Research Article

Potential Retinoid X Receptor Agonists for Treating Alzheimer's Disease from Traditional Chinese Medicine

Kuan-Chung Chen,[1] Yu-Cheng Liu,[2] Cheng-Chun Lee,[2] and Calvin Yu-Chian Chen[2,3]

[1] *School of Pharmacy, China Medical University, Taichung 40402, Taiwan*
[2] *School of Medicine, College of Medicine, China Medical University, Taichung 40402, Taiwan*
[3] *Department of Biomedical Informatics, Asia University, Taichung 41354, Taiwan*

Correspondence should be addressed to Cheng-Chun Lee; leeck@mail.cmu.edu.tw and Calvin Yu-Chian Chen; ycc929@MIT.edu

Alzheimer's disease is neurodegenerative disorder due to the accumulation of amyloid-β in the brain and causes dementia with ageing. Some researches indicate that the RXR agonist, Targretin, has also been used for treatment of Alzheimer's disease in mouse models. We investigate the potent candidates as RXR agonists from the vast repertoire of TCM compounds in TCM Database@Taiwan. The potential TCM compounds, β-lipoic acid and sulfanilic acid, had higher potent binding affinities than both 9-*cis*-retinoic acid and Targretin in docking simulation and have stable H-bonds with residues Arg316 and some equivalent hydrophobic contacts with residues Ala272, Gln275, Leu309, Phe313, Val342, Ile345, and Cys432 as Targretin. The carboxyl or sulfonyl hydroxide group can form a H-bond with key residue Arg316 in the docking pose, and the phenyl group next to the carboxyl or sulfonyl hydroxide group can form a π interaction with residue Phe313. Moreover, β-lipoic acid and sulfanilic acid have stable H-bonds with residue Gln275, Ser313, and residue Ala327, respectively, which may strengthen and stabilize TCM candidates inside the binding domain of RXR protein. Hence, we propose β-lipoic acid and sulfanilic acid as potential lead compounds for further study in drug development process with the RXR protein against Alzheimer's disease.

1. Introduction

Alzheimer's disease is serious problem which will cause huge cost for taking care of the patient. It is neurodegenerative disorder due to the accumulation of amyloid-β in the brain and causes dementia with ageing [1, 2]. The cholesterol transport protein apolipoprotein E plays the important role in the clearance of amyloid-β from the brain [3]. In addition, the transcription of apolipoprotein E expression is also regulated by the heterodimers of RXR with PPAR-γ and LXRs [4, 5]. The RXR agonist, Targretin, has also been used for treatment of Alzheimer's disease in mouse models [6].

The retinoic X receptors (RXRs) belong to a superfamily of eukaryotic transcription factors. They are ligand-dependent nuclear receptors involved in the processes of retinoid signaling in normal hematopoiesis [7, 8] and cell development such as cell patterning, organogenesis, proliferation, and differentiation [9, 10]. RXRs have three different isoforms (α, β, and γ) which form heterodimers with other nuclear receptors, such as retinoid acid receptor (RAR), peroxisome proliferator-activated receptor (PPAR), farnesoid X receptor (FXR), liver X receptor (LXR), thyroid hormone receptor (TR), and vitamin D receptor (VDR) as coregulators [11–17]. The transcription of RXRs is activated by endogenous 9-*cis*-retinoic acid, which is a vitamin A derivative for regulation of growth and morphogenesis [18–21]. The selective synthetic ligands for the RXRs have been also indicated as therapeutic agents to treat cancer and dermatological diseases [22–27].

Nowadays, more and more mechanisms of diseases had been determined to detect the useful target protein against diseases [28–33]. Previously to *in silico* drug discovery
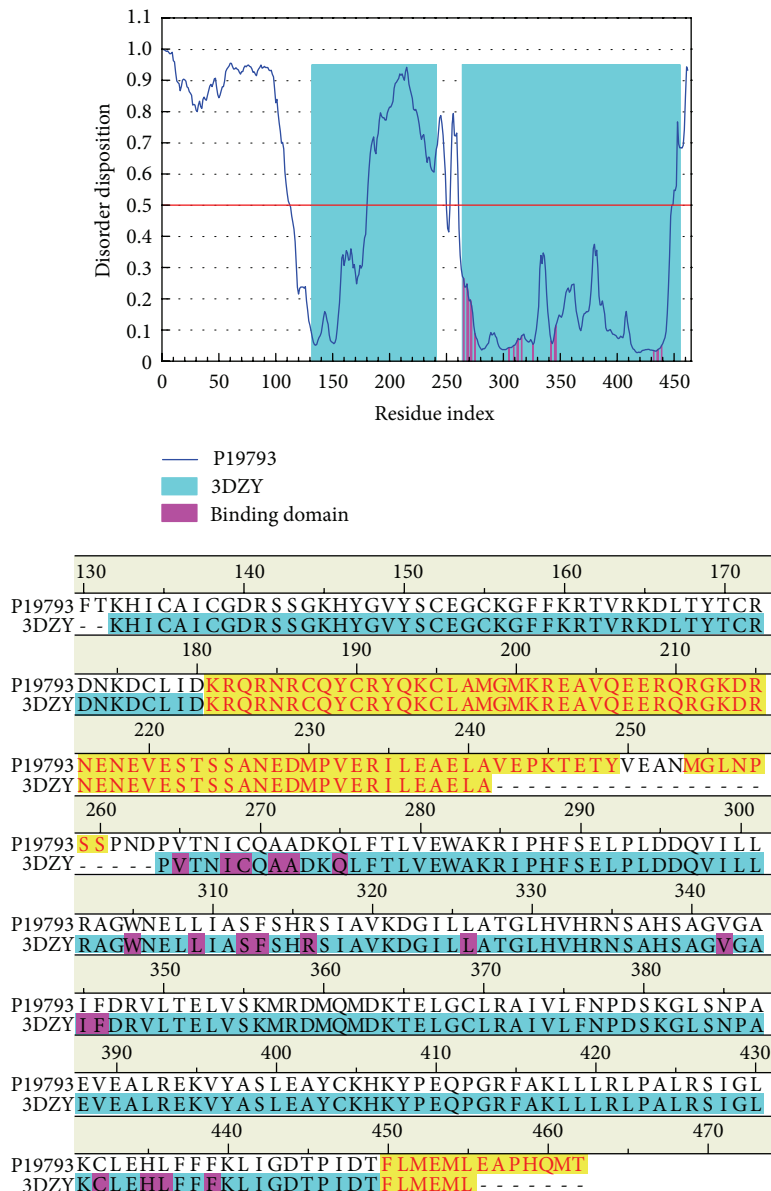
FIGURE 1: Disordered amino acids predicted by PONDR-Fit and sequence alignment with disordered residues (yellow regions) and residues in the binding domain (magenta regions).

researches, many compounds extracted from traditional Chinese medicine (TCM) have been indicated as potential lead compounds used for wide range of diseases, including metabolic syndrome [34–36], stroke [37–40], cancers [41–45], influenza [46–49], viral infection [50], diabetes [51], inflammation [52], and some other diseases [53, 54]. To improve drug discovery from TCM compounds, we aim to investigate the potent candidates as RXR agonists from the vast repertoire of TCM compounds in TCM Database@Taiwan. As the side effect and ligand binding with target protein may affect by the structural disorders of residues in the protein [55, 56], the prediction of disordered amino acids of RXR protein was performed before virtual screening. After virtual screening the TCM compounds,

the molecular dynamics (MD) simulations were then employed to study protein dynamics and analyze the stability of interactions for each docking poses of TCM candidates.

