-17-producing $\gamma\delta$ T cells is restricted to a functional embryonic wave. Immunity 2012; 37: 48–59.
- 12 Bonneville M, O'Brien RL, Born WK, $\gamma\delta$ T cell effector functions: a blend of innate programming and acquired plasticity. Nat Rev Immunol 2010; 10: 467–478.
- 13 Hayes SM, Love PE. Distinct structure and signalling potential of the $\gamma\delta$ TCR complex. Immunity 2002; 16: 1–20.
- 14 Hayes SM, Li L, Love PE. TCR signal strength influences $\alpha\beta/\gamma\delta$ lineage fate. Immunity 2005; 22: 583–593.
- 15 Rock EP, Sibbald PR, Davis MM, Chien YH. CDR3 length in antigen-specific immune receptors. J Exp Med 1994; 179: 323-328.
- 16 Born W, Cady C, Jones-Carson J, Mukasa A, Lahn M, O'Brien R. Immunoregulatory functions of $\gamma \delta$ T cells. Adv Immunol 1999; 71: 77-144.
- 17 Morita CT, Beckman EM, Bukowski JF, Tanaka Y, Band H, Bloom BR et al. Direct presentation of nonpeptide prenyl pyrophosphate antigens to human $\gamma\delta$ T cells. Immunity 1995; 3: 495–507.
- 18 Cady CT, Lahn M, Vollmer M, Tsuji M, Seo SJ, Reardon CL et al. Response of murine $\gamma\delta$ T cells to the synthetic polypeptide poly-Glu⁵⁰Tyr⁵⁰. J Immunol 2000; 165: 1790-1798.
- 19 Zhang L, Jin N, Nakayama M, O'Brien RL, Eisenbarth GS, Born WK. $\gamma\delta$ T cell receptors confer autonomous responsiveness to the insulin-peptide B:9-23. J Autoimmun 2010; 34: 478–484.
- 20 Blackman M, Yague J, Kubo R, Gay D, Coleclough C, Palmer E et al. The T cell recpertoire may be biased in favor of MHC recognition. Cell 1986; 47: 349–357.
- 21 Garcia KC, Gapin L, Adams JJ, Birnbaum ME, Scott-Browne JP, Kappler JW et al. A closer look at TCR germline recongition. Immunity 2012; 36: 887-888.
- 22 Adams EJ, Strop P, Shin S, Chien YH, Garcia KC, An autonomous CDR3 δ is sufficient for recognition of the nonclassical MHC class I molecules T10 and T22 by $\gamma\delta$ T cells. Nat Immunol 2008; 9: 777–784.
- 23 Boyden LM, Lewis JM, Barbee SD, Bas A, Girardi M, Hayday AC et al. Skint1, the prototype of a newly identified immunoglobulin superfamily gene cluster, positively selects epidermal $\gamma \delta$ T cells. Nat Genet 2008; 40: 656–662.
- 24 Born WK, O'Brien RL. Antigen-restricted $\gamma\delta$ T-cell receptors? Arch Immunol Ther Exp (Warsz) 2009; 57: 129–135
- 25 Matis LA, Cron R, Bluestone JA. Major histocompatibility complex-linked specificity of yo receptor-bearing T lymphocytes. Nature 1987; 330: 262-264
- 26 Matis LA, Fry AM, Cron RQ, Cotterman MM, Dick RF, Bluestone JA. Structure and specificity of a class II alloreactive $\gamma\delta$ T cell receptor heterodimer. Science 1989; 245: 746–749.
- Wingren C, Crowley MP, Degano M, Chien YH, Wilson IA. Crystal structure of a $\gamma\delta$ T cell receptor ligand T22: a truncated MHC-like fold. Science 2000; 287: 310–314.
- 28 Adams EJ, Chien Y-H, Garcia KC. Structure of a $\gamma\delta$ T cell receptor in complex with the nonclassical MHC T22. Science 2005; 308: 227–231.
- 29 Shin S, El-Diwany R, Schaffert S, Adams EJ, Garcia KC, Pereira P et al. Antigen recognition determinants of $v\delta$ T cell receptors. Science 2005: 308: 252–255.
