

# NIH Public Access

**Author Manuscript**

*J Clin Virol*. Author manuscript; available in PMC 2015 June 01.

#### Published in final edited form as:

*J Clin Virol*. 2014 June ; 60(2): 127–132. doi:10.1016/j.jcv.2014.03.002.

# **Reduced Human Herpesvirus-8 Oropharyngeal Shedding Associated with Protease Inhibitor-Based Antiretroviral Therapy**

**Soren Gantt**1,6,7,\* , **Ashok Cattamanchi**2,\* , **Elizabeth Krantz**7, **Amalia Magaret**3,7, **Stacy Selke**3, **Steven R. Kuntz**3, **Meei-Li Huang**3, **Lawrence Corey**2,3,7,8,9, **Anna Wald**2,3,4,7, and **Corey Casper**2,3,4,5,7,8,9

<sup>1</sup>Department of Pediatrics, Health, University of Washington

<sup>2</sup>Department of Medicine, Health, University of Washington

<sup>3</sup>Department of Laboratory Medicine, Health, University of Washington

<sup>4</sup>Department of Epidemiology, Health, University of Washington

<sup>5</sup>Department of Global, Health, University of Washington

<sup>6</sup>Seattle Children's Hospital, Fred Hutchinson Cancer Research Center, Seattle, Washington, USA

<sup>7</sup>Vaccine and Infectious Disease, Fred Hutchinson Cancer Research Center, Seattle, Washington, USA

<sup>8</sup>Public Health Sciences, Fred Hutchinson Cancer Research Center, Seattle, Washington, USA

<sup>9</sup>Clinical Research Divisions, Fred Hutchinson Cancer Research Center, Seattle, Washington, USA

# **Abstract**

**Background—**Human herpesvirus 8 (HHV-8) replication increases the risk of Kaposi sarcoma (KS). Highly-active antiretroviral therapy (HAART) reduces the incidence of KS, and regimens that contain protease inhibitors (PIs) may be particularly effective.

**Objective—**To determine whether PI-based HAART regimens may more effectively inhibit HHV-8 shedding compared to regimens without PIs.

<sup>© 2014</sup> Elsevier B.V. All rights reserved.

Correspondence to: Dr. Soren Gantt, Child & Family Research Institute, 950 West 28th Avenue, Room A5-144, Vancouver BC V5Z 4H4, Canada, sgantt@cfri.ubc.ca, Tel: 604-875-2151, Fax: 604-875-2226.

<sup>&</sup>lt;sup>®</sup>Present Affiliations: Soren Gantt, Department of Pediatrics, University of British Columbia, Vancouver, BC, Canada; Ashok Cattamanchi, Inova Fairfax Hospital, Fairfax, Virginia, USA.

**Competing Interests**: None declared.

**Ethical Approval**: Written informed consent was obtained in accordance with a protocol approved by the University of Washington Human Subjects Division.

**Publisher's Disclaimer:** This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final citable form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

**Study design—**Prospective, observational study of 142 HIV-1 and HHV-8 co-infected men conducted in Seattle, Washington. Quantitative HHV-8 PCR testing was performed on daily swabs of the oropharynx, the primary site of HHV-8 replication. Associations between antiretroviral regimen and detection of HHV-8 DNA in swabs were evaluated using generalized estimating equations.

**Results—**HHV-8 DNA was detected in 3,016 (26%) of 11,608 specimens collected. PI-based HAART was associated with a statistically significantly lower frequency of detection (RR 0.2; 95% CI 0.1 to 0.5) compared to ART-naïve persons, whereas HAART without a PI was not (RR 0.7; 95% CI 0.4 to 1.3). Compared to ART-naïve persons, there was also a trend toward lower quantities of HHV-8 detected during treatment with HAART regimens that contained a PI. These associations between PIs and measures of HHV-8 shedding could not be attributed to use of nelfinavir, which inhibits HHV-8 replication *in vitro*, and were independent of CD4 count and HIV plasma viral load (VL).

**Conclusions—**HAART regimens that contain PIs appear to decrease HHV-8 shedding compared to NNRTIs. Further study of PI-based HAART is warranted to determine the optimal regimens for prevention and treatment of KS.

