

Association between Cellular Immune Activation, Target Cell Frequency, and Risk of Human Immunodeficiency Virus Type 1 Superinfection

Catherine A. Blish,^{a,b*} Ozge C. Dogan,^a Walter Jaoko,^c R. Scott McClelland,^{b,c,d,e} Kishorchandra Mandalaya,^f Katherine Odem-Davis,^a Barbra A. Richardson,^{e,g,h} Julie Overbaugh^a

Division of Human Biology^a and Vaccine and Infectious Disease Division,^b Fred Hutchinson Cancer Research Center, Seattle, Washington, USA; Departments of Medicine,^b Epidemiology,^d Global Health,^e and Biostatistics,^g University of Washington, Seattle, Washington, USA; Department of Medical Microbiology, University of Nairobi, Nairobi, Kenya^f; Coast General Hospital, Mombasa, Kenya^f

We performed a case-control study of women at risk of HIV-1 superinfection to understand the relationship between immune activation and HIV-1 acquisition. An increase in the frequency of HIV-1 target cells, but not in other markers of T cell activation, was associated with a 1.7-fold increase in the odds of superinfection. This suggests that HIV-1 acquisition risk is influenced more by the frequency of target cells than by the generalized level of immune activation.

Inflammation and immune activation promote HIV-1 disease progression, presumably because activated target cells support high levels of viral replication (1, 2); however, their role in driving HIV-1 transmission is less clear. A number of studies have examined immune activation in HIV-exposed but uninfected individuals. The majority of these studies support the idea that low levels of immune activation at the time of exposure reduce the risk of HIV-1 infection (3–16). However, one recent study indicated that condom use was associated with lower levels of immune activation, potentially confounding some of the prior results (10). Further, in other studies, exposure to HIV-1 is associated with increased levels of immune activation (17–20) and with the development of HIV-1-specific immune responses (18, 21–26) that could play a role in protection. Differences in study design, control groups, and the type of immune response that was assessed and classified as immune activation may be partly responsible for these somewhat conflicting results from exposed, uninfected cohorts.

HIV-1 vaccine trials have done little to clarify the role of immune activation in driving HIV-1 acquisition. In the Step Study, the enhanced risk of infection among the members of a subset of vaccinees was attributed to their circumcision status and prior elevated antibody titers with respect to the Ad5 vector rather than to increased immune activation of T cells (27, 28). In the RV144 vaccine trial, generalized immune activation was not directly linked to either protection or susceptibility (29). The factors that contribute to HIV-1 acquisition in the setting of natural exposure to diverse circulating strains remain incompletely understood.

Here, we examined the association between immune activation and HIV-1 acquisition in a cohort of HIV-1-infected women at ongoing risk of HIV-1 superinfection through sex work. A subset of these women went on to become superinfected with a second HIV-1 strain (30–32). Superinfection provides a unique opportunity to evaluate correlates of HIV-1 acquisition, since both immune activation and HIV-1-specific immune responses can be evaluated. Our previous studies of this cohort found no significant differences in preexisting HIV-1-specific antibody or T cell responses in women who went on to be superinfected versus those who did not (33–35). However, individuals in other cohort studies of mostly male subjects who acquired a second HIV-1 infection

within ~1 year of initial infection were found to have weak neutralizing antibody responses to their initial infection, suggesting the possibility that neutralizing antibodies can play some role in mediating susceptibility (36–38). Superinfection has also been noted to occur despite broad CD8⁺ T cell responses (39). None of these studies explored the role of generalized immune activation in HIV-1 superinfection, though such activation could mitigate the beneficial effects of HIV-1-specific immunity.

Immune activation was assessed by examining differentiation and activation markers on T cell subsets by flow cytometry analysis of peripheral blood mononuclear cells (PBMCs) from 10 superinfected women and 29 nonsuperinfected controls as previously described (34). Cases and controls were matched based on the timing of sample collection with respect to initial infection and their HIV-1 plasma viral loads (Table 1). All analyses were performed using samples collected at the visit prior to documented superinfection in order to assess immunity at the time point most relevant in terms of exposure to, and lack of protection from, the second virus. Cases and controls did not differ significantly in their sex frequencies (mean of 1.4 versus 1.5 self-reported sex acts in the preceding week averaged over follow-up prior to sample collection; Table 1). CD4 counts were not routinely available but were >200 cells/mm³ either immediately prior to or within 2 years of superinfection in all cases (30–32). As expected based on prior results (1, 40–43), HIV-infected individuals showed higher levels of CD4⁺ and CD8⁺ T cell activation markers (Ki-67, CD38, HLA-DR, CCR5) and had perturbations in the expression of memory

Received 21 January 2014 Accepted 5 March 2014

Published ahead of print 12 March 2014

Editor: R. W. Doms

Address correspondence to Catherine A. Blish, cblish@stanford.edu.

* Present address: Catherine A. Blish, Division of Infectious Diseases and Geographic Medicine, Department of Medicine, Stanford University School of Medicine, Stanford, California, USA.