- 30 Hampl J, Schild H, Litzenberger C, Baron M, Crowley MP, Chien YH. The specificity of a weak $\gamma\delta$ TCR interaction can be modulated by the glycosylation of the ligand. J Immunol 1999; 163: 288–294.
- 31 Sugita M, Brenner MB. T lymphocyte recognition of human group 1 CD1 molecules: implications for innate and acquired immunity. Semin Immunol 2000; 12: 511-516.
- 32 Das H, Sugita M, Brenner MB. Mechanisms of V δ 1 γ δ T cell activation by microbial components. J Immunol 2004; 172: 6578–6586.
- 33 Cui Y, Kang L, Cui L, He W. Human $\gamma\delta$ T cell recognition of lipid A is predominately presented by CD1b or CD1c on dendritic cells. Biol Direct 2009; 4: 47–58.
- 34 Agea E, Russano A, Bistoni O, Mannucci R, Nicoletti I, Corazzi L et al. Human CD1restricted T cell recognition of lipids from pollens. *J Exp Med* 2005: **202**: 295–308.
- 35 Russano AM, Agea E, Corazzi L, Postle AD, de Libero G, Porcelli SA et al. Recognition of pollen-derived phosphatidyl-ethanolamine by human CD1d-restricted gamma delta T cells. J Allergy Clin Immunol 2006; 117: 1178–1184.
- 36 Dieude M, Striegl H, Tyznik AJ, Wang J, Behar SM, Piccirillo CA et al. Cardiolipin binds to CD1d and stimulates CD1d-restricted $\gamma\delta$ T cells in the normal murine repertoire. J Immunol 2011; 186: 4771–4781.
- 37 Huber S, Sartini D, Exley M. Role of CD1d in Coxsackievirus B3-induced myocarditis. J Immunol 2003; 170: 3147–3153.
- 38 Gapin L. iNKT cell autoreactivity: what is 'self' and how is it recognized? Nat Rev Immunol 2010; 10: 272–277.
- 39 Russano AM, Bassotti G, Agea E, Bistoni O, Mazzocchi A, Morelli A et al. CD1 restricted recognition of exogenous and self-lipid antigens by duodenal $\gamma\delta^+$ T lymphocytes. J Immunol 2007; 178: 3620–3626.
- 40 Bai L, Picard D, Anderson B, Chaudhary V, Luoma A, Jabri B et al. The majority of CD1d-sulfatide-specific T cells in human blood use a semiinvariant V81 TCR. Eur J Immunol 2012; 42: 2505–2510.
- Groh V, Steinle A, Bauer S, Spies T. Recognition of stress-induced MHC molecules by intestinal epithelial $\gamma \delta$ T cells. Science 1998; 279: 1737–1740.
- 42 Steinle A, Groh V, Spiess T, Diversification, expression, and $\gamma \delta$ T cell recognition of evolutionarily distant members of the MIC family of major histocompatibility complex class I-related molecules. Proc Natl Acad Sci U S A 1998; 95: 12510–12515.
- 43 Groh V, Rhinehart R, Secrist H, Bauer S, Grabstein KH, Spies T. Broad tumorassociated expression and recognition by tumor-derived gamma delta T cells of MICA and MICB. Proc Natl Acad Sci U S A 1999; 96: 6879–6884.
- 44 Bauer S, Groh V, Wu J, Steinle A, Phillips JH, Lanier LL et al. Activation of NK cells and T cells by NKG2D, a receptor for stress-inducible MICA. Science 1999; 285: 727– 729.
- 45 Das H, Groh V, Kuiji C, Sugita M, Morita CT, Spies T et al. MICA engagement by human Vy2V82 T cells enhances their antigen-dependent effector function. Immunity 2001; 15: 83–93.
- Wu J, Groh V, Spies T. T cell antigen receptor engagement and specificty in the recognition of stress-inducible MHC class I-related chains by human epithelial $\gamma \delta$ T cells. J Immunol 2002; 169: 1236–1240.
- 47 Xu B, Pizarro JC, Holmes MA, McBeth C, Groh V, Spies T et al. Crystal structure of a γ_0 T-cell receptor specific for the human MHC class I homolog MICA. Proc Natl Acad Sci USA 2011; 108: 2414–2419.