#### **Keywords**

Human herpesvirus 8; Kaposi sarcoma; antiretroviral therapy; protease inhibitor

# **Background**

Kaposi sarcoma (KS) is an AIDS-defining malignancy caused by infection with human herpesvirus 8 (HHV-8). The rising incidence of KS in the United States leveled off in 1987 shortly after approval of zidovudine for antiretroviral therapy (ART), and decreased further after the widespread use of combination "highly active" antiretroviral therapy (HAART; 3 or more drugs with at least one protease inhibitor (PI) or non-nucleoside reverse transcriptase inhibitor (NNRTI)) in 1995.<sup>1</sup> Nevertheless, KS remains the most common malignancy among HIV-infected people worldwide, and incident cases develop even among patients on HAART whose HIV infection is well controlled.<sup>2-4</sup> Furthermore, although HAART is beneficial for the treatment of AIDS-KS, nearly half of patients will have persistent disease despite receiving the current standard of care.5-9 As such, better KS prevention and treatment strategies are needed.

The mechanisms by which ART affects KS have not been fully defined. HHV-8 replication is a strong risk factor for the development of  $KS$ ,  $^{10-16}$  and the use of ART is associated with reductions of HHV-8 levels in blood.17-19 The immune reconstitution that accompanies effective ART likely improves immune control of HHV-8 replication and tumor surveillance.<sup>20, 21</sup> Additionally, ART may interfere with KS progression by reducing the levels of the HIV-1 Tat protein, which has angiogenic and tumorigenic functions<sup>22</sup> and promotes replication of HHV-8 *in vitro*. 23

Several observational studies have suggested that PI-based HAART may be superior to NNRTI-based regimens for the treatment of prevention of KS, independently of effects on HIV plasma viral load (VL) or CD4 count.<sup>15, 24-26</sup> However, this has not been found in all

 $\text{cohorts}^{27-29}$  and data from controlled trials with adequate power to address the question are not currently available.<sup>30</sup> Among their many cellular effects, various PIs display antiangiogenic and anti-tumor properties that may impair the growth and persistence of KS lesions.31-33 Furthermore, some antiretroviral drugs may have direct effects on HHV-8 replication. Among PIs, nelfinavir appeared to preferentially inhibit production of infectious HHV-8 *in vitro* at concentrations achieved in plasma with routine oral dosing.34 Though an effect on HHV-8 replication by nucleoside reverse transcriptase inhibitors (NRTIs) has not been demonstrated, the HHV-8 thymidine kinase is capable of phosphorylating both zidovudine and stavudine.<sup>35, 36</sup> As such, specific antiretroviral regimens may have activity against HHV-8 that could confer clinically important effects.

Men co-infected with HIV and HHV-8 frequently shed HHV-8 DNA in saliva, and daily collection of oropharyngeal swabs offers a detailed portrait of HHV-8 oropharyngeal replication.37 Additionally, ART use is associated with a significantly reduced risk of HHV-8 oropharyngeal shedding.38 We therefore examined HHV-8 shedding among HIV/ HHV-8 co-infected men to determine whether the type of ART regimen or use of PIs affects HHV-8 oropharyngeal replication.

# **Study Design**

#### **Study Participants**

Men in Seattle, Washington were recruited from outpatient clinics and advertisements in the community for participation in studies of the epidemiology of human herpesviruses between 1993 and 2009. All participants in one or more of these studies were included in the analyses described here if they met the inclusion criteria of: 1) a positive HIV-1 serology test, and 2) a positive test for HHV-8 infection by either serology or PCR. Participants were not assigned ART by study investigators, but rather were asked to record ART regimens prescribed by their HIV care providers.

#### **Specimen and Data Collection**

Oropharyngeal sampling was performed by participants, by swabbing the buccal, lingual, palatine and tonsillar mucosa in a standardized fashion using a Dacron swab, as previously described.39, 40 Swabs were collected during "sessions"; each session consisted of a period of consecutive days on which oral swab collection was performed. Some men participated in more than one session. The shedding rate was computed as the number of swabs in which HHV-8 DNA was detected by PCR divided by the number of swabs collected for each session. Blood was collected at the beginning of each session for measurement of HIV-1 plasma RNA and CD4 T cell counts.

#### **Laboratory Testing**

Whole virus enzyme immunoassay (EIA) or immunofluorescence assay (IFA) was used to detect serum antibodies to HHV-8 as previously described.<sup>41</sup> DNA was extracted from oral swabs HHV-8 DNA was measured quantitatively with a real-time fluorescent polymerase chain reaction (PCR) with primers to the *orf73* gene, with positive and negative controls as previously described.<sup>42, 43</sup> Oral swabs with  $\,$  150 copies were considered positive for

HHV-8.40 CD4 T cell counts were measured with flow cytometry and HIV-1 plasma RNA was quantified using the AMPLICOR Monitor HIV-1 Test (Roche, Alameda, CA).

