

Mechanistic Consequences of Chiral Radical Clock Probes: Analysis of the Mononuclear Non-Heme Iron Enzyme HppE with 2-Hydroxy-3-methylenecyclopropyl Radical Clock Substrates

Hui Huang,^{†,‡} Wei-Chen Chang,^{†,‡} Geng-Min Lin,[†] Anthony Romo,[†] Pei-Jing Pai,[§] William K. Russell,[§] David H. Russell,[§] and Hung-Wen Liu^{*,†}

[†]Division of Medicinal Chemistry, College of Pharmacy, and Department of Chemistry, University of Texas at Austin, Austin, Texas 78712, United States

[§]Department of Chemistry, Texas A&M University, College Station, Texas 77842, United States

Supporting Information

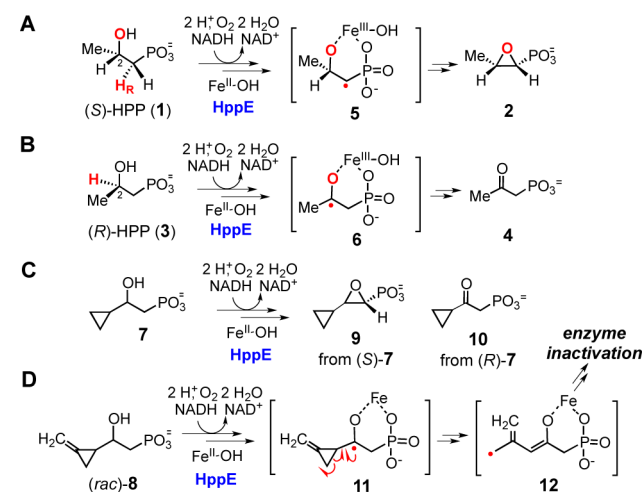
ABSTRACT: (*S*)-2-Hydroxypropylphosphonic acid [(*S*)-HPP] epoxidase (HppE) is a mononuclear iron enzyme that catalyzes the last step in the biosynthesis of the antibiotic fosfomycin. HppE also processes the (*R*)-enantiomer of HPP but converts it to 2-oxo-propylphosphonic acid. In this study, all four stereoisomers of 3-methylenecyclopropyl-containing substrate analogues, (2*R*, 3*R*)-8, (2*R*, 3*S*)-8, (2*S*, 3*R*)-8, and (2*S*, 3*S*)-8, were synthesized and used as radical probes to investigate the mechanism of the HppE-catalyzed reaction. Upon treatment with HppE, (2*S*, 3*R*)-8 and (2*S*, 3*S*)-8 were converted via a C1 radical intermediate to the corresponding epoxide products, as anticipated. In contrast, incubation of HppE with (2*R*, 3*R*)-8 led to enzyme inactivation, and incubation of HppE with (2*R*, 3*S*)-8 yielded the 2-keto product. The former finding is consistent with the formation of a C2 radical intermediate, where the inactivation is likely triggered by radical-induced ring cleavage of the methylenecyclopropyl group. Reaction with (2*R*, 3*S*)-8 is predicted to also proceed via a C2 radical intermediate, but no enzyme inactivation and no ring-opened product were detected. These results strongly suggest that an internal electron transfer to the iron center subsequent to C–H homolysis competes with ring-opening in the processing of the C2 radical intermediate. The different outcomes of the reactions with (2*R*, 3*R*)-8 and (2*R*, 3*S*)-8 demonstrate the need to carefully consider the chirality of substituted cyclopropyl groups as radical reporting groups in studies of enzymatic mechanisms.

Mononuclear non-heme iron enzymes are an important class of biocatalysts that are widespread in nature.¹ The catalytic functions of these enzymes are remarkably diverse, but the involvement of a high valent Fe(IV)-oxo (ferryl) species as a reactive intermediate is common to all their catalytic mechanisms. This reactive iron–oxygen species abstracts a hydrogen atom, typically from an unactivated carbon of the substrate, to generate a substrate radical, which then undergoes further transformations en route to the final product.² Despite the proposed intermediacy of substrate radicals in various mononuclear non-heme iron enzyme mechanisms, only a limited

number of studies provide evidence for the radical intermediates during turnover.³