Copyright © 2014, American Society for Microbiology. All Rights Reserved.

doi:10.1128/JVI.00187-14

TABLE 1 Sample timing and demographics for superinfection cases and controls^a

Case no.	SI case ID	SI testing ypi ^b	SI VL (log ₁₀ copies/ml)	SI sex frequency ^c	Cont case ID	Cont testing ypi	Cont VL (log ₁₀ copies/ml)	Cont sex frequency
1	QA013	0.73	5.12	1.4	QA465	0.62	5.05	1.8
					QC370	0.83	5.05	1.7
					QD435	0.82	4.29	1.1
2	QB008	0.83	4.52	1.1	QB099	0.77	4.93	4.3
					QC406	1.0	4.77	0.8
					QD370	0.76	4.22	1.7
3	QA413	2.0	4.95	0.8	QA509	2.1	4.61	1.0
					QC036	2.1	5.30	0.6
					QC890	1.8	4.84	0.2
4	QB045	4.6	4.50	0.3	QA281	4.7	4.98	0.8
					QA692	4.8	5.15	0.9
					QB247	4.7	4.70	0.9
5	QB726	2.8	3.77	1.0	QA874	2.9	4.68	1.6
					QB670	2.8	4.18	1.3
					QC413	2.7	3.38	1.9
6	QB850	0.14	5.22	3.0	QB424	0.086	5.52	2.5
					QB543	0.14	5.52	0.5
					QC805	0.12	5.16	2.0
7	QC885	0.16	5.56	1.4	QB461	0.16	5.58	1.2
					QB765	0.22	5.50	1.0
8	QB609	0.28	2.39	1.3	QB857	0.30	4.17	2.3
					QC048	0.20	5.05	4.7
					QF481	0.26	3.97	0
9	QA252	2.7	4.02	0.8	QC100	2.7	1.69	1.7
					QD470	3.0	4.31	0
					QD834	2.9	4.91	1.6
10	QC858	0.93	4.29	2.9	QA560	0.96	4.07	0.7
					QB936	0.59	5.12	2.4
					QC888	1.07	4.50	2.4

^a Abbreviations: Cont, control; ID, identification number; SI, superinfection; ypi, years postinfection; VL, viral load.

^b Samples were tested at the time point immediately prior to detection of superinfection in order to best assess the status of the immune response at the time point most relevant in terms of exposure to, and lack of protection from, the second virus. Controls were matched to this time postinfection.

^c Data represent averages of self-reported frequencies of sexual intercourse in the preceding week from all follow-up visits up to the time of sample collection. This measure includes both protected and unprotected visits. The median number of visits contributing to this summary was 37 (interquartile range, 20 to 64 visits) over a median of 2 years (interquartile range, 1 to 5 years).

and differentiation markers in comparison to HIV-1-uninfected controls (Fig. 1).

To determine whether higher levels of activation were associated with altered odds of superinfection, exact conditional logistic regression analysis was performed (Table 2). Increased frequencies of both CD3⁺ T cells and, more specifically, CD3⁺ CD4⁺ CCR5⁺ cells, primary targets of HIV-1 infection, were associated with increased odds of superinfection (Table 2). For every 1% increase in the frequency of CD3⁺ CD4⁺ CCR5⁺ HIV-1 target cells among total lymphocytes, the odds of superinfection were 1.69-fold higher (95% confidence interval [CI], 1.02 to 3.36), with a *P* value of 0.04. This elevation in risk was reflected only in the frequency of these cells as a percentage of total lymphocytes, as the percentage of CCR5-expressing CD4⁺ T cells was not associated with an elevated risk of superinfection (odds ratio [OR], 0.99; 95%

CI, 0.94 to 1.05). There was no association between expression of other immune activation markers such as Ki-67, CD38, or HLA-DR among T cells and the odds of superinfection (Table 2). Furthermore, the levels of activation among either CD4⁺ or CD8⁺ cells, as measured by expression of any combination of Ki-67, CD38, and HLA-DR, was not associated with the odds of superinfection (not shown). Finally, differences in the frequencies of regulatory T cells, naive cells, and various memory and effector T cell subsets were not associated with significant alterations in the odds of superinfection (Table 2). Thus, only increases in the frequency of CD3⁺ and CD3⁺ CD4⁺ CCR5⁺ target cells, but not increases in the frequencies of any other activated subsets, were associated with increased odds of superinfection.