- 48 Scotet E, Martinez LO, Grant E, Barbaras R, Jeno P, Saulquin X et al. Tumor recognition following V γ 9V δ 2 T cell receptor interactions with a surface F1-ATPaserelated structure and apolipoprotein A-1. Immunity 2005; 22: 71-80.
- Born WK, Vollmer M, Reardon C, Matsuura E, Voelker DR, Giclas PC et al. Hybridomas expressing $\gamma\delta$ T-cell receptors respond to cardiolipin and β 2-glycoprotein 1 (apolipoprotein H). Scand J Immunol 2003; 58: 374–381.
- Fisch P, Malkovsky M, Braakman E, Sturm E, Bolhuis RLH, Prieve A et al. γ/δ T-cell clones and natural killer-cell clones mediate distinct patterns of non-major histocompatibility complex-restricted cytolysis. J Exp Med 1990; 171: 1567-1579.
- 51 Fisch P, Malkovsky M, Kovats S, Sturm E, Braakman E, Klein BS et al. Recognition by human Vy9/V δ 2 T cells of a GroEL homolog on Daudi Burkitt's lymphoma cells. Science 1990; 250: 1269–1273.
- 52 Fisch P, Oettel K, Fudim N, Surfus JE, Malkovsky M, Sondel PM. MHC-unrestricted cytotoxic and proliferative responses of two distinct human γ/δ T cell subsets to Daudi cells. J Immunol 1992; 148: 2315–2323.
- 53 Aydintug MK, Roark CL, Yin X, Wands JM, Born WK, O'Brien RL. Detection of cell surface ligands for the $\gamma\delta$ TCR using soluble TCRs. J Immunol 2004; 172: 4167– 4175.
- 54 Aydintug MK, Roark CL, Chain JL, Born WK, O'Brien RL. Macrophages express multiple ligands for $\gamma\delta$ TCRs. Mol Immunol 2008; 45: 3253-3263.
- 55 Kozbor D, Trinchieri G, Monos DS, Isobe M, Russo G, Haney JA et al. Human TCR- γ^+ / δ^+ , CD8⁺ T lymphocytes recognize tetanus toxoid in an MHC-restricted fashion. J Exp Med 1989; 169: 1847–1851.
- 56 Kozbor D, Cassatella MA, Lessin S, Kagan J, Finver S, Faust J et al. Expression and function of γ ⁸- and α β -T cell receptor heterodimers on human somatic T cell hybrids. J Immunol 1990; 144: 3677–3683.
- 57 Rust CJJ, Verreck F, Vietor H, Koning F. Specific recognition of staphylococcal enterotoxin A by human T cells bearing receptors with the V γ 9 region. Nature 1990; 346: 572–574.
- Guo Y, Ziegler HK, Safley SA, Niesel DW, Vaidya S, Klimpel GR. Human T-cell recognition of Listeria monocytogenes: recognition of listeriolysin O by TcR $\alpha\beta^+$ and TcR $\gamma\delta^+$ T cells. Infect Immunol 1995; 63: 2288-2294.
- Rust CJJ, Koning F. $\gamma\delta$ T cell reactivity towards bacterial superantigens. Semin Immunol 1993; 5: 41–46.
- 60 Skeen MJ, Ziegler HK. Intercellular interactions and cytokine responsiveness of peritoneal α/β and γ/δ T cells from *Listeria*-infected mice: synergistic effects of interleukin 1 and 7 on $\sqrt{\delta}$ T cells. J Exp Med 1993; 178: 985–996.
- 61 Fu YX, Roark CE, Kelly K, Drevets D, Campbell P, O'Brien R et al. Immune protection and control of inflammatory tissue necrosis by $\gamma\delta$ T cells. J Immunol 1994; 153: 3101–3115.
- 62 O'Brien RL, Xiang Y, Huber S, Ikuta K, Born WK. Depletion of a $\gamma\delta$ T cell subset can increase host resistance to a bacterial infection. J Immunol 2000; 165: 6472–6479.
- 63 Ryan-Payseur B, Frencher J, Shen L, Chen CY, Huang D, Chen ZW. Multieffectorfunctional immune responses of HMBPP-specific V γ 2V δ 2 T cells in non-human primates inoculated with Listeria monocytogenes DeltaactA prfA. J Immunol 2012; 189: 1285–1293.