(*S*)-2-Hydroxypropylphosphonic acid epoxidase (HppE), a mononuclear non-heme iron enzyme, performs an epoxidation reaction on (*S*)-2-hydroxypropylphosphonic acid ((*S*)-HPP, **1**) to form fosfomycin (**2**) in the last step of the biosynthesis of this clinically useful antibiotic.^{4,5} Previous studies showed that the epoxide ring of **2** is generated by an initial abstraction of 1-H_R from (*S*)-HPP and subsequent C–O bond formation between C1 and the 2-OH group of (*S*)-HPP in a dehydrogenation–cyclization manner (Scheme 1A).^{4–7} When HppE is incubated with (*R*)-HPP (**3**), the 2-H is selectively abstracted, leading to direct oxidation of the alcohol moiety to give 2-oxopropylphosphonic acid (**4**) as the sole product (Scheme 1B).⁸ Crystallographic and spectroscopic experiments have shown that both

Scheme 1. Proposed Mechanism for the Oxidation of (*S*)- and (*R*)-HPP by HppE^a



^a(A) Conversion of (*S*)-HPP (**1**) to fosfomycin (**2**). (B) Conversion of (*R*)-HPP (**3**) to 2-oxopropylphosphonic acid (**4**). (C) Conversion of (*S*)-7 to **9** and (*R*)-7 to **10** by HppE. (D) Proposed mechanisms for the inactivation of HppE by **8**.

Received: October 2, 2013

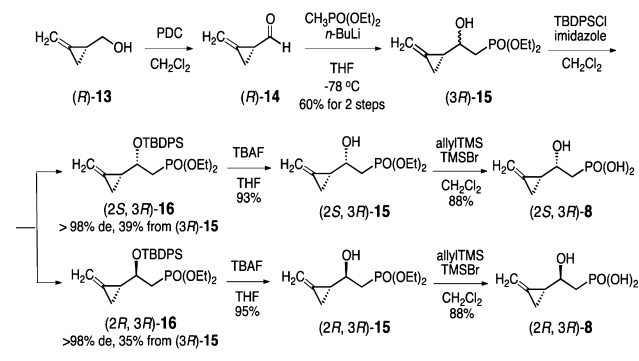
Published: February 10, 2014

(*S*)- and (*R*)-HPP bind to the iron center of the enzyme as bidentate ligands.⁹ The C2 chirality of the substrates governs the binding geometry such that regio- and stereoselective hydrogen atom abstraction (1-*H_R* for (*S*)-HPP and 2-*H* for (*R*)-HPP) is facilitated by the reactive iron–oxygen species^{10,11} giving different radical intermediates (**5** or **6**).¹²

Recently, we reported mechanistic studies of HppE with the 2-hydroxyl-3-cyclopropyl and 2-hydroxyl-3-methylenecyclopropyl compounds (*S*)-**7**, (*R*)-**7**, and (*rac*)-**8**.¹³ Both (*S*)- and (*R*)-**7** were efficiently converted by HppE to the epoxide and ketone products **9** and **10**, respectively (Scheme 1C). In contrast, when HppE was incubated with (*rac*)-**8**, only a small portion of the substrate was converted to new products before the enzyme was inactivated. It was suggested that inactivation results from ring-opening of the C3 methylenecyclopropyl group triggered by the C2 radical (**11**) generated during turnover (Scheme 1D). These studies provided preliminary evidence for a radical intermediate in the HppE-catalyzed reactions.¹³ However, (*rac*)-**8** used in the experiment was a mixture of four stereoisomers, which complicated the analysis of the reaction outcomes and thus hampered investigation of the inactivation mechanism. To gain further mechanistic insight and verify our previous observations, we synthesized all four stereoisomers of **8** and studied their reactions with HppE. Our results, reported herein, show that HppE reactivity toward different diastereomers of **8** is dictated not only by the chirality of the C2 hydroxyl group but also the methylenecyclopropyl substituent at C3. Specifically, it was found that incubation of the (*2R, 3R*)-isomer with HppE leads to enzyme inactivation, whereas reaction with the (*2R, 3S*)-isomer gives rise to complete turnover to a 2-keto product. These results are mechanistically informative and shed light on the likely formation of a carbocation intermediate in HppE catalysis.