These data leverage long-term, regular follow-up of a high-risk population and the setting of superinfection to evaluate,

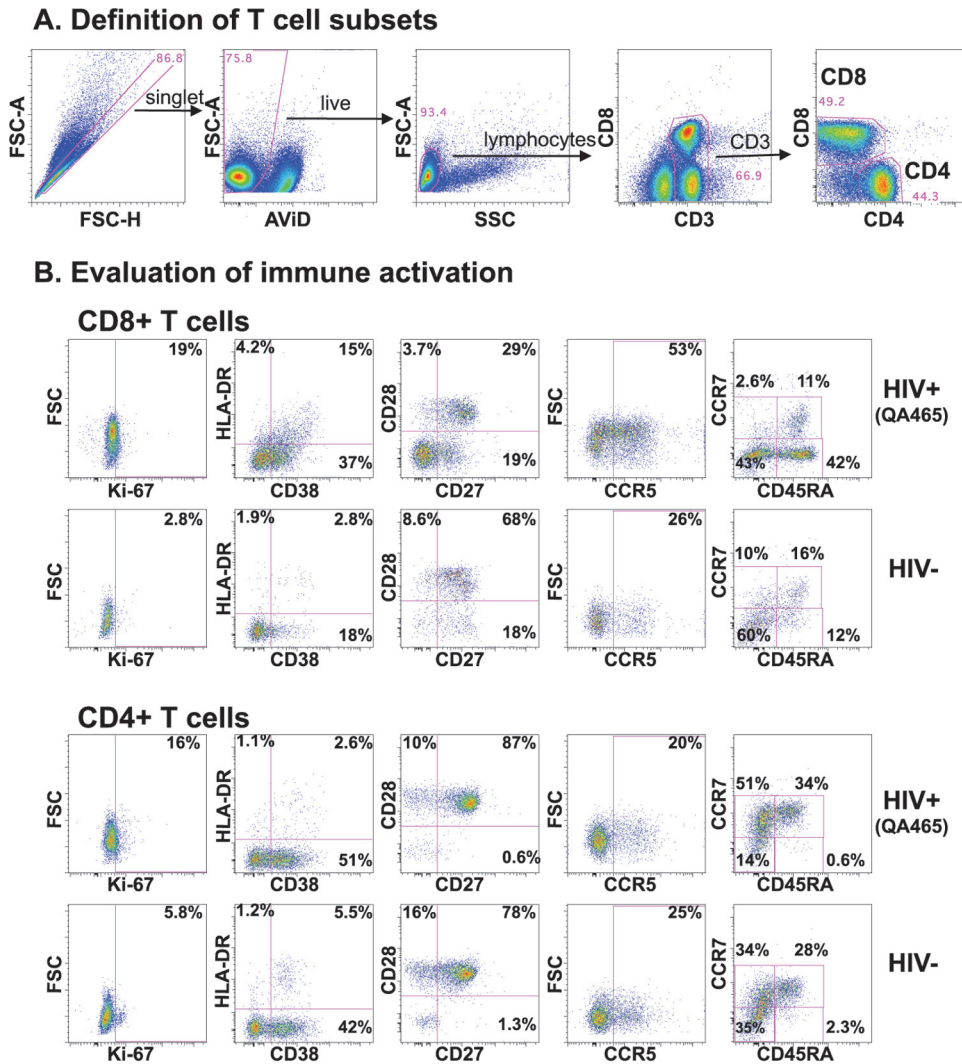


FIG 1 Representative flow cytometry data to assess T cell subsets and immune activation levels from superinfection cases and controls. (A) An example of the sequential gating tree used to identify T cell subsets. FSC, forward scatter; AViD, Live/Dead fixable aqua dead cell stain; SSC, side scatter. (B) Phenotypes of CD8⁺ (top two rows) and CD4⁺ (bottom two rows) T cells based on expression of the activation and memory markers Ki-67, HLA-DR, CD38, CD28, CD27, CCR5, CCR7, and CD45RA are shown. The top panels show the flow cytometry plots from HIV-infected subject QA465, and the bottom panels show the plots from cells from an HIV-uninfected individual that were cryopreserved and evaluated in each experiment in order to set consistent gating. The following antibodies were used: CD195 (CCR5)-phycoerythrin (PE)-Cy7, CD127-Alexa 647, CD25-PE-Cy7, CD28-PE-Cy5, CD38-PE, CD4-fluorescein isothiocyanate (FITC), CD8-peridinin chlorophyll protein (PerCP)-Cy5.5, GranzymeB-Alexa 700, and Ki-67-FITC (BD Biosciences); CD197 (CCR7)-allophycocyanin (APC) (R&D Systems); CD27-APC-eFluor780 and Fox-p3-PE (eBioscience); CD3-Qdot605 (Molecular Probes); CD3-ECD and CD45RA-electron-coupled dye (ECD) (Beckman Coulter); and HLA-DR-Pacific blue (BioLegend).

for the first time, whether the activation and/or differentiation of T cells was associated with HIV acquisition. This is an important consideration, as the contributions of HIV-1-specific immune responses to protection remain uncertain. Some immune responses may even enhance infection risk, particularly if they increase recruitment of HIV-1 target cells to sites of viral entry without containing early replication. Our finding that an increased frequency of HIV-1 primary target cells is associated with increased odds of a second infection may partially explain the observation that the risk of superinfection appears to be highest in the first 6 months following initial infection (44, 45), before target cell depletion.

A strength of this study is the fact that superinfection cases were prospectively identified, allowing us to evaluate immunity in

matched individuals with similar risk behaviors for exposure to HIV-1 but different outcomes. While this remains the largest superinfection cohort available with adequate sampling for such immunologic studies, we still had relatively modest power to detect significant differences between groups and therefore did not adjust for multiple comparisons. With the number of statistical tests, a single significant association may have been observed by chance, and our finding that increased numbers of HIV-1 target cells increase the odds of superinfection warrants follow-up in additional studies. Although the lack of association between other markers of immune activation and the risk of HIV-1 superinfection could be attributed to a lack of statistical power, this seems somewhat unlikely because the majority of observed associations were modest in magnitude. The median absolute estimated odds ratio for one

TABLE 2 Odds of superinfection based on frequency and expression of activation markers and effector and memory CD8⁺ and CD4⁺ subsets

Frequency (%) for indicated member of subset	Odds ratio ^a	95% CI ^b	P value
Total lymphocytes			
CD3 ⁺	1.15	1.01–1.35	0.03
CD3 ⁺ CD8 ⁺	1.03	0.95–1.14	0.48
CD3 ⁺ CD4 ⁺	1.11	0.98–1.27	0.10
CD3 ⁺ CD4 ⁺ CCR5 ⁺	1.67	1.02–3.29	0.04
CD3⁺ T cells			
CD8 ⁺	0.99	0.91–1.06	0.72
CD4 ⁺	1.04	0.96–1.14	0.33
CD4 ⁺ CCR5 ⁺	1.13	0.89–1.45	0.33
Ki-67 ⁺	0.97	0.92–1.01	0.20
CD38 ⁺	0.96	0.89–1.03	0.33
HLA-DR ⁺	1.02	0.90–1.15	0.82
Ki-67 ⁺ CD38 ⁺ HLA-DR ⁺	0.97	0.75–1.24	0.83
CD8⁺ T cells			
CCR5 ⁺	0.99	0.94–1.05	0.85
Ki-67 ⁺	0.98	0.92–1.02	0.34
CD38 ⁺	0.96	0.89–1.02	0.24
HLA-DR ⁺	1.00	0.92–1.09	0.98
CD38 ⁺ HLA-DR ⁺	0.97	0.87–1.08	0.63
Ki-67 ⁺ CD38 ⁺ HLA-DR ⁺	0.97	0.86–1.06	0.53
GranzymeB ⁺	0.98	0.93–1.03	0.48
Naive (CD45RA ⁺ CCR7 ⁺)	1.03	0.87–1.22	0.7
Central memory (CD45RA ⁻ CCR7 ⁺)	1.06	0.88–1.29	0.51
Effector memory (CD45RA ⁻ CCR7 ⁻)	1.02	0.96–1.08	0.56
Terminally differentiated (CD45RA ⁺ CCR7 ⁻)	0.96	0.89–1.03	0.32
CD4⁺ T cells			
CCR5 ⁺	0.99	0.93–1.04	0.83
Ki-67 ⁺	0.97	0.91–1.01	0.18
CD38 ⁺	0.97	0.9–1.04	0.41
HLA-DR ⁺	1.02	0.91–1.13	0.67
CD38 ⁺ HLA-DR ⁺	1.00	0.8–1.22	0.91
Ki-67 ⁺ CD38 ⁺ HLA-DR ⁺	0.99	0.71–1.27	0.96
Treg (foxp3 ⁺ CD25 ⁺ CD127 ⁻)	1.04	0.63–1.67	0.79
Naive (CD45RA ⁺ CCR7 ⁺ CD27 ⁺)	0.99	0.91–1.08	0.91
Transitional memory (CD45RA ⁻ CCR7 ⁻ CD27 ⁺)	1.02	0.92–1.14	0.64
Central memory (CD45RA ⁻ CCR7 ⁺ CD27 ⁺)	1.04	0.96–1.13	0.36
Effector memory (CD45RA ⁻ CCR7 ⁻ CD27 ⁻)	1.00	0.94–1.06	1.00
Terminally differentiated (CD45RA ⁺ CCR7 ⁻ CD27 ⁻)	0.95	0.80–1.09	0.48

^a Data represent odds of superinfection for every 1% increase in the frequency of the indicated cell subset.

^b CI, confidence interval.

unit increase in frequency was 1.03, equivalent in magnitude to the observed association (OR = 0.97) for percent CD38⁺ HLA-DR⁺ of CD8 T cells. Using cluster-level bootstrap sampling of the observed data for percent CD38⁺ HLA-DR⁺ of CD8 T cells, we would need approximately 300 case-control clusters (1,200 subjects with 1:3 case:control matching) to have 80% power to detect this observed association. Another potential limitation is that we were able to measure only reported risk behavior, and background variations in actual exposure to HIV-1 could have attenuated observed relationships between immune responses and HIV-1 acquisition.

This report highlights the complex interplay of different fac-

tors that influence the risk of HIV-1 acquisition in an HIV-1-infected individual. It is likely that both HIV-1-specific immunity and the availability of CD3⁺ CD4⁺ CCR5⁺ HIV-1 primary target cells contribute to the risk of superinfection. Larger studies will be needed to confirm and fully define this dynamic. Here, we show that target cell availability appears to be a more significant contributor to superinfection risk than the generalized immune activation state.

ACKNOWLEDGMENTS

We thank the clinical, laboratory, and administrative staff in Mombasa for their tremendous efforts to recruit and retain the women in the sex worker cohort and for collection and storage of the samples. We also express our gratitude to Stephen De Rosa, Evan Thomas, and Helen Horton for assistance with flow cytometry and to Dara Strauss-Albee and Keshet Ronen for critical comments on the manuscript. We gratefully acknowledge the women who participated in the study.

This work was supported by NIH R37-AI038518 to J.O. and by NIH K08 AI068424 and a CTSA UL1 RR025014 University of Washington Institute of Translational Health Sciences Technology Access Grant to C.A.B. The Mombasa study site infrastructure was supported in part by the University of Washington Center for AIDS Research (P30 AI027757).

We declare that we have no conflicts of interest.

The sponsors of the study had no role in the study design, data collection, data analysis, data interpretation, or in the writing of the report.

REFERENCES

- Giorgi JV, Hultin LE, McKeating JA, Johnson TD, Owens B, Jacobson LP, Shih R, Lewis J, Wiley DJ, Phair JP, Wolinsky SM, Detels R. 1999. Shorter survival in advanced human immunodeficiency virus type 1 infection is more closely associated with T lymphocyte activation than with plasma virus burden or virus chemokine coreceptor usage. *J. Infect. Dis.* 179:859–870. <http://dx.doi.org/10.1086/314660>.
- Lawn SD, Butera ST, Folks TM. 2001. Contribution of immune activation to the pathogenesis and transmission of human immunodeficiency virus type 1 infection. *Clin. Microbiol. Rev.* 14:753–777. <http://dx.doi.org/10.1128/CMR.14.4.753-777.2001>.
- Card CM, Ball TB, Fowke KR. 2013. Immune quiescence: a model of protection against HIV infection. *Retrovirology* 10:141. <http://dx.doi.org/10.1186/1742-4690-10-141>.
- Salkowitz JR, Purvis SF, Meyerson H, Zimmerman P, O'Brien TR, Aledort L, Eyster ME, Hilgartner M, Kessler C, Konkle BA, White GC, II, Goedert JJ, Lederman MM. 2001. Characterization of high-risk HIV-1 seronegative hemophiliacs. *Clin. Immunol.* 98:200–211. <http://dx.doi.org/10.1006/clim.2000.4969>.
- Bégaud E, Chartier L, Marechal V, Ipero J, Léal J, Versmisse P, Breton G, Fontanet A, Capoulade-Metay C, Fleury H, Barré-Sinoussi F, Scott-Algara D, Pancino G. 2006. Reduced CD4 T cell activation and in vitro susceptibility to HIV-1 infection in exposed uninfected Central Africans. *Retrovirology* 3:35. <http://dx.doi.org/10.1186/1742-4690-3-35>.
- Koning FA, Otto SA, Hazenberg MD, Dekker L, Prins M, Miedema F, Schuitemaker H. 2005. Low-level CD4⁺ T cell activation is associated with low susceptibility to HIV-1 infection. *J. Immunol.* 175:6117–6122.
- Chege D, Chai Y, Huibner S, Kain T, Wachihhi C, Kimani M, Barasa S, McKinnon LR, Muriuki FK, Kariri A, Jaoko W, Anzala O, Kimani J, Ball TB, Plummer FA, Kaul R. 2012. Blunted IL17/IL22 and pro-inflammatory cytokine responses in the genital tract and blood of HIV-exposed, seronegative female sex workers in Kenya. *PLoS One* 7:e43670. <http://dx.doi.org/10.1371/journal.pone.0043670>.
- Legrand FA, Nixon DF, Loo CP, Ono E, Chapman JM, Miyamoto M, Diaz RS, Santos AM, Succi RC, Abadi J, Rosenberg MG, De Moraes-Pinto MI, Kallas EG. 2006. Strong HIV-1-specific T cell responses in HIV-1-exposed uninfected infants and neonates revealed after regulatory T cell removal. *PLoS One* 1:e102. <http://dx.doi.org/10.1371/journal.pone.0000102>.
- Card CM, McLaren PJ, Wachihhi C, Kimani J, Plummer FA, Fowke KR. 2009. Decreased immune activation in resistance to HIV-1 infection is associated with an elevated frequency of CD4(+)CD25(+)FOXP3(+)

- regulatory T cells. *J. Infect. Dis.* 199:1318–1322. <http://dx.doi.org/10.1086/597801>.
10. Camara M, Dieye TN, Seydi M, Diallo AA, Fall M, Diaw PA, Sow PS, Mboup S, Kestens L, Jennes W. 2010. Low-level CD4+ T cell activation in HIV-exposed seronegative subjects: influence of gender and condom use. *J. Infect. Dis.* 201:835–842. <http://dx.doi.org/10.1086/651000>.
 11. Lajoie J, Juno J, Burgener A, Rahman S, Mogk K, Wachih C, Mwanjewe J, Plummer FA, Kimani J, Ball TB, Fowke KR. 2012. A distinct cytokine and chemokine profile at the genital mucosa is associated with HIV-1 protection among HIV-exposed seronegative commercial sex workers. *Mucosal Immunol.* 5:277–287. <http://dx.doi.org/10.1038/mi.2012.7>.
 12. McLaren PJ, Ball TB, Wachih C, Jaoko W, Kelvin DJ, Danesh A, Kimani J, Plummer FA, Fowke KR. 2010. HIV-exposed seronegative commercial sex workers show a quiescent phenotype in the CD4+ T cell compartment and reduced expression of HIV-dependent host factors. *J. Infect. Dis.* 202:S339–S344. <http://dx.doi.org/10.1086/655968>.
 13. Naranbhai V, Abdool Karim SS, Altfeld M, Samsunder N, Durgiah R, Sibeko S, Abdool Karim Q, Carr WH, CAPRISA004 TRAPS team. 2012. Innate immune activation enhances HIV acquisition in women, diminishing the effectiveness of tenofovir microbicide gel. *J. Infect. Dis.* 206:993–1001. <http://dx.doi.org/10.1093/infdis/jis465>.
 14. Songok EM, Luo M, Liang B, McLaren P, Kaefer N, Apidi W, Boucher G, Kimani J, Wachih C, Sekaly R, Fowke K, Ball BT, Plummer FA. 2012. Microarray analysis of HIV resistant female sex workers reveal a gene expression signature pattern reminiscent of a lowered immune activation state. *PLoS One* 7:e30048. <http://dx.doi.org/10.1371/journal.pone.0030048>.
 15. Yao X-D, Omenge RW, Henrick BM, Lester RT, Kimani J, Ball TB, Plummer FA, Rosenthal KL. 26 June 2013. Acting locally: innate mucosal immunity in resistance to HIV-1 infection in Kenyan commercial sex workers. *Mucosal Immunol.* <http://dx.doi.org/10.1038/mi.2013.44>.
 16. Jennes W, Evertse D, Borget M-Y, Vuylsteke B, Maurice C, Nkengasong JN, Kestens L. 2006. Suppressed cellular alloimmune responses in HIV-exposed seronegative female sex workers. *Clin. Exp. Immunol.* 143:435–444. <http://dx.doi.org/10.1111/j.1365-2249.2006.03017.x>.
 17. Tran HK, Chartier L, Troung LX, Nguyen NN, Fontanet A, Barré-Sinoussi FE, Pancino G, Scott-Algara D. 2006. Systemic immune activation in HIV-1-exposed uninfected Vietnamese intravascular drug users. *AIDS Res. Hum. Retroviruses* 22:255–261. <http://dx.doi.org/10.1089/aid.2006.22.255>.
 18. Biasin M, Caputo SL, Speciale L, Colombo F, Racioppi L, Zagliani A, Blé C, Vichi F, Cianferoni L, Masci AM, Villa ML, Ferrante P, Mazzotta F, Clerici M. 2000. Mucosal and systemic immune activation is present in human immunodeficiency virus-exposed seronegative women. *J. Infect. Dis.* 182:1365–1374. <http://dx.doi.org/10.1086/315873>.
 19. Restrepo C, Rallón NI, Carrillo J, Soriano V, Blanco J, Benito JM. 2011. Host factors involved in low susceptibility to HIV infection. *AIDS Rev.* 13:30–40.
 20. Suy A, Castro P, Nomdedeu M, García F, López A, Fumero E, Gallart T, Lopalco L, Coll O, Gatell JM, Plana M. 2007. Immunological profile of heterosexual highly HIV-exposed uninfected individuals: predominant role of CD4 and CD8 T-cell activation. *J. Infect. Dis.* 196:1191–1201. <http://dx.doi.org/10.1086/521193>.
 21. Rowland-Jones SL, Dong T, Fowke KR, Kimani J, Krausa P, Newell H, Blanchard T, Ariyoshi K, Oyugi J, Ngugi E, Bwayo J, Macdonald KS, Mcmichael AJ, Plummer FA. 1998. Cytotoxic T cell responses to multiple conserved HIV epitopes in HIV-resistant prostitutes in Nairobi. *J. Clin. Invest.* 102:1758–1765. <http://dx.doi.org/10.1172/JCI4314>.
 22. Fowke KR, Kaul R, Rosenthal KL, Oyugi J, Kimani J, Rutherford WJ, Nagelkerke NJ, Ball TB, Bwayo JJ, Simonsen JN, Shearer GM, Plummer FA. 2000. HIV-1-specific cellular immune responses among HIV-1-resistant sex workers. *Immunol. Cell Biol.* 78:586–595. <http://dx.doi.org/10.1046/j.1440-1711.2000.00944.x>.
 23. Lo Caputo S, Trabattini D, Vichi F, Piconi S, Lopalco L, Villa ML, Mazzotta F, Clerici M. 2003. Mucosal and systemic HIV-1-specific immunity in HIV-1-exposed but uninfected heterosexual men. *AIDS* 17:531–539. <http://dx.doi.org/10.1097/00002030-200303070-00008>.
 24. Pattacini L, Murnane PM, Kahle EM, Bolton MJ, Delrow JJ, Lingappa JR, Katabira E, Donnell D, McElrath MJ, Baeten JM, Lund JM. 2013. Differential regulatory T cell activity in HIV type 1-exposed seronegative individuals. *AIDS Res. Hum. Retroviruses* 29:1321–1329. <http://dx.doi.org/10.1089/aid.2013.0075>.
 25. Carrillo J, Restrepo C, Rallón NI, Massanella M, del Romero J, Rodríguez C, Soriano V, Clotet B, Benito JM, Blanco J. 2013. HIV exposed seronegative individuals show antibodies specifically recognizing native HIV envelope glycoprotein. *AIDS* 27:1375–1385. <http://dx.doi.org/10.1097/QAD.0b013e32835fac08>.
 26. Restrepo C, Rallón NI, del Romero J, Rodríguez C, Hernando V, López M, Peris A, Lozano S, Sempere-Ortells JM, Soriano V, Benito JM. 2010. Low-level exposure to HIV induces virus-specific T cell responses and immune activation in exposed HIV-seronegative individuals. *J. Immunol.* 185:982–989. <http://dx.doi.org/10.4049/jimmunol.1000221>.
 27. Buchbinder SP, Mehrotra DV, Duerr A, Fitzgerald DW, Mogg R, Li D, Gilbert PB, Lama JR, Marmor M, Rio CD, McElrath MJ, Casimiro DR, Gottesdiener KM, Chodakewitz JA, Corey L, Robertson MN. 2008. Efficacy assessment of a cell-mediated immunity HIV-1 vaccine (the Step Study): a double-blind, randomised, placebo-controlled, test-of-concept trial. *Lancet* 372:1881–1893. [http://dx.doi.org/10.1016/S0140-6736\(08\)61591-3](http://dx.doi.org/10.1016/S0140-6736(08)61591-3).
 28. McElrath MJ, De Rosa SC, Moodie Z, Dubey S, Kierstead L, Janes H, Defawe OD, Carter DK, Hural J, Akondy R, Buchbinder SP, Robertson MN, Mehrotra DV, Self SG, Corey L, Shiver JW, Casimiro DR. 2008. HIV-1 vaccine-induced immunity in the test-of-concept Step Study: a case-cohort analysis. *Lancet* 372:1894–1905. [http://dx.doi.org/10.1016/S0140-6736\(08\)61592-5](http://dx.doi.org/10.1016/S0140-6736(08)61592-5).
 29. Haynes BF, Gilbert PB, McElrath MJ, Zolla-Pazner S, Tomaras GD, Alam SM, Evans DT, Montefiori DC, Karnasuta C, Sutthent R, Liao H-X, DeVico AL, Lewis GK, Williams C, Pinter A, Fong Y, Janes H, Decamp A, Huang Y, Rao M, Billings E, Karasavvas N, Robb ML, Ngauy V, de Souza MS, Paris R, Ferrari G, Bailer RT, Soderberg KA, Andrews C, Berman PW, Frahm N, De Rosa SC, Alpert MD, Yates NL, Shen X, Koup RA, Pitisuttithum P, Kaewkungwal J, Nitayaphan S, Rerks-Ngarm S, Michael NL, Kim JH. 2012. Immune-correlates analysis of an HIV-1 vaccine efficacy trial. *N. Engl. J. Med.* 366:1275–1286. <http://dx.doi.org/10.1056/NEJMoa1113425>.
 30. Chohan B, Lavreys L, Rainwater SMJ, Overbaugh J. 2005. Evidence for frequent reinfection with human immunodeficiency virus type 1 of a different subtype. *J. Virol.* 79:10701–10708. <http://dx.doi.org/10.1128/JVI.79.16.10701-10708.2005>.
 31. Piantadosi A, Chohan B, Chohan V, McClelland RS, Overbaugh J. 2007. Chronic HIV-1 infection frequently fails to protect against superinfection. *PLoS Pathog.* 3:e177. <http://dx.doi.org/10.1371/journal.ppat.0030177>.
 32. Piantadosi A, Ngayo MO, Chohan B, Overbaugh J. 2008. Examination of a second region of the HIV type 1 genome reveals additional cases of superinfection. *AIDS Res. Hum. Retroviruses* 24:1221. <http://dx.doi.org/10.1089/aid.2008.0100>.
 33. Blish CA, Dogan OC, Derby NR, Nguyen M-A, Chohan B, Richardson BA, Overbaugh J. 2008. Human immunodeficiency virus type 1 superinfection occurs despite relatively robust neutralizing antibody responses. *J. Virol.* 82:12094–12103. <http://dx.doi.org/10.1128/JVI.01730-08>.
 34. Blish CA, Dogan OC, Jaoko W, McClelland RS, Mandaliya KK, Odeh-Davis KSK, Richardson BA, Overbaugh J. 2012. Cellular immune responses and susceptibility to HIV-1 superinfection: a case-control study. *AIDS* 26:643–646. <http://dx.doi.org/10.1097/QAD.0b013e3283509a0b>.
 35. Forthal DN, Landucci G, Chohan B, Richardson BA, McClelland RS, Jaoko W, Blish C, Overbaugh J. 2013. Antibody-dependent cell-mediated virus inhibition activity does not correlate with risk of HIV-1 superinfection. *J. Acquir. Immune Defic. Syndr.* 63:31–33. <http://dx.doi.org/10.1097/QAI.0b013e3182874d41>.
 36. Smith DM, Strain MC, Frost SDW, Pillai SK, Wong JK, Wrin T, Liu Y, Petropoulos CJ, Daar ES, Little SJ, Richman DD. 2006. Lack of neutralizing antibody response to HIV-1 predisposes to superinfection. *Virology* 355:1–5. <http://dx.doi.org/10.1016/j.viro.2006.08.009>.
 37. Chaillan G, Wagner GA, Hepler NL, Little SJ, Kosakovsky Pond SL, Caballero G, Pacold ME, Phung P, Wrin T, Richman DD, Wertheim JO, Smith DM. 2013. Dynamics of viral evolution and neutralizing antibody response after HIV-1 superinfection. *J. Virol.* 87:12737–12744. <http://dx.doi.org/10.1128/JVI.02260-13>.
 38. Basu D, Kraft CS, Murphy MK, Campbell PJ, Yu T, Hrabner PT, Irene C, Pinter A, Chomba E, Mulenga J, Kilembe W, Allen SA, Derdeyn CA, Hunter E. 2012. HIV-1 subtype C superinfected individuals mount low autologous neutralizing antibody responses prior to intra-subtype superinfection. *Retrovirology* 9:76. <http://dx.doi.org/10.1186/1742-4690-9-76>.
 39. Altfeld M, Allen TM, Yu XG, Johnston MN, Agrawal D, Korber BT,

- Montefiori DC, O'Connor DH, Davis BT, Lee PK, Maier EL, Harlow J, Goulder PJR, Brander C, Rosenberg ES, Walker BD. 2002. HIV-1 superinfection despite broad CD8+ T-cell responses containing replication of the primary virus. *Nature* 420:434–439. <http://dx.doi.org/10.1038/nature01200>.
40. Appay V, Dunbar PR, Callan M, Klenerman P, Gillespie GMA, Papagno L, Ogg GS, King A, Lechner F, Spina CA, Little S, Havlir DV, Richman DD, Gruener N, Pape G, Waters A, Easterbrook P, Salio M, Cerundolo V, McMichael AJ, Rowland-Jones SL. 2002. Memory CD8+ T cells vary in differentiation phenotype in different persistent virus infections. *Nat. Med.* 8:379–385. <http://dx.doi.org/10.1038/nm0402-379>.
 41. Deeks SG. 2004. Immune activation set point during early HIV infection predicts subsequent CD4+ T-cell changes independent of viral load. *Blood* 104:942–947. <http://dx.doi.org/10.1182/blood-2003-09-3333>.
 42. Giorgi JV, Lyles RH, Matud JL, Yamashita TE, Mellors JW, Hultin LE, Jamieson BD, Margolick JB, Rinaldo CR, Phair JP, Detels R, Detels R, Multicenter AIDS Cohort Study. 2002. Predictive value of immunologic and virologic markers after long or short duration of HIV-1 infection. *J. Acquir. Immune Defic. Syndr.* 29:346–355.
 43. Liu ZZ, Cumberland WG, Hultin LEL, Kaplan AHA, Detels RR, Giorgi JV. 1998. CD8+ T-lymphocyte activation in HIV-1 disease reflects an aspect of pathogenesis distinct from viral burden and immunodeficiency. *J. Acquir. Immune Defic. Syndr. Hum. Retrovirol.* 18:332–340. <http://dx.doi.org/10.1097/00042560-199808010-00004>.
 44. Ronen K, McCoy CO, Matsen FA, Boyd DE, Emery S, Odem-Davis K, Jaoko W, Mandaliya K, McClelland RS, Richardson BA, Overbaugh J. 2013. HIV-1 superinfection occurs less frequently than initial infection in a cohort of high-risk Kenyan women. *PLoS Pathog.* 9:e1003593. <http://dx.doi.org/10.1371/journal.ppat.1003593>.
 45. Redd AD, Quinn TC, Tobian AA. 2013. Frequency and implications of HIV superinfection. *Lancet Infect. Dis.* 13:622–628. [http://dx.doi.org/10.1016/S1473-3099\(13\)70066-5](http://dx.doi.org/10.1016/S1473-3099(13)70066-5).