#### **Statistical Analysis**

Participant characteristics, HHV-8 oropharyngeal detection patterns, and ART use were reviewed using descriptive statistics. The distribution of copies of HHV-8 DNA was highly skewed and thus  $log_{10}$  –transformed prior to analyses. Correlates of HHV-8 shedding frequency were examined using generalized estimating equation (GEE) models with a Poisson link and robust standard errors to account for overdispersion, and correlation among multiple sessions belonging to the same participant.<sup>44</sup> Analyses of the quantity of HHV-8 detected among sessions with at least one day with HHV-8 detected were performed using GEE models for normal outcomes.44 HAART was defined as at least a three-drug regimen that included either a PI or a non-nucleoside reverse transcriptase inhibitor (NNRTI). For models, ART use was categorized using the following 6 categories: 1) ART-naïve; 2) No current ART but previous ART use; 3) current non-HAART ART; 4) HAART with no PI (i.e., only NNRTI-based HAART); 5) current HAART containing any PI; and 6) current HAART containing the PI nelfinavir. Stata version 8.2 (College Station, Texas) and SAS version 8.2 (SAS Institute) statistical software was used for all analyses.

# **Results**

#### **Study participants**

142 HIV-1 and HHV-8 co-infected men participated in the study; 115 (81%) reported their race as white (Table 1). The median time since HIV diagnosis was 10 years (range 0 - 24 years). The median CD4 count at first session was 385 cells/uL, with a wide range (1 to 1240). Of 132 participants with valid measures at first session, 61 (46%), 24 (18%) and 47  $(36\%)$  had a HIV VL of <50, 50 to 10,000, and >10,000 copies/mL, respectively.

#### **Oropharyngeal swabs**

The 142 participants collected a total of 11,608 oral swabs for HHV-8 DNA quantification (Table 2). A median of 40 daily swabs (range 14 - 129) was obtained during 262 sessions, and 74 (52%) participants collected more than one session over time for a total of 262 sessions. Among those with multiple sessions, the median time between sessions was 6 months (range <1 month to 29 months).

#### **Antiretroviral therapy use**

A wide variety of ART use was observed among study participants (Table 2). While 128 participants (90%) either used the same combination of ART classes for all sessions or were not taking ART during all sessions, 14 participants changed ART regimen classifications between sessions and thus appear in multiple categories. Forty-six participants collected oropharyngeal swabs (3,111 (27%) of 11,608 total swabs) at a time when they were naïve to ART, 20 participants (1,272 (11%) of swabs) were not taking ART at the time of swab collection but had previously taken ART, 87 participants (6,564 (57%) of swabs) were taking HAART, and 9 (661 (6%) of swabs) were taking ART that did not meet the definition of HAART.

Of persons on HAART, 32 participants (2,216 (34%) of 6,564 swabs) were taking a NNRTIbased regimen without a PI, and 55 (4,348 (66%) of swabs) were taking a PI-based regimen, of whom 9 (587 (14%) of 4,348 swabs) were taking a regimen that included both a NNRTI and a PI. NRTI combinations included zidovudine/lamivudine, stavudine/lamivudine, didanosine/lamivudine and abacavir/lamivudine. The most commonly used PI was nelfinavir (1,139 (26%) of 4,348 swabs). Other PIs used were indinavir, saquinavir, ritonavir, ritonavir-boosted lopinavir, amprenavir, fosamprenavir, and atazanavir.

#### **HHV-8 oropharyngeal shedding frequency and ART use**

HHV-8 DNA was detected from the oropharynx in 3,016 (26%) of 11,608 swabs collected (Table 2). The frequency of HHV-8 shedding was 18.9% (1243 of 6,564 swabs) among participants using HAART and 39.6% in those who were ART-naïve. The lowest rates of shedding were observed persons taking PI-containing HAART (13.2%, or 572 of 4348 swabs). Participants receiving regimens specifically containing the PI nelfinavir also showed a relatively low rate of shedding (19.1%, or 217 of 1,139 swabs).

PI-based HAART was associated with a 80% lower rate of shedding (RR 0.2; 95% CI 0.1 to 0.5) in univariate analysis, whereas previous ART, non-HAART ART, or HAART without a PI did not significantly decrease shedding frequency compared to ART-naïve persons (Table 3). Nelfinavir-based HAART was not significantly associated with a reduction in HHV-8 shedding frequency (RR 0.7; 95% CI 0.2 to 1.9). HHV-8 shedding frequency was not significantly associated with low CD4 T cell count, high HIV VL, time since HIV diagnosis, or year of participation, but was greatly increased by the presence of KS (RR 2.5; 95% CI 1.5 to 4.1). The associations between ART regimens and HHV-8 shedding frequency were not markedly changed by adjusting for KS status in multivariate analysis (Table 3).