The syntheses of all four diastereomers of **8** began by addressing the chirality of the C3 methylenecyclopropyl group. This was achieved by following the previously reported enantioselective synthetic method for alcohols (*S*)- and (*R*)-**13**.¹⁴ Oxidation of (*R*)-**13** produced aldehyde (*R*)-**14**, and subsequent nucleophilic addition with methyl-diethylphosphonate gave (*2S, 3R*)- and (*2R, 3R*)-**15** as a pair of diastereomeric alcohols (Scheme 2). Direct separation or enzymatic resolution

Scheme 2. Synthetic Scheme for the Preparation of (*2S, 3R*)- and (*2R, 3R*)-8****

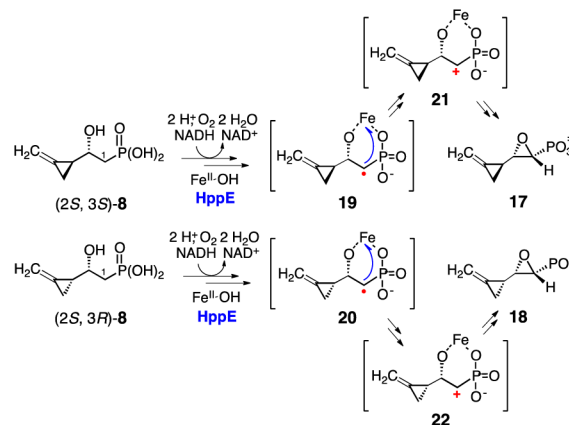


(using lipases or esterases) of these two diastereomers did not give satisfactory results. Consequently, we focused on chemical resolution by derivatizing the C2 hydroxyl group with a TBDPS group to inflict internal steric hindrance and to introduce a UV chromophore for product detection during purification. It was hoped that the attachment of such a bulky substituent would

limit the free rotation of the C2–C3 bond connecting the two neighboring chiral centers and, therefore, magnify the diastereomeric difference. This strategy generated two TBDPS-ethers, (*2S, 3R*)- and (*2R, 3R*)-**16**, which were successfully separated by silica gel chromatography. Removal of the TBDPS group yielded pure (*2S, 3R*)- and (*2R, 3R*)-**15**, enabling chirality and enantiomeric excess determination of the C2 alcohol by the Mosher ester method.¹⁵ Subsequent deprotection by TMSBr gave the desired substrate analogues (*2S, 3R*)- and (*2R, 3R*)-**8**. Following the same procedure, (*2S, 3S*)- and (*2R, 3S*)-**8** were obtained using (*S*)-**13** as the starting material.

The four purified diastereomers of **8** were each incubated with HppE, and the reactions were monitored using real-time ¹H NMR spectroscopy. All four compounds were substrates for HppE, but each reaction gave a different result. The 1,2-epoxide **17** was produced when (*2S, 3S*)-**8** was employed as the substrate, whereas formation of a different 1,2-epoxide, **18**, was detected in the reaction with (*2S, 3R*)-**8** (see Figures S3, S4).¹⁶ These results are consistent with what was previously observed for (*rac*)-**8** with HppE¹³ and support the regio- and stereoselective conversions of (*2S, 3S*)- and (*2S, 3R*)-**8** proposed in Scheme 1A. Specifically, when these substrate analogues bearing C2-(*S*) hydroxyl group react with HppE, the putative C1 centered radicals (**19** and **20**) are produced by selective hydrogen abstraction with the high-valent Fe(IV)-oxo species. These C1 radicals are not affected by the methylenecyclopropyl group at C3 position and undergo cyclization to the corresponding epoxides **17** and **18** in a manner similar to the formation of **2** from **1** (Scheme 3). Our more

Scheme 3. Conversion of (*2S, 3S*)-8** to **17** and (*2S, 3R*)-**8** to **18** by HppE**

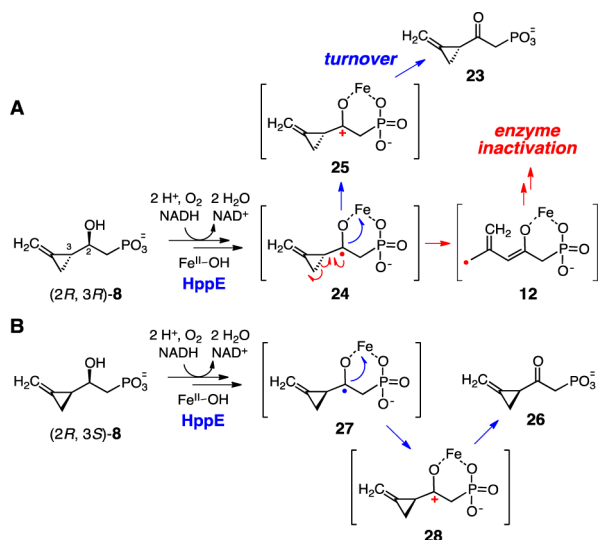


recent results suggest that cyclization of the resulting radical intermediate (**19** and **20**) to the epoxide product likely involves a carbocation intermediate (**21** and **22**) via internal electron transfer to reduce the iron center.¹⁷