2. Materials and Methods

2.1. Data Collection. The X-ray crystallography structure of the intact PPAR-γ-RXR-nuclear receptor complex on DNA with 9-*cis*-retinoic acid was obtained from RCSB Protein Data Bank with PDB ID: 3DZY [57]. To protonate the structure of protein with Chemistry at HARvard Macromolecular Mechanics (CHARMM) force field [58] and remove crystal water, the crystal structure of RXR protein was prepared by
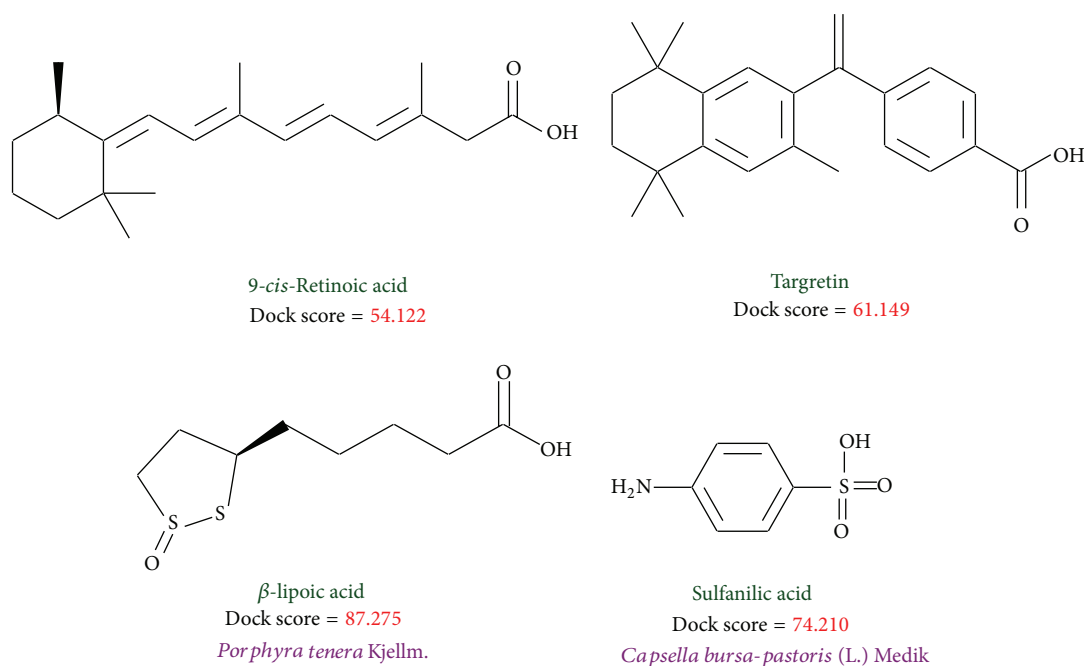
FIGURE 2: Chemical scaffold of controls and two TCM candidates with their scoring function and sources.

Prepare Protein module in Discovery Studio 2.5 (DS2.5). The binding site for virtual screening was defined by the volume and location of 9-*cis*-retinoic acid. A total of 9,029 nonduplicate TCM compounds from TCM Database@Taiwan [59] were protonating the structure by Prepare Ligand module in DS2.5 after filtering by Lipinski's Rule of Five [60]. The sequence of RXR protein from Swiss-Prot (UniProtKB: P19793) was employed to predict the disordered amino acids using PONDR-Fit [61].

2.2. Docking Simulation. The TCM compounds were virtually screened by LigandFit protocol [62] in DS2.5. The LigandFit docking procedure was performed by five major steps. Firstly, it generates candidate ligand conformations using Monte Carlo for docking. Then it positions each ligand conformation in the binding site by multiple orientation or permutation sampling of the ligand principal moments with the principal moments of the site. The docking poses were then optionally minimized with CHARMM force field [58] and calculated the score using the dock score energy function as follows:

$$
\begin{aligned}
\text{Dock Score} = &- (\text{ligand receptor interaction energy} \\
&+ \text{ligand internal energy}).
\end{aligned}
\tag{1}
$$

A pose-saving algorithm was employed to compare the candidate poses and reject the similar poses.

2.3. Molecular Dynamics (MD) Simulation. Gromacs [63] is employed to study protein dynamics using classical molecular dynamics theory. The global MD algorithm is defined by four major parts. (1) We input the initial conditions with potential interaction (V), position (r), and velocities (v) for all atoms in the system in this part. (2) The program computes the forces on each atom in the system as follows:

$$
F_i = \frac{\partial V}{\partial r_i},
\tag{2}
$$

where F_i is obtained by calculating the forces between nonbonded atom pairs, bonded interactions, and restraining and external forces.

The potential and kinetic energies and the pressure tensor are also computed in this part. (3) The MD program updates the configuration by simulating the movement of the atoms using Newton's equations of motion as follows:

$$
\frac{\partial r_i}{\partial t} = v_i; \qquad \frac{\partial v_i}{\partial t} = \frac{F_i}{m_i}.
\tag{3}
$$

(4) The MD program outputs the information of positions, velocities, energies, temperature, pressure, and so forth. Finally, the MD program repeats the 2–4 parts for the required number of steps.

The molecular dynamics simulations (MD) are performed by Gromacs. To obtain the initial conditions of each protein-ligand complex, the topology of RXR protein, including charmm27 force fields, was reprepared by Gromacs. The topology and parameters of each ligand for use with Gromacs were provided by SwissParam program [64]. The program has employed a cubic box with a minimum distance of 12 Å from the molecules periphery and solvated by a water model of TIP3P. Firstly, a maximum of 5,000 steps energy minimization were performed using Steepest Descent
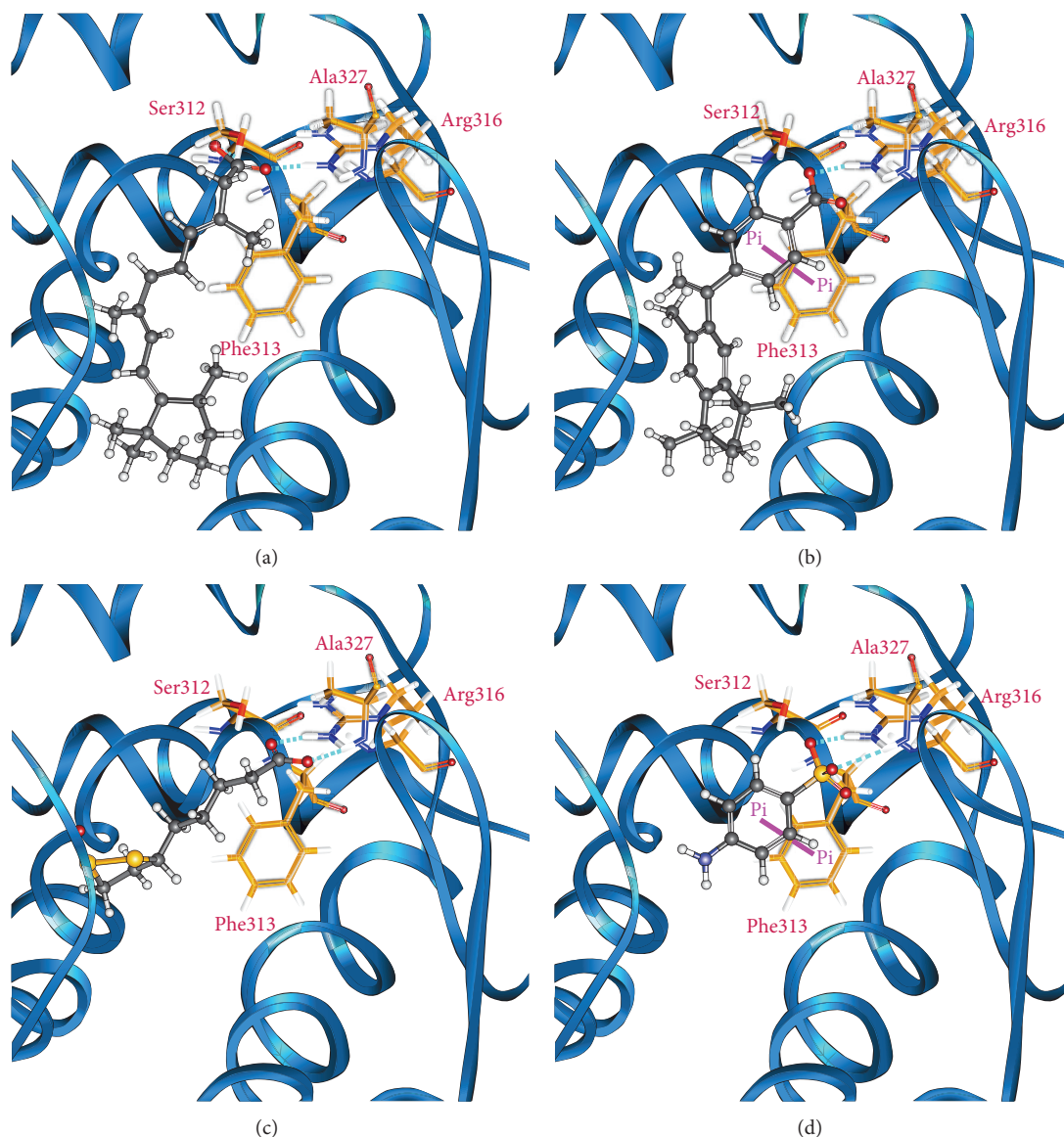
(a)

(b)

(c)

(d)

Figure 3: Docking pose of RXR protein complexes with (a) 9-*cis*-retinoic acid, (b) Targretin, (c) β-lipoic acid, and (d) sulfanilic acid.

algorithm [65]. Then a single constant temperature (NVT ensemble) equilibration was performed using Berendsen weak thermal coupling method. The MD program repeats 2–4 parts in a time step unit of 2 fs under the particle mesh Ewald (PME) option to obtain a total of 20 ns production simulation. The 20 ns MD trajectories were analyzed by a series of protocols in Gromacs.