- 64 Holoshitz J, Koning F, Coligan JE, de Bruyn J, Strober S. Isolation of CD4⁻CD8⁻ mycobacteria-reactive T lymphocyte clones from rheumatoid arthritis synovial fluid. Nature 1989; 339: 226–229.
- 65 Janis EM, Kaufmann SHE, Schwartz RH, Pardoll DM, Activation of $\gamma\delta$ T cells in the primary immune response to *Mycobacterium tuberculosis. Science* 1989; 244: 713– 716.
- 66 O'Brien RL, Happ MP, Dallas A, Palmer E, Kubo R, Born WK. Stimulation of a major subset of lymphocytes expressing T cell receptor $\gamma\delta$ by an antigen derived from Mycobacterium tuberculosis. Cell 1989; 57: 667–674.
- 67 Happ MP, Kubo RT, Palmer E, Born WK, O'Brien RL. Limited receptor repertoire in a mycobacteria-reactive subset of $\gamma\delta$ T lymphocytes. Nature 1989; 342: 696–698.
- 68 Bitter W, Houben ENG, Bottai D, Brodin P, Brown EJ, Cox JS et al. Systematic genetic nomenclature for type VII secretion systems. PLOS Pathog 2009: 5: 1-6, e1000507.
- Bottai D, Mailessi L, Simeone R, Friqui W, Laurent C, Lenormand P et al. ESAT-6 secretion-independent impact of ESX-1 genes espF and espG1 on virulence of Mycobacterium tuberculosis. J Infect Dis 2011; 203: 1155–1164.
- Rhodes SG, Buddle BM, Hewinson RG, Vordermaier HM. Bovine tuberculosis: immune responses in the peripheral blood and at the site of active disease. Immunology 2000; 99: 195–202.
- 71 Welsh MD, Kennedy HE, Smyth AJ, Girvin RM, Andersen P, Pollock JM. Responses of bovine WC1⁺ γ δ T cells to protein and nonprotein antigens of *Mycobacterium bovis*. Infect Immun 2002; 70: 6114–6120.
- 72 Maue AC, Waters WR, Davis WC, Palmer MV, Minion FC, Estes DM. Analysis of immune responses directed toward a recombinant early secretory antigenic target six-kilodalton protein-culture filtrate protein 10 fusion protein in Mycobacterium bovis infected cattle. Infect Immun 2005; 73: 6659–6667.
- 73 Gioia C, Agrati C, Goletti D, Vincenti D, Carrara S, Amicosante M et al. Different cytokine production and effector/memory dynamics of $\alpha\beta^+$ or $\gamma\delta^+$ T-cell subsets in the peripheral blood of patients with active pulmonary tuberculosis. Int J Immunopathol Pharmacol 2003; 16: 247–252.
- 74 Li L, Wu CY. CD4+CD25+ Treg cells inhibit human memory $\gamma\delta$ T cells to produce IFN- γ in response to M. tuberculosis antigen ESAT-6. Blood 2008; 111: 5629-5636.
- 75 Casetti R, Martino A, Sacchi A, Agrati C, Goletti D, Martini F. Do human $\gamma\delta$ T cells respond to M. tuberculosis protein antigens? Blood 2008; 112: 4776.
- 76 Li L, Wu CY. Human memory but not naive $\gamma\delta$ T cells from TST-positive individuals respond to M. tuberculosis antigen. Blood 2008; 112: 4777.
- 77 Cendron D, Ingoure S, Martino A, Casetti R, Horand F, Romagne F et al. A tuberculosis vaccine based on phosphoantigens and fusion proteins induces distinct $\gamma\delta$ and $\alpha\beta$ T cell responses in primates. Eur J Immunol 2007; 37:549–565.
- 78 Sciammas R, Kodukula P, Tang Q, Hendricks RL, Bluestone JA. T cell receptor- γ/δ cells protect mice from herpes simplex virus type 1 -induced lethal encephalitis. $JExp$ Med 1997; 185: 1969–1975.
- Johnson RM, Lancki DW, Sperling AI, Dick RF, Spear PG, Fitch FW et al. A murine CD4⁻, CD8⁻ T cell receptor- γ ⁸ T lymphocyte clone specific for Herpes simplex virus glycoprotein I. J Immuno/1992; 148: 983-988.