Based on the mechanism proposed in Scheme 1B, incubation of the C2-(*R*) analogue, (*2R, 3R*)-**8**, with HppE is expected to proceed with regioselective C-2 hydrogen atom abstraction, where the resulting C2 radical will trigger ring-opening of the neighboring methylenecyclopropyl group¹⁸ leading to enzyme inactivation. Interestingly, incubation of (*2R, 3R*)-**8** with HppE produces a small amount of new product as detected by ¹H NMR spectroscopy (see Figure S1). However, product formation stopped with no further consumption of the substrate after ~10 min. Moreover, no fosfomycin could be detected when the natural substrate, (*S*)-HPP (**1**), was incubated with HppE that had been preincubated with (*2R, 3R*)-**8**, suggesting complete

inactivation of HppE by the pretreatment. The turnover product of this process was later determined to be ketone **23**, which had been previously shown not to be an inactivator of HppE.¹³ Thus, a logical explanation for the inactivation of HppE by (2*R*, 3*R*)-**8** is a mechanism initiated by a radical-triggered ring-opening of the C3-(*R*)-methylene cyclopropyl group (Scheme 4A). The fact that

Scheme 4. (A) HppE Inactivation by (2*R*, 3*R*)-8** and (B) Conversion of (2*R*, 3*S*)-**8** to **26** by HppE**



(2*R*, 3*R*)-**8** is an inactivator and also a substrate for HppE reveals a competition between the radical-induced ring-opening (24 → 12) and the internal electron-transfer (24 → 25) reactions.

For the other C2-(*R*) diastereomer, (2*R*, 3*S*)-**8**, a similar C2 radical-triggered ring-opening reaction with subsequent enzyme inactivation was also expected. However, HppE remained active after incubation with (2*R*, 3*S*)-**8**, and NMR analysis of the incubation showed that (2*R*, 3*S*)-**8** was completely transformed into ketone **26** without formation of ring-opened products (Scheme 4B and Figure S2). Given that the methylenecyclopropyl group is a highly sensitive radical probe,^{18,19} the different outcomes of these two reactions may lie in the orientation of the methylenecyclopropyl group of **8** in the enzyme active site. Specifically, the three-membered ring is more readily opened when the C3–C4 bond of the methylenecyclopropyl group and the singly occupied orbital of the radical adopt an anticoplanar conformation¹⁸ (the ring-opening rate of a methylenecyclopropyl carbinyl radical in free solution is $\sim 6 \times 10^9 \text{ s}^{-1}$).¹⁹ It is possible that such an anticoplanar conformation is inaccessible for (2*R*, 3*S*)-**8** due to steric interference between the methylenecyclopropyl moiety and the groups at the enzyme active site. Thus, electron transfer from the radical intermediate **27** to the iron center (27 → 28) becomes the path of choice leading to a 2-keto product (**26**) instead of the ring-opened product(s).

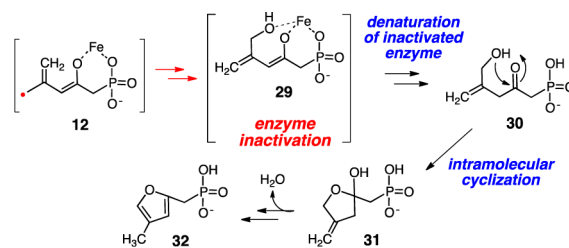
To gain more insight into the mechanism of inhibition by (2*R*, 3*R*)-**8**, the kinetics of the reaction were determined using HPLC. When HppE was incubated with 10-fold excess of (2*R*, 3*R*)-**8**, time-dependent inactivation was observed, and the enzyme completely lost activity within 3 min (Figure S6). When (*S*)-HPP (**1**) was coinubated with (2*R*, 3*R*)-**8** (molar ratio of 1:8 = 20:1), fosfomycin production was still discernible after 8 min, indicating a reduction of the rate of inactivation in the presence of native substrate. The observed substrate protection implies that the inhibition is active-site directed. When HppE was treated with

different amounts of (2*R*, 3*R*)-**8**, a plot of the residual enzyme activity versus the equivalents of (2*R*, 3*R*)-**8** employed resulted in a partition ratio of 0.5 (Figure S6), reflecting that ~ 1.5 equiv of (2*R*, 3*R*)-**8** are required to fully inactivate 1.0 equiv of HppE. Moreover, the inactivation appears to be irreversible since enzyme activity could not be regenerated after extensive dialysis.