3. Results and Discussion

3.1. Disordered Protein Prediction. The disordered amino acids of RXR protein were predicted by PONDR-Fit with the protein sequence from Swiss-Prot (UniProtKB: P19793). The sequence alignment and result of disordered amino acids prediction were displayed in Figure 1. The residues in

the binding domain do not deposit in the disordered region. It has shown that the RXR protein may have a stable structure of binding domain in protein folding.

3.2. Docking Simulation. Figure 2 displays the chemical scaffold of 9-*cis*-retinoic acid, Targretin, and the top TCM compounds ranked by dock score with their scoring function and sources. The scoring function of dock score indicates that the TCM compounds β-lipoic acid and sulfanilic acid have higher binding affinities than both 9-*cis*-retinoic acid and Targretin. The TCM compound β-lipoic acid is extracted from *Porphyra tenera* Kjellm., whereas sulfanilic acid is extracted from *Capsella bursa-pastoris* (L.) Medik. The docking pose of RXR protein complexes with 9-*cis*-retinoic acid, Targretin, and two top TCM candidates was illustrated in
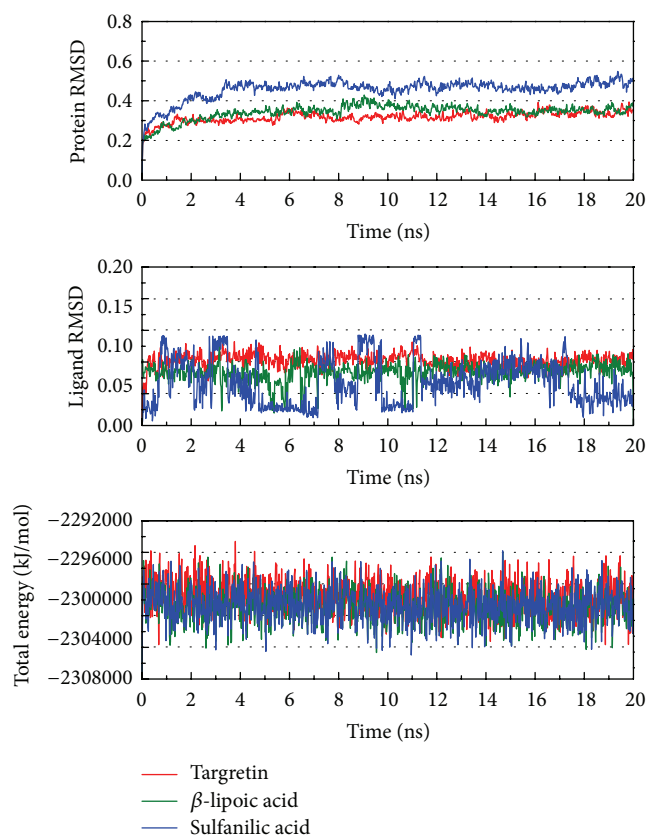
FIGURE 4: Root-mean-square deviations in units of nm and total energies over 20 ns MD simulation for RXR protein complexes with Targretin, β-lipoic acid, and sulfanilic acid.
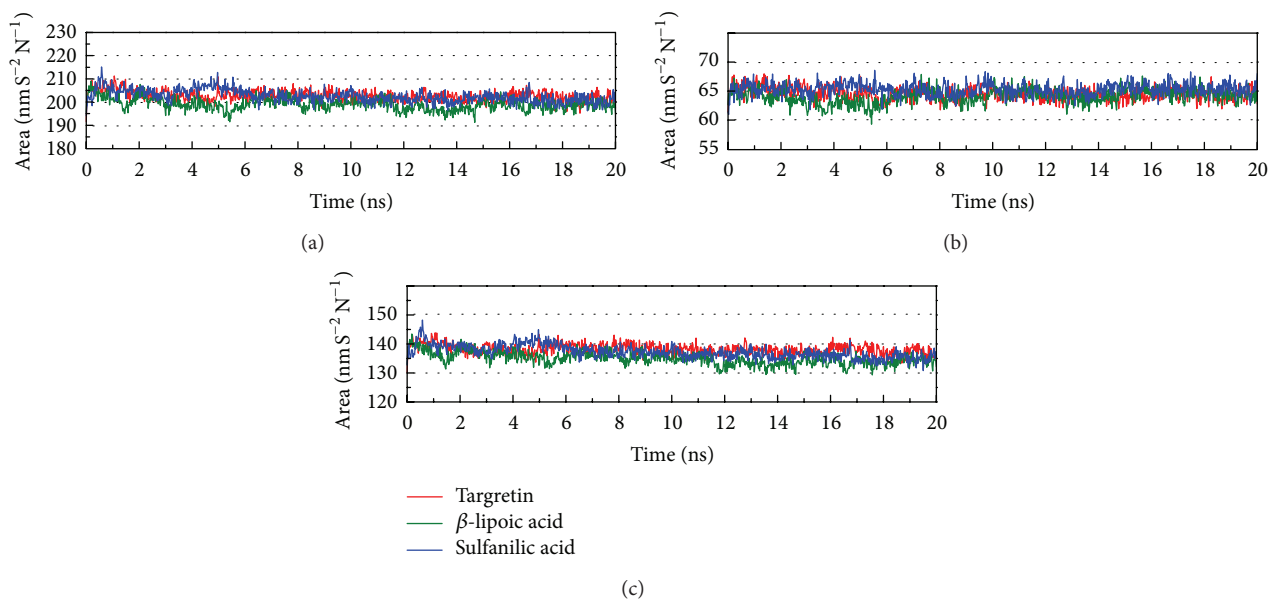
(a)

(b)

(c)

FIGURE 5: Variation of (a) total solvent accessible surface area, (b) hydrophobic surface area, and (c) hydrophilic surface area over 20 ns MD simulation for RXR protein complexes with Targretin, β-lipoic acid, and sulfanilic acid.
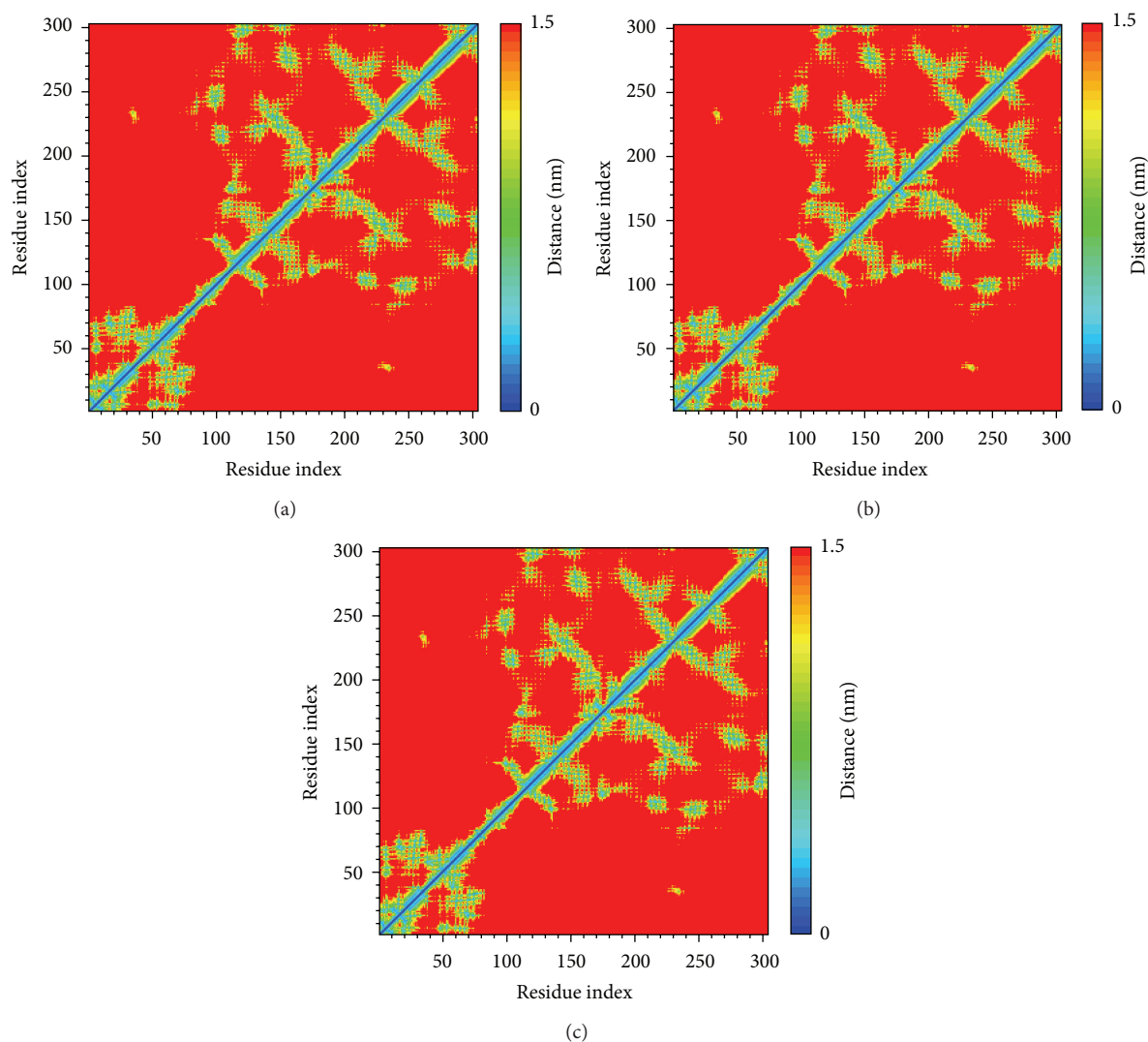
(a)

