

Original Article

2-Amino-nonyl-6-methoxyl-tetralin muriate inhibits sterol C-14 reductase in the ergosterol biosynthetic pathway

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Aim: To investigate the action mechanism of a novel chemical structural aminotetralin derivate, 2-Amino-Nonyl-6-Methoxyl-Tetralin Muriate (10b), against *Candida albicans* (*C. albicans*) in the ergosterol biosynthetic pathway.

Methods: Antifungal susceptibility test of 10b was carried out using broth microdilution method, the action mechanism of 10b against *C. albicans* was investigated by GC-MS spectrometry and real-time RT-PCR assay, and cytotoxicity of 10b *in vitro* was assessed by MTS/PMS reduction assay.

Results: 10b reduced the ergosterol content markedly, and the 50% ergosterol content inhibitory concentration (ECIC₅₀ value) was 0.08 µg/mL. Although the sterol composition of 10b-grown cells was completely identical with that of *erg24* strain, the content of ergosta-8,14,22-trienol in 10b-grown cells was much higher than that in *erg24* strain. Real-time RT-PCR assay revealed a global upregulation of sterol metabolism genes. In addition, the 50% inhibitory concentration (IC₅₀ value) of 10b was 11.30 µg/mL for murine embryonic fibroblasts and 35.70 µg/mL for human normal liver cells.

Conclusion: 10b possessed a mode of action different from that of azoles and morpholines, whose targets were sterol C-14 reductase (encoded by *ERG24* gene) and sterol C-5 desaturase (encoded by *ERG3*) related enzyme. Although 10b seemed to reduce MTS/PMS reduction in a dose dependent manner, IC₅₀ value for mammalian cells was much higher than 50% minimum inhibitory concentration (MIC₅₀) value for *C. albicans*. This indicates that the formulation is preliminarily safe and warrants further study for possible human applications.

Keywords: *Candida albicans*; sterol C-14 reductase; ergosterol; 2-amino-nonyl-6-methoxyl-tetralin muriate

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Introduction

Current therapy for invasive mycoses uses a relatively reduced number of antifungal drugs such as polyenes, azoles and echinocandins. The polyene antibiotic amphotericin B (AMB) remains to be the “gold standard” for the treatment of severe mycoses, but its severe acute and chronic toxicities such renal function impairment limit its clinical use^[1, 2]. Azoles are the most common agents for the treatment of *Candida* infections owing to their fungistatic and inhibitory effects on cytochrome P450 enzyme lanosterol demethylase (encoded

by *ERG11* gene), a key enzyme in ergosterol biosynthesis. Of them, fluconazole (FLC) and itraconazole (ITZ) are the drugs of choice for the treatment of severe systemic mycoses. Ketoconazole (KCZ) is widely used in the treatment of superficial mycoses. Nevertheless, azole resistance and cross-resistance have been documented in clinical *Candida* species^[3, 4]. Despite continuous expansion in the arsenal of antifungal drugs, antifungal drugs available cannot meet the increasing requirements for managing infections in medically complex patients.

The potential loss of efficacy of azoles has prompted concerted efforts to discover new drugs that might block fungal growth at different metabolic sites^[5]. Additional steps in ergosterol biosynthesis may be effective targets to be explored in this search. The reaction catalyzed by sterol C-14 reductase

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may be a good target for an antifungal suitable for human use. sterol C-14 reductase ends C-14 modification reactions initiated by cytochrome P-450 lanosterol 14 α -demethylase. An antifungal attack at this target by morpholines such as fenpropimorph (Fm) and amorolfine in *Saccharomyces cerevisiae* resulted in accumulation of ergosterol (ergosta-8,14-dienol), thus perturbing yeast membrane function and inhibiting the uptake of glucose and pyridines^[6]. Morpholines have been proved to be effective against fungal plant pathogens^[7], of which amorolfine has been successfully applied to the treatment of human tinea pedis, onychomycosis and other foot mycoses owing to its potent fungistatic and fungicidal activity *in vitro* against clinical isolates^[8–10].

2-Aminotetralin derivatives were synthesized as novel chemical structural antifungal agents by virtue of their high antifungal activity, broad antifungal spectra and potentially low toxicity^[11]. Of them, 10b (Figure 1) was found to have the strongest antifungal activity *in vitro*, especially against FLC-resistant *C albicans* strains. The aim of the present work was to further investigate the mechanism of 10b action against *C albicans* in the ergosterol biosynthetic pathway.

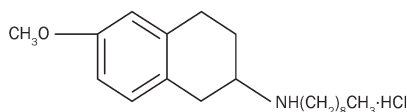


Figure 1. Chemical structure of 2-amino-nonyl-6-methoxy-tetralin muriate (10b).