Tritiated (*rac*)-[3-³H]-**8** was also synthesized and incubated with HppE to determine the stoichiometry of inactivation.¹⁶ After HppE was completely inactivated by (*rac*)-[3-³H]-**8**, the reaction mixture was subjected to size exclusion chromatography followed by dialysis to remove unbound small molecules. About 1.1 equiv of tritiated compound per enzyme were found to be associated with the inactivated HppE.¹⁶ Interestingly, when this protein sample was denatured by boiling, over 80% of the radioactivity was lost from the protein fraction after size exclusion centrifugal filtration.¹⁶ In addition, no covalently modified peptide fragment was detected by mass spectrometry analysis of the inactivated HppE after denaturation and tryptic digestion (see Figure S7). Taken together, these results strongly suggest that the inactivation is not due to covalent modification of HppE, as previously proposed,¹³ but is likely due to tight binding of a ring-opened product to the enzyme active site.

It is conceivable that the causative agent for the observed inactivation is compound **29**, which arises from canonical hydroxyl group rebound to the initially formed acyclic radical intermediate (**12**, Scheme 5). The tridentate coordination of **29**

Scheme 5. Proposed HppE Inactivation by the Ring-Opened Product Derived From (2*R*, 3*R*)-8****



to the iron center could render **29** to bind strongly in the active site of HppE, resulting in enzyme inactivation. However, efforts to isolate and identify this putative tight binding species after protein denaturation were unsuccessful. It was later found during our attempt to chemically synthesize **30** (the keto tautomer of **29**) that this compound readily undergoes intramolecular cyclization to give **32** as the main product.¹⁶ The fact that a low abundance signal at m/z 175 [$M - 1$]⁻ was detected by LC-MS analysis in the denatured enzymatic sample (but not in the control) is consistent with the presence of **32** in the protein-free solution after enzyme denaturation. Unfortunately, the low abundance of this signal prevented MS-MS analysis to further verify its structure.

The results presented herein are significant for three reasons. First, a highly effective inhibitor for HppE, (2*R*, 3*R*)-**8**, has been identified. The inactivation is mechanism-based, since assays using NMR, HPLC, MS, and radioactive methods indicate that the inactivation arises from the methylenecyclopropyl ring-opening triggered by the C2-centered radical. However, unlike most known suicide inhibitors, the inactivation by (2*R*, 3*R*)-**8** does not involve covalent modification of the targeted enzyme but is likely due to tight binding of a ring-opened product in the active site.

Second, to “capture” radical intermediate(s) in a reaction with transient lifetimes, the sensitivity of the cyclopropylcarbinyl radical probe can be fine-tuned by adding substituent(s) to the cyclopropyl ring.¹⁸ Such modifications usually introduce new chiral center(s) and inflict steric demands upon binding to the chiral environment of the active site. However, the effect of the chirality of the substituted cyclopropyl ring of the probe on the outcomes of the radical triggered ring-opening reaction has rarely been addressed.^{20,21} In this study, enzymatic reactions with (2R, 3R)- and (2R, 3S)-**8** led to different results, even though both are believed to involve a C2 radical intermediate. The distinct outcomes may simply reflect different binding geometries of (2R, 3R)- and (2R, 3S)-**8** in the enzyme active site, and underscores the need to consider the chirality of substituted cyclopropyl moiety in a radical probe and its binding mode in the active site in studies of enzyme mechanisms. One also needs to be cautious to interpret the experimental data when no predicated ring-opening/rearrangement happens.

Third, the intermediacy of a substrate-derived cation in the catalytic cycle of HppE was recently proposed based on the observed 1,2-phosphono migration with an alternative substrate (R)-1-HPP.¹⁷ The observation of both enzyme inactivation and 2-keto product formation upon incubation of HppE with (2R, 3S)-**8** is mechanistically significant, as it reveals that an internal electron transfer (leading to the 2-keto product **23**) is a competing pathway with ring-opening (leading to enzyme inactivation) in the processing of the C2 radical intermediate. Thus, these results provide additional evidence supporting the formation of a cation intermediate in HppE catalysis and permit estimation of the rate constant for internal electron transfer to be on the order of $\sim 6 \times 10^9 \text{ s}^{-1}$, which is the reported rate constant for ring-opening of the methylenecyclopropylcarbinyl radical.¹⁹

■ ASSOCIATED CONTENT

Supporting Information

Details of experimental conditions and procedures. This material is available free of charge via the Internet at <http://pubs.acs.org>.