(b)

(c)

FIGURE 6: Distance matrices consisting of the mean smallest distance between residue pairs for RXR protein complexes with (a) Targretin, (b) β-lipoic acid, and (c) sulfanilic acid. Residues 1–110 in y-axis correspond to residues 132–241. Residues 111–302 in y-axis correspond to residues 264–455.

Figure 3. All compounds have a hydrogen bond (H-bond) with key residue Arg316. Moreover, both TCM candidates, β-lipoic acid and sulfanilic acid, has a H-bond with residue Ala327. Both Targretin and sulfanilic acid, which have a phenyl group, have a π interaction with residue Phe313. It shows that the carboxyl group or the group of sulfonyl hydroxide can form a H-bond with key residue Arg316 in the docking pose. The phenyl group next to the carboxyl group or the group of sulfonyl hydroxide can form a π interaction with residue Phe313.

3.3. Molecular Dynamics Simulation. As the docking simulations are perform in the condition of rigid body of RXR protein, the molecular dynamics (MD) simulations were then employed to study of protein dynamics and analyze the stability of interactions for each docking poses. The root-mean-square deviations (RMSDs) and total energies over 20 ns MD simulation for RXR protein complexes

with Targretin, β-lipoic acid, and sulfanilic acid are illustrated in Figure 4. The complexes with Targretin, β-lipoic acid, and sulfanilic acid tend to be stable after 16 ns, 16.2 ns, and 17.4 ns MD simulation, respectively. The analysis of solvent accessible surface area for complexes under dynamics condition in Figure 5 indicates that the RXR protein complexes with Targretin, β-lipoic acid, and sulfanilic acid have similar hydrophobic and hydrophilic surface area and the MD simulation tends to be stable. Figure 6 also shows that the RXR protein has similar mean smallest distance between residue pairs for protein complex with Targretin and two top TCM compounds. They indicate that the protein structure of RXR protein complex with top two TCM compounds, β-lipoic acid and sulfanilic acid, may not cause the significant differences from docking with Targretin.

Root-mean-square deviation value and graphical depiction of the clusters with cutoff 0.11 nm for each RXR protein complexes are employed to display the RMSD values
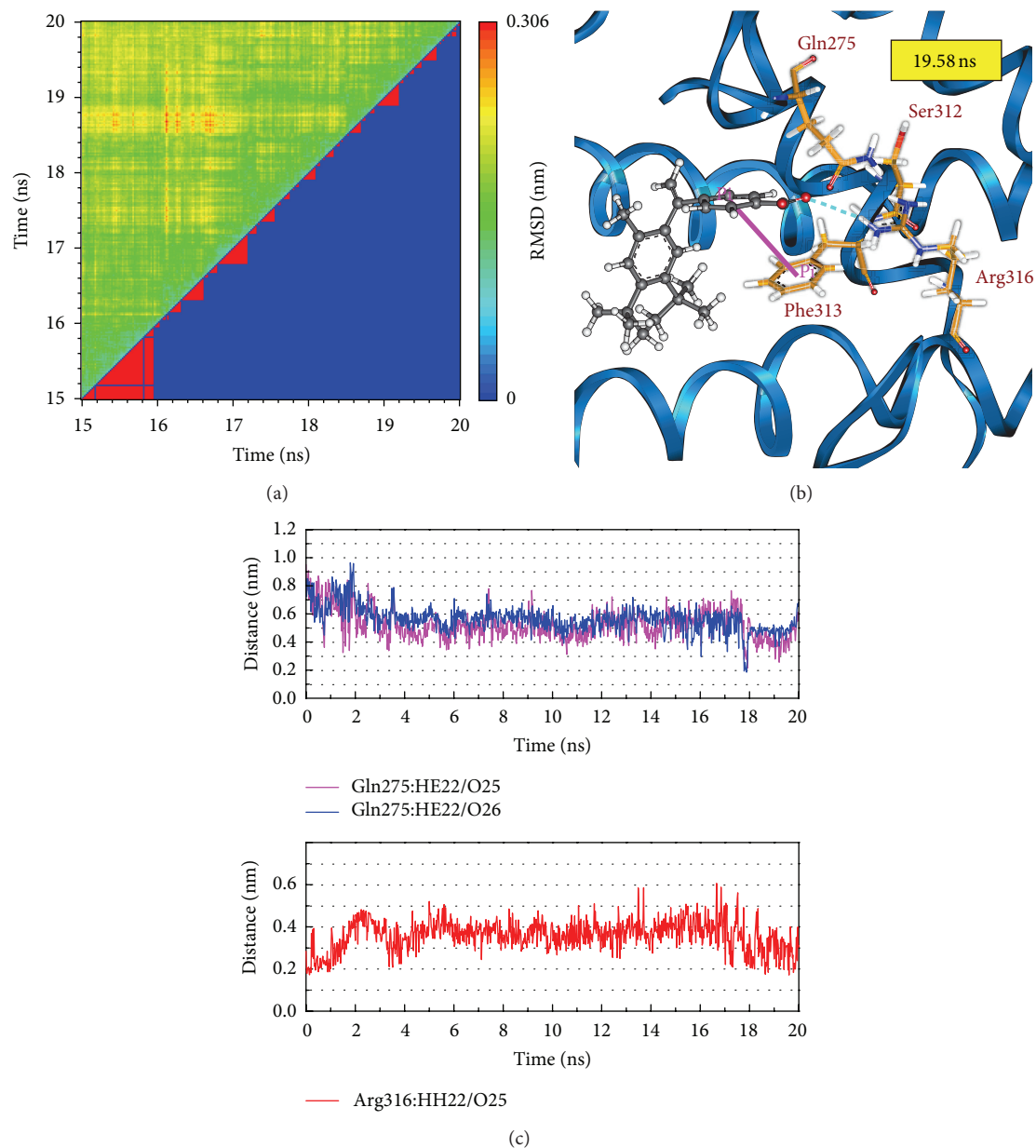
(a)

(b)

(c)

FIGURE 7: (a) Root-mean-square deviation value (upper left half) and graphical depiction of the clusters with cutoff 0.11 nm (lower right half) for RXR protein complexes with Targretin. (b) Docking poses of middle RMSD structure in the major cluster for RXR protein complexes with Targretin. (c) Distances of hydrogen bonds with common residues during 20 ns MD simulation.

