

**David F. Meaney<sup>1</sup>**

Departments of Bioengineering  
and Neurosurgery,  
University of Pennsylvania,  
Philadelphia, PA 19104-6392  
e-mail: dmeaney@seas.upenn.edu

**Barclay Morrison**

Department of Biomedical Engineering,  
Columbia University,  
New York, NY 10027

**Cameron Dale Bass**

Department of Biomedical Engineering,  
Duke University,  
Durham, NC 27708-0281

# The Mechanics of Traumatic Brain Injury: A Review of What We Know and What We Need to Know for Reducing Its Societal Burden

*Traumatic brain injury (TBI) is a significant public health problem, on pace to become the third leading cause of death worldwide by 2020. Moreover, emerging evidence linking repeated mild traumatic brain injury to long-term neurodegenerative disorders points out that TBI can be both an acute disorder and a chronic disease. We are at an important transition point in our understanding of TBI, as past work has generated significant advances in better protecting us against some forms of moderate and severe TBI. However, we still lack a clear understanding of how to study milder forms of injury, such as concussion, or new forms of TBI that can occur from primary blast loading. In this review, we highlight the major advances made in understanding the biomechanical basis of TBI. We point out opportunities to generate significant new advances in our understanding of TBI biomechanics, especially as it appears across the molecular, cellular, and whole organ scale. [DOI: 10.1115/1.4026364]*

## Introduction

Traumatic brain injury (TBI) presents a significant public challenge in today's society. Approximately  $1.7 \times 10^6$  people in the U.S. each year suffer some form of TBI that requires at least a visit to the hospital [1]. The relative fraction of severe TBI patients—estimated as 3% of the total annual cases—requires the most intensive medical care and is often complicated by injuries to other body regions. Moderate brain injury represents 22% of the annual incidence and commonly leaves patients with persistent deficits that they carry for the remainder of life. Mild TBI has the highest incidence rate in the population (75% of total injuries) and may be even higher than reported in epidemiological studies, owing to substantial underreporting in young populations, athletes, and members of the military [2,3]. In aggregate, the socioeconomic toll is significant—TBI remains the most prevalent cause of death in adults aged less than 45 years and is also the highest cause of long-term disability [4,5]. As the population ages, we are also seeing the relative incidence rise in the elderly population, where it is now only second to cancer as a cause of death in people aged 65 and over [4]. With the growing awareness of TBI in both the civilian and military population, TBI is no longer a silent epidemic.

With the longstanding knowledge of neurodegenerative changes in boxers now expanding to include repeated TBI in other professional athletes and soldiers [6–8], it is also clear that TBI is both an acute disorder and, for some patients, a chronic neurological disease. This awareness, however, also exposes the need for far more clarification on many issues in the scientific literature because it brings to light many questions that center on defining the exposure for possible at-risk populations that include civilians (e.g., athletes in contact sports) and members of the military. As we answer these questions, we will have the opportunity to use

some of the same tools we describe below to tackle the looming challenge of mitigating the chronic effects of trauma to the brain.

The focus of this review is to provide a summary of past efforts to understand key points in the TBI neurophysiological injury cascades, which include:

- defining the environments in which these injuries occur
- understanding how mechanical loads are transferred to the brain structures during the loading conditions that occur in these environments
- using the mechanical, physiological, and pathophysiological response of the brain—at multiple length scales—to identify the critical mechanisms for damage, including necessary interspecies scaling between animal models and humans
- identifying the key acute mechanisms of injury that cause the most significant functional impairments, and testing if these primary mechanisms contribute to the long-term changes associated with injury

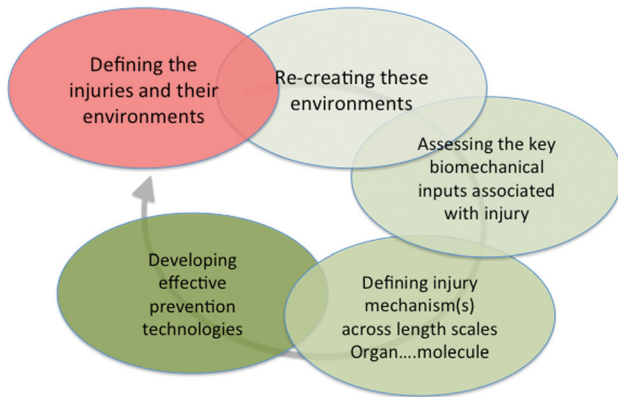
We present both past and ongoing work addressing these central questions. We also identify several areas that need more study. Several excellent reviews provide some historical context and additional information on efforts across the length scale; we refer the reader to these articles for further detail [9–14]. We structure the review by (a) reviewing the existing knowledge to define when traumatic brain injuries occur in the civilian and military environments and (b) the separate work identifying how immediate mechanism(s) of injury mediate acute impairments after injury.

## An Integrated, Multiscale Approach for Understanding Traumatic Injury to the Brain

Drawing on several decades of research with applications primarily in producing protective headgear, developing head protection standards, and designing of safety systems in motor vehicles, there is a considerable literature that has defined the scenarios causing TBI. The work spans multiple scales and forms a natural research cycle (Fig. 1)—from population-based surveys

<sup>1</sup>Corresponding author.

Contributed by the Bioengineering Division of ASME for publication in the JOURNAL OF BIOMECHANICAL ENGINEERING. Manuscript received September 20, 2013; final manuscript received December 20, 2013; accepted manuscript posted December 27, 2013; published online February 5, 2014. Editor: Victor H. Barocas.



**Fig. 1 The research cycle of reducing the societal burden of traumatic brain injury. Epidemiological evidence collected from clinical studies, and analysis of motor vehicles crashes, forms part of the first tier for defining where the most significant brain injuries occur and if these injuries change over time (red). The work transitions to the research laboratory (green) for defining how these injuries occur, establishing key relationships between the physical inputs in these environments and their resulting injuries. The inevitable translation of this new knowledge into the next generation of protection technologies completes the cycle and also triggers the next research cycle for focusing efforts on the most significant injuries in the population. One broad research cycle has already occurred for moderate and severe brain injuries, resulting in advances in helmet protection technologies and passive safety systems. Emerging efforts have now shifted to include more focus on mild TBI, which occurs across both the civilian and military population.**

identifying significant injuries [15–17] down to tissue/single cell-based work for detecting key molecular signatures of the injury [18–20]. The work begins with the clinical environment to define injury incidence, transitions to the laboratory environment to replicate and study both injury mechanisms and tolerance, and proceeds to the translational environment for developing effective countermeasures to reduce or eliminate injury incidence. A central component in this past work is using experimental and analytical tools to draw relationships between the physical input (acceleration, impact force, duration, etc.), the resulting mechanical response of the brain/skull, and to integrate thresholds for damage to the brain and its coverings to identify the mechanical loading scenarios most often associated with injury. Ongoing work to study blast-induced traumatic brain injury is following a similar trajectory, although the tolerance criteria for blast loading are in the early developmental stages [21–23]. A defining characteristic of this research cycle is to address the most significant injuries occurring in the population, to implement new technologies for reducing the incidence of these injuries, and generate new surveys of the population for focusing future research efforts.

