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## Butyrophilin 3A1 presents phosphoantigens to human $\gamma\delta$ T cells: the fourth model of antigen presentation in the immune system

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T lymphocytes are major players in immunity, providing protection against pathogenic microorganisms and tumors. They achieve this goal because they express clonally distributed receptors (T-cell receptor, TCR) on their surface capable of recognizing antigens displayed on the surface of specialized antigen-presenting cells, in association with antigen presenting molecules.<sup>1</sup>

The situation for  $\gamma\delta$  T cells, an unconventional subset of so called innate memory T cells, has long remained obscure.<sup>2</sup> In the blood of most healthy individuals, the predominant  $\gamma\delta$  T cell population consists of T cells expressing the V $\delta$ 2 gene paired with the V $\gamma$ 9 chain.

 $V\gamma 9V\delta 2$  T cells recognize host- and microbe-derived phosphorylated prenyl metabolites (phosphoantigens, PAgs),<sup>3</sup> but the molecular basis for PAg recognition has long remained a major unanswered question in the biology of  $V\gamma 9V\delta 2$  T cells.

In a study published in the September issue of *Nature Immunology*, De Libero and colleagues<sup>4</sup> shed light on the mechanism of  $V\gamma9V\delta2$  T-cell recognition of PAgs and identify butyrophilin (BTN) 3A1 as the PAg-presenting molecule. Many previous studies have been consistent with the existence of a PAg-presenting molecule:

- The small size of the stimulatory PAgs and the requirement for cellto-cell contact have suggested that PAgs bind a specialized antigen presenting molecules. Such a putative molecule should be species specific, ubiquitously expressed and functionally nonpolymorphic.<sup>5</sup>
- None of the known human presenting molecules (major histocompatibility complex (MHC) class I and class II, MR1 or CD1) is involved in PAg presentation<sup>5,6</sup> and antigen processing is not required, likely explaining why PAgs can be presented by non professional antigen-presenting cell, including Vγ9Vδ2 T cells themselves.<sup>5,6</sup>
- 3. The findings that PAgs fail to bind directly to the soluble  $V\gamma9V\delta2$  TCR and that the putative antigen-binding groove of the  $V\gamma9V\delta2$  TCR is much larger than that occupied by the PAg alone, further suggest interactions with a complex formed by the PAg and a PAg-presenting molecule.<sup>7,8</sup>

Therefore, all these observations strongly support the requirement for a surface protein(s) distinct from all known antigen-presenting molecules for PAg presentation.

Identification of a PAg-presenting molecule, however, has been complicated by the fact the human  $V\gamma 9V\delta 2$  T

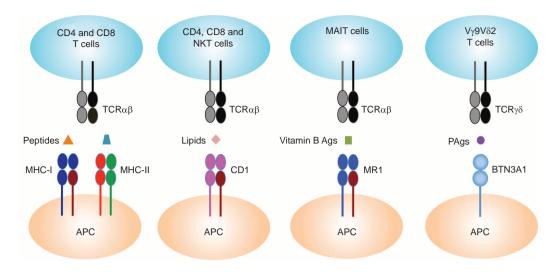
cells are able to 'autopresent' PAgs to themselves, thus preventing results' interpretation.<sup>5,9</sup> De Libero and colleagues<sup>4</sup> have developed a new strategy to avoid the autopresentation phenomenon: they have generated mice with transgenic expression of the V $\gamma$ 9V $\delta$ 2 TCR and have tested mouse–human hybrid lines for their ability to present PAgs to V $\gamma$ 9V $\delta$ 2 T cells from the transgenic mice.

Using this approach, De Libero and colleagues<sup>4</sup> have found that the gene coding for a candidate PAg-presenting molecule is located in the 3-27.4 telomeric megabases region (p25.2-p22.1) of human chromosome 6. Exploiting the ubiquitous expression and the presence of two immunolgobulin-like extracellular domains of the putative PAg-binding molecule to narrow the search, De Libero and colleagues<sup>4</sup> identify several members of the BTN (B7related molecules of the immunoglobulin superfamily) family as potential candidate. Finally, using transfection and small interfering RNA, they finally demonstrate that BTN3A1 is the key PAg-presenting molecule.