- 80 Sciammas R, Johnson RM, Sperling AI, Brady W, Linsley PS, Spear PG et al. Unique antigen recognition by a herpsevirus-specific TCR- $\gamma\delta$ cell. J Immunol 1994; 152: 5392–5397.
- 81 Born W, Hall L, Dallas A, Boymel J, Shinnick T, Young D et al. Recognition of a peptide antigen by heat shock reactive $\gamma\delta$ T lymphocytes. Science 1990; 249: 67–69.
- Sciammas R, Bluestone JA. HSV-1 glycoprotein I-reactive TCR $\gamma\delta$ cells directly recognize the peptide backbone in a conformationally dependent manner. J Immunol 1998; 161: 5187–5192.
- 83 Sciammas R, Bluestone JA. TCR γ ₈ cells and viruses. Microbes Infect 1999; 1: 203– 212.
- 84 Wright A, Lee JE, Link MP, Smith SD, Carroll W, Levy R et al. Cytotoxic T lymphocytes specific for self tumor immunoglobulin express T cell receptor δ chain. J Exp Med 1989; 169: 1557–1564.
- 85 Kim HT, Nelson EL, Clayberger C, Sanjanwala M, Sklar J, Krensky AM. $\gamma\delta$ T cell recognition of tumor Ig peptide. J Immunol 1995; 154: 1614-1623.
- 86 Pluschke G, Rüegg D, Hohlfeld R, Engel AG. Autoaggressive myocytotoxic T lymphocytes expressing an unusual γ/δ T cell receptor. J Exp Med 1992; 176: 1785–1789.
- 87 Wiendl H, Malotka J, Holzwarth B, Weltzien HU, Wekerle H, Hohlfeld R et al. An autoreactive $\gamma\delta$ TCR derived from a polymyositis lesion. J Immunol 2002; 169: 515–521.
- 88 Dornmair K, Schneider CK, Malotka J, Dechant G, Wiendl H, Hohlfeld R. Antigen recongition properties of a V γ 1.3V δ 2-T-cell receptor from a rare variant of polymyositis. J Neuroimmunol 2004; 152: 168–175.
- 89 Bruder J, Siewert K, Obermeier B, Malotka J, Scheinert P, Kellermann J et al. Target specificity of an autoreactive pathogenic human $\gamma\delta$ -T cell receptor in myositis. J Biol Chem 2012; 287: 20986–20995.
- 90 Born WK, Zhang L, Nakayama M, Jin N, Chain JL, Huan Y et al. Peptide antigens for $\gamma\delta$ T cells. Cell Mol Life Sci 2011; 68: 2335–23343.
- Fu YX, Kersh G, Vollmer M, Kalataradi H, Heyborne K, Reardon C et al. Structural requirements for peptides that stimulate a subset of $\sqrt{\delta}$ T cells. *J Immunol* 1994; 152: 1578–1588.
- 92 O'Brien RL, Fu YX, Cranfill R, Dallas A, Reardon C, Lang J et al. Heat shock protein Hsp-60 reactive $\gamma\delta$ cells: a large, diversified T lymphocyte subset with highly focused specificity. Proc Natl Acad Sci U S A 1992; 89: 4348-4352.
- 93 Blond-Elguindi S, Cwirla SE, Dower WJ, Lipshutz RJ, Sprang SR, Sambrook JF et al. Affinity panning of a library of peptides displayed on bacteriophages reveals the binding specificity of BiP. Cell 1993; 75: 717-728.
- Mohan JF, Levisetti MG, Calderon B, Herzog JW, Petzold SJ, Unanue ER. Unique autoreactive T cells recognize insulin peptides generated within the islets of Langerhans in autoimmune diabetes. Nat Immunol 2010; 4: 350–354.
- 95 Zhang L, Nakayama M, Eisenbarth GS. Insulin as an autoantigen in NOD/human diabetes. Curr Opin Immunol 2008; 20:111-118.
- 96 Holoshitz J, Vila LM, Keroack BJ, McKinley DR, Bayne NK. Dual antigenic recognition by cloned human $\gamma\delta$ T cells. J Clin Invest 1992; 89: 308-314.