In general, the primary mechanical response of the brain to either impact, impulsive, or blast loading is driven on the macro-scale by the brain/skull geometry, the partitioning of the brain within the skull, and the material properties of the cranial tissues. Although general principles of the brain response to mechanical loading can be developed, an important caveat is that every traumatic brain injury occurs under unique mechanical loading conditions. Perhaps one of the greatest challenges is to consider how the unique mechanical inputs associated with each injury can be coalesced into a single, universal approach for determining when injuries occur in the population.

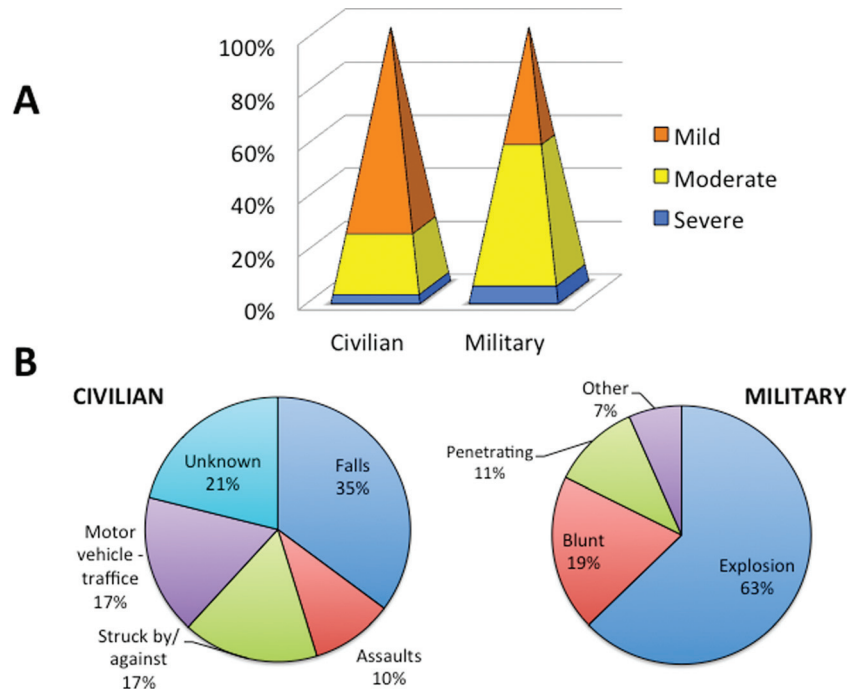
To date, the most complete approach for predicting the incidence of TBI in humans requires several key steps: (1) defining the external mechanical loads experienced by the head during situations that cause injury, (2) using models of the brain (either physical, analytical, or computational) to estimate how these external mechanical loads transfer to mechanical conditions (e.g.,

stress, strain, etc.) in the brain at the tissue and cellular scale, and (3) using tissue and cellular tolerance criteria to determine the regions of the brain that will be injured or impaired as a result of the external applied loading. In the remainder of this paper, we will review the work in support of this approach and discuss emerging areas of research that will significantly extend the ability to predict TBI incidence in the future.

**Characterizing the Causal Environments of TBI.** Historically, motor vehicle crashes were a primary environmental focus for TBIs because they consistently ranked as the most frequent cause of TBI-related deaths in civilians [24,25]. However, a broader view of TBI across the severity spectrum (Fig. 2(a)) shows that falls are the leading cause of emergency department visits and hospitalization stays related to TBI; the second leading cause is where the individual is struck by or strikes another object [1]. For visits that require either hospitalization or only an emergency department visit, motor vehicles are the third leading cause of injury [1,2]. The loading scenarios for head impacts occurring in the automotive environment are well developed and embedded into vehicle safety testing protocols used in the U.S. and elsewhere [26]. These testing procedures specify collision speeds and impact directions that are linked to common accident scenarios associated with death and disability in motor vehicle crashes. As the focus of the field shifts to more moderate and mild TBIs, we must continue to expand our test conditions and include more scenarios associated with the injuries that occur in nonvehicular environments (Fig. 2(b)) [27]. In particular, concussions in sports, including youth, need better biomechanical testing scenarios.

Recent evidence of a large risk of mild/moderate TBI in military scenarios associated with blasts (bTBI; Fig. 2(a)) [22,28,29] (i.e., blast-induced traumatic brain injury (bTBI)) lacks a comparable level of epidemiological detail. For example, recent work shows many of the scenarios causing TBI in the military are from the motor vehicle crashes, falls, and the head striking another object, similar to the civilian population (e.g., Ref. [30]). Of course, the initiating event for injury is different from the civilian population: ~67% of the TBI injuries requiring hospitalization in U.S. military operations in Iraq and Afghanistan were from explosions (Fig. 2(b)), with direct blunt trauma contributing ~19% and penetrating injuries contributing ~11% of the injuries. Even within the injuries attributable to explosions, many are linked with low rate blunt trauma following the blast event [31].

Although this may initially downplay the importance of primary blast in military TBI, there is other evidence showing a substantial number of injuries occur with direct blast exposure, including primary blast exposure of dismounted service members (Fig. 2(b)) [31,32]. Recent evidence suggests that civilians exposed to large blasts also have the potential for sustaining bTBI without pulmonary injury [22,33]. Together, these reports demonstrate that our perspective on the mechanisms of TBI in the military population is still evolving. Some confusion on the relative role of blast-induced traumatic brain injury may arise because the classification system for bTBI was designed to encompass any physical phenomena that could cause brain injury [34]. Primary blast injury, defined as the damage occurring as the blast wave travels through the brain, is unique to bTBI. However, the secondary and tertiary forms of bTBI—in which the injury is caused by direct laceration of the brain from fragments or shrapnel (secondary) or the head moves suddenly and may strike another object (tertiary)—shares a common mechanistic base with injuries observed in the civilian population. Therefore, mechanisms of blast-induced TBI may have a mixture of mechanisms from primary, secondary, and tertiary blast injury. Unknown, though, is how effects of the primary blast wave interact with the injury mechanisms caused by the secondary and tertiary phases.



**Fig. 2 (a)** The relative incidence of TBI in the civilian and military population, and their causes. Excluding penetrating TBI and unclassified injuries, the relative incidence rates for the military and civilian population appear distinct. However, the possible underreporting of mild TBI in the military may alter the relative incidence rates significantly. **(b)** Within each population, the causes of TBI span a broad range. Primary blast TBI is unique to the military environment.