■ AUTHOR INFORMATION

Corresponding Author

h.w.liu@mail.utexas.edu

Author Contributions

[‡]These authors contributed equally.

Notes

The authors declare no competing financial interest.

■ ACKNOWLEDGMENTS

We thank Steve Sorey for his help in acquiring the NMR spectra. This work is supported in part by grants from the National Institutes of Health (GM040541 to H.-W.L.) and the Welch Foundation (F-1511 to H.-W.L. and A-1176 to D.H.R.).

■ REFERENCES

- (1) Buijninx, P. C. A.; van Koten, G.; Klein Gebbink, R. J. M. *Chem. Soc. Rev.* **2008**, *37*, 2716.
- (2) (a) Fitzpatrick, P. F. *Annu. Rev. Biochem.* **1999**, *68*, 355. (b) Costas, M.; Mehn, M. P.; Jensen, M. P.; Que, L., Jr. *Chem. Rev.* **2004**, *104*, 939. (c) Bollinger, J. M., Jr.; Krebs, C. *Curr. Opin. Chem Biol.* **2007**, *11*, 151. (d) Kovaleva, E. G.; Lipscomb, J. D. *Nat. Chem. Biol.* **2008**, *4*, 186. (e) van der Donk, W. A.; Krebs, C.; Bollinger, J. M., Jr. *Curr. Opin. Struct. Biol.* **2010**, *20*, 673.
- (3) For examples: (a) Baldwin, J. E.; Adlington, R. M.; Dornayehayman, B. P.; Knight, G.; Ting, H. H. *J. Chem. Soc., Chem.*

Commun. **1987**, 1661. (b) Ryle, M. J.; Liu, A.; Muthukumar, R. B.; Ho, R. Y. N.; Koehntop, K. D.; McCracken, J., Jr.; Que, L.; Hausinger, R. P. *Biochemistry* **2003**, *42*, 1854. (c) Howard-Jones, A. R.; Elkins, J. M.; Clifton, I. J.; Roach, P. L.; Adlington, R. M.; Baldwin, J. E.; Rutledge, P. J. *Biochemistry* **2007**, *46*, 4755.

(4) Hidaka, T.; Goda, M.; Kuzuyama, T.; Takei, N.; Hidaka, M.; Seto, H. *Mol. Gen. Genet.* **1995**, *249*, 274.

(5) (a) Liu, P.; Murakami, K.; Seki, T.; He, X.; Yeung, S. M.; Kuzuyama, T.; Seto, H.; Liu, H.-w. *J. Am. Chem. Soc.* **2001**, *123*, 4619. (b) Liu, P.; Liu, A.; Yan, F.; Wolfe, M. D.; Lipscomb, J. D.; Liu, H.-w. *Biochemistry* **2003**, *42*, 11577.

(6) Thibodeaux, C. J.; Chang, W.-c.; Liu, H.-w. *Chem. Rev.* **2012**, *112*, 1681.

(7) (a) Hammerschmidt, F.; Bovermann, G.; Bayer, K. *Liebigs Ann. Chem.* **1990**, 1055. (b) Hammerschmidt, F. *J. Chem. Soc. Perkin Trans. 1* **1991**, 1993.

(8) Zhao, Z.; Liu, P.; Murakami, K.; Kuzuyama, T.; Seto, H.; Liu, H.-w. *Angew. Chem., Int. Ed. Engl.* **2002**, *41*, 4529.

(9) (a) Higgins, L. J.; Yan, F.; Liu, P.; Liu, H.-w.; Drennan, C. L. *Nature* **2005**, *437*, 838. (b) Yun, D.; Dey, M.; Higgins, L. J.; Yan, F.; Liu, H.-w.; Drennan, C. L. *J. Am. Chem. Soc.* **2011**, *133*, 11262.