between MD trajectories and identify the middle RMSD structure in the major cluster as the representative structures of each protein-ligand complex after MD simulation. For RXR protein complexes with Targretin, the docking poses of middle RMSD structure in the major cluster after 16 ns MD simulation are illustrated in Figure 7(b). Targretin has the similar docking pose as docking simulation and maintains the H-bond with key residue Arg316 and π interaction with residue Phe313. According to the occupancies of H-bonds for common residues of RXR protein listed in Table 1 and the fluctuation of distances for H-bonds displayed in Figure 7(c), they show that the carboxyl group of Targretin can form

stable interactions with residue Arg316. For RXR protein complexes with β-lipoic acid, the representative structures of docking pose after MD simulation are illustrated in Figure 8(b). β-lipoic acid maintains the H-bond with key residue Arg316 and forms the other H-bonds with residue Gln275 and Ser313, which can stabilize the docking pose in the binding domain. From the occupancies of H-bonds for common residues of RXR protein listed in Table 1 and the fluctuation of distances for H-bonds displayed in Figure 8(c), they indicate that the H-bonds between β-lipoic acid and residue Ala241 existing in the initial period of MD simulation were not stable. However, the carboxyl group of β-lipoic

(a)

(b)

(c)

FIGURE 8: (a) Root-mean-square deviation value (upper left half) and graphical depiction of the clusters with cutoff 0.11 nm (lower right half) for RXR protein complexes with β-lipoic acid. (b) Docking poses of middle RMSD structure in the major cluster for RXR protein complexes with β-lipoic acid. (c) Distances of hydrogen bonds with common residues during 20 ns MD simulation.
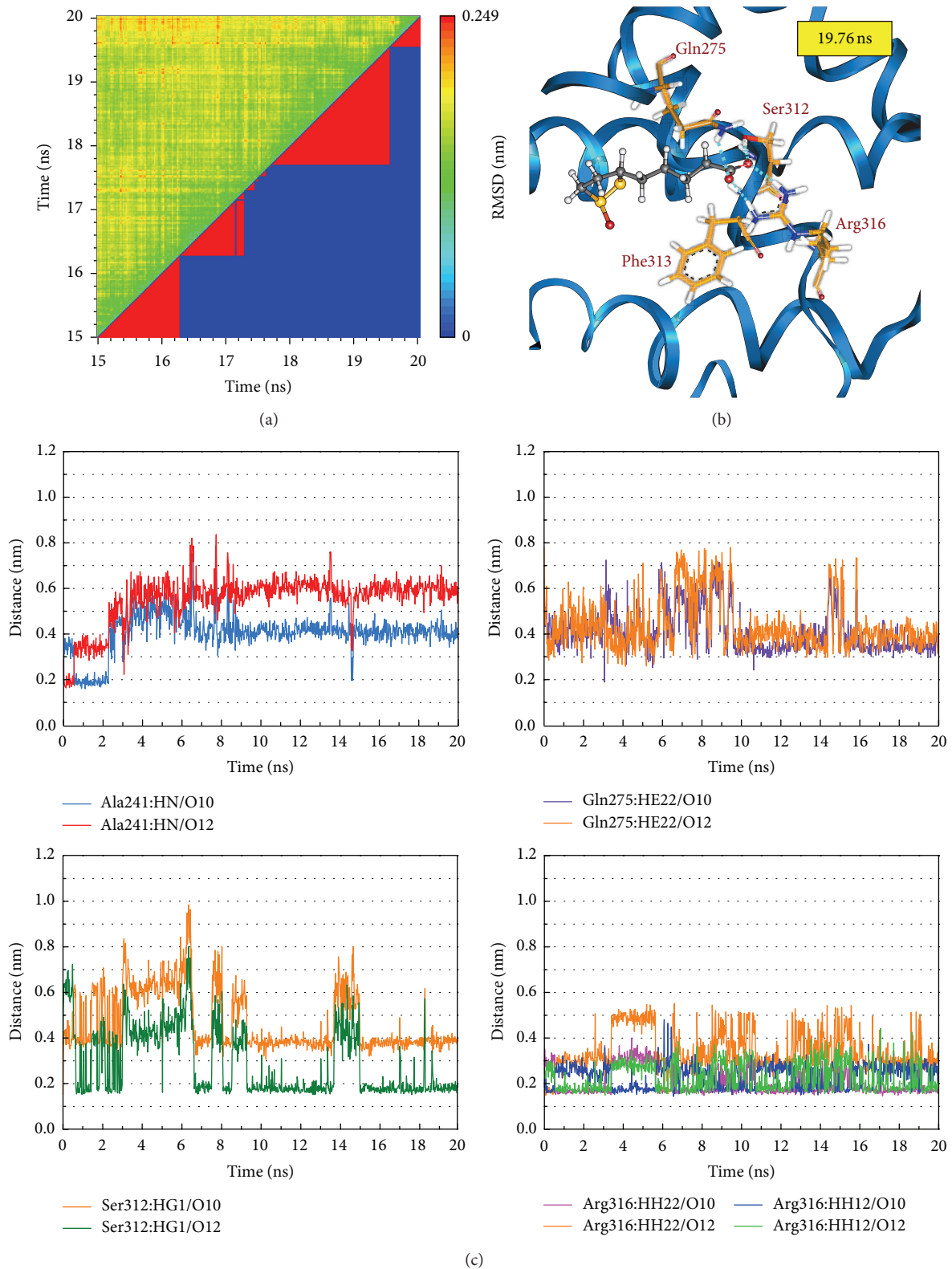
(a)

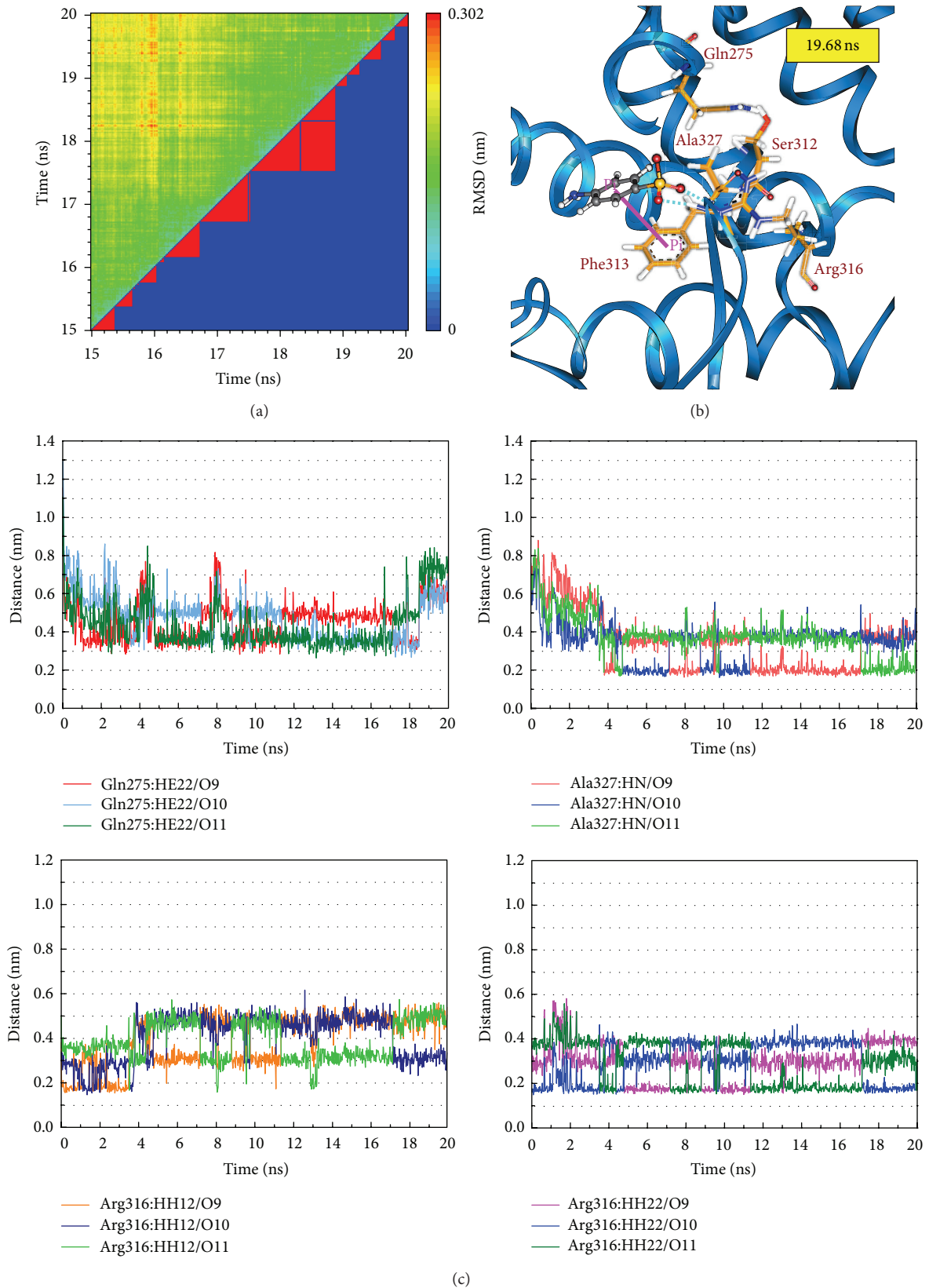
(b)