Materials and methods

Antifungal agents

Antifungal reagents used in the present experiment included FLC solution (Pfizer, Inc, New York, NY, USA); ITZ and KCZ (Sigma, St Louis, MO, USA); Fm (Dr Ehrenstorfer, Inc, Augsburg, Germany); 10b (Department of Medicinal chemistry, School of Pharmacy, Shanghai, China); stock solutions of various concentrations in dimethyl sulfoxide (DMSO) (Sigma); and cell proliferation kit (MTS/PMS) (Promega Co, Madison, USA).

Organisms

Except for BWP17, NJ51-2, SC5314, and SC5314R, 24 *C albicans* strains were from the Dermatology Department of the First Hospital of Beijing University (Beijing, China). *C albicans* SC5314 was kindly provided by Dr William A FONZI from the Department of Microbiology and Immunology Georgetown University (Washington, USA). SC5314R was induced from SC5314 by FLC in our laboratory. *C albicans* strain BWP17 (*ura3 Δ ::imm434/ura3 Δ ::imm434/his1 Δ /his1 Δ /arg4 Δ /arg4 Δ) and NJ51-2 (*erg24/erg24*) were kindly provided by Dr Joseph HEITMAN from the Department of Molecular Genetics and Microbiology, Duke University Medical Center (Durham, NC, USA). NJ51-2 was derived from BWP17 with *C albicans* *ERG24* gene disrupted^[12]. The two strains (BWP17 and NJ51-2) were used*

for GC-MS assay. Uridine (80 μ g/mL) was added to ensure the growth of BWP17.

All strains were cultivated at 30 °C under constant shaking (200 r/min) in a liquid complete medium (YPD) consisting of 1% (*w/v*) yeast extract, 2% (*w/v*) peptone and 2% (*w/v*) dextrose. The synthetic dextrose medium containing 0.67% (*w/v*) yeast nitrogen base without amino acids and 2% (*w/v*) dextrose was buffered with 0.165 mol/L morpholinepropanesulfonic acid (MOPS; Sigma), with pH adjusted to 7.0 with NaOH.

BALB/C 3T3 murine embryo fibroblast and L-02 (HL-7702) human normal liver cells were obtained from the Chinese Academy of Sciences Cell Bank (Shanghai, China). All tissue culture plastics were purchased from TPP (Switzerland). 3T3 cells were routinely grown in the medium (Dulbecco's modified Eagle's medium (DMEM) supplemented with *L*-glutamine (4 mmol/L), penicillin (100 U/mL), streptomycin (100 μ g/mL) and 10% (*v/v*) calf bovine serum (CBS). L-02 human normal liver cells were routinely grown in the medium (RPMI 1640 supplemented with *L*-glutamine (4 mmol/L), penicillin (100 U/mL), streptomycin (100 μ g/mL) and 20% (*v/v*) CBS. The cultures were maintained in a 37 °C incubator in a humidified atmosphere of 95: 5 air: CO₂. The cells were grown to confluency in 75-cm² flasks, incubated at 37 °C with trypsin/EDTA solution for 2–5 min, pipetted to 96-well plates at the desired density (100 μ L), and allowed to attach for 24 h prior to exposure to 10b.

Antifungal susceptibility test

Drug sensitivities were assayed by microbroth dilution in 96-well plates according to methods of the Clinical and Laboratory Standards Institute (CLSI, formerly NCCLS) (M27-A2)^[13]. Briefly, cells were inoculated into successive wells of a 96-well microtiter plate containing serial two-fold dilutions of antifungal drugs in RPMI-1640 medium buffered to pH 7.0 with MOPS. The initial concentration of the fungal suspension in RPMI-1640 medium was 1 \times 10³ colony forming unit (CFU)/mL. After incubation at 30 °C for 24 h, absorbance at 630 nm was determined in an ELISA reader (Multiskan MK3). According to CLSI, we adopted *Candida parapsilosis* strain ATCC18062 as the quality control strain. The MIC₈₀ result that falls within the requested limits is 0.25–1.0 μ g/mL for fluconazole. Each isolate was tested in triplicate. Drug concentrations inhibiting growth to 50% and 80% of control levels (MIC₅₀ and MIC₈₀) were estimated by interpolation.

Sterol extraction

Cells were grown to the stationary phase in YPD liquid medium at 30 °C under constant shaking (200 r/min). Total sterols were extracted from whole cells based on the previous report^[14] with slight modifications. Briefly, *Candida* cells grown at 30 °C to 0.2 OD₆₀₀ unit/mL were treated with or without antifungal agents, and incubated for 16 h. Cells were then harvested by centrifugation at 3000 \times g at 4 °C for 5 min and washed three times with phosphate buffer saline (PBS). The net weight of the cell pellet (approximate 0.5 g) was determined. PBS 2.5 mL and 6 mL 15% alcoholic sodium

hydroxide solution (15 g NaOH and 10 mL sterile distilled water, brought to 100 mL with 100% ethanol) were added to each pellet and mixed by vortex for 1 min. Cell suspensions were then incubated in an 80 °C water bath for 1 h. Following incubation, tubes were allowed to cool to room temperature. Total sterols were then extracted (three times) with 6 mL petroleum ether. Upper phases were combined and transferred to a clean borosilicate glass screw-cap tube. Sterile distilled water 6 mL was added to each tube and swirled for 1 min. With water removed, the tubes were incubated in a 60 °C water bath until the petroleum ether layer volatilized completely. The resulting samples were stored at -20 °C for use after they were dissolved by 0.5 mL hexamethylene.