- 97 Vidovic D, Roglic M, McKune K, Guerder S, MacKay C, Dembic Z. Qa-1 restricted recognition of foreign antigen by a $\gamma\delta$ T-cell hybridoma. Nature 1989; 340: 646–650.
- 98 Modlin RL, Pirmez C, Hofman FM, Torigian V, Uyemura K, Rea TH et al. Lymphocytes bearing antigen-specific $\gamma\delta$ T-cell receptors in human infectious disease lesions. Nature 1989; 339: 544–548.
- 99 Kabelitz D, Bender A, Schondelmaier S, Schoel B, Kaufmann SHE. A large fraction of human peripheral blood $v\delta^+$ T cells is activated by *Mycobacterium tuberculosis* but not by its 65 kD heat shock protein. J Exp Med 1990; 171: 667–679.
- 100 Pfeffer K, Schoel B, Gulle H, Kaufmann SHE, Wagner H. Primary responses to human T cells to mycobacteria: a frequent set of $\sqrt{\delta}$ T cells are stimulated by proteaseresistant ligands. Eur J Immunol 1990; 20: 1175–1179.
- 101 Constant P, Davodeau F, Peyrat MA, Poquet Y, Puzo G, Bonneville M et al. Stimulation of human $\sqrt{6}$ T cells by nonpeptidic mycobacterial ligands. Science 1994; 264: 267– 270.
- 102 Tanaka Y, Sano S, Nieves E, de Libero G, Rosa D, Modlin RL et al. Nonpeptide ligands for human $v\delta$ T cells. Proc Natl Acad Sci U S A 1994: 91: 8175–8179.
- 103 Tanaka Y, Morita CT, Tanaka Y, Nieves E, Brenner MB, Bloom BR. Natural and synthetic non-peptide antigens recognized by human $\gamma\delta$ T cells. Nature 1995; 375: 155–158.
- 104 Hintz M, Reichenberg A, Altincicek B, Bahr U, Gschwind RM, Kollas AK et al. Identification of (E)-4-hydroxy-3-methyl-but-2-enyl pyrophosphate as a major activator for human $\gamma\delta$ T cells in Escherichia coli. FEBS Lett 2001; 509: 317–322.
- 105 Wang H, Fang Z, Morita C. V γ 2V δ 2 T cell receptor recognition of prenyl pyrophosphates is dependent on all CDRs. J Immunol 2010; 184: 6209–6222.
- 106 Sarikonda G, Wang H, Puan KJ, Liu XH, Lee HK, Song Y et al. Photoaffinity antigens for human $\gamma\delta$ T cells. J Immunol 2008; 181: 7738-7750.
- 107 Wei H, Huang D, Lai X, Chen M, Zhong W, Wang R et al. Definition of APC presentation of phosphoantigen (E)-4-hydroxy-3-methyl-but-2-enyl pyrophosphate to V γ 2V δ 2 TCR. J Immunol 2008; 181: 4798–4806.
- 108 Gober HJ, Kistowska M, Angman L, Jeno P, Mori L, de Libero G. Human T cell receptor $\gamma\delta$ cells recognize endogenous mevalonate metabolites in tumor cells. J Exp Med 2003; 197:163–168.
- 109 Thompson K, Rojas-Navea J, Rogers MJ. Alkylamines cause V γ 9V δ 2 T cell activation and proliferation by inhibiting the mevalonate pathway. Blood 2006; 107: 651-654.
- 110 Kunzmann V, Bauer E, Wilhelm M. γ/δ T-cell stimulation by pamidronate. New Engl J Med 1999; 340: 737–738.
- 111 Bukowski JF, Morita CT, Brenner MB. Human $\gamma\delta$ T cells recognize alkylamines derived from microbes, edible plants, tea: implications for innate immunity. Immunity 1999; 11: 57–65.
- 112 Matsuura E, Lopez LR, Shoenfeld Y, Ames PRJ. β 2 glycoprotein I and oxidative inflammation in early atherogenesis: a progression from innate to adaptive immunity? Autoimmun Rev; e-pub ahead of print 2012 Apr 27; doi: 10.1016/ j.autrev.2012.04.003.