(10) Wang, C.; Chang, W.-c.; Guo, Y.; Huang, H.; Peck, S. C.; Pnadelia, M. E.; Lin, G.-m.; Liu, H.-w.; Krebs, C.; Bollinger, J. M., Jr. *Science* **2013**, *342*, 991.

(11) HppE had previously been considered an oxidase that uses Fe(III)-superoxo for H• abstraction. However, a recent study revealed that the preferred cosubstrate of HppE is H₂O₂ instead of O₂.¹⁰ Hence, HppE should now be classified as a non-heme iron peroxidase that uses Fe(IV)-oxo complex for hydrogen atom abstraction. It was also shown that slow infusion of H₂O₂ into the assay mixture is essential for high turnover and minimization of HppE inactivation during catalysis. Since H₂O₂ is generated *in situ* when O₂ is used with the NADH/FMN reducing system as described herein (and in our early work), the slow evolution of H₂O₂ from O₂ by the chemical reducing system remains an effective approach to assay HppE activity.

(12) (a) Woschek, A.; Wuggenig, F.; Peti, W.; Hammerschmidt, F. *ChemBioChem* **2002**, *3*, 829. (b) Yan, F.; Moon, S.-J.; Liu, P.; Zhao, Z.; Lipscomb, J. D.; Liu, A.; Liu, H.-w. *Biochemistry* **2007**, *46*, 12628.

(13) Huang, H.; Chang, W.-c.; Pai, P.-j.; Romo, A.; Mansoorabadi, S.; Russell, D.; Liu, H.-w. *J. Am. Chem. Soc.* **2012**, *134*, 16171.

(14) (a) Okuma, K.; Tanaka, Y.; Yoshihara, K.; Ezaki, A.; Koda, G.; Ohta, H.; Hara, K.; Kashimura, S. *J. Org. Chem.* **1993**, *58*, 5915. (b) Le Corre, M.; Hercouet, A.; Bessieres, B. *J. Org. Chem.* **1994**, *59*, 5483. (c) Chen, X.; Zemlicka, J. *J. Org. Chem.* **2002**, *67*, 286.

(15) (a) Ohtani, I.; Kusumi, T.; Ishitsuka, M. O.; Kakisawa, H. *Tetrahedron Lett.* **1989**, 3147. (b) Hoye, T. R.; Jeffrey, C. S.; Shao, F. *Nat. Protoc.* **2007**, *2*, 2451.

(16) See SI for experimental details.

(17) (a) Chang, W.-c.; Dey, M.; Liu, P.; Mansoorabadi, S. O.; Moon, S.-J.; Ahao, Z. K.; Drennan, C. L.; Liu, H.-w. *Nature* **2013**, *496*, 114.

(b) Chang, W.-c.; Mansoorabadi, S. O.; Liu, H.-w. *J. Am. Chem. Soc.* **2013**, *135*, 11457.

(18) (a) Griller, D.; Ingold, K. U. *Acc. Chem. Res.* **1980**, *13*, 317. (b) Nonhebel, D. C. *Chem. Soc. Rev.* **1993**, 347. (c) Newcomb, M. *Tetrahedron* **1993**, *49*, 1151. (d) Silverman, R. B. *Acc. Chem. Res.* **1995**, *28*, 335. (e) Newcomb, M.; Toy, P. H. *Acc. Chem. Res.* **2000**, *33*, 449. (f) Ortiz de Montellano, P. R. *Chem. Rev.* **2010**, *110*, 932.

(19) Horner, J. H.; Johnson, C. C.; Lai, M. T.; Lin, H.-w.; Martinesker, A. A.; Newcomb, M.; Oh, E. *Bioorg. Med. Chem. Lett.* **1994**, *4*, 2693.

(20) Spence, E. L.; Langley, G. J.; Bugg, T. D. H. *J. Am. Chem. Soc.* **1996**, *118*, 8336.

(21) For examples of covalent enzyme modification by a ring-opened butenyl radical: (a) Silverman, R. B.; Zhou, J. P.; Eaton, P. E. *J. Am. Chem. Soc.* **1993**, *115*, 8841. (b) Lai, M. T.; Liu, L. D.; Liu, H.-w. *J. Am. Chem. Soc.* **1991**, *113*, 7388. (c) Ziering, D. L.; Pascal, R. A. *J. Am. Chem. Soc.* **1990**, *112*, 834. (d) Bishwajit Paul, B.; Das, D.; Ellington, B.; Marsh, E. N. G. *J. Am. Chem. Soc.* **2013**, *135*, 5234.