GC-MS spectrometry analyses of sterols were done with a Thermo Focus DSQ gas chromatograph interfaced to a FactorFour VF-5MS Mass Spectrometer. GC separation was performed on a fused silica column (VF-5MS; 30 m by 0.25 mm by 0.25 µm [film thickness]; Varian Inc). The injector temperature was 250 °C. The oven temperature was programmed to constant 100 °C for 1 min, followed by a temperature increase of 10 °C/min to a final temperature of 300 °C. The final temperature was maintained for 10 min. Helium was used as the carrier gas. The linear velocity was 1 ml/min with a split ratio 10:1. The mass spectrometer was in the electron impact ionization mode at 70 eV electron energy. The ion source temperature was 250 °C. Scanning was done from 41 to 460 atomic mass units at 0.6-s intervals. The concentration of 10b showing 50% reduction in ergosterol content (ECIC₅₀ value) was then calculated.

Quantitative real-time RT-PCR assay

Total RNA was extracted from *C. albicans* SC5314 in synthetic dextrose medium using Fungal RNAout kit (TIANDS, china). Reverse transcription was performed in a total volume of 20 µL with Avian Myeloblastosis Virus Reverse Transcriptase (TaKaRa), Random Primer (6-mer) (TaKaRa) and 1 µg total RNA, followed by the condition of 30 °C for 10 min, 45 °C for 15 min, and 99 °C for 2 min, as recommended by the manufacturer. Real-time PCR reactions were performed with SYBR Green I (TaKaRa), using LightCycler Real-Time PCR system (Roche Molecular Biochemical). Gene-specific primers were designed using Discovery Studio Gene software (Accelrys, Inc). The thermal cycling conditions comprised an initial step at 95 °C for 10 s, followed by 40 cycles at 95 °C for 10 s, 62 °C for 20 s, and 72 °C for 15 s. Change in SYBR Green I fluorescence in every cycle was monitored by the system software, and the threshold cycle (C_T) was measured. Using 18S rRNA as the internal control, gene expression of SC5314 strain treated by 10b relative to SC5314 strain without treatment was calculated using the formula $2^{-\Delta\Delta C_T}$, where ΔC_T was the C_T value of genes of interest minus that of the internal control, and $\Delta\Delta C_T$ was the mean ΔC_T value of SC5314 strain treated by 10b minus that of SC5314 strain without treatment. Primer sequences used in real-time RT-PCR assay are listed in Table 1.

Table 1. List of primers used for real-time RT-PCR.

Target gene	Primer pairs (5'-3') ^a	Amplicon size (bp)
18s	(F) TCTTCTTGATTTGTGGGTGG (R) TCGATAGTCCCTAAGAAGTG	150
ERG1	(F) TTAGAATCATGCCAAACC (R) CCAACTGCATACCACCC	127
ERG2	(F) TAATAATGCTGGTGGTGC (R) CAGGATAAGCTGCTCTTT	167
ERG3	(F) GTCTAATGACCCAGTTGT (R) TCTTCTTGCCCTTTGCA	162
ERG4	(F) TATACGCCAATGCTTGTG (R) AGTAACTGAATGGAACCC	120
ERG5	(F) AGATACCGTCCACCAGTC (R) TGCAAAGCAGGATACAAT	119
ERG6	(F) GCTACCGTTCATGCTCCA (R) CCATCACCAGCTCAATA	164
ERG7	(F) GCTTGGGCTTTGATAGGG (R) TCCACTCACCAGTCGGTA	106
ERG10	(F) TGCCCTGGGTCATCCTCT (R) CCGTTACAACACCAGCA	108
ERG11	(F) GAATCCCTGAAACCAAT (R) AGCAGCAGTATCCCATC	131
ERG13	(F) TGGAACACGCTTACGATT (R) CACATGGAAGGCACTGAA	191
ERG24	(F) GGTGACTTAGCGTGGGT (R) GCTGAGCGGAAGATGTA	143

^a F, forward; R, reverse.