Extensive biophysical and structural experiments provide evidence for a direct role for BTN3A1 in PAg presentation. By mass spectroscopy and surface plasmon resonance, De Libero and colleagues<sup>4</sup> demonstrate binding of PAgs to the immunoglobulin V domain of BTN3A1 and crystal structures of the immunoglobulin V domain of BTN3A1 with PAgs identify a shallow groove on

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**Figure 1** TCR recognition of peptides, lipids, vitamin B Ags and PAgs. MHC class I and II molecules present peptides to CD8 and CD4 T cells, respectively; MHC class I-like molecules CD1 present lipids to CD4, CD8 and NKT cells; the MHC class I-related protein MR1 presents vitamin B metabolites to MAIT cells and BTN3A1 presents PAgs to  $V\gamma$ 9V $\delta$ 2 T cells. Ag, antigen; BTN, butyrophilin; MHC, major histocompatibility complex; NKT, natural killer T; PAg, phosphoantigen; TCR, T-cell receptor.

the protein surface that accommodates a single PAg.

Finally, by measuring surface-enhanced Raman scattering, the authors show that the V $\gamma$ 9V $\delta$ 2 TCR interacts directly, but weakly, with BTN3A1, and that PAg enhances this interaction.

Despite the study by De Libero *et al.* makes key advances in antigen recognition by  $V\gamma 9V\delta 2$  T cells, several questions remain unanswered. The major one concerns the role of the intracellular B30.2 domain of BTN3A1, which is required for  $V\gamma 9V\delta 2$  T-cell activation.

Earlier study suggested that PAgs modify the BTN3A1 molecule *via* interaction with the B30.2 cytoplasmic region, which would potentially involve dimerization and clustering.<sup>10,11</sup>

Another study by Morita and colleagues<sup>12</sup> found that a predicted PAg-binding pocket only exists on the intracellular B30.2 domain of the BTN3A1. Nonetheless, BTN3A1 did not bind a photo affinity-labeled PAg, thus indicating that neither extracellular nor the cytoplasmic B30.2 domains actually was capable to bind PAgs.<sup>12</sup>

De Libero and colleagues<sup>4</sup> now find that only the extracellular V domain of BTN3A1 contains a PAg-binding pocket and suggest that the B30.2 domain might be involved in the trafficking of BTN3A1 to the proper intracellular compartments where it is loaded with PAgs. In support to this, De Libero *et al.*<sup>4</sup> show that BNT3A2, which shares a similar V domain with BTN3A1, but lacks B30.2, fails to stimulate  $V\gamma9V\delta2$  T cells.

Unfortunately however, the authors do not provide evidence of the capability of BTN3A2 to bind PAg, which leaves unsolved the question of whether the intracytoplasmic B30.2 domain influences PAg binding or PAg/BTN3A1 complexes are necessary, but not sufficient to activate  $V\gamma 9V\delta 2$  T cells.

Solving this issue may help in understanding how PAgs are trafficked to and loaded onto BTN3A1, and whether or not this requires specialized enzymes. Many different chaperons promote loading of peptides into MHC class I and II, and of glycolipids into CD1 molecules: specialized enzymes are also capable of covalent addition of isoprenoids to proteins (a process known as protein penylation), but whether or not similar mechanisms also promote PAgs loading into BTN3A1 is worth of additional studies.

Finally, De Libero *et al.*<sup>4</sup> show that the V $\gamma$ 9V $\delta$ 2 TCR has a very weak affinity for BTN3A1 and PAg only modestly enhances it: thus, additional structural studies are needed to understand how PAg–BTN3A1 complexes are recognized by the V $\gamma$ 9V $\delta$ 2 TCRs.

The thymus 'educates' T lymphocytes to recognize foreign antigens in a genetically

restricted fashion. It is already known that there are three families of molecules involved in this:1 MHC class I and II molecules present peptides to CD8 and CD4 T cells, respectively; another MHClike family of proteins called CD1 present glycolipids to CD4, CD8 and natural killer T cells; a third MHC-related protein called MR1 (MHC-related 1) presents vitamin B metabolites to a class of immune T cells called MAIT (mucosaassociated invariant T) cells:<sup>13</sup> the paper by De Libero et al.4 now provides evidence that BTN3A1 presents PAg to  $V\gamma 9V\delta 2$  T cells (Figure 1), and it therefore defines a fourth model of antigen presentation to immune cells.

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