#### **HHV-8 oropharyngeal shedding quantity and ART use**

The mean quantity of HHV-8 detected among subjects receiving different ART regimens in shown in Table 2. Use of HAART with or without a PI showed a non-significant trend toward lower quantity with PI use (Table 4). Previous use of ART was associated with a mean decrease of 0.7 log copies (95% CI -1.3 to 0.0 log) and current non-HAART ART was associated with a mean increase of 0.3 log copies (95% CI 0.0 to 0.6 log) in the quantity of HHV-8 detected compared to ART-naïve persons in multivariate analysis. Compared to those participants without KS, the presence of KS was also associated with a mean increase of 0.9 log copies of HHV-8 (95% CI 0.2 to 1.6 log) in multivariate analysis. No clear trend was observed between HIV VL and the quantity of HHV-8 detected in oropharyngeal swabs. Those participants with a HIV VL between 500 and 10,000 copies/ml had a mean quantity of HHV-8 that was 0.6 log higher (95% CI 0.1 to 1.0 logs) compared to persons with HIV VL <500 copies/mL, while the mean quantity of HHV-8 among those with >100,000 copies of HIV/ml tended to be lower (coefficient -0.8; 95% CI -2.0 to 0.3 log) compared to those with HIV VL <50 copies/mL. Similarly, a direct association was observed between quantity of HHV-8 and CD4 count, in which the mean copy number was 0.4 log lower (95% CI -0.6 to -0.2 log) among those participants with a CD4 count <200, compared to those with a CD4 count 200. Neither time since HIV diagnosis nor the year of participation was associated with the HHV-8 quantity detected in the oropharynx.

# **Discussion**

In our study that evaluated the effect of different ART regimens on HHV-8 shedding, in a cohort of 142 men with HIV-1 and HHV-8 co-infection who had daily sampling for quantitative HHV-8 PCR, on >11,000 days, PI-based HAART was associated with a significantly lower frequency of oral viral shedding compared to ART-naïve persons. In contrast, ART that did not contain a PI did not appear to decrease shedding frequency. A trend for reduced HHV-8 shedding quantity among PI-based HAART users was also observed. This study is limited by its observational design and inability to evaluate the effects of ART combinations and other risk factors in greater detail. For example, the duration of the ART regimens used and the level of medication adherence were unknown, but could affect HHV-8 shedding. A trend toward less frequent HHV-8 detection as well as significantly reduced HHV-8 quantity was observed among subjects with previous ART use but who were not on treatment during the study, but it is unknown what regimens they had received or when they were discontinued. Choice of drug regimen is influenced by clinical factors, potentially leading to confounding by indication, where the observed relationship between HHV-8 shedding patterns and antiretroviral use is related not to the use of a specific ART regimen but rather to its indication.<sup>45</sup> For example, if clinicians prescribed PIbased therapy to individuals with a common characteristic (previous resistance to NNRTIs, significant comorbidities, etc.), then the observed relationship between PI use and lower HHV-8 shedding could instead be attributable to the factor influencing the choice of ART. We were also limited in our ability to evaluate the effect of any individual antiretroviral agents on HHV-8 shedding due to the large number of drug combinations used. Finally, although HHV-8 shedding reflects viral replication in the oropharynx and appears to occur prior to systemic replication,  $40, 46$  our study did not have the ability to evaluate other measures of HHV-8 replication such as viremia.

Prevention of KS is an important goal given that KS resolution rates range between only 44%-60% despite HAART and chemotherapy treatment.5-9 The optimal strategy to prevent KS is not clear, but there is evidence that suppression of HHV-8 replication may be highly advantageous.30, 47 Remarkably, *in vitro* studies have shown that some PIs decrease inflammatory cytokine production implicated in HHV-8 reactivation<sup>48, 49</sup> and one, nelfinavir, inhibits HHV-8 replication directly.<sup>34</sup> An inhibitory effect of nelfinavir on HHV-8 shedding was not observed in this cohort. While this may indicate that nelfinavir is inactive against HHV-8 replication *in vivo*, it is also possible that small numbers and the presence of residual confounding in the analyses obscured finding an effect. Alternatively, measurement of HHV-8 DNA alone may not accurately reflect the antiviral activity of nelfinavir; although the drug blocks generation of infectious HHV-8 virions *in vitro*, it appears to act late in virus production after DNA replication and may actually induce reactivation of latent virus.50 These findings together with the *in vivo* results reported here lend additional support to the hypothesis that PIs may have anti-HHV-8 activities that impede progression to KS. On the other hand, observational cohort studies of KS development in Western countries have reported similar risk reduction with PI- and NNRTIbased HAART.27, 51 This may be explained in part by the varying anti-tumor and anti-

HHV-8 activities of different  $PIs$ ,  $32, 48, 49$  such that when lumped together the beneficial effects of some agents could be obscured.