(c)

FIGURE 9: (a) Root-mean-square deviation value (upper left half) and graphical depiction of the clusters with cutoff 0.11 nm (lower right half) for RXR protein complexes with sulfanilic acid. (b) Docking poses of middle RMSD structure in the major cluster for RXR protein complexes with sulfanilic acid. (c) Distances of hydrogen bonds with common residues during 20 ns MD simulation.
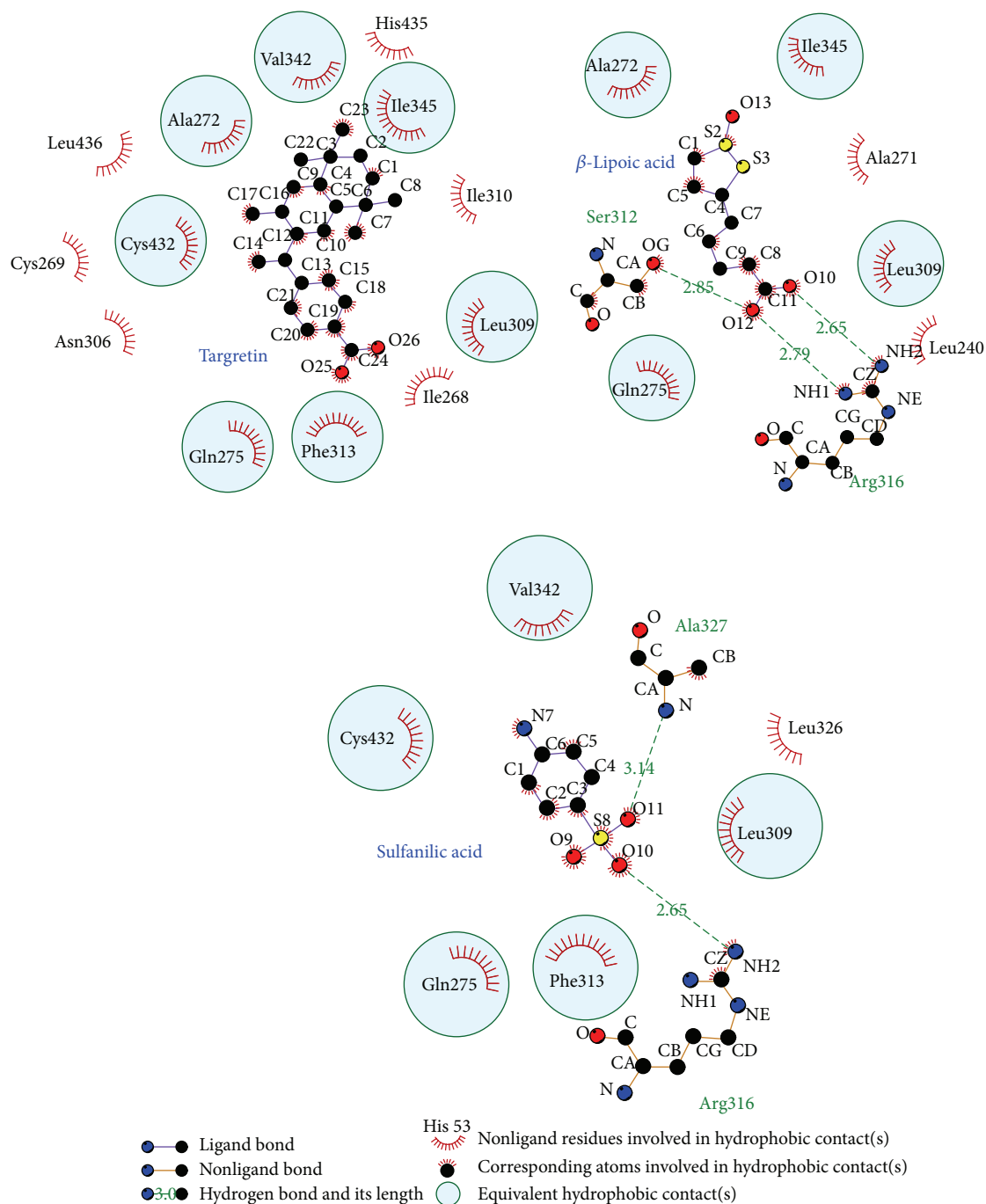
FIGURE 10: Docking poses of middle RMSD structure in the major cluster for RXR protein complexes drawn by LIGPLOT program.

acid has stable H-bonds with residues Arg316, Gln275, and Ser313. From the representative structures of RXR protein complex with sulfanilic acid after MD simulation displayed in Figure 9(b), it is illustrated that the group of sulfonyl hydroxide of sulfanilic acid maintains the H-bonds with key residues Arg316 and Ala327 and the phenyl group next to sulfonyl hydroxide keeps the π interaction with residue Phe313. The fluctuation of distances for H-bonds displayed in Figure 9(c) indicates that sulfanilic acid had the H-bonds with residue

Gln275, but it misses those H-bonds when the system tends to be stable after 18 ns MD simulation. Figure 10 displays the docking poses of the representative structures for RXR protein complexes with Targretin, β-lipoic acid, and sulfanilic acid. It indicates that the TCM candidates, β-lipoic acid and sulfanilic acid, have some equivalent hydrophobic contacts with residues Ala272, Gln275, Leu309, Phe313, Val342, Ile345, and Cys432 as Targretin. These hydrophobic contacts hold the compounds in the binding domain.

TABLE 1: H-bond occupancy for key residues of RXR protein with Targretin and two candidates overall 20 ns molecular dynamics simulation.

Name	H-bond Interaction	Occupancy
Targretin	Gln275:HE22/O25	1%
	Gln275:HE22/O26	1%
	Arg316:HH22/O25	17%
β-lipoic acid	Ala241:HN/O10	10%
	Ala241:HN/O12	3%
	Gln275:HE22/O10	2%
	Gln275:HE22/O12	2%
	Ser312:HG1/O12	60%
	Arg316:HH22/O10	89%
	Arg316:HH22/O12	39%
	Arg316:HH12/O10	96%
	Arg316:HH12/O12	86%
Sulfanilic acid	Gln275:HE22/O9	1%
	Gln275:HE22/O10	1%
	Gln275:HE22/O11	2%
	Arg316:HH22/O9	53%
	Arg316:HH22/O10	45%
	Arg316:HH22/O11	49%
	Arg316:HH12/O9	25%
	Arg316:HH12/O10	23%
	Arg316:HH12/O11	16%
	Ala327:HN/O9	39%
	Ala327:HN/O10	24%
	Ala327:HN/O11	17%

H-bond occupancy cutoff: 0.3 nm.

4. Conclusion

The cholesterol transport protein apolipoprotein E plays the important role in the clearance of amyloid-β from the brain, and the transcription of apolipoprotein E expression is also regulated by the heterodimers of RXR with PPAR-γ and LXRs. In this study, we aim to investigate the potent TCM candidates of agonists for RXR protein, and the prediction of disordered amino acids of RXR protein was performed to discuss the stability of residues for RXR protein before virtual screening. The top TCM candidates, β-lipoic acid and sulfanilic acid, had higher potent binding affinities than both 9-*cis*-retinoic acid and Targretin. After optimizing the result of docking simulation to validate the stability of H-bonds between each ligand and RXR protein under dynamic conditions, the top TCM compounds, β-lipoic acid and sulfanilic acid, have stable H-bonds with residues Arg316 and some equivalent hydrophobic contacts with residues Ala272, Gln275, Leu309, Phe313, Val342, Ile345, and Cys432 as Targretin. The carboxyl group or the group of sulfonyl hydroxide can form a H-bond with key residue Arg316 in the docking pose, and the phenyl group next to the carboxyl group or the group of sulfonyl hydroxide can form a π interaction with residue Phe313. In addition, β-lipoic acid has stable H-bonds with residues Gln275 and Ser313, and sulfanilic acid has stable H-bonds with residue Ala327. These stable H-bonds may strengthen and stabilize TCM candidates inside the binding domain of RXR protein. Hence, we propose β-lipoic acid and sulfanilic acid as potential lead compounds for further study in drug development process with the RXR protein against Alzheimer's disease.