**Defining the Injuries in the Clinical Context.** From a clinical standpoint, brain injuries are often categorized as either focal or diffuse [35]. These general descriptions apply for both the civilian and military environment. Focal injuries are readily visible using standard imaging techniques (CT; MR). Primary vascular injuries that cause bleeding within the brain (intracerebral hematomas; tissue tears), on the surface of the brain (acute subdural hematoma; subarachnoid hemorrhage; extradural hematoma), or in the cortical gray matter (cerebral contusion) are common examples of focal brain injuries that appear in the severely and moderately head-injured population. With the exception of subarachnoid hemorrhage, these focal injuries do not appear in the mild TBI population. Moreover, the number of mild TBI cases with subarachnoid hemorrhage is rare. Therefore, diffuse injuries are considered the predominant category of injury in mild TBI.

Diffuse brain injuries are, as the name implies, not localized to one area of the brain but are more distributed throughout the brain. Diffuse brain swelling is one form of injury that can appear over time following the injury and is not often the focus of studies for predicting how the mechanical forces can cause subsequent injury throughout the brain. An ongoing discussion in the bTBI literature suggests that diffuse brain swelling may be a more common component of the injury pattern after bTBI [36,37].

The most common diffuse brain injury receiving attention for TBI biomechanics is diffuse axonal injury (DAI), which is the appearance of axonal injury at the microscopic scale in selected regions of the brain [38]. The mechanisms and progressive changes to the cytoskeleton, organelles and membrane within the axonal compartment is an ongoing area of study with DAI, as this would point towards possible therapeutic intervention [39]. Strictly defined as an entity that appears in humans, DAI is often the subject of study in mild traumatic brain injury (mTBI) patients because of the widespread disruption of brain networks that can appear in these patients without any other sign of brain damage. The continuum of DAI in humans is well described, and the general conditions that cause DAI in the human are providing a

template for studying these same types of injuries in animal models. There is some evidence of diffuse injury to axons in primary blast TBI models [21,22,40,41], but a complete description of DAI in human primary blast TBI is not yet available. The closest demonstration of DAI in blast TBI shows alterations in directional diffusion within white matter measured by diffusion tensor imaging [42], changes that are presumed to reflect areas of microscopic axonal injury based on earlier work in animals [43,44]. However, the distribution of DAI patterns in humans following blast exposure, similar to the definition of DAI in human patients after falls, assaults, and motor vehicle accidents [45], would help shape future biomechanical studies to understand primary blast TBI more completely.

**Emerging Key Issues.** With the dramatic change in the passive safety technologies for motor vehicles, there is a shift in the distribution of the types of specific brain injuries observed in the moderate and severely brain injured population [46,47]. The most extensive clinical study that detailed the distribution of severe head injuries is over three decades old [15], and an updated set of data would point toward specific populations that are potentially at risk for different types of brain injuries, and also different scenarios leading to these injuries. Perhaps most significantly, the distribution of these injuries in the military population is extremely vital as it can shape priorities for the next decade, yet this distribution is not completely defined. Moreover, there is a well-recognized paucity of assessment techniques for mild TBI that achieve both good sensitivity and specificity over the time course of TBI. Without these specific and sensitive measures to record the accurate incidence of mild TBI, the overall fraction of mild, moderate, and severe TBI in either the civilian or military population remains to be defined.

For military injuries associated with blast, there are potentially large numbers of exposed personnel, especially for mild bTBI. It is unclear for mild severities how to differentiate between bTBI



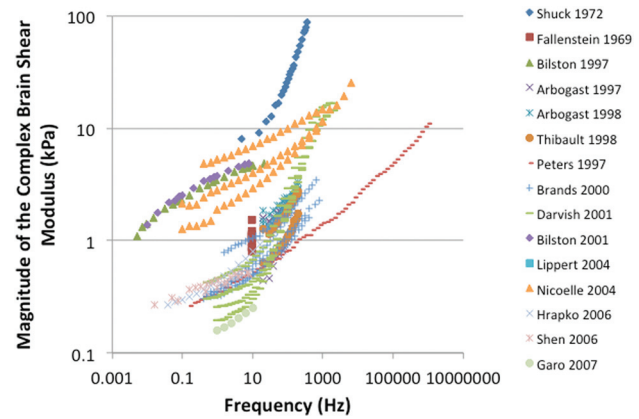
and common, potentially comorbid, syndromes such as posttraumatic stress syndrome (PTSD) with similar symptomology [48].

**Estimating the Primary Mechanical Response: Modeling the Structure.** Knowing the accurate size and shape of the brain, the brain deformation, and the brain movement relative to the skull during an exposure is a critical factor in developing an ability to accurately predict when injuries will occur. The human brain varies in size across the population, across age, and across gender. In particular, the brain size decreases over the latter part of the lifespan and, therefore, the subdural space may increase to increase the risk of TBI with age [49]. The variation in size and shape alone means there is a broad range in the primary brain mechanical response to blunt, acceleration-based, or blast loading. Although there is a well-developed anatomical description of the human brain, less clear is how these anatomical regions vary among individuals. Recently, magnetic resonance imaging (MRI) technologies make it possible to explore the variation in size, shape, and organization of the brain across any desired population, especially using imaging techniques without ionizing radiation (i.e., MRI). Alternatively, existing public data sets<sup>2</sup> provide high-resolution images that can be used for any subsequent biomechanical examination.

Perhaps the most important new element to consider in the brain structure is its organization across regions, i.e., the connectome. Imaging technologies to examine the direction of white matter tracts, in combination with techniques to measure local blood flow changes in the brain, provides a way to connect brain regions and assess how the brain connection map changes following TBI [50]. Public data sets of the human connectome are available<sup>3</sup>. This technology presents interesting opportunities for understanding the biomechanics of TBI, as it could provide insight into the regions of the brain damaged in mild TBI patients. Not only will we need to consider the distribution of stress and strain throughout the brain during an impact/blast exposure, we will also need to predict if these patterns of acute changes in the brain may be influenced greatly by the initial connection map in the brain at the moment of injury [50]. Therefore, understanding the evolving relationship between functional networks and structure is critical to making progress in understanding neurotrauma.

The physical properties of the brain tissue within the cranial vault are a critical determinant of how the brain moves and deforms during impact. Some physical properties are either well characterized and do not appear to vary across the population [51] or they likely do not contribute to the motion of the brain during blast/acceleration/impact conditions. The most critical factors contributing to the mechanical response are the mechanical properties of skull, brain parenchyma, brain coverings, and the supporting vessels. Estimates of the scalp properties stem back several decades and include estimates for the failure limits during impact [52]. The mechanical impedance of the scalp to an incoming blast wave, though, is not defined. Similarly, the mechanical properties of dura are also known [53–57] but the dynamic properties are not well described, especially for high rate loading. Although there is some discussion on the relative importance of the cerebrovascular network providing mechanical integrity to the brain [58], only the mechanical properties and failure limits of the parasagittal bridging veins are known [58–63].