Cell viability assay

Cell viability was tested using MTS [3-(4,5-dimethylthiazol-2-yl)-5-(3-carboxymethoxyphenyl)-2-(4-sulfophenyl)-2H-tetrazolium] assay based on the reduction of MTS tetrazolium to a formazan product by dehydrogenase enzymes found in metabolically active cells in the presence of PMS (phenazine methosulfate), an electron-coupling agent, which was quantitated using an ELISA reader. According to the manufacturer's instructions, MTS/PMS assay was performed with appropriate controls. Cells were seeded in 96-well microtitre plates (for BALB/C 3T3 cells, 4×10³ cells/100 µL growth medium/well; for L-02 cells, 1×10⁴ cells/100 µL growth medium/well) followed by 24 h incubation. The desired concentrations of 10b (dissolved in methanol) were added. After 24 h, MTS/PMS reagent (20 µL) was added in each well and incubated for 2 h. A₄₉₂ was recorded using an ELISA reader (Multiskan MK3). The 10b concentration showing 50% reduction in cell viability (IC₅₀ value) was then calculated.

Statistics

All the experiments were carried out in triplicate independently. The data obtained were expressed in terms of mean±SD. Wherever appropriate, the data were also subjected to unpaired two tailed Student's *t*-test. A value of *P*<0.05 was considered statistically significant.

Table 2. *In vitro* antifungal activities of 10b, FLC, ITZ, and KCZ against *C. albicans*^a.

Strains	10b		FLC		ITZ		KCZ		Fm	
	MIC ₅₀	MIC ₈₀	MIC ₅₀	MIC ₈₀	MIC ₅₀	MIC ₈₀	MIC ₅₀	MIC ₈₀	MIC ₅₀	MIC ₈₀
SC5314R	0.031	0.125	128	256	128	256	0.5	16	0.031	0.063
0511655	0.063	0.25	0.25	128	0.125	128	0.063	8	0.063	0.125
01010	0.063	0.5	0.25	128	0.063	128	0.031	2	0.125	0.125
901	0.5	4	0.25	128	0.125	128	0.5	32	0.125	0.5
385	0.125	8	8	256	16	256	0.125	8	0.063	2
333	0.031	0.063	0.25	128	0.063	128	0.063	8	ND	ND
300	0.125	1	0.25	256	0.063	256	0.063	8	0.031	0.125
271	0.125	8	32	256	32	256	0.125	32	0.063	2
255	0.063	0.25	0.063	128	0.063	128	0.063	0.063	ND	ND
103	0.063	2	0.25	256	1	256	0.031	0.063	0.031	0.125
25	0.125	0.25	0.25	128	0.5	256	0.063	32	0.031	0.063
24	0.063	0.25	1	128	0.063	128	0.063	0.063	ND	ND
18	0.031	0.031	0.5	128	1	128	0.063	4	ND	ND
12	0.031	0.063	0.25	128	0.5	2	0.063	8	ND	ND
8	0.063	0.063	0.25	256	0.063	0.063	0.031	0.063	ND	ND
SC5314	0.031	0.25	0.125	0.5	0.031	0.063	0.031	0.063	0.063	0.063
BWP17	0.125	1	0.063	0.5	0.031	0.063	0.031	0.063	0.031	0.063
NJ51-2	0.25	2	0.125	0.5	0.063	0.125	0.031	0.063	0.031	0.063
Y0109	0.031	0.063	0.5	2	0.063	0.25	0.031	0.063	0.031	0.063
687	0.125	0.5	0.25	0.5	0.063	0.063	0.063	0.063	0.063	0.25
0303052	0.031	0.063	0.25	0.5	0.25	0.5	0.031	0.031	ND	ND
0512644	0.063	1	8	32	0.25	4	0.063	8	ND	ND
0601523	0.125	0.5	16	16	0.5	2	0.031	0.063	0.063	0.125
0511479	0.25	0.5	4	16	0.25	8	0.063	4	0.125	0.25

^a) µg/mL; "ND", not determined.

Results

Antifungal activity *in vitro*

As shown in Table 2, 10b possessed potent activities against almost all *C. albicans* isolates, with MIC₈₀ value ranging from 0.063 to 8 µg/mL, which were similar to or more powerful than that of KCZ, and 4096 times lower than that of FLC. Despite high resistance to FLC and ITZ (both MIC₅₀ value 128 µg/mL) for isolate SC5314R, 10b, and Fm displayed powerful antifungal activity against this strain, MIC₅₀ value being 0.031 µg/mL.

Sterol C-14 reductase might be the most important target of 10b

GC-MS analyses showed that 10b reduced the content of ergosterol in a dose-dependent manner, where EC₅₀ value was 0.08 µg/mL (Figure 2). Gas chromatography showed that KCZ-grown cells produced a profile with 24-methylene-ergosterol (73.55%) as the predominant sterol (Table 3), while 10b-grown cells produced a profile similar to that of homozygous *erg24* strain^[12], in which ergosta-8, 14-dienol (ignosterol) was a major sterol accompanied by large amounts of ergosta-8,14,22-trienol. To see whether 10b affected sterol C-14 reductase, sterols were extracted from the wild-type strain (BWP17) and *erg24* strain (NJ51-2) before and after treatment with 10b and Fm. Table 3 shows that the sterols composition of 10b-grown cells was completely identical to that of *erg24* strain except a different proportion in ergosta-8,14,22-

trienol to ignosterol. The ratio was 0.63: 1 in 10b-grown wild-type strain and 0.89: 1 in 10b-grown *erg24* strain. However, the ratio was 1: 6.82 in *erg24* strain without treatment. The wild-type strain treated by Fm produced a profile with ignos-

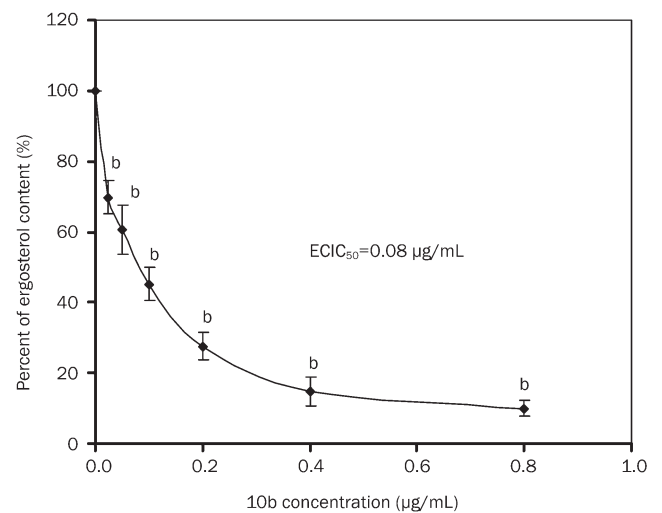


Figure 2. Percentage of the ergosterol content in *C. albicans* BWP17 strain treated with different concentrations of 10b. Ergosterol biosynthesis is expressed as the percentage of that of control cells (unexposed cells). ^b) *P*<0.05 vs control.

Table 3. Analyses of the sterol composition and relative abundance of sterols within wild-type (BWP17), *erg24* (NJ51-2), 10b-grown cells and Fm-grown cells^a.

Strains ^a	Sterol	Percent sterol	Mass
BWP17 (wild-type strain)	Squalene	1.16	410
	Zymosterol	3.46	384
	Ergosterol	80.19	396
	Fecosterol	8.71	398
	Lanosterol	2.57	426
	4,4-dimethyl zymosterol	2.61	412
	Identification of sterols uncertain	4.30	
BWP17 (treated by KCZ)	Squalene	0.65	410
	Ergosterol	3.54	396
	4,14-dimethyl-ergosta-8,24(28)-dienol	10.28	426
	Lanosterol	10.52	426
	24-methylene-lanost-8-en-3-ol	73.55	440
	Identification of sterols uncertain	1.46	
	BWP17 (treated by 10b)	Squalene	0.82
Ergosta-5,8,14,22-tetraenol		9.94	394
Ergosterol		23.91	396
Ergosta-8,14,22-trienol ³		22.35	396
Ergosta-8,14-dinol(ignosterol)		35.54	398
4,4-Dimethylcholesta-8,14,22-trien-ol		3.01	410
Identification of sterols uncertain		4.43	
BWP17 (treated by Fm)	Squalene	2.27	410
	Ergosta-5,8,14,22-tetraenol	5.11	394
	Ergosterol	6.40	396
	Ergosta-8,14-dienol(ignosterol)	68.30	398
	Ergosta-8-en-3-ol	7.32	400
	4,4-Dimethylcholesta-8,14,22-trien-ol	2.88	410
	Identification of sterols uncertain	7.72	
NJ51-2 (<i>erg24</i> strain)	Squalene	1.24	410
	Ergosta-5,8,14,22-tetraenol	5.52	394
	Ergosta-8,14,22-trienol	10.46	396
	Ergosta-8,14-dinol(ignosterol)	71.36	398
	4,4-Dimethylcholesta-8,14,22-trien-ol	6.40	410
	Identification of sterols uncertain	5.02	
	NJ51-2 (treated by 10b)	Squalene	0.84
Ergosta-5,8,14,22-tetraenol		2.60	394
Ergosta-8,14,22-trienol		40.79	396
Ergosta-8,14-dienol(ignosterol)		45.83	398
4,4-Dimethylcholesta-8,14,22-trien-ol		6.49	410
Identification of sterols uncertain		3.45	
NJ51-2 (treated by Fm)		Squalene	0.58
	Ergosta-5,8,14,22-tetraenol	2.59	394
	Ergosta-8,14,22-trienol	9.90	396
	Ergosta-8,14-dienol(ignosterol)	78.70	398
	4,4-Dimethylcholesta-8,14,22-trien-ol	4.44	410
	Identification of sterols uncertain	3.79	

^a The concentrations of KCZ, 10b, and Fm are 3 µg/mL.

sterol as predominate sterol (68.3%) accompanied with a new sterol, ergosta-8-en-3-ol (7.32%) without a trace of ergosta-8,14,22-trienol. Fm-grown *erg24* strain produced a profile completely identical to that of *erg24* strain, whose ratio of ergosta-8,14,22-trienol to ignosterol was 1:7.95. In addition, a negative correlation was observed between 10b concentrations and the ratios of ergosta-8,14,22-trienol to ignosterol (Table 4). The ratio of ergosta-8,14,22-trienol to ignosterol was from 7.63 to 0.76 when the concentration of 10b increased from 0.025 to 0.8 µg/mL.

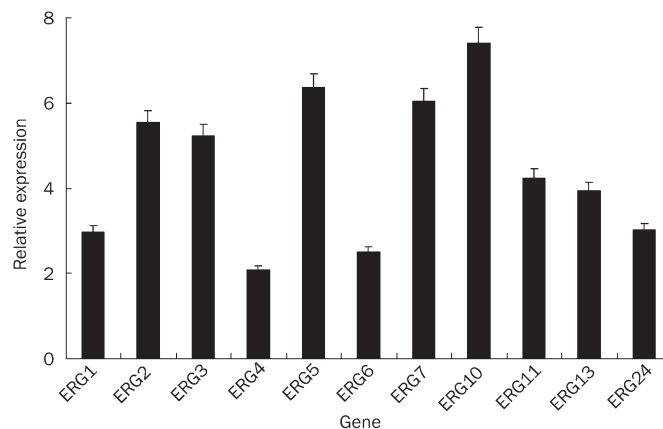
Table 4. The content of ergosta-8,14,22-trienol and ignosterol as well as the ratio of ergosta-8,14,22-trienol to ignosterol in BWP17 strain treated with different concentrations of 10b.

Concentra- tions of 10b (µg/mL ⁻¹)	Content (µg)		Ratio ^a
	Ergosta-8,14,22- trienol	Ignostronol	
0.025	156.23±15.71	20.47±7.76	7.63
0.05	152.43±4.17	50.65±7.70	3.01
0.1	95.36±13.06	54.74±5.93	1.74
0.2	66.40±12.06	53.67±10.69	1.24
0.4	73.92±6.95	75.04±9.70	0.99
0.8	65.39±14.80	85.77±14.63	0.76

^a The ratio of Ergosta-8,14,22-trienol to Ignostronol.

10b increases mRNA levels of sterol metabolism genes

To study cellular responses to perturbation of ergosterol biosynthesis caused by 10b, real-time RT-PCR analysis was performed, and the results revealed a global upregulation of sterol metabolism genes in response to 10b treatment. Figure 3 shows that the expression of gene *ERG5*, *ERG10*, and *ERG7* significantly increased by 6.36, 7.41, and 6.05 fold respectively,

**Figure 3.** Gene expression changes of sterol metabolism genes in 10b-treated SC5314 strain. The concentration of 10b is 6 µg/mL. All genes were examined by real-time RT-PCR with gene-specific primers. Relative fold change was calculated with C_T value (see details in Materials and Methods). Mean±SD. $n=3$.

and that gene *ERG6*, *ERG1*, *ERG24*, *ERG13*, *ERG11*, *ERG3*, and *ERG2* upregulated intermediately (2.5 fold to 5.54 fold).

Dose-dependent toxicity of 10b on BALB/C 3T3 and L-02 cells

Based on investigation of the mechanism of 10b against *C albicans*, its cytotoxicity *in vitro* was further assessed using MTS/PMS reduction assay. Figure 4 shows that the cytotoxic response of BALB/C 3T3 and L-02 cells to 10b appeared to be dose dependent, where IC_{50} value was 11.31 $\mu\text{g}/\text{mL}$ and 35.70 $\mu\text{g}/\text{mL}$, respectively. In addition, no cytotoxic response was found in the two mammalian cells when the concentration of 10b was 6 $\mu\text{g}/\text{mL}$.

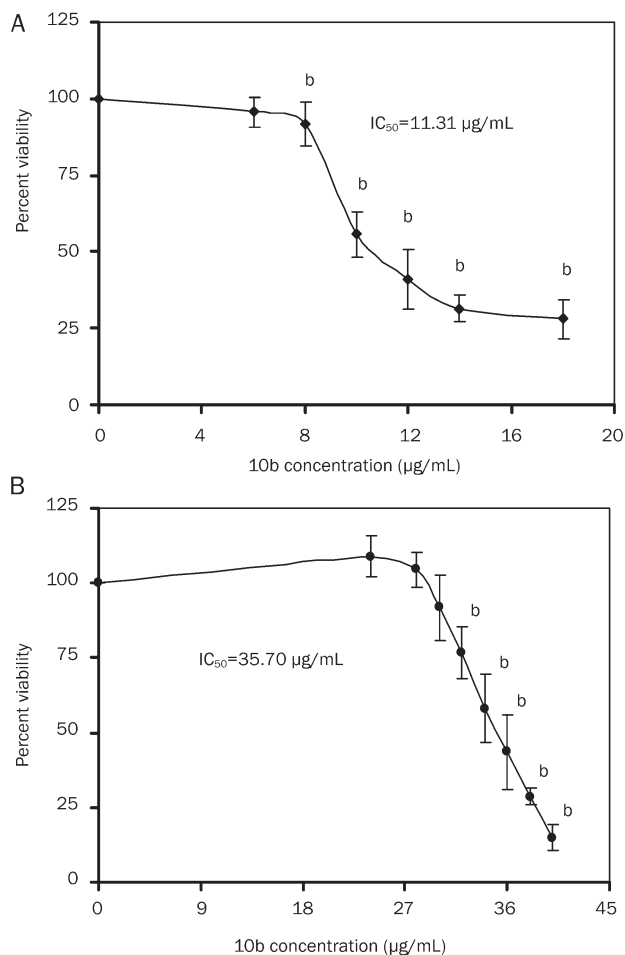


Figure 4. Percentage of viability measured by MTS/PMS assay in murine embryo fibroblast (A) and human liver cells (B) after treatment with 10b for 24 h. Mean \pm SD. $n=3$. An OD value of control cells (unexposed cells) was taken as 100% viability (0% cytotoxicity). ^b $P<0.05$ vs control.

Discussion

On the basis of the three-dimensional model of lanosterol 14 α -demethylase (CYP51) from *C albicans* constructed in previous studies^[15-18], a series of 2-aminotetralin compounds was designed and synthesized as antifungal agents. We hope that

these compounds are able to interact with amino acid residues in the active site of CYP51 and avoid connection with the iron atom of heme, and therefore they can not only generate potent antifungal activity but attenuate severe toxicities of azoles. Experiments^[11] has confirmed that 2-aminotetralin derivatives possessed potential antifungal activities, especially against FLC-resistant *C albicans*, *Candida parapsilosis* and *Trichophyton rubrum*. Theoretically, the most important action mechanism of 10b should inhibit CYP51 as other azoles do.

Unexpectedly, we found that 10b had an action mode completely different from other azoles in the ergosterol biosynthetic pathway. Cells treated with 10b produced a profile similar to that of homozygous *erg24* strain, in which ignosterol and ergosta-8,14,22-trienol were major sterols and no trace of 24-methylene-lanost-8-en-3-ol was found. However, KCZ-grown cells had a profile with 24-methylene-lanost-8-en-3-ol as the predominant sterol. We therefore postulate that sterol C-14 reductase, encoded by *ERG24*, instead of lanosterol 14 α -demethylase, encoded by *ERG11*, is an important target for 2-aminotetralin derivatives in this pathway.

10b also showed an action mechanism different from that of morpholines in the ergosterol biosynthetic pathway, although both of them are sterol C-14 reductase inhibitors. Morpholines inhibit sterol C-8 isomerase, encoded by *ERG2* gene, resulting in ergosta-8-en-3-ol accumulation. As sterol C-8 isomerase is situated in the downstream of sterol C-14 reductase, Fm-grown *erg24* strain produced a profile completely identical to that of *erg24* strain. Although the sterol composition of 10b-grown cells was completely identical to that of *erg24* strain, the ratio of ergosta-8,14,22-trienol to ignosterol was different. The content of ergosta-8,14,22-trienol in 10b-treated *erg24* strain was much more than that in *erg24* strain without 10b treatment, indicating that there must be another important target upstream sterol C-14 reductase. We deduced that another target was an enzyme related to sterol C-5 desaturase, which catalyzes the conversion of ergosta-8,14,22-trienol to ergosta-5,8,14,22-tetraenol as presented in Figure 5. In addition, we found that the content of ignosterol built up in the wake of increase in the concentration of 10b, indicating that sterol C-5 desaturase related enzyme was the dominant target of 10b at a low concentration, while the affinity of 10b with sterol C-14 reductase augmented along with the increase in the concentration of 10b and finally changed into the fundamental target at a higher concentration.

Ergosterol is an important constituent of cellular membranes and plays a fundamental role in many biological processes^[19-21]. 10b significantly reduced the ergosterol content by inhibiting the activity of sterol C-14 reductase and C-5 desaturase related enzyme, destructing the integration of the fungal membrane and finally leading to fungal death. To further investigate cellular responses to perturbation of ergosterol biosynthesis stimulated by 10b, we detected the expression of sterol metabolism genes by real-time RT-PCR analyses. The results revealed a global upregulation of sterol metabolism genes, including gene *ERG24*, *ERG5*, and *ERG11*, which is consistent with previous reports^[22, 23]. This was presumed as

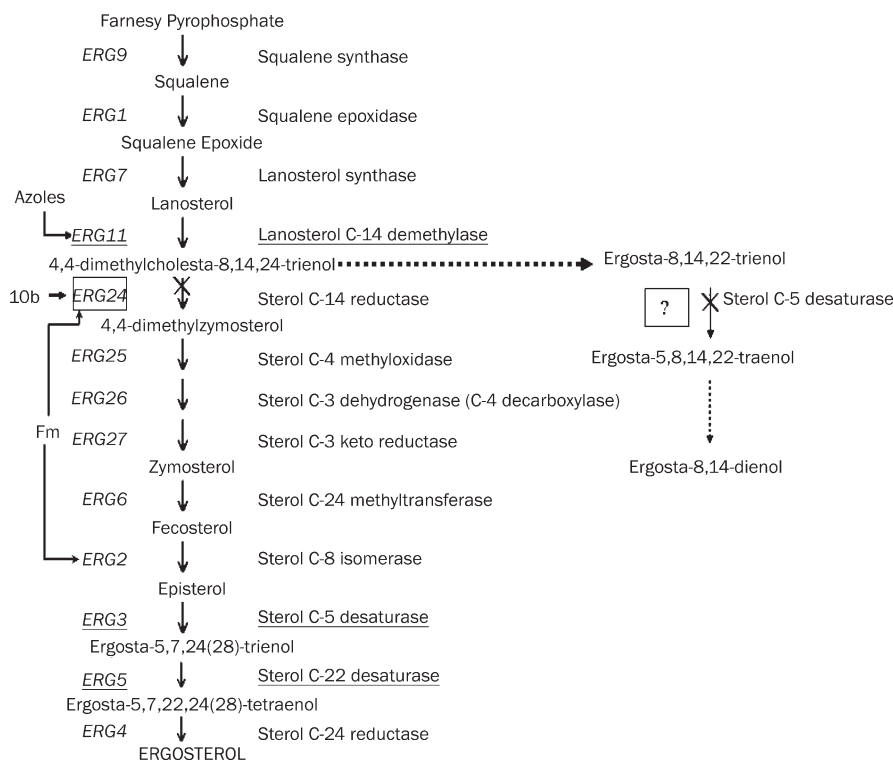


Figure 5. Proposed mechanism action of 10b against *C. albicans* in the ergosterol biosynthetic pathway.

a result of feedback control^[24]. However, which mechanism is responsible for the global upregulation of sterol metabolism genes in response to ergosterol biosynthesis inhibitors remains unclear. One theory postulates that depletion of ergosterol or another sterol formed late in the pathway increases the expression of global sterol metabolism genes. Others argue that accumulation of an early substrate or toxic sterol by-products induces the expression of sterol metabolism genes. Upregulation of sterol metabolism genes in 10b or azole-treated *C. albicans* suggests that change in transcript levels is not directly related to ergosterol biosynthesis in *C. albicans*.

The Interagency Coordinating Committee on the Validation of Alternative Methods (ICCVAM) recommends two standard cytotoxicity assays in order to better predict acute systemic toxicity on *in vitro* methods: one using a human cell system and the other using a rodent cell system (see website address: <http://iccvam.niehs.nih.gov/methods/acutetox/acutetox.htm>). BALB/C 3T3 murine embryo fibroblasts and L-02 human normal liver cells were used. These cell lines have been well characterized for their relevance for toxicity studies^[25, 26]. Our data clearly suggest that although 10b seemed to reduce MTS/PMS reduction in a dose dependent manner, IC₅₀ value for these cells was much higher than MIC₅₀ value for *C. albicans* (~0.5 µg/mL). This indicates that the formulation is preliminarily safe and warrants further study for possible human applications.

The present study indicates that tetralin derivatives are potentially applicable to the field of Psychiatry, of which the most effective are dialkylated tetralin derivatives^[27]. Amino-tetralins, including 8-hydroxy-2-(di-n-propylamino) tetralin

(8-OH-DPAT) and 7-hydroxy-2-(di-n-propylamino) tetralin (7-OH-DPAT) behave as preferential agonists at serotonin (5-HT)1A and dopamine D3 and D2 receptors^[28]. The former affects intracranial self-stimulation (ICSS) and the latter possesses anxiolytic properties. However, there are few reports from other laboratories describing the antifungal activities of 2-aminotetralin derivatives. In conclusion, our findings suggest that 2-aminotetralin compounds deserve further study by virtue of their novel chemistry structures, definite antifungal activities and a distinctive mode of action.

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Author contribution

Yuan-ying JIANG, Yong-bing CAO, and Kai-hua FAN designed the research protocol; Rong-mei LIANG performed the research and wrote the paper; You-jun ZHOU and Hui TANG synthesized 10b; Ping-hui GAO, Hong-tao LIU, Yi XU, Bao-di DAI, Yong-hong TAN, and Shi-hua WANG made a critical reading of the manuscript.

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