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

Authors' Contribution

Kuan-Chung Chen and Yu-Cheng Liu contributed equally to this work.

Acknowledgments

References

[1] S. Crunkhorn, "Neurodegenerative disease: RXR agonist reverses Alzheimer's disease," *Nature Reviews Drug Discovery*, vol. 11, no. 4, p. 271, 2012.

[2] L. Ghezzi, E. Scarpini, and D. Galimberti, "Disease-modifying drugs in Alzheimer's disease," *Drug Design Development and Therapy*, vol. 7, pp. 1471–1479, 2013.

[3] F. M. LaFerla, "Preclinical Success against Alzheimer's Disease with an old drug," *New England Journal of Medicine*, vol. 367, no. 6, pp. 570–572, 2012.

[4] H. Kölsch, D. Lütjohann, F. Jessen et al., "RXRA gene variations influence Alzheimer's disease risk and cholesterol metabolism," *Journal of Cellular and Molecular Medicine*, vol. 13, no. 3, pp. 589–598, 2009.

[5] G. Aicardi, "New hope from an old drug: fighting Alzheimer's disease with the cancer drug bexarotene (targretin)?" *Rejuvenation Research*, vol. 16, no. 6, pp. 524–528, 2013.

[6] P. E. Cramer, J. R. Cirrito, D. W. Wesson et al., "ApoE-directed therapeutics rapidly clear β-amyloid and reverse deficits in AD mouse models," *Science*, vol. 335, no. 6075, pp. 1503–1506, 2012.

[7] M. Mark, N. B. Ghyselinck, O. Wendling et al., "A genetic dissection of the retinoid signalling pathway in the mouse," *Proceedings of the Nutrition Society*, vol. 58, no. 3, pp. 609–613, 1999.

[8] S. A. Ross, P. J. McCaffery, U. C. Drager, and L. M. De Luca, "Retinoids in embryonal development," *Physiological Reviews*, vol. 80, no. 3, pp. 1021–1054, 2000.

[9] N. J. McKenna and B. W. O'Malley, "SnapShot: nuclear receptors I," *Cell*, vol. 142, no. 5, p. 822, 2010.

[10] M. I. Dawson and Z. Xia, "The retinoid X receptors and their ligands," *Biochimica et Biophysica Acta. Molecular and Cell Biology of Lipids*, vol. 1821, no. 1, pp. 21–56, 2012.

[11] P. Chambon, "A decade of molecular biology of retinoic acid receptors," *FASEB Journal*, vol. 10, no. 9, pp. 940–954, 1996.

[12] M. Leid, P. Kastner, R. Lyons et al., "Purification, cloning, and RXR identity of the HeLa cell factor with which RAR or TR heterodimerizes to bind target sequences efficiently," *Cell*, vol. 68, no. 2, pp. 377–395, 1992.

[13] P. L. Hallenbeck, M. S. Marks, R. E. Lippoldt, K. Ozato, and V. M. Nikodem, "Heterodimerization of thyroid hormone (TH) receptor with H-2RIIBP (RXRβ) enhances DNA binding and TH-dependent transcriptional activation," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 89, no. 12, pp. 5572–5576, 1992.

[14] V. C. Yu, C. Delsert, B. Andersen et al., "RXRβ: a coregulator that enhances binding of retinoic acid, thyroid hormone, and vitamin D receptors to their cognate response elements," *Cell*, vol. 67, no. 6, pp. 1251–1266, 1991.

[15] T. H. Bugge, J. Pohl, O. Lonnoy, and H. G. Stunnenberg, "RXRα, a promiscuous partner of retinoic acid and thyroid hormone receptors," *EMBO Journal*, vol. 11, no. 4, pp. 1409–1418, 1992.

[16] S. A. Kliewer, K. Umesono, D. J. Mangelsdorf, and R. M. Evans, "Retinoid X receptor interacts with nuclear receptors in retinoic acid, thyroid hormone and vitamin D3 signalling," *Nature*, vol. 355, no. 6359, pp. 446–449, 1992.

[17] K. L. Gearing, M. Gottlicher, M. Teboul, E. Widmark, and J.-A. Gustafsson, "Interaction of the peroxisome-proliferator-activated receptor and retinoid X receptor," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 90, no. 4, pp. 1440–1444, 1993.

[18] C. Thaller and G. Eichele, "Identification and spatial distribution of retinoids in the developing chick limb bud," *Nature*, vol. 327, no. 6123, pp. 625–628, 1987.

[19] S. Keidel, M. Szardenings, and W. H. Mueller, "In vivo biological activity of retinoids partially correlates to their affinity to recombinant retinoic-acid receptor α and recombinant-cellular retinoic-acid-binding protein I," *European Journal of Biochemistry*, vol. 212, no. 1, pp. 13–26, 1993.

[20] D. J. Mangelsdorf, U. Borgmeyer, R. A. Heyman et al., "Characterization of three RXR genes that mediate the action of 9-cis retinoic acid," *Genes and Development*, vol. 6, no. 3, pp. 329–344, 1992.

[21] M. A. Kane, "Analysis, occurrence, and function of 9-cis-retinoic acid," *Biochimica et Biophysica Acta. Molecular and Cell Biology of Lipids*, vol. 1821, no. 1, pp. 10–20, 2012.

[22] M. F. Boehm, L. Zhang, L. Zhi et al., "Design and synthesis of potent retinoid X receptor selective ligands that induce apoptosis in leukemia cells," *Journal of Medicinal Chemistry*, vol. 38, no. 16, pp. 3146–3155, 1995.

[23] M. F. Boehm, L. Zhang, B. A. Badea et al., "Synthesis and structure-activity relationships of novel retinoid X receptor-selective retinoids," *Journal of Medicinal Chemistry*, vol. 37, no. 18, pp. 2930–2941, 1994.

[24] E. Pérez, W. Bourguet, H. Gronemeyer, and A. R. De Lera, "Modulation of RXR function through ligand design," *Biochimica et Biophysica Acta. Molecular and Cell Biology of Lipids*, vol. 1821, no. 1, pp. 57–69, 2012.

[25] R. Lotan, "Retinoids and apoptosis: implications for cancer chemoprevention and therapy," *Journal of the National Cancer Institute*, vol. 87, no. 22, pp. 1655–1657, 1995.

[26] N. Clarke, P. Germain, L. Altucci, and H. Gronemeyer, "Retinoids: potential in cancer prevention and therapy," *Expert Reviews in Molecular Medicine*, vol. 6, no. 25, pp. 1–23, 2004.

[27] P. Shankaranarayanan, A. Rossin, H. Khanwalkar et al., "Growth factor-antagonized rexinoid apoptosis involves permissive PPAR gamma/RXR heterodimers to activate the intrinsic death pathway by NO," *Cancer Cell*, vol. 16, no. 3, pp. 220–231, 2009.

[28] S. P. Mahamuni, R. D. Khose, F. Menaa, and S. L. Badole, "Therapeutic approaches to drug targets in hyperlipidemia," *BioMedicine*, vol. 2, no. 4, pp. 137–146, 2012.

[29] C.-H. Wang, W.-D. Lin, D.-T. Bau et al., "Appearance of acanthosis nigricans may precede obesity: an involvement of the insulin/IGF receptor signaling pathway," *BioMedicine*, vol. 3, no. 2, pp. 82–87, 2013.

[30] T. Yamamoto, W.-C. Hung, T. Takano, and A. Nishiyama, "Genetic nature and virulence of community-associated methicillin-resistant Staphylococcus aureus," *BioMedicine*, vol. 3, no. 1, pp. 2–18, 2013.

[31] Y.-M. Chang, B. K. Velmurugan, W. W. Kuo et al., "Inhibitory effect of alpinate Oxyphyllae fructus extracts on Ang II-induced cardiac pathological remodeling-related pathways in H9c2 cardiomyoblast cells," *BioMedicine*, vol. 3, no. 4, pp. 148–152, 2013.

[32] Y. M. Leung, K. L. Wong, S. W. Chen et al., "Down-regulation of voltage-gated Ca2+ channels in Ca2+ store-depleted rat insulinoma RINm5F cells," *BioMedicine*, vol. 3, no. 3, pp. 130–139, 2013.

[33] I. C. Chou, W.-D. Lin, and C.-H. Wang, "Association analysis between Tourette's syndrome and two dopamine genes (DAT1, DBH) in Taiwanese children," *BioMedicine*, vol. 3, no. 2, pp. 88–91, 2013.