Brain tissue, by far, is the most extensively characterized of these tissues, but unfortunately has large unresolved differences among reported values (overview of the range in material properties shown in Fig. 3). Early work on brain tissue stiffness showed that it was primarily elastic and nearly incompressible under cyclic, dilatational loading up to 100 kHz. Although initial estimates of shear properties revealed a moderately compliant, viscoelastic material with a complex modulus of  $\sim 20$  kPa at loading



**Fig. 3 Large variance in reported white matter and brain material properties by study. Early work estimated both bulk and shear modulus. In the past two decades, work has shown that brain is one of the softest biological tissues, more than ten times more compliant than the earliest measurements.**

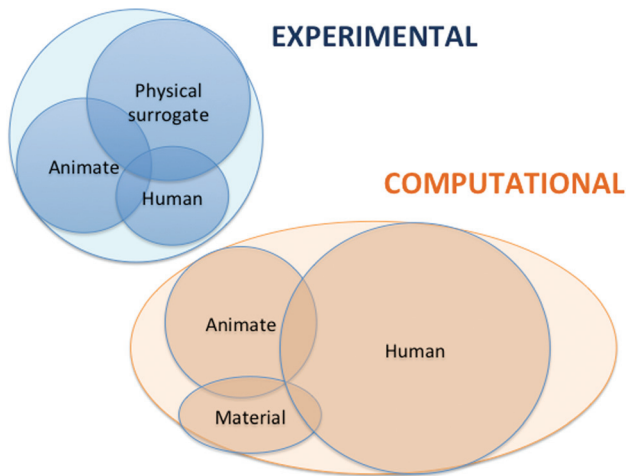
frequencies up to 120 Hz [64,65], these studies are now replaced with a larger complement of studies that demonstrates brain is one of the softest biological solid tissues measured (complex modulus  $\sim 3$ – $2$  kPa), can be nonlinear viscoelastic, and varies across species (recent reviews: [66,67]). A remarkable characteristic is that the brain material softens at finite strains and that this softening response is repeatable across many consecutive loading cycles. Less clear, though, are the properties of brain, including the criteria for functional failure of brain tissue, at the much higher strain rates associated with blast loading [68,69]. Measures of bulk elastic properties at ultrasonic frequencies produce estimates that are nearly 1000x stiffer than shear properties at much lower loading frequencies, and the stiffness of the brain at these loading rates is under ongoing examination [70]. Where possible, direct measures of brain material properties in vivo are complementing past studies. For example, recent results using brain MR elastography provides estimates of the changes that occur in vivo and are in the range of properties derived from previous in vivo and in situ measurements [11,71–75].

In selected studies, the regional and local anisotropic variation in brain material properties was examined [76–81]. At small strains, the relative stiffness of highly oriented brain stem samples showed modest anisotropy [81]. Gray matter properties show less directional dependence but more heterogeneity than previously appreciated, even within anatomical structures with several fold differences at large strains [82]. Moreover, white matter and gray matter show some significant differences in their relative shear properties, although these changes are within twofold to threefold. The regional properties for blast loading conditions are virtually absent from the literature, although it remains an active area of study. In work motivated by blast-TBI, a key concern is how to measure these material properties under high strain rate conditions. Hopkinson bar-based methods are now scaled to examine shear properties at very high rates, but the soft nature of brain tissue makes this a very challenging set of experiments [83–85]. Though values are reported at high strain levels (to 50% engineering strain, an overwhelmingly destructive loading condition), the experiments lack sufficient resolution to estimate the response at more realistic strain levels associated with primary blast injury ( $\sim 1\%$  strain).

The development of macroscopic material properties, combined with recent advances in computer modeling capabilities and a desire to know which components of the brain are injured in response to a macroscopic loading conditions, now provide an opportunity to develop more precise material models of the brain that reflect both the underlying cellular structure of the material and the unique macroscopic material behavior. Anatomical descriptions of

<sup>2</sup>Allen Brain Atlas: <http://www.brain-map.org/>; Visible Human Project: <http://www.nlm.nih.gov/research/visible/>

<sup>3</sup><http://humanconnectome.org>



**Fig. 4 Multimodal modeling approaches for defining the structural response of the brain to applied mechanical loading. Historically, experimental approaches led to insight into the most important types of mechanical loading associated with severe brain injuries. These experimental approaches span both human and animate models and use physical surrogates to complement either scale. The most significant development in the past decade is the growth of computational approaches to examine the biomechanics of TBI in both experimental models and humans. However, the need to validate these models for numerical issues (e.g., mesh convergence, mesh quality) as well as biofidelic output is even higher given their increased complexity and proliferation.**

brain tissue are easily obtained using standard histopathological techniques. In some cases, existing material models gleaned from the composite materials community can help assist in interpreting anisotropic material behavior [79,80]. Some models of the white matter already suggest how different cell types may couple to each other and propose how structural elements of the tissue interact in complex manner to expose a subpopulation of axons to high mechanical loading [86–90]. Continuum-based nonlinear material descriptions are also available to correlate with these structurally-based material models [87,89]. With their continuum formulation, the integration of these models into existing finite element software packages is possible. Currently, though, there are no efforts to model the complex mixture of cell types found in other areas of the brain (e.g., hippocampus) and the relative change in the macroscopic/microscopic transformation that underlies any injury patterns that occur in these areas. A recent set of measurements show that material properties of the hippocampus *in situ* differ significantly, yet the cellular mechanism(s) for this difference is not clear [76–78]. Potential reasons for this difference include a change in the relative balance of excitatory and inhibitory neurons, each of which possess a different morphology that could contribute to a difference in the kinematics of deformation at the cellular level. Additionally, this difference may also reflect a change in the neuron/glial ratio among the regions, given the material properties of these two cell types are distinct. An inability to track different cell types in living tissue over the microseconds to milliseconds time scale remains a major technical impediment to validating any structure-based model of the brain.

*Emerging Key Issues.* In comparison to our current knowledge of the geometry and the multiscale structure of the brain, the material properties are better known but vary in their magnitude over a broad range (Fig. 4). The consistent use of these material property measures, and not stiffer values from very early studies, remains problematic. In addition, the specific material properties in some, but not all, brain regions are known and the expansion of this data set to include more brain regions and more anisotropic properties would significantly help in the predictive ability of

models. Extending these properties to blast loading conditions must be done, but we already have information that can be applied to both the secondary and tertiary forms of bTBI. The bulk dilatational response as the primary blast wave transmits through the tissue is key, but may already be described with early work in the field [52]. The associated deviatoric response under the high loading rates associated with blast, however, is not fully described. More insightful findings could come from studying the underlying role of the vasculature in contributing towards the macroscopic material properties of the brain conditions across the loading spectrum; early works suggest intriguing changes that could map the macroscopic response to the underlying structural failure of the vascular elements [58]. Additionally, there is evidence that material properties of individual cell types within the brain are distinct [91–96], and these variations may contribute greatly to our understanding of cell types within specific brain regions that would be mechanically vulnerable. Extending the structural descriptions of brain tissue to reflect the transmission of the macroscopic mechanical input during blast loading may provide unique insights into how the extremely rapid events during blast extend to the cellular scale. Moreover, the heterogeneity of the macroscopic to microscopic transformation will inevitably extend to the subcellular scale—e.g., do synapses from the same neuron (or same dendrite from within a neuron) show the same local deformation that is applied macroscopically? Undoubtedly changes at the synaptic scale will be important in decoding how the network function can be compromised after injury.