Whether PI-based HAART should be preferentially used for treatment of KS is a matter of debate. Observational studies of the effect of HAART type on KS response have yielded variable conclusions.7, 24, 27, 28, 49, 52-54 One uncontrolled trial of indinavir suggested a clinical benefit for classic (HIV-negative) KS.<sup>55</sup> Indinavir, ritonavir, lopinavir and other PIs have anti- angiogenic and anti-tumor properties, though they appear to be less potent than nelfinavir and lack anti-HHV-8 activity *in vitro*. 33, 34 As such, KS treatment and prevention trials should be considered to specifically evaluate nelfinavir or other agents with inhibitory activity against HHV-8.30, 47

In summary, our data suggest that PI-based HAART may suppress HHV-8 infection more effectively than regimens without PIs, and may therefore confer particular benefits for individuals at high-risk for KS development and disease progression. Controlled trials are needed to definitively determine whether specific PI-based regimens differentially suppress HHV-8 oropharyngeal replication or have other beneficial effects for the prevention and treatment of KS, particularly in areas with a high burden of KS.

# **Acknowledgments**

The authors sincerely thank the study participants and staff of the Virology Research Clinic.

**Funding**: National Institutes of Health R01 CA138165, K24 AI-071113, P01 AI030731 KL2 RR025015, and P30 AI027757.

# **References**

- 1. Eltom MA, Jemal A, Mbulaiteye SM, Devesa SS, Biggar RJ. Trends in Kaposi's sarcoma and non-Hodgkin's lymphoma incidence in the United States from 1973 through 1998. J Natl Cancer Inst. 2002; 94:1204–10. [PubMed: 12189223]
- 2. Maurer T, Ponte M, Leslie K. HIV-associated Kaposi's sarcoma with a high CD4 count and a low viral load. N Engl J Med. 2007; 357:1352–3. [PubMed: 17898112]
- 3. Krown SE, Lee JY, Dittmer DP. More on HIV-associated Kaposi's sarcoma. N Engl J Med. 2008; 358:535–6. author reply 36. [PubMed: 18234764]
- 4. Casper C. The increasing burden of HIV-associated malignancies in resource-limited regions. Annu Rev Med. 2011; 62:157–70. [PubMed: 20868276]
- 5. Dupont C, Vasseur E, Beauchet A, Aegerter P, Berthe H, de Truchis P, et al. Long-term efficacy on Kaposi's sarcoma of highly active antiretroviral therapy in a cohort of HIV-positive patients. CISIH 92. Centre d'information et de soins de l'immunodeficience humaine. Aids. 2000; 14:987–93. [PubMed: 10853980]
- 6. Dupin N, Rubin De Cervens V, Gorin I, Calvez V, Pessis E, Grandadam M, et al. The influence of highly active antiretroviral therapy on AIDS-associated Kaposi's sarcoma. Br J Dermatol. 1999; 140:875–81. [PubMed: 10354025]
- 7. Lebbe C, Blum L, Pellet C, Blanchard G, Verola O, Morel P, et al. Clinical and biological impact of antiretroviral therapy with protease inhibitors on HIV-related Kaposi's sarcoma. Aids. 1998; 12:F45–9. [PubMed: 9619797]
- 8. Bihl F, Mosam A, Henry LN, Chisholm JV 3rd, Dollard S, Gumbi P, et al. Kaposi's sarcomaassociated herpesvirus-specific immune reconstitution and antiviral effect of combined HAART/ chemotherapy in HIV clade C-infected individuals with Kaposi's sarcoma. Aids. 2007; 21:1245–52. [PubMed: 17545700]