[34] H.-J. Huang, K.-J. Lee, H. W. Yu, H.-Y. Chen, F.-J. Tsai, and C. Y.-C. Chen, "A novel strategy for designing the selective PPAR agonist by the "Sum of activity" model," *Journal of Biomolecular Structure & Dynamics*, vol. 28, no. 2, pp. 187–200, 2010.

[35] P.-C. Chang, J.-D. Wang, M.-M. Lee et al., "Lose weight with traditional Chinese medicine? Potential suppression of fat mass and obesity-associated protein," *Journal of Biomolecular Structure & Dynamics*, vol. 29, no. 3, pp. 471–483, 2011.

[36] K. C. Chen, S. S. Chang, H. J. Huang et al., "Three-in-one agonists for PPAR-alpha, PPAR-gamma, and PPAR-delta from traditional Chinese medicine," *Journal of Biomolecular Structure & Dynamics*, vol. 30, no. 6, pp. 662–683, 2012.

[37] K.-C. Chen and C. Yu-Chian Chen, "Stroke prevention by traditional Chinese medicine? A genetic algorithm, support vector machine and molecular dynamics approach," *Soft Matter*, vol. 7, no. 8, pp. 4001–4008, 2011.

[38] K.-C. Chen, K.-W. Chang, H.-Y. Chen, and C. Y.-C. Chen, "Traditional Chinese medicine, a solution for reducing dual stroke risk factors at once?" *Molecular BioSystems*, vol. 7, no. 9, pp. 2711–2719, 2011.

[39] T.-T. Chang, K.-C. Chen, K.-W. Chang et al., "In silico pharmacology suggests ginger extracts may reduce stroke risks," *Molecular BioSystems*, vol. 7, no. 9, pp. 2702–2710, 2011.

[40] W. Ieongtou, S. S. Chang, D. Wu et al., "Molecular level activation insights from a NR2A/NR2B agonist," *Journal of Biomolecular Structure & Dynamics*, 2013.

[41] C.-Y. Chen and C. Y.-C. Chen, "Insights into designing the dual-targeted HER2/HSP90 inhibitors," *Journal of Molecular Graphics and Modelling*, vol. 29, no. 1, pp. 21–31, 2010.

[42] H.-J. Huang, K.-J. Lee, H. W. Yu et al., "Structure-based and ligand-based drug design for HER 2 receptor," *Journal of Biomolecular Structure & Dynamics*, vol. 28, no. 1, pp. 23–37, 2010.

[43] S.-C. Yang, S.-S. Chang, H.-Y. Chen, and C. Y.-C. Chen, "Identification of potent EGFR inhibitors from TCM Database@ Taiwan," *PLoS Computational Biology*, vol. 7, no. 10, Article ID e1002189, 2011.

[44] Y. A. Tsou, K. C. Chen, H. C. Lin, S. S. Chang, and C. Y. C. Chen, "Uroporphyrinogen decarboxylase as a potential target for specific components of traditional Chinese medicine: a virtual screening and molecular dynamics study," *PLoS ONE*, vol. 7, no. 11, Article ID e50087, 2012.

[45] Y. A. Tsou, K. C. Chen, S. S. Chang, Y. R. Wen, and C. Y. Chen, "A possible strategy against head and neck cancer: in silico investigation of three-in-one inhibitors," *Journal of Biomolecular Structure & Dynamics*, vol. 31, no. 12, pp. 1358–1369, 2013.

[46] C.-Y. Chen, Y.-H. Chang, D.-T. Bau et al., "Ligand-based dual target drug design for H1N1: swine flu—a preliminary first study," *Journal of Biomolecular Structure & Dynamics*, vol. 27, no. 2, pp. 171–178, 2009.

[47] C.-H. Lin, T.-T. Chang, M.-F. Sun et al., "Potent inhibitor design against H1N1 swine influenza: structure-based and molecular dynamics analysis for M2 inhibitors from traditional Chinese medicine database," *Journal of Biomolecular Structure & Dynamics*, vol. 28, no. 4, pp. 471–482, 2011.

[48] T.-T. Chang, M.-F. Sun, H.-Y. Chen et al., "Screening from the world's largest TCM database against H1N1 virus," *Journal of Biomolecular Structure & Dynamics*, vol. 28, no. 5, pp. 773–786, 2011.

[49] S.-S. Chang, H.-J. Huang, and C. Y.-C. Chen, "Two birds with one stone? Possible dual-targeting H1N1 inhibitors from traditional Chinese medicine," *PLoS Computational Biology*, vol. 7, no. 12, Article ID e1002315, 2011.

[50] H. J. Huang, Y. R. Jian, and C. Y. Chen, "Traditional Chinese medicine application in HIV: an in silico study," *Journal of Biomolecular Structure & Dynamics*, vol. 32, no. 1, pp. 1–12, 2014.

[51] K. C. Chen, S. S. Chang, F. J. Tsai, and C. Y. Chen, "Han ethnicity-specific type 2 diabetic treatment from traditional Chinese medicine?" *Journal of Biomolecular Structure & Dynamics*, vol. 31, no. 11, pp. 1219–1235, 2013.

[52] K.-C. Chen, M.-F. Sun, S.-C. Yang et al., "Investigation into potent inflammation inhibitors from traditional chinese medicine," *Chemical Biology and Drug Design*, vol. 78, no. 4, pp. 679–688, 2011.

[53] W. I. Tou, S. S. Chang, C. C. Lee, and C. Y. Chen, "Drug design for neuropathic pain regulation from traditional Chinese medicine," *Scientific Reports*, vol. 3, p. 844, 2013.

[54] K. C. Chen, Y. R. Jian, M. F. Sun et al., "Investigation of silent information regulator 1 (Sirt1) agonists from Traditional Chinese Medicine," *Journal of Biomolecular Structure & Dynamics*, vol. 31, no. 11, pp. 1207–1218, 2013.

[55] W. I. Tou and C. Y. Chen, "May disordered protein cause serious drug side effect?" *Drug Discovery Today*, 2013.

[56] C. Y. Chen and W. I. Tou, "How to design a drug for the disordered proteins?" *Drug Discovery Today*, vol. 18, no. 19-20, pp. 910–915, 2013.

[57] V. Chandra, P. Huang, Y. Hamuro et al., "Structure of the intact PPAR-γ-RXR-α nuclear receptor complex on DNA," *Nature*, vol. 456, no. 7220, pp. 350–356, 2008.

[58] B. R. Brooks, R. E. Bruccoleri, B. D. Olafson et al., "CHARMM: a program for macromolecular energy minimization and dynamics calculations," *Journal of Computational Chemistry*, vol. 4, pp. 187–217, 1983.

[59] C. Y.-C. Chen, "TCM Database@Taiwan: the world's largest traditional Chinese medicine database for drug screening In Silico," *PLoS ONE*, vol. 6, no. 1, Article ID e15939, 2011.

[60] C. A. Lipinski, F. Lombardo, B. W. Dominy, and P. J. Feeney, "Experimental and computational approaches to estimate solubility and permeability in drug discovery and development settings," *Advanced Drug Delivery Reviews*, vol. 46, no. 1–3, pp. 3–26, 2001.

[61] B. Xue, R. L. Dunbrack, R. W. Williams, A. K. Dunker, and V. N. Uversky, "PONDR-FIT: a meta-predictor of intrinsically disordered amino acids," *Biochimica et Biophysica Acta. Proteins and Proteomics*, vol. 1804, no. 4, pp. 996–1010, 2010.

[62] C. M. Venkatachalam, X. Jiang, T. Oldfield, and M. Waldman, "LigandFit: a novel method for the shape-directed rapid docking of ligands to protein active sites," *Journal of Molecular Graphics and Modelling*, vol. 21, no. 4, pp. 289–307, 2003.

[63] B. Hess, C. Kutzner, D. Van Der Spoel, and E. Lindahl, "GRGMACS 4: algorithms for highly efficient, load-balanced, and scalable molecular simulation," *Journal of Chemical Theory and Computation*, vol. 4, no. 3, pp. 435–447, 2008.

[64] V. Zoete, M. A. Cuendet, A. Grosdidier, and O. Michielin, "SwissParam: a fast force field generation tool for small organic molecules," *Journal of Computational Chemistry*, vol. 32, no. 11, pp. 2359–2368, 2011.

[65] R. Fletcher, *Optimization*, Academic Press, London, UK, 1969.