#### **Estimating the Primary Mechanical Response: Role of Physical Models, Human Surrogate Models, and Computational Models.**

With a primary goal of linking an external mechanical input to injury patterns within the brain, investigators have commonly used tools from both experimental and computational mechanics (Fig. 4). With such a complex geometry, though, it is not surprising that the earliest efforts to achieve this overall goal were experimentally based. Seminal studies by Holbourn demonstrated the value of using simple photoelastic materials to illuminate areas of a brain surrogate that experience high shear strains during rapid rotational motions [97]. Holbourn’s models highlighted that cortical regions were most vulnerable to injury when the head was rapidly rotated about the sagittal plane (anterior-posterior head motion), while structures deeper within with the brain were more vulnerable with rotations along the coronal plane. Subsequent studies using similar technologies highlighted how high stresses can also appear at the craniocervical junction [98]. Direct visualization of grid patterns or embedded markers within a transparent silicone gel also provided direct evidence for the unique patterns of deformations that occur with accelerations imposed in different directions and the influence that different skull/gel boundary conditions and ventricular structures have on intracranial strains, showing that the ventricles can redistribute and, in some regions, reduce the strains appearing within the brain after impact [99–102]. In some instances, these models have been used to assess the effectiveness of different animal models to recreate the deformation patterns that appear during impact and have led to a redesign of animal models to produce deformation patterns that more closely resemble the strains within the hemispheres during injury [103–105]. These same techniques are now extended to blast loading conditions [102–109], where the efforts will yield significant information on the manner that external blast waves transfer to the brain simulant, how these pressures are distributed throughout the surrogate, and how these pressures dissipate over time. Although providing a direct window into the possible response of the brain to any external mechanical loading condition, it is worth noting that the highly elastic material properties of brain tissue surrogates will need to be considered in extending or interpreting these results for the viscoelastic, nonlinear brain tissue.

Some of the disadvantages associated with physical models of the brain within the skull are mitigated with human surrogate

tests. By using cadaveric material with minimal material degradation, neutrally buoyant markers placed within the brain, and high speed X-ray imaging technologies, one can track motions at any point within the brain during an impact event [110]. Depending on the location of the markers within the brain, these data can provide direct measures of the relative motion between the cortical surface and the skull, the relative motion within the deep white matter, and the differential motion across the hemispheres [110–116]. Key results from these studies include the demonstration that the cortical surface of the brain can move relative to the inner skull surface, thereby creating conditions that can cause bridging vein rupture. Perhaps most significantly, Hardy and colleagues provided a well-characterized data set of neutrally buoyant markers in the brain that illustrate how many points in the brain move simultaneously during impact. In blast loading, these same approaches can provide estimates of how the blast wave transmits throughout the parenchyma. These data are critical in bridging the physical models with more realistic in situ measurements and linking computational models with the in vivo environment.

To date, the only direct visualization of living brain motion in humans has been accomplished with cyclical, noninjury motions. Using a well-developed MR technology to place magnetic “lines” within an anatomic plane, it is striking to see how much the brain can move under a simple, repeated rotation in the horizontal plane [117,118]. Estimates of the intracranial strains that appear during physiological head rotations in volunteers show the shear strains can reach 0.05 mm/mm, which is near the thresholds for axonal damage but at much lower deformation rates. Until this work appeared, it was not clear if there were any methods to cross-correlate the experimental database on tolerance criteria for the tissue with deformations that occur in vivo. These data provide yet more validation data for finite element models of the brain within the skull, especially since it is the only data available for the in vivo brain response. Although these data are not for injurious situations, they will also provide an opportunity to match skull/brain boundary conditions and assess the need for anatomic detail and proper material descriptions for the brain.

The experimental work across physical models, human surrogates, and human volunteers forms an extensive database of information for developing computational models of the brain response to impact or blast exposure. An excellent set of reviews details the early development of these models, which focused on modeling impact testing in human surrogates and developing estimates of tolerance under simple impact conditions [119,120]. These models quickly expanded into efforts for modeling the brain mechanical response under more complex conditions, and the need to accurately model the structure, material properties, and developing new material formulations for the soft brain material properties became evident. The extensive array of computational models developed in the past decade for the study of injury in the human brain is nothing short of remarkable [109,113,121–158].

Existing models can be roughly grouped into human and non-human species. Human-based models contain different level of complexity, depending on their purpose. A significant effort by the National Highway Traffic Safety Administration to build a finite element model of the human brain/skull structure aims to compute a solution to any impact condition within hours on a desktop computer [159]. For this reason, the geometry is less refined than other existing models. The reduction in complexity suits the long-term goal of the model, which is to provide a tool for evaluating brain injury specific risk in motor vehicle collisions and, ultimately, inclusion of this tool in assessing passenger car safety [157]. In comparison, models that use high resolution images and details of the anatomy require more solution time but offer more ability to interpret the predicted response and match the multiscale aspects of TBI. For example, accounting for the highly complex cortical gray matter and the underlying white matter structure yields insight into how the stress patterns can match the exact patterns of injury observed in animal models and

patients [158,160]. In general, the material properties used in these human-based models have migrated over the past decade into estimates more consistent with the soft material characteristics measured experimentally. A continuing effort to use accurate material properties in these models is challenging, as the resulting deformations can be large and the algorithms for computing the forces at interfaces must be monitored carefully. Moreover, it is well known that the soft material properties of the brain, when coupled with its nearly incompressible dilatational behavior, present significant computational challenges. Unanticipated mesh warping must be carefully considered to avoid error propagation in these models.

Investigators continue to use experimental data to validate the models, complementing early data showing the pressures during blunt impact with more recent data showing motions within the brain during impact. Currently, publicly available models show an increasing sophistication in their anatomical detail and their correlation with available validation data. In the past five years, these same models were extended to study blast exposure [105,109,124,128,131,134–136,138–140,143,144,157,161–164]. In many cases, though, the absence of validation data remains a key concern and must be addressed with each model before the models can be meaningfully used to correlate blast exposure with specific injury risk.

Given its importance as a linking process to accurately predict the incidence of TBI, the *process* of validation needs better definition. Although the intent of validation is centered on the goal of building virtual, computationally based models that accurately describe the human mechanical response to impact, the specific levels of validation for a model should always be considered in its use for predicting injury. For example, finite element models developed over two decades ago often used impact response data from human cadaveric testing conducted by Nahum and colleagues to validate the model. This validation process is best suited to evaluate the dilatational response of the finite element model and is, therefore, a key step in evaluating finite element models that use pressure as a metric for predicting injury. One could view this as an initial validation level in the model development process. However, injury mechanisms caused by deformation, and not pressure, would be more difficult to study with these models validated at this level because the deformation response can vary widely over a range of deviatoric material properties that would not significantly influence the pressure distribution in the brain during impact. The data on the displacements of points within the brain during impact or, alternatively, the strains within the living human brain during repeated, slower rotations form the basis for a second validation level that concentrates on matching the motion of the brain during impact. Given the relationship between the displacement and resulting deformations in the brain, models that achieve this validation level would improve the confidence that the model could be used to study injuries in the brain caused by different deformation mechanisms. However, there is no standardized performance specification for a model matching the data in this second validation level. Currently, the motion of several points within the brain and the comparison to model predictions leads to a more generalized statement on the performance of the model instead of a specific performance parameter. The clear addition of more experimental data on the movement of points within the brain under different impact conditions, directions, and with different-sized brains would significantly improve the process of validation at this second level. A third validation level to consider in the future would match patterns of damage observed in human surrogate studies with predictions of damage from the computational model. This validation level would test the accuracy of applying a model to predicting some injuries appearing in the moderate and severely head injured population. This validation level, much like the first two levels, would need a standardized scoring metric to assess the performance of the model. Similar to how performance test criteria for protective headgear resulted in a continuing improvement in performance



over decades of development, including this validation level would eventually ensure the improving correlation between computational models and the injury patterns they are designed to predict.

For nonhuman models, much of the work was an examination of existing experimental models of TBI. Species include rodents [109,128,135,138,140,143,144,161–167], pigs [168–173], and sheep [174–176]. These experimental models offered a measure that human physical models and human surrogate tests often could not provide—an estimate of the injuries that occurred throughout the brain as a result of the mechanical loading. Although some vascular injuries can be captured in human surrogate tests, much of the underlying pathobiology of TBI at the microscopic scale must be examined directly in the living brain. Similar to the human models, an extensive series of models appeared in the past decade to provide insight into the relative risk of injury to the living brain. Spatial descriptions of material properties may be necessary to explain the relative injury risk in the hippocampus that occurs commonly in rodent models of TBI [76,77,82]. Moreover, the distribution of stress and deformation is key for predicting areas of blood-brain barrier compromise, an injury often overlooked in biomechanical investigations [177]. These models also provide to opportunity to design new features into experimental models of TBI—e.g., the shape of an impounder tip or the speed of impact in the well described cortical impact model of TBI can significantly change the cortical lesion volume in this TBI model [177]. These models offer a direct translational path for studying blast exposure, and early results indicate that these models are transferrable when attention to details such as mesh size and material properties are made [22,164]. However, a systematic correlation of model results with the histopathology of injury is warranted to assess the efficacy of these models. Perhaps most importantly, these models lack the extensive model validation data sets that exist for the impact/acceleration-based models.

**Linking the Physical Response to the Biological Response: The Eventual Definition of Human Tolerance.** With efforts to identify the most common environments that cause TBI, the relevant mechanical loading scenarios that occur in these environments, and the subsequent physical response of the brain during these loading scenarios, one is faced with the next logical question—when will these conditions cause injury to the brain? And, relatedly, where will the injury occur? In some instances, the direct correlation between the mechanical input and the resulting pathobiological response will be made possible through a close comparison of the computational/physical model and the histopathological description of the injury pattern. However, the direct comparisons between model and injury patterns will provide little insight into the functional consequences of the injury patterns presented in any TBI. For example, although one may correlate strains at the tissue level to different levels of axonal damage at the microscopic level [178–182], an unanswered question is the threshold of damage needed to cause impairment of electrophysiological activity in the cell body, in the pathways connecting these circuits, or any alterations in circuit plasticity that would be the basis for impairments in learning and memory. Moreover, these correlations do not provide insight into the direct mechanisms of injury, an element that is critical for successful treatment.

**Simplifying the Physical Inputs of the Injury for in Vitro Study or “Reduced” in Vivo Models to Determine Injury Mechanisms.** With the clear need for coupling mechanical input into functional consequences, work in the past decade has responded and provided more direct insight into the mechanisms that cause the resulting functional changes. Motivated by the early work using physical models and finite element simulations, several investigators developed microscope-based systems to study directly the relationship between the mechanical deformation and resultant biochemical signaling [183–189]. As a result, we now

know that both neural and glial cells respond to mechanical deformation, that synaptically localized receptors are uniquely mechanosensitive, show immediate alterations in their physiological properties, and changes occur across both excitatory and inhibitory neurons [190–195]. At higher loading conditions, an additional mechanism of injury appears, which is the nonspecific, transient opening of pores within the membrane [183,186,196–202] after cellular deformation. In contrast to our knowledge on the effects of mechanical deformation on neural and glial cells of the central nervous system (CNS), the role of dynamic pressures in affecting cellular function is not well described. In nearly all cases of deformation-based mechanisms, the in vivo evidence matches the in vitro observations. New models to mimic only the blast wave transmission in cell cultures open up an entirely new opportunity for discovery in the blast loading regime in which several potential mechanisms of injury can be tested precisely with in vitro analogues [14,23,203–211].

Perhaps the most informative and relevant in vitro model for directly coupling mechanical inputs into brain tolerance and injury mechanisms will be the organotypic, in vitro models or the reduced in vivo models [18,23,212–219]. Organotypic brain cultures are sections of the brain isolated from the postnatal rodent brain and cultured over days to weeks. With the isolation from a living brain and without dissociation of the tissue common to other culturing methods, the in vivo architecture is well preserved in this model. Moreover, the combination of cell types within the brain is also maintained. Although organotypic cultures can be generated from different regions of the brain (cortex, thalamus, hippocampus, cerebellum), the most complete data for tolerance exists for the cortex and hippocampus [173,207,209,213,214,217]. Because these cultures are not vascularized, however, they do not provide an estimate of the selective change in the tolerance in cases where blood flow is compromised (ischemia; relative ischemia) or vascular damage occurs (blood-brain barrier breakdown; vasospasm). The use of in vitro models to study the effects of blast exposure is in its early stages, and estimates for blood-brain barrier opening, alterations in glial signaling, and the recovery of function are starting to appear in the literature [23,203]. A key issue that will need more clarification is the correlation of these loading conditions used in vitro to the loading environment in situ during blast.

Reduced in vivo models are the next most informative method for establishing links between input and resulting functional impairment. The optic nerve is a highly aligned cranial nerve that is part of the CNS, is accessible and can be injured directly to estimate thresholds for tissue tolerance [181,182]. Similarly, dorsal nerve roots are also accessible and provide a method to measure directly the electrophysiological impairment after tensile stretch, and data show that injury is linked to both the strain and strain rate applied to the nerve roots [178–180].

Interspecies scaling to translate experimental model results to the human from in vitro and in vivo testing plays a role on both the macroscale and the microscale. Biomechanical scaling on the macroscale is well established (e.g., Ref. [220].), but it is unclear how brain scaling works on the microscale. Scaling principles for bTBI are in their infancy (e.g., Refs. [22,221].), but investigation is crucial to establish realistic exposures in models and scalable endpoints for correlation with human clinical outcomes.

*Key Emerging Issues.* A growing concentration of efforts are aimed towards understanding the tolerance of mild TBI, and these efforts are critically reliant on defining conditions that will cause some change in either the wiring of a neural network, a compromise in the network’s mechanisms to adapt in response to an incoming signal (plasticity), and the ability of the network to shape or control activation patterns. Therefore, the mechanisms of injury over the mild spectrum will span the cellular level—e.g., the direct changes to the plasma membrane, channels, and receptors on the neuronal surface, the accompanying changes to surrounding glial cells and the vascular cells—and the network level

that includes both the neural circuit formed by neuronal ensembles within a brain region and the coordination of signaling among these brain regions. Based on current published work, the extension of these tolerance criteria and injury mechanisms in vitro to functional impairment is limited [218,219,222,223]. These predictions of network impairment will be facilitated with better descriptions of the material behavior that embed neuronal and glial connectivity and will also be very reliant upon measures of impairment made in experimental models of TBI. In this way, the prediction of function will be a significant and natural extension of ongoing activities in the field.

Scaling network results, especially functional microscale behavior from the models to humans, remains uncertain. Investigation of this unexplored territory may increase relevance of models and lead to insights into the interface of structure, network and functional outcome.

#### **Estimating Human Tolerance Through the Playing Field.**

Rather than using a coordinated series of physical surrogates, computational simulations, and in vitro models of traumatic brain injury, a new concept has surfaced in the past decade that uses a profoundly different approach—using the sports playing field and other sources of exposure to TBI as a “passive” biomechanics laboratory where one collects data to eventually estimate the human tolerance to mild TBI. Rather than relying on approximations across several steps in the laboratory, accurate measures of individual exposure will yield a direct estimate of the human tolerance over time. In one approach, the effort is made possible by novel monitoring technologies that allow one to estimate the key mechanical exposures that an athlete experiences over a practice or game [224]. When a concussion occurs, the exposure that led directly to the concussion would be archived and a distribution of loading inputs would emerge over time to yield the aggregate human tolerance. The most widely used monitoring technology (head impact telemetry system, or HITS) allows investigators to record the estimated exposures in an American football game [225]. As of this writing, nearly  $2 \times 10^6$  exposures have been recorded with this technology, and over 200 concussion events are contained in this data set [226–228]. New recording technologies are under active development, especially as the advantages of microelectronic fabrication technologies make these sensors smaller, less expensive, and potentially more widely available.

An alternative approach reconstructs the scenarios causing concussion, as captured on videotape, using anthropomorphic test dummies in a testing laboratory. Based on the pre- and postimpact positions of the striking and struck test dummies, the most likely loading conditions are estimated [149,152,229–231]. A comparison of the average peak accelerations associated with concussion in this data set, compared to the concussion data set collected with the helmet-based recording technologies, show reasonable agreement, especially given the uncertainties involved in both approaches.

A review of these approaches best puts into relief the challenges presented with the “human laboratory.” At a broad level, neither approach is designed to measure the unique tolerance for each individual. Alternatively, measuring a range of conditions causing concussion will inevitably raise the question of whether we can conclusively assign a concussion risk function for an individual, based on data from a population. From a simple biomechanical viewpoint, normal variations in brain shape, material properties, and loading direction can each produce significant variations in the deformations at any point within the brain. With this variability, even in the absence of any biological variability, the corridor of conditions associated with concussion can be large. Even if this concussion risk curve were constrained to a single individual, the role that previous impacts occurring minutes, hours, or even days prior to a given impact has given rise to great speculation about the potential for repeated impacts leading to increased vulnerability. Recently, the uncertainty of the measurements from the helmet

recording systems has shown to exceed earlier estimates, which would further contribute to the range of conditions recorded for concussions in the field [232,233].

Separate from developing human-based concussion thresholds, one may choose to use exposure measurements to take players out of a game or practice for medical evaluation or simple rest. Already, evidence shows that allowing players to self-report concussion leads to a significant underestimate of the actual concussions occurring in a game. Therefore, this monitoring system would provide a possible approach to better capture participants that should receive medical evaluation. Once again, though, the uncertainty of a unique concussion threshold and the potential uncertainty in the measurement accuracy could lead to both false positive and false negative events.

Is this key concept of the human laboratory useful for other injury situations, like blast? Technology is already developed for detecting threshold blast overpressures in the field [234–236]. Acting as a sensor for deciding if a soldier warrants medical evaluation, this application is not designed as a precise recording technology. Even if such precise monitoring for blast overpressure was available, though, one must also consider simultaneously recording key mechanical parameters that contribute to secondary and tertiary blast injury (e.g., linear and rotational acceleration) so that a recording of the complete blast exposure is recorded. Many of the same caveats applied to the use of helmet recording systems in sports would apply equally to the blast environment. Helmet-mounted systems present even more critical challenges for use in assessing exposure in the military environment. Blast waves are highly directional [22] and produce helmet motions with small peak displacements with very high accelerations ( $>1000$  g or more in the helmet) [237,238] with much lower resultant accelerations of the head ( $\sim 200$ – $300$  g). It is not clear that a helmet to head transfer function is even possible for omnidirectional blast exposure. Understanding blast biomechanics of neurotrauma is even more complicated because we are only beginning to understand how these mechanical input conditions contribute to the primary injury response.

#### **Using These Efforts to Reduce the Societal Burden of TBI.**

With this collection of tools to examine how traumatic brain injury occurs in both the civilian and military environment, it is worth considering the broader impact of how new knowledge will eventually ease the burden of this disease on society. Some of the general benefits are clear, as a more detailed understanding of injury causality will inevitably lead to better protective headgear, automobiles designed to reduce TBI incidence, and even safer sporting environments. With the current projections of the economic consequences of traumatic brain injury and disability in the U.S., these benefits can become more specific. For example, even a 25% reduction in the incidence of TBI would translate to an economic savings of  $25 \times 10^9$  U.S. dollars per year. The same reduction in incidence, if applied equally over the severity spectrum, would save 10,000 lives annually and result in a decrease of 250,000 emergency department visits each year. The number of lives saved would compare to almost halving the deaths due to prostate cancer in the U.S., or reducing the overall accident-related deaths by more than 10%.

Perhaps equally compelling is the potential long-term effects of providing a safer environment. The potential link between TBI and Alzheimer’s disease (AD) provides a useful case study. If there is significant increase in the risk for developing AD in people with a history of TBI, we could see a meaningful decline in the incidence of AD over the ensuing decades with better protection against TBI. Developing a specific estimate of the benefit is difficult, as there are a range of studies that show a clear link between TBI and enhanced risk for AD, while others show no significant increase in the risk [239]. Clearly, the net benefit of better protection would be a product of the decreased incidence rate of TBI and the relative enhanced risk of developing AD in people



with a history of TBI. For example, assume 1/6 of a population has a history of head injury and that the relative increase in risk for developing AD in the TBI population is twice the risk for a population with no prior TBI. If protection technologies led to a 50% reduction in the incidence rate of TBI, then 1/12 of the future population would have a history of head injury and we would see an approximate 7% reduction in the incidence of AD in society. Although this may seem modest, the growing economic burden of Alzheimer's disease means this reduction in incidence would save  $11\text{--}15 \times 10^9$  U.S. dollars per year in healthcare costs. As the exact relative risk for AD in patients with a history of TBI becomes more fully developed, this illustrative case study will be replaced with more specific estimates of how better protection technologies in the future will not only save lives but also contribute towards lowering the burden of diseases that could be triggered or accelerated with a history of TBI.

**Areas of Opportunity in the Future.** Although foundations of the mechanics of neurotrauma are over 70 years old, we still face significant challenges in merging the structural mechanical response and human pathophysiological response across the length scales. This gap is especially true for mild TBI, in part because we are just beginning to understand the mechanisms responsible for acute and long-term impairment for mild TBI. Using the outline presented in this review, we identify several critical unanswered questions that would accelerate our understanding in the next decade.

For defining the environments associated with TBI, we see a need to maintain a current working knowledge of the incidence rates for focal and diffuse brain injuries, and to critically define injury subgroups that are either declining in incidence or significantly increasing in incidence in specific environments. Achieving this goal would keep research foci relevant as the injurious environment changes either through new threats or the consequences of improved safety systems.

Similarly, we see a significant shortcoming in our clinicopathological understanding of primary blast injury to the human brain. Our definition of research priorities for brain injuries suffered in motor vehicle crashes was made possible by a systematic description of the injuries in the human condition (e.g., skull fracture, intracerebral hematoma, diffuse axonal injury), leading to the replication and careful study of these injuries in the laboratory. A similar, systematic description of the key injury features for primary bTBI in the human would significantly focus research efforts and consequently accelerate our understanding of their causation as well as how to protect against these types of injuries.

In estimating the primary mechanical response, several open areas of opportunity exist along the length scales:

- At the tissue scale, the continuum descriptions of material behavior are maturing but the deviatoric properties at high loading rates ( $>500 \text{ s}^{-1}$ ) are lacking.
- At the cellular scale, the nonlinearity of material behavior is nearly absent. Although we know some key transduction events, we know far less about how these force transducers and cellular inhomogeneities will affect the circuit function.
- At the molecular scale, some evidence shows key molecular domains within receptors can control their mechanosensitivity, but detailed molecular-level study across all force-responsive receptors and channels is lacking. Knowing these key molecular and atomic scale interactions would reveal potentially new insights into how forces are transduced across the mechanical loading spectrum
- At the organ and organismal scale, there remains a strong need to develop rational interspecies scaling relationships for bTBI that account for the primary mechanical response, the interspecies differences in the connectome, and any resulting changes in behavior for networks across the phylogenetic spectrum

Finally, we also see an opportunity in the far term to move these efforts for exploring two interrelated questions—how does the acute injury progress into a chronic disease, and can we better identify individual risk-curves instead of relying on estimates for the population? As we learn more about the key biological events or, alternatively, key brain regions that are important in the progression of some acute injuries into chronic impairments, we will be positioned to develop more specific tolerance criteria and protection strategies to reduce the long-term burden of TBI. Additionally, as more data become available from the human laboratory, we will see an opportunity to identify how the individual features—e.g., brain size, shape, the unique exposure profile, etc.—can lead to a better estimate for customizing protection technologies for an individual rather than relying on one design for an entire population, akin to the emerging efforts to customize treatment options based on genetic profiles in cancer patients. These efforts, although admittedly in the distant future, would represent an important evolution in our efforts to reduce the burden of TBI on the population by understanding it in each of us.

## References

- [1] Faul, M., Xu, L., and Coronado, V. G., 2010, *Traumatic Brain Injury in the United States: Emergency Department Visits, Hospitalizations, and Deaths*, CDC, Atlanta, GA.
- [2] Laker, S. R., 2011, "Epidemiology of Concussion and Mild Traumatic Brain Injury," *PM R*, 3(10 Suppl. 2), pp. S354–S358.
- [3] Tanielian, T., and Jaycox, L. H., 2008, "Invisible Wounds of War: Psychological and Cognitive Injuries, Their Consequences, and Services to Assist Recovery," Paper No. MG-720-CCF, Santa Monica, CA.
- [4] Coronado, V. G., Xu, L., Basavaraju, S. V., McGuire, L. C., Wald, M. M., Faul, M. D., Guzman, B. R., and Hemphill, J. D., 2011, "Surveillance for Traumatic Brain Injury-related Deaths—United States, 1997–2007," *MMWR Surveill. Summ.*, 60(5), pp. 1–32.
- [5] Wright, D. W., Kellerman, A., McGuire, L. C., Chen, B., and Popovic, T., 2013, "CDC Grand Rounds: Reducing Severe Traumatic Brain Injury in the United States," *MMWR Morb. Mortal. Wkly. Rep.*, 62(27), pp. 549–552.
- [6] Omalu, B. I., DeKosky, S. T., Minster, R. L., Kambou, M. I., Hamilton, R. L., and Wecht, C. H., 2005, "Chronic Traumatic Encephalopathy in a National Football League Player," *Neurosurgery*, 57(1), pp. 128–134.
- [7] McKee, A. C., Cantu, R. C., Nowinski, C. J., Hedley-Whyte, E. T., Gavett, B. E., Budson, A. E., Santini, V. E., Lee, H. S., Kubilus, C. A., and Stern, R. A., 2009, "Chronic Traumatic Encephalopathy in Athletes: Progressive Tauopathy After Repetitive Head Injury," *J. Neuropathol. Exp. Neurol.*, 68(7), pp. 709–735.
- [8] Goldstein, L. E., Fisher, A. M., Tagge, C. A., Zhang, X. L., Velisek, L., Sullivan, J. A., Upreti, C., Kracht, J. M., Ericsson, M., Wojnarowicz, M. W., Golettiani, C. J., Maglakelidze, G. M., Casey, N., Moncaster, J. A., Minaeva, O., Moir, R. D., Nowinski, C. J., Stern, R. A., Cantu, R. C., Geiling, J., Blusztajn, J. K., Wolozin, B. L., Ikezu, T., Stein, T. D., Budson, A. E., Kowall, N. W., Chargin, D., Sharon, A., Saman, S., Hall, G. F., Moss, W. C., Cleveland, R. O., Tanzi, R. E., Stanton, P. K., and McKee, A. C., 2012, "Chronic Traumatic Encephalopathy in Blast-Exposed Military Veterans and a Blast Neurotrauma Mouse Model," *Sci. Transl. Med.*, 4(134), p. 134–160.
- [9] Goldsmith, W., 2001, "The State of Head Injury Biomechanics: Past, Present, and Future: Part 1," *Crit. Rev. Biomed. Eng.*, 29(5–6), pp. 441–600.
- [10] Meaney, D. F., and Smith, D. H., 2011, "Biomechanics of Concussion," *Clin. Sports Med.*, 30(1), pp. 19–31.
- [11] Bayly, P. V., