Inflammation in neurodegenerative diseases - an update

Sandra Amor,^{1,2} Laura A. N. Peferoen,¹ Daphne Y. S. Vogel,^{1,3} Marjolein Breur,³ Paul van der Valk,¹ David Baker² and Johannes M. van Noort⁴

¹Department of Pathology, VU University Medical Centre, Amsterdam, the Netherlands, ²Neuroimmunology Unit, Blizard Institute of Cell and Molecular Science, Barts and The London School of Medicine and Dentistry, London, UK, ³Department of MCBI, VU University Medical Centre, Amsterdam, the Netherlands and ⁴Delta Crystallon BV, Leiden,

doi:10.1111/imm.12233

Received 19 November 2013; revised 09 December 2013; accepted 11 December 2013.

Correspondence: Professor Sandra Amor, Department of Pathology, VU University Medical Centre, De Boelelaan 1117, Amsterdam 1081 HV, the Netherlands. Email: s.amor@vumc.nl Senior author: Sandra Amor

Summary

Neurodegeneration, the progressive dysfunction and loss of neurons in the central nervous system (CNS), is the major cause of cognitive and motor dysfunction. While neuronal degeneration is well-known in Alzheimer's and Parkinson's diseases, it is also observed in neurotrophic infections, traumatic brain and spinal cord injury, stroke, neoplastic disorders, prion diseases, multiple sclerosis and amyotrophic lateral sclerosis, as well as neuropsychiatric disorders and genetic disorders. A common link between these diseases is chronic activation of innate immune responses including those mediated by microglia, the resident CNS macrophages. Such activation can trigger neurotoxic pathways leading to progressive degeneration. Yet, microglia are also crucial for controlling inflammatory processes, and repair and regeneration. The adaptive immune response is implicated in neurodegenerative diseases contributing to tissue damage, but also plays important roles in resolving inflammation and mediating neuroprotection and repair. The growing awareness that the immune system is inextricably involved in mediating damage as well as regeneration and repair in neurodegenerative disorders, has prompted novel approaches to modulate the immune system, although it remains whether these approaches can be used in humans. Additional factors in humans include ageing and exposure to environmental factors such as systemic infections that provide additional clues that may be human specific and therefore difficult to translate from animal models. Nevertheless, a better understanding of how immune responses are involved in neuronal damage and regeneration, as reviewed here, will be essential to develop effective therapies to improve quality of life, and mitigate the personal, economic and social impact of these diseases.

Keywords: central nervous system; inflammation; innate; neurodegeneration; neuroprotection; repair.

Introduction

The demographics of dementia published by the World Health Organization (WHO) predict that neurodegenerative diseases will reach over 70 million in 2030 and 106 million in 2050.¹ As in dementia, neurodegeneration associated with immune activation is observed in many other disorders including stroke, spinal cord injury, multiple sclerosis (MS), brain injury (Table 1), and the animal models of these diseases (Table 2). Hence, while the central nervous system (CNS) is an immune-privileged site, it is clear that innate and adaptive immune responses do take place in the CNS to limit neurotrophic viral and bacterial infections, limit tumour growth, remove necrotic cells following ischaemia, shape the brain during development, and promote regeneration and repair following damage. For example, both macrophages and microglia clear debris as a result of damage in the CNS and when such phagocytosis is impeded, regeneration is delayed. Likewise, inhibiting migration of adaptive immune cells into the brain renders the CNS susceptible to devastating infections.

The field of neuroimmunology has uncovered many examples of cross-talk between the nervous and immune systems, some of which have been exploited to design therapeutic strategies. Given the major steps this field has taken in recent years we provide an update of our original paper on immune responses in the CNS,² incorporating

Table 1.	Immune	responses	in	neurodegenerative	diseases
rubic i.	minune	responses	111	neurouegeneruure	anocaoco

Disease	Clinical and neuropathology	Innate immunity	Adaptive immunity	References
AD	Cognitive decline Atrophy of hippocampus and neocortex Neuronal loss with amyloid plaques and neurofibrillary tangles	Activated microglia ↑ Cytokines, chemokines and complement ↑ TLRs and PRRs	↑ Antibody and T-cell responses to Aβ peptide	3
FTD	Also called Pick's disease Changes in personality behaviour, and aphasia Microvacuolation, neuronal loss, demyelination, astrocytic gliosis Mutations in the progranulin gene present in familial FTD	Activated microglia ↑ Macrophages ↑ IL-1β and COX-2	Not reported	4
PD and LBD	Resting tremor, bradykinesia, rigidity, gait dysfunction Degeneration of dopaminergic neurons due to aggregates of α-synuclein and ubiquitin	 Microglia activation ↑ NK cells ↑ IL-1β, IL-6, TNF-α in CSF ↑ TLR2, TLR5, CD14 in CNS ↑ CD14 and TLR4 in substantia nigra of MPTP animal model 	 Antibodies to neuronal antigens In LBD autoantibodies serum directed to Aβ and S100B, α-synuclein ↑ T cells in periphery and CNS 	5,6
HD	Chorea, dystonia, atrophy of caudate and putamen, cognitive changes Diffuse atrophy of neostriatum Protein aggregates in cytoplasm and nucleus of neurons	 ↑ Microglia proliferation ↑ Complement factor 3 and 9 ↑ Inflammatory factors ↑ Kynurenine pathway activity 	Not reported	7
SMA	Autosomal-recessive disorder Hypotonia and weakness in newborn Degeneration of anterior horn cells in spinal cord and lower brain stem	↑ IL-6, IL-1 β	Not reported	8
SCA	Polyglutamine disease Progressive ataxia and neurodegeneration Mutant protein expressed in glia	↑ Microglia proliferation↑ C3 and C9 expression	Not reported	9
PNND	Heterogeneous clinical symptoms Immune response to antigens expressed on tumours	↑ Microglia activation ↑ Immune regulatory molecules	Antibodies to neuronal antigens ↑ Regulatory, memory T cells, B cells	10,11
Lysoso- mal storage disorders	Congenital metabolic disorders, due to defects in enzymes required for lysosomal acid hydrolysis, resulting in heterogeneous clinical symptoms	 ↑ Activation of glia before neuronal damage ↑ Inflammatory genes and factors 	↑ Genes encoding CD3, CD8, CD28, CTLA-4, and CD22	12
ALS	Progressive paralysis Progressive severe motor neuron loss in anterior horn, ubiquitin aggregation and axonopathy	 ↑ Complement, TLRs, CD14 RAGE, HMGB1 ↑ Genes for TRAF6, CNTF, HGF and GDNF 	Antibodies to neurons in blood ↑ CD4 ⁺ , CD8 ⁺ T cells	13,14
MS	Heterogeneous neurological symptoms, depending on lesion location Chronic inflammation Demyelination and progressive neurodegeneration Different lesion staging	 ↑ Microglial activation ↑ NK cells ↑ Innate receptors Phagocytosis of neuronal debris ↑ Cytokines, chemokines, complement, growth factors 	Antibodies and T-cells to neurons, myelin and myelin associated antigens ↑ Heat-shock proteins Neurotrophins secreted by T cells	15–18
NMO	Autoimmune disease with demyelination of spinal cord and optic nerve in which astrocytes are targeted	↑ Microglia activation ↑ Chemokines, cytokines	Pathogenic antibodies to AQP4	19,20
Prion diseases	Progressive dementia, ataxia, monoclonus, seizures	↑ Microglia activation ↑ IL-1B, IL-6, ROS ↑ Complement Mast cell express PrP	B cells aid transport of PrP	6,21

Disease	Clinical and neuropathology	Innate immunity	Adaptive immunity	References
Infection	Neurotrophic and peripheral pathology varies with pathogen	↑ Microglia activation astrocytosis ↑ Innate receptors, cytokines,	↑ (Auto) antibodies ↑ T cells ↑ B cells	2
Toxins and metals	For example, manganese induces neurodegeneration Diesel exhaust particles	complement ↑ Microglia activation ↑ MMP production ↑ TNF-α, IL-6	↑ Autoantibodies ↑ T cells ↑ B cells	22
CVD	Heavy metals Brain ischaemia due to several	↑ Activated microglia, astrocytes	T cells contribute to	23,24
	pathological processes involving the blood vessels	Phagocytic macrophages	early post-stroke neuronal injury	
Epilepsy and Epileptic syndro-mes	Seizures associated with cognitive and psychological sequelae. Occurs in about 1% of the population	Microglia, astrocytes activation ↑ CCL3, CCL4 genes in CNS ↑ IL-1β, IL-6 in the CSF ↑ TLR2 and TLR4, RAGE, HMGB1 ↑ COX-2 in neurons, astrocytes	↑ Antibodies ↑ CD8 ⁺ granzyme B, T cells in parenchyma	25
TBI	Mechanical injuries of brain or spinal cord resulting in inflammatory responses enhancing tissue damage	↑ C1q, C3b, d, C4 and C5b-9 ↑ Proinflammatory cytokines ↑ TLRs	↑ T cells	26

Table 1 (Continued)

Aβ, amyloid-β; AD, Alzheimer's disease; ALS, amyotrophic lateral sclerosis; AQP4, Aquaporin 4; COX-2, cyclooxygenase-2; CTLA-4, cytotoxic T-lymphocyte antigen 4; CVD, cerebrovascular diseases; FTD, frontotemporal dementia; GluR3, glutamate receptor 3; HD, Huntington's disease; IL, interleukin; LBD, Lewy body disease; MHC, major histocompatibility complex; MMP, matrix metalloproteinase; MS, multiple sclerosis; NK, natural killer; NMO, neuromyelitis optica; PD, Parkinson's disease; PNND, paraneoplastic neurological disorders; PrP, prion protein; PRR, pattern recognition receptor; ROS, reactive oxygen species; SCA, spinocerebellar ataxia; SMA, spinal muscle atrophy; TBI, traumatic brain injury; TNF, tumour necrosis factor; TSI, traumatic spinal cord injury.

recent knowledge of the delicate balance between pathogenic and reparative processes. We also review emerging evidence showing that immune responses differ in the ageing brain, or following peripheral infections and other previous insults, indicating that a so-called 'primed' environment may make the CNS more susceptible to damage. These factors may be key in explaining why neurodegenerative diseases are more prevalent in the elderly, and why even peripheral infections are linked to chronic CNS diseases such as MS. A better understanding of the crosstalk between the innate and adaptive immune response and the CNS will be crucial to harness natural beneficial responses for therapeutic strategies.

Immune privilege in the CNS

That the CNS enjoys a state of 'immune privilege' was recognized by Sir Peter Medawar who, with Sir Frank Macfarlane Burnet was awarded the Nobel Prize in 1960. Medawar's idea of 'immune privilege' led to the notion that immune responses are tightly regulated in the brain, such that antigens within the brain do not elicit immune responses but can only be targets of an immune response initiated in the periphery. The CNS lacks recognizable parenchymal dendritic cells and is devoid of a classical lymphatic system. While several studies have revealed that CNS antigens can be drained to cervical lymph nodes, it is generally believed that this leads to immune tolerance rather than priming.²⁷ The 'immune privilege' status of the CNS is dependent on several elements, notably including the physical blood-brain barrier and bloodspinal cord barrier. The unusually tight vessel walls within the CNS are designed to limit the entry of solutes and ions into the CNS. These barriers, together with neurons, glia and extracellular matrix, form the neurovascular unit that regulates immune responses in the CNS, in concert with local production of neuroimmunoregulatory molecules.²⁸ The majority of these local cues come from neurons, although microglia,²⁹ astrocytes and oligodendrocytes³⁰ also play important roles in maintaining this integrity. In concert, these 'don't eat me' cell-contactdependent signals and soluble mediators maintain a muted immune response that inhibits microglia activation and maturation of antigen-presenting cells, and restricts survival, especially of activated lymphocytes. Such regulation is mediated by several factors, including CX3CL1 (fractalkine), CD22, CD27, CD47, CD200 and neuronal cell adhesion molecule, known to be expressed by neurons and glia (Table 3). Neurons also produce a range of soluble mediators including chemokines, neuropeptides, neurotransmitters and neurotrophins, all of which contribute to the control and tight regulation of local

Disease	Model	Clinical features	References
AD Ageing	APP transgenic mice: Tg2576, PDAPP, TgAPP23 Tg-APPswe/PS1dE9, 5XFAD, 3xTg-AD mice, which carry APP and PS1 Tau transgenic mice Senescence accelerated mice	Behavioural and cognitive changes, amyloid pathology, accumulation of $A\beta$, loss of neuronal subpopulations, mitochondrial oxidative damage, synaptic loss, proliferation and activation of astrocytes and microglia Age-related changes in the brain. Brain	42 43
		atrophy, loss of cortical neurons, and dendritic spines, loss of synapses, impaired learning and memory	
ALS	SOD-1 mutation Wobbler mouse (point mutation in the Vps54 gene) Immunization with motor neurons ALS in knockout mouse hTDP-43-associated familial ALS (transgenic)	ER stress-related toxicity, neurofilament aggregation, neuronal hyper-excitability, neuroinflammation, antibodies to neurons, reactive astrocytes, microglia activation and proliferation, increased TDP-43 expression	44
HD and other polyglutamine disorders	Many transgenic mice in which each disease possesses unique features as a result of the polyQ mutation	Dependent on model. Cognitive deficits, ataxia seizures, neuronal loss, microglia activation and reactive astrocytes	45
PD	Over-expression of α-synuclein Neurotoxic models, e.g. MPTP; 6-OHDA, rotenone	Microglial activation, adaptive immunity directed to neurons expressing α-synuclein, CD4 ⁺ T-cell-mediated damage	46
Prion	Tg(Hu-PrP)110, Tg(HuPrP)152 transgenic mice Tg(MHuPrP)-segment of mouse PrP gene replaced with human PrP	Expression of PrP plaques, neuronal loss, and spongiosis depends on transgenic mouse	47
Stroke	Middle cerebral artery occlusion model, photothrombotic model (non-invasive); thrombin injections, vessel occlusion	Leucocytes, macrophages, activated microglia and astrocytes are considered to contribute to secondary damage.	48
MS	Viral models: SFV, TMEV, MHV infectionAutoimmune models: acute, chronic andsecondary progressive EAE immunized withmyelin antigens. Spasticity in mice immunizedwith NF-L.Toxin models: cuprizone, lysolecithin,ethidium bromide	SFV induces T-cell-mediated inflammatory demyelination, TMEV induces neurological symptoms depending route and strain, autoimmune models immunized with spinal cord homogenate, myelin protein or neuronal antigens in strong adjuvants	49,50

Table 2. Animal models of neurodegenerative diseases and involvement of immune responses

AD, Alzheimer's disease; ALS, amyotrophic lateral sclerosis; APP, amyloid precursor protein; ER, endoplasmic reticulum; EAE, experimental autoimmune encephalomyelitis; HD, Huntington's disease; MHV, mouse hepatitis virus; MTPT, 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine; MS, multiple sclerosis; NF-L, neurofilaments light; PD, Parkinson's disease; PrP, prion protein; TDP-43, TAR DNA-binding protein 43 kd; TMEV, Theiler's murine encephalomyelitis virus; SFV, Semliki Forest virus; SOD, superoxide dismutase; 6-OHDA, 6-hydroxydopamine.

immune responses. For example, TREM2 ligands on neurons help preserve the 'immune privilege' environment,³¹ and neuronal semaphorins additionally assist in immune regulation and repair by mediating oligodendrocyte precursor cell migration (Table 3). Glial cells, and especially astrocytes, can additionally produce an array of molecules to inhibit or kill activated lymphocytes, for example by Fas–Fas ligand (FasL) interactions. Defects in such regulatory circuits are known to contribute to inflammatory neurodegeneration. Some semaphorins are aberrantly expressed in neurons during Alzheimer's disease (AD), at the neuromuscular junction in amyotrophic lateral sclero-

sis (ALS) and in MS lesions where they control oligodendrocyte precursor cell migration whereas others regulate inflammation suppressing disease.^{32,33} When neurons are lost altogether, immunoregulatory signals otherwise delivered by these cells are impaired, rendering the CNS less able to maintain its protective shield.

Innate and adaptive immune responses in the CNS

Local innate immune responses play a key role in the first line of defence against bacterial, viral, fungal and parasitic

Microglia	Neurons	Astrocytes	Oligodendrocytes	References
CD200R	CD200	CD200	CD200	28,29
CX3CR1	CX3CL1	CX3CL1	Unknown	28,29
CD172a/Sirp1α	CD47	CD47	CD47	28,29
thrombospondin				
TREM2	HSP60	HSP60	HSP60	28,30,31
	TREM2L			
Plexins	Semaphorins	Unknown	Plexin4 assists in OPC migration but semaphorins likely to be pathogenic	32,33
ICAM	sICAM5 secreted	Unknown	Unknown	34
Csflr	IL-34	IL-34	Unknown	35
CD45	CD22	Not reported	Unknown	36
NCAM/CD56	NCAM/CD56	NCAM/CD56	Unknown	37
Complement, MAC, C3, C5,	Regulators of complement activation, CD46, factor H	CD46, factor H	Unknown	28
Soluble factors: IL-4, IL-1β, IFN-γ, TGF-β, BDNF, GDNF, TIMPS, IDO	CX3CL1, TGF- β , C22, NT3 neuropeptides, sCD95L, IL-10, Sema-3A dopamine, GABA	IL-4, IL-10, IFN-γ, TGF-β, Proteoglycans, factor H	TGF- β isoforms expressed <i>in vitro</i>	28,29,37

Table 3. Molecules expressed or secreted imparting immune regulation in the central nervous system

BDNF, brain-derived neurotrophic factor; Csf1R, colony-stimulating factor 1; GABA, γ -aminobutyric acid; GDNF, glial cell-derived neurotrophic factor; HSP, heat-shock protein; ICAM, intercellular adhesion molecule 1; IDO, indoleamine-pyrrole 2,3-dioxygenase; IFN- γ , interferon- γ ; IL, interleukin; MAC, membrane attack complex; NCAM, neural cell adhesion molecule; NT, neurotrophin; Sirp1 α , signal regulatory protein α ; TGF, transforming growth factor; TIMPS, tissue inhibitors of metalloproteinases; TREM, triggering receptor expressed on myeloid cells 2.

infections in the CNS, and they are also essential to clear apoptotic cells, misfolded or aggregated proteins. In this way, innate responses pave the way for tissue repair and full restoration of homeostasis. Yet, exaggerated or prolonged innate responses may cause damage. Such damaging responses may result not only from activation of microglia, astrocytes or oligodendrocytes,^{29,30} but also of peripheral innate immune cells including natural killer cells, natural killer T cells, mast cells,³⁸ granulocytes and $\gamma\delta$ T cells. Such cells can contribute significantly to local inflammatory processes (Table 1, Fig. 1).

Innate immune responses are generally initiated following recognition of pathogen-associated molecular patterns (PAMPS), conserved structures expressed by infectious agents. Equally important are endogenous signals for innate responses, known as danger-associated molecular patterns (DAMPS). DAMPS include a plethora of different molecules such as nucleic acids, heat-shock proteins, ATP, high mobility group box chromosomal protein 1 (HMGB-1), fibrinogen and aggregated and modified or misfolded proteins. They tend to appear as the result of stress or tissue damage. Through expression of conserved pattern-recognition receptors (PRRs), local CNS cells are triggered by the appearance of such PAMPS and DAMPS. The PRRs include C-type lectins, Toll-like receptors (TLRs), retinoic acid inducible gene I-like receptors, nucleotide-binding domain, leucine-rich repeat-containing proteins (NLRs), HIN200/PYHIN family members

designated as in the 'Absent-In-Melanoma' receptors, as well as interferon-induced proteins called IFITs.39,40 For many of these PPRs, their expression pattern and role in neurodegenerative disorders are still under investigation. The best studied PRRs are the TLRs, which are expressed by many cells in the CNS. Activation of TLR induces several signalling pathways via the intracellular adaptor proteins Myd88 and Toll/interleukin-1 receptor-domaincontaining adapter-inducing interferon- β , and production of a wide range of immunoregulatory mediators. The involvement of TLRs in neurodegenerative diseases is evidenced by pathology studies of human disorders, as well as by data from experimental animal models. For example, TLR9-deficient mice produce less pro-inflammatory cytokines and exhibit reduced neuronal damage, whereas TLR4-knockout mice are resistant to the induction of experimental autoimmune encephalomyelitis (EAE), an animal model of MS (Table 2). Activation of TLR3 suppresses EAE and leads to the production of several neuroprotective mediators in vitro.41 Several TLR-deficient mice also exhibit milder clinical disease following brain injury, and middle cerebral artery occlusion, indicating a pathogenic role for TLR activation in these models. On the other hand, TLRs aid uptake of $A\beta$ indicative of beneficial effects in other situations. As is so often the case in the context of immune regulation, TLR-mediated responses can both aggravate inflammation and contribute to its control.

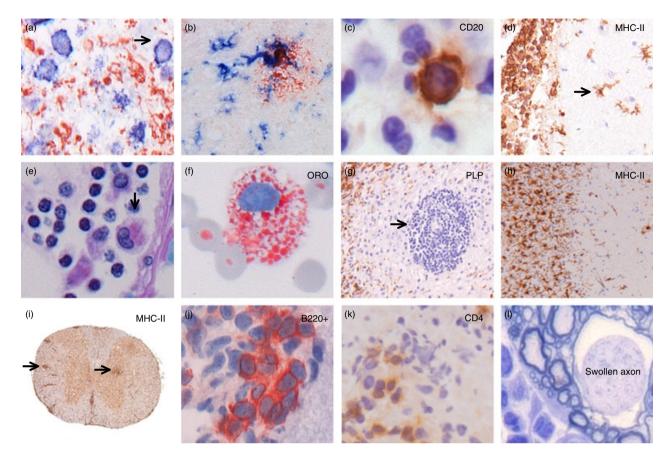


Figure 1. Immune responses in human and experimental inflammatory neurodegenerative disorders. In an ischaemic area in stroke, HLA class II^+ cells (blue, arrow) can be seen phagocytosing myelin basic protein (red) (a). In Alzheimer's disease, activated microglia (HLA class II^+ ; blue) cluster around neurons and amyloid deposits (red) (b). A meningeal infiltrate in acute bacterial meningitis contains a single CD20⁺ B cell (brown; c), yet large numbers of activated microglia (HLA class II^+ ; brown) in the meninges as well as in the brain parenchyma (d). In multiple sclerosis (MS), macrophages and microglia phagocytose myelin, and turn into so-called foam cells that are found in the perivascular space (e) as well as in the cerebrospinal fluid (f). Large numbers of inflammatory cells around a blood vessel are found in the centre of MS lesions that show extensive loss of myelin (g), as well as at the rim of the demyelinating zone (h). In the autoimmune mouse model of MS, perivascular infiltrates of HLA-class II positive cells can be observed in the white and grey matter of the spinal cord (i). In the meninges of mice that have been immunized with the neuronal antigen NF-L, the presence of B220⁺ B cells (j) and CD4⁺ T cells (k) is closely associated with neuronal damage (l).

Another group of PRRs, NLRs initiate the assembly of inflammasomes that activate caspases and in this way, promote production and secretion of mature interleukin- 1β and interleukin-18.⁵¹ A β and prion proteins are potent activators of the NLRP3 inflammasome. NLRP1 and NLRP5 are known to contribute to neuronal death while Nlrp3 or ASC (apoptosis-associated speck-like protein containing a CARD)-deficient microglia secrete significantly reduced levels of pro-inflammatory cytokines following A β phagocytosis. Consistent with these findings, the presence of ASC and NLRP3 are necessary for progression of EAE in mice. Yet other receptors important in neuron-glia interactions are the purinergic receptors P1 (adenosine) and P2 (ATP) receptors.⁵¹ Adenosine receptors can mediate both potentially neuroprotective and potentially neurotoxic effects. Their roles in different neurodegenerative diseases are not elucidated yet. In Parkinson's disease (PD), AD and ALS, the expression of both P1 and P2 receptors is altered. In attempts to clarify the functional significance of these changes, much attention has been devoted to the role of the adenosine A2A receptorin PD because antagonists for this receptor improve clinical symptoms as well as protection against toxin-induced neuronal degeneration.⁵² Hence, PRRs play an ambivalent role in neurodegenerative diseases, depending on the mode of activation.

Complement, a major effector arm of the innate immune system, is often considered a link between the innate and adaptive immune responses. During development, synapses destined for removal are tagged with complement proteins and so may be eliminated by microglial cells expressing complement. Such expression in neurodegenerative diseases is important for elimination of aggregated proteins that are found for example in AD, or necrotic cells in neurological complications of systemic lupus erythematous. However, the broad and often profoundly unregulated expression of complement components indicates that complement activation may equally contribute to the demise of neurons and axons. That neurons and oligodendrocytes express low levels of complement regulatory proteins renders these cells particularly vulnerable to complement-associated death.⁵³

As emphasized above, the anti-inflammatory environment afforded by the immune privilege status is very effective in dampening any adaptive immune response within the CNS. Despite the microenvironment being hostile to T cells, large numbers of T cells still routinely patrol the perivascular space in a healthy CNS, and activated lymphocytes readily cross the blood-brain barrier. The extent and nature of any adaptive T-cell-mediated immune response that may occur in neurodegenerative diseases is largely determined by the type of local innate immune response. This, in turn, is determined by the initial trigger that activates such responses: the collection of PAMPs and DAMPs simultaneously acting on innate immune cells. In CNS infections, adaptive immunity is primarily directed to the infectious agent. If the infected cell is a neuron, direct cytolysis and neuronal damage may ensue.

Evidence that T cells may be involved in neurodegenerative disorders comes from studies on alterations of T-cell subsets in the periphery of patients during disease and evidence for a direct association between the presence of T cells and neurodegeneration in the CNS. For example, the decline in T-cell responses to $A\beta$ during ageing, and the absence of such responses in AD suggests that in AD, T cells may play a role in clearing plaques. Evidence for T-cell mediated plaque removal in mice supports this notion.54 In PD, CD4+ T cells are observed in the substantia nigra whereas in MS, T cells are clearly associated with demyelinating lesions and neuronal damage. T-cell derived granzyme B and perforin, as well as apoptosis-inducing signals, may be operational. Although the major histocompatibility recognition elements for T cells are generally down-regulated within the healthy nervous system, MHC class II antigens become up-regulated on microglia and perivascular microglia/pericytes during inflammation and can serve to facilitate antigen presentation to T cells. In some instances neural cells can express MHC class I antigens, opening the potential for CD8-mediated killing. Direct neuronal killing of CD4-expressing T helper type 17 cells has been noted, suggesting the potential of MHC class II independent killing.55

That the adaptive immune system can contribute to neuronal damage in the CNS is shown by paraneoplastic neurological disorders. Such disorders are associated with the emergence of both T cells and antibodies against antigens expressed on neuronal tumours. For example, limbic encephalitis is associated with autoantibodies to the *N*-methyl D-aspartate receptor and voltage-gated potassium channel, whereas in stiff person syndrome, antibodies are directed to GAD65 and amphiphysin.^{10,11} Removing the tumour or performing plasmapheresis is often beneficial, particularly when the autoantibodies contribute to neuronal dysfunction.

Autoantibodies to neuronal proteins are also present in other neurodegenerative diseases such as MS, during which antibodies are produced intrathecally. That at least some of these anti-neuronal antibodies may be pathogenic is suggested by the finding that in animal models, antibodies against neuronal antigens can induce neuronal damage, or are associated with progression of neurological disease.^{56–59} That some of these antibodies are pathogenic is supported by the findings that plasma exchange is effective in a subgroup of patients. In neuromyelitis optica (NMO), the findings are clearer in that direct evidence for pathogenicity has also been obtained for serum antibodies to aquaporin-4, an abundant water channel expressed by astrocytes. Such antibodies are pathogenic in animals. Although these antibodies may not trigger a direct attack on neurons, the loss of trophic support afforded by the astrocytes that are targeted, may well contribute to neurodegeneration in NMO patients. In both MS and NMO the exact mechanism of antibody-mediated neuronal damage, or the origin of autoreactive antibodies, remains to be fully clarified. Although it has been suggested that in MS, activation of an intrathecal B-cell pool may be caused by Epstein-Barr virus infection, this explanation remains controversial.60,61 In some movement disorders, antibodies to group A β -haemolytic streptococcal infections are known to cross-react with basal ganglia tissue, inducing motor and psychiatric symptoms. In these cases, treatment with antibiotics is very effective. In addition to pathogenic responses both play critical roles in mediating damage to neurons. Yet, and as stated above already, both innate and adaptive immune responses also play critical roles in down-regulating inflammatory processes and actively participate in the repair process. These beneficial roles are discussed further below.

Infectious agents and peripheral infections

Infections with viruses, bacteria and prions induce neurodegeneration by a variety of mechanisms. As well as inducing a direct cytolytic effect, neurotrophic viruses may trigger apoptosis, necrosis or autophagy of infected neurons because of virus-specific immunity. Infections may also induce the dying back of axons and secondary myelin damage, induce protein aggregation, or trigger neurotoxic pathways. Although not a direct target themselves, neurons can be subject to collateral damage through an immune-mediated attack on nearby, infected oligodendrocytes or astrocytes, or as a result of released pro-inflammatory factors (Fig. 2).

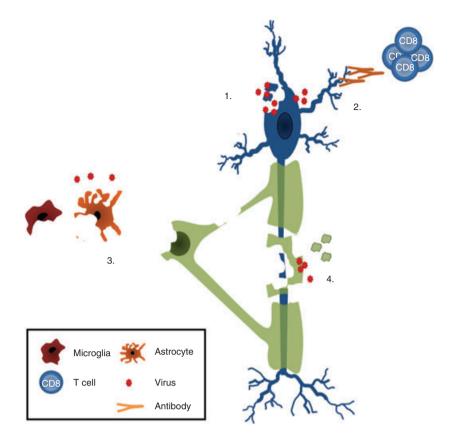


Figure 2. Proposed mechanisms of viral-induced neuronal damage. Infection of neurons with viruses leads to apoptosis, necrosis, autophagy or dying back of axons (1). Immune-mediated attack of neurons by virus-specific CD8⁺ T cells or autoantibodies induces damage (2). Infection of endothelial cells, ependymal cells, glia (astrocytes, oligodendrocytes, microglia) leads to so-called bystander damage as the result of release of cytokines or reactive oxygen species that damage neurons in a variety of ways (see text for details) (3). Secondary neuronal degeneration as a consequence of trophic support, when oligodendrocytes and myelin are infected with viruses (4).

Serological and epidemiological studies have revealed marked associations between infections and many neurodegenerative diseases. In some cases, viruses are considered to act as trigger, or to contribute to progression of disease by exacerbating clinical symptoms, for example in MS (Table 1). The association of viruses with neurodegenerative diseases in humans is supported by studies in animal models (Table 2). Not only can viruses or microbial components like lipopolysaccharide directly cause neuronal damage, they also influence the severity of disease. Peripheral infections are also known to precipitate as well as exacerbate ongoing neurodegeneration.⁶² Evidence for this comes from studies in experimental models in which peripheral inflammation is associated with exacerbation of experimental AD, MS, PD, prion disease and stroke. For example, an increase in $A\beta$ in the CNS of animals that survive sepsis correlates with a decrease in synaptophysin levels and cognitive impairment. Transgenic mice susceptible to neurodegeneration develop less severe disease when raised under pathogen-free conditions.63 That such associations also exists in humans is supported by the fact that upper respiratory tract infections are associated with the occurrence of clinical relapses during MS while AD has been associated with spirochetes and other pathogens such as *Chlamydia pneumoniae* and herpes simplex virus type 1; whether this is cause or effect remains to be determined (Table 4).

Metals trigger neurodegeneration

Exposure to metals and environmental pollutants has been postulated as a risk factor for neurodegenerative disorders.²² Also, as several of these agents can trigger or modulate immune responses, their possible involvement in inflammatory neurodegeneration should be considered. Indeed, an animal model of PD uses the neurotoxin 1-methyl-4-phenyl-1,2,5,6-tetrahydropyridine (MPTP) to selectively injure nigrostriatal dopaminergic neurons. The experimental use of MPTP was originally inspired by the PD-like syndrome in drug addicts who accidentally injected themselves with MPTP as a contaminant of pethidine. MPTP is metabolized by astrocytes to form the active metabolite 1-methyl-4-phenylpyridinium (MPP⁺) that enters cells via the dopamine transporter and inhibits complex 1 of the mitochondrial electron transport chain. As a result, depletion of striatal dopamine neurons

Tuble 1. Infectious agents milled to neurodegenerative aborder	agents linked to neurodegenerative	disorders
--	------------------------------------	-----------

Disease	Infectious agent	Association	References
AD	HSV-1	HSV-1 genome present in brain tissues. Genes related to HSV-1 reactivation in familial AD. Virus associated with protein aggregates. HSV-1 induces protein aggregation	64
	Chlamydia pneumoniae	90% of AD brains infected with <i>C. pneumoniae</i> in plaques Infection of mice induces AD pathology with $A\beta$ aggregation	65
ALS	Borrelia burgdorferi Retroviruses	Organism isolated from plaques Analysis of serum and brain tissues reveals increased expression of HERV-K	66 67
PD	Influenza	Risk of developing Parkinsonian symptoms increases with number of influenza attacks. Outbreaks of encephalitis lethargica (von Ecomo's disease) in 1918 influenza pandemic caused by type A H1N1. In PD influenza infections associated with transient tremor and gait disturbances. H5N1 induces α-synuclein deposits	68,69
MS	EBV HHV6 Pseudomonas aeruginosa, Retroviruses	Infectious mononucleosis associated with 2-3-fold higher risk to develop MS. Heightened antibody responses to EBV. Controversy over expression of EBV in the brain Antibodies to acinetobacteria cross-react with myelin Relapses associated with upper respiratory tract infections HHV6 present in MS lesions	60,61,70
HIV infection	HIV	HIV6 present in MS lesions HIV-associated neurocognitive disorder; HIV mild neurocognitive disorder; HIV-associated dementia. Replication of HIV and degeneration of synapses and neurons. Activation of microglia, macrophages and astrocytes. HIV increases amyloid plaque and $A\beta$ accumulation	71

AD, Alzheimer's disease; ALS, amyotrophic lateral sclerosis; EBV, Epstein–Barr virus; HIV, human immunodeficiency virus; HSV, herpes simplex virus; HERV-K, human endogenous retrovirus type K; HHV6, human herpes virus 6; MS, multiple sclerosis.

occurs, and recruitment and activation of microglia ensues. Similar effects are observed following administration of rotenone, or 6-hydroxydopamine infusion. Likewise, the deleterious effects on the CNS of psychostimulant use,⁷² exposure to pesticides,⁷³ mercury intoxication⁷⁴ and diesel exhaust particles involve at least in part neuroinflammatory mechanisms. One mechanism by which toxic agents could augment neurodegenerative diseases is through the induction of protein aggregation, or by activation of pathogenic innate immune responses, both common findings in neurodegenerative diseases of the peripheral and central nervous systems.

Ageing and immune responses in the CNS

Ageing is a major risk factor for many neurodegenerative disorders. In addition to the environmental factors discussed above, genetic mutations, oxidative or metabolic stress including endoplasmic reticulum and mitochondrial dysfunction are known to act as triggers for protein misfolding and aggregation. It is also known that these triggers accumulate with age. Another characteristic of ageing is the accumulation of advanced glycation end-products (AGE), which is accelerated in MS, AD and PD.⁷⁵ This is mirrored by an increase in the receptor for AGE (RAGE),

which induces pro-inflammatory cytokines and free radicals following engagement of RAGE, so perpetuating a cycle of damage. RAGE is increased in the AD-affected CNS, where it is expressed on neurons and astrocytes. During MS, RAGE is expressed on oligodendrocytes in response to stress, and levels of soluble RAGE are now under examination as a potential biomarker of disease.⁷⁵ One known ligand of RAGE is HMGB1, a DNA-binding protein with pro-inflammatory properties. Levels of HMGB1 are increased in AD and PD as well as in MS, and accordingly HMGB1 has been suggested to amplify the inflammatory response that causes the disease, possibly as a consequence of its ability to trigger TLR4-mediated inflammatory responses⁷⁶ in contrast to Huntington's disease where HMGB1 has been reported to protect neurons.77

Several factors contribute to an age-related increase in the risk of neurodegeneration,⁷⁸ In demyelinating diseases such as MS, it has become apparent that the ability to repair damage in the CNS declines with age. Concomitant with this is a reduction in the anti-oxidant decline, antioxidant defence mechanisms weaken, leaving axons and neurons more vulnerable to damage, part of which is due to the capacity of the innate immune system to facilitate repair by glia.⁷⁹ In addition, immune senescence the process by which immune responses become less efficient with age. This phenomenon manifests itself by an increased susceptibility to infections, a reduced ability to mount adaptive immune responses following vaccinations, and an increased susceptibility to autoimmune diseases. These phenomena all reflect a reduced ability to adequately control inflammatory responses.⁸⁰ Aging is associated with altered innate immune responses such as dysregulation of TLR responses and production of cytokines and other inflammatory mediators. In aged mice, dysmorphic microglia appear, which display a more reactive and activated phenotype. In addition, the levels of immune suppressive neuronal factors decrease, such as CX3CL1, its receptor CX3CR1 and CD200. This suggests that the ability of neurons to maintain microglia in a quiescent state becomes gradually less effective in an aged brain.²⁹

Much of what is known about the impact of ageing on neuroinflammatory processes has been derived from experimental intervention in aged animals such as the senescence-accelerated mouse.⁴³ In addition, systemic injection of lipopolysaccharide in aged mice leads to higher levels of microglial cytokines compared with young mice, reflecting a more sensitized state or 'primed' state of these aged microglia. That age impacts neuroinflammation is shown by studies using parabiosis between old and young mice, showing the importance of immune senescence in repair and synaptogenesis.^{79,81,82} However, it is still unclear which factors in humans lead to such priming. As discussed above systemic infections may fulfil that role.

Immune-mediated neurodegeneration

Necrosis of neurons is observed following acute brain injury and neurotrophic infections, or as a result of the release of damaging chemicals such as glutamate, nitric oxide and reactive oxygen species (Fig. 3). Damaged neurons themselves may also exacerbate immune-mediated disease by release of chemokines such as CCL21,⁸³ a ligand of the chemokine receptor CCR7 present on T cells, macrophages and microglia.84,85 Consequences of neuronal loss also include a reduction in the inhibitory signals that control production of pro-inflammatory cytokines and neurotoxic substances including nitric oxide, oxidative radicals and proteolytic enzymes by activated microglia. Disturbance in glutamate homeostasis as a result of astrocyte damage, for example, leads to excitotoxicity and is observed in acute viral encephalomyelitis, AD, PD and acute brain injury.

As explained above, activation of macrophages and microglia is observed during infections, trauma, following exposure to toxic compounds or, indeed, upon ageing. Aggregated, altered or misfolded proteins that accumulate during ageing or as a cause or consequence of pathology are known to represent DAMPs and bind to PRRs, key molecules that drive the innate immune responses. Although not all of these pathways are acutely detrimental to neurons, chronic microglia activation may compromise blood–brain barrier integrity, and lead to a markedly increased influx of peripheral immune cells as well as potentially neuron-reactive antibodies.

Cytotoxic CD8⁺ and CD4⁺ T cells can contribute to neuronal damage or destruction by directly targeting neurons. This is for example observed in Rasmussen syndrome. Under pathological conditions, neurons express MHC class I antigens, which makes them susceptible to CD8⁺ T-cell-mediated damage. Neurons and axons also express non-classical MHC molecules, and are susceptible to perforin- and granzyme B-mediated damage and Fas-FasL interactions. Neuronal damage may also occur as a consequence of an immune-mediated attack on myelin, oligodendrocytes or astrocytes, as observed during NMO.

Antibodies to neurons correlate with CNS injury in systemic lupus erythematous, ALS, MS and paraneoplastic neurological disorders. These antibodies may well be directed to receptors such as *N*-methyl D-aspartate receptors, inducing neuronal excitotoxicity, or to intracellular antigens such as neurofilaments. Autoantibodies to neurofascin, KIR4, contactin-2 and neurofilaments are present in MS patients, and such antibodies can mediate axonal injury in mice.^{10,86} The autoantibodies could activate the complement system, inducing damage as a result of membrane attack complex formation. In AD, one risk factor is the presence of polymorphisms in complement receptor 1 that may reduce the ability of microglia to clear $A\beta$.⁸⁷ In vitro, aggregated amyloid and prion

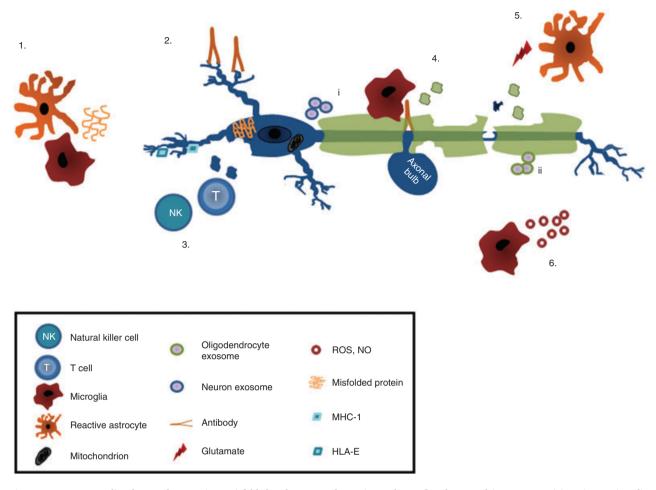


Figure 3. Immune-mediated neurodegeneration. Misfolded and aggregated proteins such as $A\beta$ and α -synuclein aggregates (1), activate microglia and suppress brain derived neuronal growth factor production by astrocytes. Neuronal specific antibodies (2) activate the complement system or FcR-mediated damage. Natural killer and T cells damage neurons either via MHC class-I or non-classical MHC molecules (3). Damage to neurons reduces the levels of neuroimmunoregulatory molecules, decreasing the immune suppressive environment (4). Excessive production of glutamate by, for example, stressed neurons and activated immune cells, together with reduced glutamate uptake causes excitotoxic damage of neurons (5). Macrophage and microglia activation triggers release of reactive oxygen and nitrogen species, matrix metalloproteinases, chemokines and cytokines known to damage axons and neurons and to cause mitochondrial dysfunction. Neurons (i) and oligodendrocytes (ii) release exosomes that may trigger pathogenic immune responses (6). NO, nitric oxide; ROS, reactive oxygen species.

proteins bind C1q directly and activate complement in the absence of antibodies, thereby aiding clearance. The role of complement in ALS is somewhat different and depends on the model.^{88,89} In addition, deficiency of the complement regulator exacerbates Wallerian degeneration, clearly showing a role for complement in neurodegeneration.⁹⁰

Apart from being pathogenic in some cases, immune responses can also be helpful in controlling and limiting pathogenic responses (Fig. 4). As discussed above, the CNS is equipped with an arsenal of neuroimmunoregulatory molecules that help to maintain immune privilege. Despite the well-known anti-inflammatory properties of regulatory microglia in several neurodegenerative disorders,^{7,91} astrocytes also contribute to protection from immune-mediated damage by preferentially inducing apoptosis of infiltrating T cells by Fas–FasL interactions.

Astrocytes also produce interleukin-1 and encourage influx of regulatory T cells by producing interleukin-27. In addition, astrocytes are the major source of nerve growth factor and glial cell line-derived neurotrophic factor in the CNS, thereby aiding neuronal regeneration.^{92,93} Such neuronal growth factors are also secreted by T cells⁹⁴ that have been shown to aid neurite outgrowth *in vitro*, reduce secondary neurodegeneration *in vivo*, and contribute to learning behaviour. These findings indicate a likely neuroreparative role at least for some T cells during neurodegenerative disorders.⁹⁵

Cross-talk between the immune system and the CNS is known to occur via exosomes, small vesicles released from multiple cell types that contain protein, lipids or RNA species including viral RNA.⁹⁶ Following their secretion, exosomes are taken up by neighbouring cells and deliver

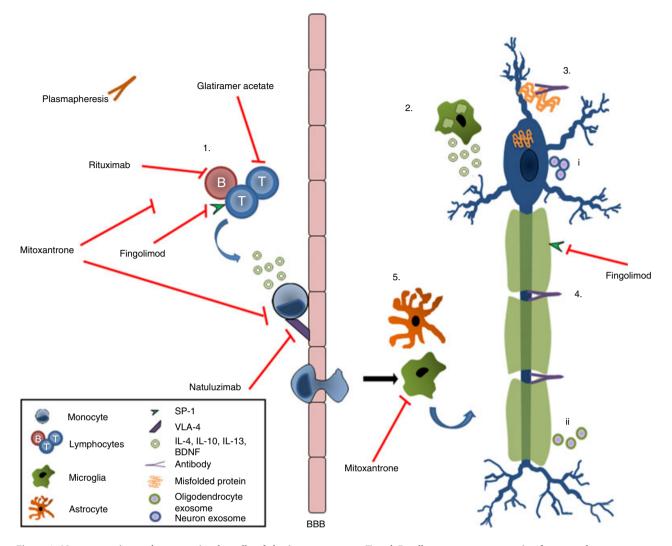


Figure 4. Neuroprotection and regeneration by cells of the immune system. T and B cells secrete neuroprotective factors and suppress proinflammatory responses (1), since macrophages/microglia that phagocytose myelin stimulate growth and repair via glial cell-derived neuronal growth factors (2). Antibodies aid remyelination thereby restoring trophic support of neurons (3) and help to remove the aggregated proteins (4). Reparative microglia and astrocytes produce neuronal growth factors (5) neurons (i) and oligodendrocytes (ii) release exosomes that dampen the pathogenic immune responses. BBB, blood–brain barrier; BDNF, brain derived neural growth factor ; IL-4, interleukin-4; SP-1, sphingosine-1-phosphate; VLA-4, very-late antigen 4.

their cargo to those recipient cells. It has been well established that neurons, microglia, astrocytes and oligodendrocytes secrete exosomes, and stressed oligodendrocytes do so even more. In the latter case, myelinating oligodendrocytes transport RNA and proteins to axons, and such support helps to maintain neuronal integrity.⁹⁷ In PD for example, the protein α -synuclein, a mediator of neurodegeneration, is released by cultured neurons via exosomes. Addition of such exosomes to cultured neurons induces neuronal cell death, again pointing to a role of exosomes in disease.⁹⁸ Given these emerging examples of how exosomes participate in the communication between neural cells under neurodegenerative conditions, it seems rather likely that they will turn out to have an important role in many neurodegenerative diseases by providing trophic support, by transferring pathogenic aggregated proteins or aiding repair by providing growth factors. They are also likely to play an important role in antigen presentation within the CNS by transferring antigens in the CNS parenchyma to perivascular antigen-presenting cells that meet up with T cells engaged in immune surveillance. Exosome-mediated signalling is clearly an intriguing new field of study in neurodegenerative disorders.

Immunotherapy in neurodegenerative disorders

Evidence for the involvement of immunity in the development and progression of neurodegenerative disorders has inspired immunotherapeutic approaches to prevent neuronal loss and to aid neuronal growth. In paraneoplastic neurological disorders, for example, the reduction of autoantibodies by plasmapheresis is usually beneficial in suppressing the immune response and improving the neurological status, at least in the short term. For longterm treatment combined therapies targeting the tumour as well are often necessary.

The use of systemic anti-inflammatory pharmacological agents may contribute to the management of neurodegenerative diseases. Epidemiological studies have shown a lower prevalence of AD in people who have regularly taken non-steroidal anti-inflammatory drugs⁹⁹ and in MS there is a prominent role for glucocorticosteroids during the treatment of exacerbations.

Inhibiting T-cell trafficking into the CNS appears a rational approach for some neurodegenerative diseases. However, such approaches not only block the entry of potentially pathogenic T cells, but also of reparative regulatory T cells as well as T cells required for control of tumours and infections. Inhibition with antibodies directed to T cells and or monocytes (trafficking) is one of the therapeutic targets in neurodegenerative diseases.

Natalizumab, a humanized monoclonal antibody against very late activating antigen-4 is an example of a therapeutic agent that acts by blocking lymphocytes going into the CNS. Alternative agents include Fingolimod, which is designed to entrap lymphocytes in lymphoid tissue. Another mechanism of action of Fingolimod is promoting oligodendrocyte proliferation and differentiation and therefore repair. Rituximab, a chimeric monoclonal antibody against the CD20 antigen, present on B cells, induces a transient depletion of B cells, resulting in a substantial effect on the inflammatory disease activity in MS.¹⁰⁰ Due to the susceptibility of the CNS for infections, long-term treatment with these therapies needs to be seriously considered.

Supplementation with intravenous immunoglobulins has gained increasing interest as a therapeutic approach to neuroinflammatory disorders. Although their actions remain to be fully clarified, intravenous immunoglobulins are likely to promote the development of regulatory T cells with beneficial reparative effects when recruited into the CNS. Active and passive immunotherapies to provide naturally occurring antibodies to remove $A\beta$ plaques have also been used. Such approaches have also been applied to manage prion diseases and PD, in which antibodies target α -synuclein that accumulates as Lewy bodies in dopaminergic neurons.

The idea that innate immune responses, in particular by microglia, are inextricably involved in neurodegeneration has inspired approaches to inhibit or modulate microglial activation for therapeutic purposes. Certain agents that inhibit microglial responses, such as minocycline or nicergoline, have been tested in several models of human disorders¹⁰¹ (Fig. 4). However, many have not yet been screened in patients, or have been found to be less effective in humans than in the corresponding animal models. Other ways to modulate the intimate interactions between microglia and neurons have been exploited for therapeutic approaches as well. For example, application of exogenous CXCL1 (generally expressed on neurons) has been shown to be neuroprotective, probably by controlling microglial activation. Likewise, the heat-shock protein hsp B5 exerts neuroprotective effects. hsp B5 is a chaperokine produced by oligodendrocytes, and it activates regulatory microglial responses⁴⁰ during MS. In several animal models of neurodegeneration, hsp B5 has been found to be neuroprotective, indicating that modulating innate microglial responses within the CNS can be an effective approach in neurodegenerative diseases. Other approaches targeting innate responses include blocking RAGE, supplying glutamate antagonists, and targeting oxidative stress or the complement pathway.²

Sodium-channel-blocking drugs have been reported to inhibit microglial activity and may have neuroprotective activity,^{102,103} during neuroinflammation. Finally, CBderived drugs are promising neuroprotective strategies and have been shown to not only exert neuroprotective, but also some immunosuppressive, activity.¹⁰⁴ When activated, neuronal CB1 receptors have been shown to attenuate excitotoxic glutamatergic neurotransmission,¹⁰⁴ this triggers signalling pathways that are effective in inhibiting signs of degeneration, although benefit was noted in only a subset of people with progressive MS¹⁰⁵ neurodegeneration in animal models.^{106,107}

Of particular interest is the use of neural stem and progenitor cell replacement therapy. These cells are no longer believed to primarily replace damaged cells, but rather to modulate immune responses and repair processes in the CNS¹⁰⁸ by secreting a variety of soluble factors. However, endogenous, and transplanted neural stem and progenitor cells are also influenced by the immune environment, which may either augment migration and proliferation, or inhibit their effects.¹⁰⁹ As a result, their impact is influenced by the nature of the inflammatory microenvironment in which they end up.

Conclusions

The nervous and immune systems are inextricably interlinked. In several cases, mediators used by the immune system to combat infections and promote tissue repair are also used by the CNS for growth and development. This common language is exemplified by the role of microglia in synaptic pruning, and the role of cytokines during development of the brain. Despite the fact that the CNS is an immune-privileged site, innate and adaptive immune responses do regularly take place in the CNS. They are essential to eliminate infectious agents and tumours, as well as for clearing debris and promoting tissue repair. Despite these beneficial roles of immune responses, such responses must remain under tight control to prevent any damage to the CNS. The bloodbrain barrier and the powerful immune-regulatory functions of all types of neural cells act together to secure such tight control. Microglia are actively maintained in a quiescent state, and the influx and local activation of peripheral immune cells is severely limited. Despite these measures, chronic immune activation within the CNS is a pathological hallmark of a wide number of neurodegenerative disorders. The factors that drive chronic inflammation include the accumulation of misfolded and aggregated proteins, heat-shock proteins and other DAMPs that are all known to trigger local innate responses. Viral and bacterial infections clearly play a role as well, as do dietary substances, or the lack of them, and toxic compounds in the environment. It appears that with aging the threshold for immune activation within the CNS goes down. This so-called priming of immune responses may play a major role in the increased risk of neurodegenerative diseases in the elderly. To develop novel therapeutic approaches to control these diseases, a closer look at the way immune regulation is organized in the CNS is useful. A better understanding of the endogenous protective pathways will undoubtedly reveal ways to harness reparative processes, and so improve control over chronic inflammatory neurodegenerative disorders.

Acknowledgements

For our research discussed in this review the authors gratefully acknowledge the financial support of the Dutch Multiple Sclerosis (MS) Research Foundation and the MS Society of Great Britain and Northern Ireland. We are also very grateful to all our colleagues for their views and contributions to the topic during many discussions.

Disclosures

JMvN holds equity in Delta Crystallon BV. The other authors have no conflicts of interest to report.

References

- World Health Organization. Dementia a public health priority. [Geneva]:: World Health Organization 2012. http://site.ebrary.com/lib/ucmerced/Doc?id=10718026 [accessed 14 November 2013].
- 2 Amor S, Puentes F, Baker D et al. Inflammation in neurodegenerative diseases. Immunology 2010; 129:154–69.
- 3 Town T, Tan J, Flavell RA et al. T-cells in Alzheimer's disease. Neuromolecular Med 2005; 7:255–64.
- 4 Bellucci A, Bugiani O, Ghetti B et al. Presence of reactive microglia and neuroinflammatory mediators in a case of frontotemporal dementia with P301S mutation. Neurodegener Dis 2011; 8:221–9.
- 5 Mosley RL, Hutter-Saunders JA, Stone DK et al. Inflammation and adaptive immunity in Parkinson's disease. Cold Spring Harb Perspect Med 2012; 2:a009381.
- 6 Maetzler W, Berg D, Synofzik M et al. Autoantibodies against amyloid and glialderived antigens are increased in serum and cerebrospinal fluid of Lewy body-associated dementias. J Alzheimers Dis 2011; 26:171–9.

- 7 Ellrichmann G, Reick C, Saft C et al. The role of the immune system in Huntington's disease. Clin Dev Immunol 2013; 2013:541259.
- 8 Papadimitriou D, Le Verche V, Jacquier A et al. Inflammation in ALS and SMA: sorting out the good from the evil. Neurobiol Dis 2010; 37:493–502.
- 9 Lobsiger CS, Cleveland DW. Glial cells as intrinsic components of non-cell-autonomous neurodegenerative disease. Nat Neurosci 2007; 10:1355–60.
- 10 Melzer N, Meuth SG, Wiendl H. Neuron-directed autoimmunity in the central nervous system: entities, mechanisms, diagnostic clues, and therapeutic options. *Curr Opin Neurol* 2012; 25:341–8.
- 11 Zuliani L, Graus F, Giometto B et al. Central nervous system neuronal surface antibody associated syndromes: review and guidelines for recognition. J Neurol Neurosurg Psychiatr 2012; 83:638–45.
- 12 Archer LD, Langford-Smith KJ, Bigger BW et al. Mucopolysaccharide diseases: a complex interplay between neuroinflammation, microglial activation and adaptive immunity. J Inherit Metab Dis 2014; 37:1–12.
- 13 Sta M, Sylva-Steenland RMR, Casula M et al. Innate and adaptive immunity in amyotrophic lateral sclerosis: evidence of complement activation. *Neurobiol Dis* 2011; 42:211–20.
- 14 Puentes F, Topping J, Kuhle J et al. Immune reactivity to neurofilament proteins in the clinical staging of amyotrophic lateral sclerosis. J Neurol Neurosurg Psychiatr 2013. doi:10.1136/jnnp-2013-305494.
- 15 Podda G, Nyirenda M, Crooks J et al. Innate immune responses in the CNS: role of toll-like receptors, mechanisms, and therapeutic opportunities in multiple sclerosis. J Neuroimmune Pharmacol 2013; 8:791–806.
- 16 Van Noort JM, Baker D, Amor S. Mechanisms in the development of multiple sclerosis lesions: reconciling autoimmune and neurodegenerative factors. CNS Neurol Disord Drug Targets 2012; 11:556–69.
- 17 Van der Valk P, Amor S. Preactive lesions in multiple sclerosis. Curr Opin Neurol 2009; 22:207–13.
- 18 Van Noort JM, Bsibsi M, Gerritsen WH et al. α-B-crystallin is a target for adaptive immune responses and a trigger of innate responses in preactive multiple sclerosis lesions. J Neuropathol Exp Neurol 2010; 69:694–703.
- 19 Saji E, Arakawa M, Yanagawa K et al. Cognitive impairment and cortical degeneration in neuromyelitis optica. Ann Neurol 2013; 73:65–76.
- 20 Mitsdoerffer M, Kuchroo V, Korn T. Immunology of neuromyelitis optica: a T cell–B cell collaboration. Ann N Y Acad Sci 2013; 1283:57–66.
- 21 Bradford BM, Mabbott NA. Prion disease and the innate immune system. Viruses 2012; 4:3389–419.
- 22 Kraft AD, Harry GJ. Features of microglia and neuroinflammation relevant to environmental exposure and neurotoxicity. Int J Environ Res Public Health 2011; 8:2980–3018.
- 23 Chamorro Á, Meisel A, Planas AM et al. The immunology of acute stroke. Nat Rev Neurol 2012; 8:401–10.
- 24 Brait VH, Arumugam TV, Drummond GR et al. Importance of T lymphocytes in brain injury, immunodeficiency, and recovery after cerebral ischemia. J Cereb Blood Flow Metab 2012; 32:598–611.
- 25 Bauer J, Vezzani A, Bien CG. Epileptic encephalitis: the role of the innate and adaptive immune system. *Brain Pathol* 2012; 22:412–21.
- 26 Das M, Mohapatra S, Mohapatra SS. New perspectives on central and peripheral immune responses to acute traumatic brain injury. J Neuroinflammation 2012; 9:236.
- 27 Locatelli G, Wörtge S, Buch T et al. Primary oligodendrocyte death does not elicit anti-CNS immunity. Nat Neurosci 2012; 15:543–50.
- 28 Hoarau J-J, Krejbich-Trotot P, Jaffar-Bandjee M-C et al. Activation and control of CNS innate immune responses in health and diseases: a balancing act finely tuned by neuroimmune regulators (NIReg). CNS Neurol Disord Drug Targets 2011; 10:25–43.
- 29 Kierdorf K, Prinz M. Factors regulating microglia activation. Front Cell Neurosci 2013; 7:44.
- 30 Peferoen L, Kipp M, van der Valk P et al. Oligodendrocyte-microglia cross-talk in the CNS. Immunology 2013; 141:302–13.
- 31 Goldbaum O, Richter-Landsberg C. Stress proteins in oligodendrocytes: differential effects of heat shock and oxidative stress. J Neurochem 2001; 78:1233–42.
- 32 Toguchi M, Gonzalez D, Furukawa S et al. Involvement of Sema4D in the control of microglia activation. Neurochem Int 2009; 55:573–80.
- 33 Okada A, Tominaga M, Horiuchi M et al. Plexin-A4 is expressed in oligodendrocyte precursor cells and acts as a mediator of semaphorin signals. Biochem Biophys Res Commun 2007; 352:158–63.
- 34 Tian L, Lappalainen J, Autero M et al. Shedded neuronal ICAM-5 suppresses T-cell activation. Blood 2008; 111:3615–25.
- 35 Wang Y, Szretter KJ, Vermi W et al. IL-34 is a tissue-restricted ligand of CSF1R required for the development of Langerhans cells and microglia. Nat Immunol 2012; 13:753–60.
- 36 Mott RT, Ait-Ghezala G, Town T *et al.* Neuronal expression of CD22: novel mechanism for inhibiting microglial proinflammatory cytokine production. *Glia* 2004; 46:369–79.

- 37 Chavarría A, Cárdenas G. Neuronal influence behind the central nervous system regulation of the immune cells. *Front Integr Neurosci* 2013; 7:64.
- 38 Skaper SD, Facci L, Giusti P. Mast cells, glia and neuroinflammation: partners in crime? Immunology 2013; 141:314–27.
- 39 Broz P, Monack DM. Newly described pattern recognition receptors team up against intracellular pathogens. Nat Rev Immunol 2013; 13:551–65.
- 40 Bsibsi M, Holtman IR, Gerritsen WH et al. α-B-crystallin induces an immune-regulatory and antiviral microglial response in preactive multiple sclerosis lesions. J Neuropathol Exp Neurol 2013; 72:970–9.
- 41 Bsibsi M, Persoon-Deen C, Verwer RWH *et al.* Toll-like receptor 3 on adult human astrocytes triggers production of neuroprotective mediators. *Glia* 2006; 53:688–95.
- 42 Lee J-E, Han P-L. An update of animal models of Alzheimer disease with a reevaluation of plaque depositions. *Exp Neurobiol* 2013; 22:84–95.
- 43 Shimada A, Hasegawa-Ishii S. Senescence-accelerated mice (SAMs) as a model for brain aging and immunosenescence. Aging Dis 2011; 2:414–35.
- 44 Moser JM, Bigini P, Schmitt-John T. The wobbler mouse, an ALS animal model. Mol Genet Genomics 2013; 288:207–29.
- 45 Figiel M, Szlachcic WJ, Switonski PM et al. Mouse models of polyglutamine diseases: review and data table. Part I. Mol Neurobiol 2012; 46:393–429.
- 46 Blesa J, Phani S, Jackson-Lewis V et al. Classic and new animal models of Parkinson's disease. J Biomed Biotechnol 2012; 2012:845618.
- 47 Wadsworth JDF, Asante EA, Collinge J. Review: contribution of transgenic models to understanding human prion disease. *Neuropathol Appl Neurobiol* 2010; 36:576–97.
- 48 Macrae IM. Preclinical stroke research–advantages and disadvantages of the most common rodent models of focal ischaemia. Br J Pharmacol 2011; 164:1062–78.
- 49 Van der Star BJ, Vogel DYS, Kipp M et al. In vitro and in vivo models of multiple sclerosis. CNS Neurol Disord Drug Targets 2012; 11:570–88.
- 50 Amor S, Smith PA, Hart B' et al. Biozzi mice: of mice and human neurological diseases. J Neuroimmunol 2005; 165:1–10.
- 51 Bajramovic JJ. Regulation of innate immune responses in the central nervous system. CNS Neurol Disord Drug Targets 2011; 10:4–24.
- 52 Shook BC, Jackson PF. Adenosine A(2A) receptor antagonists and Parkinson's disease. ACS Chem Neurosci 2011; 2:555–67.
- 53 Brennan FH, Anderson AJ, Taylor SM et al. Complement activation in the injured central nervous system: another dual-edged sword? J Neuroinflammation 2012; 9:137.
- 54 Fisher Y, Nemirovsky A, Baron R et al. T cells specifically targeted to amyloid plaques enhance plaque clearance in a mouse model of Alzheimer's disease. PLoS One 2010; 5: e10830.
- 55 Zhang J, Ke K-F, Liu Z et al. Th17 cell-mediated neuroinflammation is involved in neurodegeneration of aβ1-42-induced Alzheimer's disease model rats. PLoS One 2013; 8:e75786.
- 56 Puentes F, van der Star BJ, Victor M et al. Characterization of immune response to neurofilament light in experimental autoimmune encephalomyelitis. J Neuroinflammation 2013; 10:118.
- 57 Huizinga R, Gerritsen W, Heijmans N et al. Axonal loss and gray matter pathology as a direct result of autoimmunity to neurofilaments. *Neurobiol Dis* 2008; 32: 461–70.
- 58 Srivastava R, Aslam M, Kalluri SR et al. Potassium channel KIR4.1 as an immune target in multiple sclerosis. N Engl J Med 2012; 367:115–23.
- 59 Kusunoki S. Autoantibodies in neuroimmunological diseases; relevance of fine specificity. Exp Neurol 2013; 250C:219–20.
- 60 Lassmann H, Niedobitek G, Aloisi F et al. Epstein–Barr virus in the multiple sclerosis brain: a controversial issue – report on a focused workshop held in the Centre for Brain Research of the Medical University of Vienna, Austria. Brain 2011; 134:2772–86.
- 61 Peferoen LAN, Lamers F, Lodder LNR et al. Epstein–Barr virus is not a characteristic feature in the central nervous system in established multiple sclerosis. Brain 2010; 133:e137.
- 62 Perry VH. Contribution of systemic inflammation to chronic neurodegeneration. Acta Neuropathol 2010; 120:277–86.
- 63 Capsoni S, Carucci NM, Cattaneo A. Pathogen free conditions slow the onset of neurodegeneration in a mouse model of nerve growth factor deprivation. J Alzheimers Dis 2012; 31:1–6.
- 64 Bearer EL. HSV, axonal transport and Alzheimer's disease: in vitro and in vivo evidence for causal relationships. *Future Virol* 2012; 7:885–99.
- 65 Honjo K, van Reekum R, Verhoeff NPLG. Alzheimer's disease and infection: do infectious agents contribute to progression of Alzheimer's disease? *Alzheimers Dement* 2009; 5:348–60.
- 66 Miklossy J. Alzheimer's disease a neurospirochetosis. Analysis of the evidence following Koch's and Hill's criteria. J Neuroinflammation 2011; 8:90.
- 67 Alfahad T, Nath A. Retroviruses and amyotrophic lateral sclerosis. Antiviral Res 2013; 99:180–7.

- 68 Hack N, Jicha GA, Abell A et al. Substantia nigra depigmentation and exposure to encephalitis lethargica. Ann Neurol 2012; 72:912–7.
- 69 De Chiara G, Marcocci ME, Sgarbanti R et al. Infectious agents and neurodegeneration. Mol Neurobiol 2012; 46:614–38.
- 70 Virtanen JO, Jacobson S. Viruses and multiple sclerosis. CNS Neurol Disord Drug Targets 2012: 11:528–44.
- 71 Del Palacio M, Alvarez S, Muñoz-Fernández MÁ. HIV-1 infection and neurocognitive impairment in the current era. Rev Med Virol 2012; 22:33–45.
- 72 Clark KH, Wiley CA, Bradberry CW. Psychostimulant abuse and neuroinflammation: emerging evidence of their interconnection. *Neurotox Res* 2013; 23:174–88.
- 73 Lewis JA, Gehman EA, Baer CE et al. Alterations in gene expression in Caenorhabditis elegans associated with organophosphate pesticide intoxication and recovery. BMC Genomics 2013; 14:291.
- 74 Kern JK, Geier DA, Audhya T et al. Evidence of parallels between mercury intoxication and the brain pathology in autism. Acta Neurobiol Exp (Wars) 2012; 72:113–53.
- 75 Harris RA, Amor S. Sweet and sour oxidative and carbonyl stress in neurological disorders. CNS Neurol Disord Drug Targets 2011; 10:82–107.
- 76 Laird MD, Shields JS, Sukumari-Ramesh S et al. High mobility group box protein-1 promotes cerebral edema after traumatic brain injury via activation of toll-like receptor 4. Glia 2014; 62:26–38.
- 77 Min HJ, Ko EA, Wu J et al. Chaperone-like activity of high-mobility group box 1 protein and its role in reducing the formation of polyglutamine aggregates. J Immunol 2013; 190:1797–806.
- 78 Streit WJ, Xue Q-S. The brain's aging immune system. Aging Dis 2010; 1:254-61.
- 79 Miron VE, Boyd A, Zhao J-W et al. M2 microglia and macrophages drive oligodendrocyte differentiation during CNS remyelination. Nat Neurosci 2013; 16:1211–8.
- 80 Goronzy JJ, Li G, Yang Z et al. The janus head of T cell aging autoimmunity and immunodeficiency. Front Immunol 2013; 4:131.
- 81 Ajami B, Bennett JL, Krieger C et al. Infiltrating monocytes trigger EAE progression, but do not contribute to the resident microglia pool. Nat Neurosci 2011; 14:1142–9.
- 82 Villeda SA, Luo J, Mosher KI et al. The ageing systemic milieu negatively regulates neurogenesis and cognitive function. Nature 2011; 477:90–4.
- 83 Biber K, Tsuda M, Tozaki-Saitoh H et al. Neuronal CCL21 up-regulates microglia P2X4 expression and initiates neuropathic pain development. EMBO J 2011; 30:1864– 73.
- 84 Durafourt BA, Moore CS, Zammit DA et al. Comparison of polarization properties of human adult microglia and blood-derived macrophages. Glia 2012; 60:717–27.
- 85 Melief J, Koning N, Schuurman KG et al. Phenotyping primary human microglia: tight regulation of LPS responsiveness. Glia 2012; 60:1506–17.
- 86 Elliott C, Lindner M, Arthur A et al. Functional identification of pathogenic autoantibody responses in patients with multiple sclerosis. Brain 2012; 135:1819–33.
- 87 Crehan H, Hardy J, Pocock J. Microglia, Alzheimer's disease, and complement. Int J Alzheimers Dis 2012; 2012:983640.
- 88 Lee JD, Kamaruzaman NA, Fung JN et al. Dysregulation of the complement cascade in the hsod1G93A transgenic mouse model of amyotrophic lateral sclerosis. J Neuroinflammation 2013; 10:119.
- 89 Lobsiger CS, Boillée S, Pozniak C et al. C1q induction and global complement pathway activation do not contribute to ALS toxicity in mutant SOD1 mice. Proc Natl Acad Sci USA 2013; 110:E4385–92.
- 90 Ramaglia V, King RHM, Morgan BP et al. Deficiency of the complement regulator CD59a exacerbates Wallerian degeneration. Mol Immunol 2009; 46:1892–6.
- 91 Schwartz M, Kipnis J, Rivest S et al. How do immune cells support and shape the brain in health, disease, and aging? J Neurosci 2013; 33:17587–96.
- 92 Gleichman AJ, Carmichael ST. Astrocytic therapies for neuronal repair in stroke. *Neurosci Lett* 2013. doi:10.1016/j.neulet.2013.10.055
- 93 White RE, Jakeman LB. Don't fence me in: harnessing the beneficial roles of astrocytes for spinal cord repair. *Restor Neurol Neurosci* 2008; 26:197–214.
- 94 Chisholm SP, Cervi AL, Nagpal S et al. Interleukin-17A increases neurite outgrowth from adult postganglionic sympathetic neurons. J Neurosci 2012; 32:1146–55.
- 95 Kipnis J, Gadani S, Derecki NC. Pro-cognitive properties of T cells. Nat Rev Immunol 2012; 12:663–9.
- 96 Carson JH, Gao Y, Tatavarty V et al. Multiplexed RNA trafficking in oligodendrocytes and neurons. Biochim Biophys Acta 2008; 1779:453–8.
- 97 Frühbeis C, Fröhlich D, Kuo WP et al. Neurotransmitter-triggered transfer of exosomes mediates oligodendrocyte-neuron communication. PLoS Biol 2013; 11: e1001604.
- 98 Rajendran L, Honsho M, Zahn TR et al. Alzheimer's disease β-amyloid peptides are released in association with exosomes. Proc Natl Acad Sci USA 2006; 103:11172–7.
- 99 Jaturapatporn D, Isaac MG, McCleery J et al. Aspirin, steroidal and non-steroidal anti-inflammatory drugs for the treatment of Alzheimer's disease. Cochrane Database Syst Rev 2012; 2:CD006378.
- 100 Castillo-Trivino T, Braithwaite D, Bacchetti P et al. Rituximab in relapsing and progressive forms of multiple sclerosis: a systematic review. PLoS One 2013; 8:e66308.

Inflammation and neurodegeneration

- 101 Li C, Yuan K, Schluesener H. Impact of minocycline on neurodegenerative diseases in rodents: a meta-analysis. *Rev Neurosci* 2013; 24:553–62.
- 102 Morsali D, Bechtold D, Lee W et al. Safinamide and flecainide protect axons and reduce microglial activation in models of multiple sclerosis. Brain 2013; 136:1067–82.
- 103 Raftopoulos RE, Kapoor R. Neuroprotection for acute optic neuritis—Can it work? Mult Scler Relat Disord 2013; 2:307–11.
- 104 Pryce G, Ahmed Z, Hankey DJR et al. Cannabinoids inhibit neurodegeneration in models of multiple sclerosis. Brain 2003; 126:2191–202.
- 105 Zajicek J, Ball S, Wright D et al. Effect of dronabinol on progression in progressive multiple sclerosis (CUPID): a randomised, placebo-controlled trial. Lancet Neurol 2013; 12:857–65.
- 106 Butti E, Bacigaluppi M, Rossi S et al. Subventricular zone neural progenitors protect striatal neurons from glutamatergic excitotoxicity. Brain 2012; 135:3320–35.
- 107 Pryce G, Baker D. Potential control of multiple sclerosis by cannabis and the endocannabinoid system. CNS Neurol Disord Drug Targets 2012; 11:624–41.
- 108 Laterza C, Merlini A, De Feo D et al. iPSC-derived neural precursors exert a neuroprotective role in immune-mediated demyelination via the secretion of LIF. Nat Commun 2013; 4:2597.
- 109 Kokaia Z, Martino G, Schwartz M et al. Cross-talk between neural stem cells and immune cells: the key to better brain repair? Nat Neurosci 2012; 15:1078– 87.