- 9. Nguyen HQ, Magaret AS, Kitahata MM, Van Rompaey SE, Wald A, Casper C. Persistent Kaposi sarcoma in the era of highly active antiretroviral therapy: characterizing the predictors of clinical response. Aids. 2008; 22:937–45. [PubMed: 18453853]
- 10. Whitby D, Howard MR, Tenant-Flowers M, Brink NS, Copas A, Boshoff C, et al. Detection of Kaposi sarcoma associated herpesvirus in peripheral blood of HIV-infected individuals and progression to Kaposi's sarcoma. Lancet. 1995; 346:799–802. [PubMed: 7674745]
- 11. Lorenzen T, Albrecht D, Paech V, Meyer T, Hoffmann C, Stoehr A, et al. HHV-8 DNA in blood and the development of HIV-associated Kaposi's sarcoma in the era of HAART--a prospective evaluation. Eur J Med Res. 2002; 7:283–6. [PubMed: 12117665]
- 12. Campbell TB, Borok M, Gwanzura L, MaWhinney S, White IE, Ndemera B, et al. Relationship of human herpesvirus 8 peripheral blood virus load and Kaposi's sarcoma clinical stage. Aids. 2000; 14:2109–16. [PubMed: 11061651]
- 13. Cannon MJ, Dollard SC, Black JB, Edlin BR, Hannah C, Hogan SE, et al. Risk factors for Kaposi's sarcoma in men seropositive for both human herpesvirus 8 and human immunodeficiency virus. Aids. 2003; 17:215–22. [PubMed: 12545082]
- 14. Broccolo F, Bossolasco S, Careddu AM, Tambussi G, Lazzarin A, Cinque P. Detection of DNA of lymphotropic herpesviruses in plasma of human immunodeficiency virus-infected patients: frequency and clinical significance. Clin Diagn Lab Immunol. 2002; 9:1222–8. [PubMed: 12414753]
- 15. Gill J, Bourboulia D, Wilkinson J, Hayes P, Cope A, Marcelin AG, et al. Prospective study of the effects of antiretroviral therapy on Kaposi sarcoma--associated herpesvirus infection in patients with and without Kaposi sarcoma. J Acquir Immune Defic Syndr. 2002; 31:384–90. [PubMed: 12447008]
- 16. Engels EA, Biggar RJ, Marshall VA, Walters MA, Gamache CJ, Whitby D, et al. Detection and quantification of Kaposi's sarcoma-associated herpesvirus to predict AIDS-associated Kaposi's sarcoma. Aids. 2003; 17:1847–51. [PubMed: 12891072]
- 17. Pellet C, Chevret S, Blum L, Gauville C, Hurault M, Blanchard G, et al. Virologic and immunologic parameters that predict clinical response of AIDS-associated Kaposi's sarcoma to highly active antiretroviral therapy. J Invest Dermatol. 2001; 117:858-63. [PubMed: 11676823]
- 18. Leao JC, Kumar N, McLean KA, Porter SR, Scully CM, Swan AV, et al. Effect of human immunodeficiency virus-1 protease inhibitors on the clearance of human herpesvirus 8 from blood of human immunodeficiency virus-1-infected patients. J Med Virol. 2000; 62:416–20. [PubMed: 11074468]
- 19. Borok M, Fiorillo S, Gudza I, Putnam B, Ndemera B, White IE, et al. Evaluation of Plasma Human Herpesvirus 8 DNA as a Marker of Clinical Outcomes during Antiretroviral Therapy for AIDS-Related Kaposi Sarcoma in Zimbabwe. Clin Infect Dis. 2010
- 20. Wilkinson J, Cope A, Gill J, Bourboulia D, Hayes P, Imami N, et al. Identification of Kaposi's sarcoma-associated herpesvirus (KSHV)-specific cytotoxic T-lymphocyte epitopes and evaluation of reconstitution of KSHV-specific responses in human immunodeficiency virus type 1-Infected patients receiving highly active antiretroviral therapy. J Virol. 2002; 76:2634–40. [PubMed: 11861829]
- 21. Sirianni MC, Vincenzi L, Topino S, Giovannetti A, Mazzetta F, Libi F, et al. NK cell activity controls human herpesvirus 8 latent infection and is restored upon highly active antiretroviral therapy in AIDS patients with regressing Kaposi's sarcoma. Eur J Immunol. 2002; 32:2711–20. [PubMed: 12355422]
- 22. Barillari G, Ensoli B. Angiogenic effects of extracellular human immunodeficiency virus type 1 Tat protein and its role in the pathogenesis of AIDS-associated Kaposi's sarcoma. Clin Microbiol Rev. 2002; 15:310–26. [PubMed: 11932235]
- 23. Varthakavi V, Smith RM, Deng H, Sun R, Spearman P. Human immunodeficiency virus type-1 activates lytic cycle replication of Kaposi's sarcoma-associated herpesvirus through induction of KSHV Rta. Virology. 2002; 297:270–80. [PubMed: 12083825]
- 24. Bani-Sadr F, Fournier S, Molina JM. Relapse of Kaposi's sarcoma in HIV-infected patients switching from a protease inhibitor to a non-nucleoside reverse transcriptase inhibitor-based highly active antiretroviral therapy regimen. Aids. 2003; 17:1580–1. [PubMed: 12824806]

NIH-PA Author Manuscript

NIH-PA Author Manuscript

- 25. Leitch H, Trudeau M, Routy JP. Effect of protease inhibitor-based highly active antiretroviral therapy on survival in HIV-associated advanced Kaposi's sarcoma patients treated with chemotherapy. HIV Clin Trials. 2003; 4:107–14. [PubMed: 12671778]
- 26. Rey D, Schmitt MP, Partisani M, Hess-Kempf G, Krantz V, de Mautort E, et al. Efavirenz as a substitute for protease inhibitors in HIV-1-infected patients with undetectable plasma viral load on HAART: a median follow-up of 64 weeks. J Acquir Immune Defic Syndr. 2001; 27:459–62. [PubMed: 11511822]
- 27. Portsmouth S, Stebbing J, Gill J, Mandalia S, Bower M, Nelson M, et al. A comparison of regimens based on non-nucleoside reverse transcriptase inhibitors or protease inhibitors in preventing Kaposi's sarcoma. Aids. 2003; 17:F17–22. [PubMed: 12853764]
- 28. Martinez V, Caumes E, Gambotti L, Ittah H, Morini JP, Deleuze J, et al. Remission from Kaposi's sarcoma on HAART is associated with suppression of HIV replication and is independent of protease inhibitor therapy. Br J Cancer. 2006; 94:1000–6. [PubMed: 16570046]
- 29. Crum-Cianflone NF, Hullsiek KH, Marconi V, Weintrob A, Ganesan A, Barthel RV, et al. The impact of nelfinavir exposure on cancer development among a large cohort of HIV-infected patients. J Acquir Immune Defic Syndr. 2009; 51:305–9. [PubMed: 19412116]
- 30. Gantt S, Casper C. Human herpesvirus 8-associated neoplasms: the roles of viral replication and antiviral treatment. Curr Opin Infect Dis. 2011; 24:295–301. [PubMed: 21666458]
- 31. Bernstein WB, Dennis PA. Repositioning HIV protease inhibitors as cancer therapeutics. Curr Opin HIV AIDS. 2008; 3:666–75. [PubMed: 19373040]
- 32. Chow WA, Jiang C, Guan M. Anti-HIV drugs for cancer therapeutics: back to the future? Lancet Oncol. 2009; 10:61–71. [PubMed: 19111246]
- 33. Gantt S, Casper C, Ambinder RF. Insights Into the Broad Cellular Effects of Nelfinavir and the HIV Protease Inhibitors Supporting Their Role in Cancer Treatment and Prevention Curr Opin Oncol. 2013; 25
- 34. Gantt S, Carlsson J, Ikoma M, Gachelet E, Gray M, Geballe AP, et al. The HIV protease inhibitor nelfinavir inhibits Kaposi's sarcoma-associated herpesvirus replication in vitro. Antimicrob Agents Chemother. 2011; 55:2696–703. [PubMed: 21402841]
- 35. Gustafson EA, Schinazi RF, Fingeroth JD. Human herpesvirus 8 open reading frame 21 is a thymidine and thymidylate kinase of narrow substrate specificity that efficiently phosphorylates zidovudine but not ganciclovir. J Virol. 2000; 74:684–92. [PubMed: 10623730]
- 36. Lock MJ, Thorley N, Teo J, Emery VC. Azidodeoxythymidine and didehydrodeoxythymidine as inhibitors and substrates of the human herpesvirus 8 thymidine kinase. J Antimicrob Chemother. 2002; 49:359–66. [PubMed: 11815580]
- 37. Corey L, Brodie S, Huang ML, Koelle DM, Wald A. HHV-8 infection: a model for reactivation and transmission. Rev Med Virol. 2002; 12:47–63. [PubMed: 11787083]
- 38. Casper C, Redman M, Huang ML, Pauk J, Lampinen TM, Hawes SE, et al. HIV infection and human herpesvirus-8 oral shedding among men who have sex with men. J Acquir Immune Defic Syndr. 2004; 35:233–8. [PubMed: 15076237]
- 39. Ryncarz AJ, Goddard J, Wald A, Huang ML, Roizman B, Corey L. Development of a highthroughput quantitative assay for detecting herpes simplex virus DNA in clinical samples. J Clin Microbiol. 1999; 37:1941–7. [PubMed: 10325351]
- 40. Johnston C, Orem J, Okuku F, Kalinaki M, Saracino M, Katongole-Mbidde E, et al. Impact of HIV infection and Kaposi sarcoma on human herpesvirus-8 mucosal replication and dissemination in Uganda. PLoS One. 2009; 4:e4222. [PubMed: 19156206]
- 41. Casper C, Krantz E, Taylor H, Dalessio J, Carrell D, Wald A, et al. Assessment of a combined testing strategy for detection of antibodies to human herpesvirus 8 (HHV-8) in persons with Kaposi's sarcoma, persons with asymptomatic HHV-8 infection, and persons at low risk for HHV-8 infection. J Clin Microbiol. 2002; 40:3822–5. [PubMed: 12354890]
- 42. Pauk J, Huang ML, Brodie SJ, Wald A, Koelle DM, Schacker T, et al. Mucosal shedding of human herpesvirus 8 in men. N Engl J Med. 2000; 343:1369–77. [PubMed: 11070101]
- 43. Casper C, Krantz E, Selke S, Kuntz SR, Wang J, Huang ML, et al. Frequent and Asymptomatic Oropharyngeal Shedding of Human Herpesvirus 8 among Immunocompetent Men. J Infect Dis. 2007; 195:30–6. [PubMed: 17152006]

- 44. Diggle, P.; Heagerty, P.; Liang, KY.; Zeger, S. Analysis of Longitudinal data. 2nd. New York: Oxford University Press; 2002.
- 45. Hernan MA, Brumback B, Robins JM. Marginal structural models to estimate the causal effect of zidovudine on the survival of HIV-positive men. Epidemiology. 2000; 11:561–70. [PubMed: 10955409]
- 46. Casper C, Krantz EM, Corey L, Kuntz SR, Wang J, Selke S, et al. Valganciclovir for suppression of human herpesvirus-8 replication: a randomized, double-blind, placebo-controlled, crossover trial. J Infect Dis. 2008; 198:23–30. [PubMed: 18491970]
- 47. Ganem D. KSHV and the pathogenesis of Kaposi sarcoma: listening to human biology and medicine. J Clin Invest. 120:939–49. [PubMed: 20364091]
- 48. Sgadari C, Barillari G, Toschi E, Carlei D, Bacigalupo I, Baccarini S, et al. HIV protease inhibitors are potent anti-angiogenic molecules and promote regression of Kaposi sarcoma. Nat Med. 2002; 8:225–32. [PubMed: 11875492]
- 49. Pati S, Pelser CB, Dufraine J, Bryant JL, Reitz MS Jr, Weichold FF. Antitumorigenic effects of HIV protease inhibitor ritonavir: inhibition of Kaposi sarcoma. Blood. 2002; 99:3771–9. [PubMed: 11986235]
- 50. Kalu, N.; Shirley, C.; Ambinder, RF. International Congress on Oncogenic Herpesviruses and Associated Diseases. Philadelphia, PA: Infectious Agents and Cancer; 2012. ER stress activates lytic gene expression in KSHV associated tumor cell lines.
- 51. Grabar S, Abraham B, Mahamat A, Del Giudice P, Rosenthal E, Costagliola D. Differential impact of combination antiretroviral therapy in preventing Kaposi's sarcoma with and without visceral involvement. J Clin Oncol. 2006; 24:3408–14. [PubMed: 16849755]
- 52. Conant MA, Opp KM, Poretz D, Mills RG. Reduction of Kaposi's sarcoma lesions following treatment of AIDS with ritonovir. Aids. 1997; 11:1300–1. [PubMed: 9256954]
- 53. Stebbing J, Portsmouth S, Nelson M, Mandalia S, Kandil H, Alexander N, et al. The efficacy of ritonavir in the prevention of AIDS-related Kaposi's sarcoma. Int J Cancer. 2004; 108:631–3. [PubMed: 14696132]
- 54. Ridolfo AL, Corbellino M, Tosca N, Capelletti A, Scalamogna C, Galli M, et al. Is switching protease inhibitor-based effective antiretroviral therapy safe in patients with AIDS-associated Kaposi's sarcoma? Aids. 2004; 18:1224–6. [PubMed: 15166546]
- 55. Monini P, Sgadari C, Grosso MG, Bellino S, Di Biagio A, Toschi E, et al. Clinical course of classic Kaposi's sarcoma in HIV-negative patients treated with the HIV protease inhibitor indinavir. Aids. 2009; 23:534–8. [PubMed: 19169139]

# **Table 1 Characteristics of Study Population**







*\** Among sessions with ≥ 1 positive day

Abbreviations: HHV-8, human herpesvirus 8; ART, antiretroviral therapy; HAART, highly active ART; PI, protease inhibitor; SD, standard deviation

#### **Table 3**

Associations between variables and frequency of HHV-8 detection in oropharyngeal swabs by PCR.



Multivariate models are adjusted for all of the variables listed.

#### **Table 4**

Associations between variables and quantity of HHV-8 detected in oropharyngeal swabs by PCR.



Multivariate models are adjusted for all of the variables listed.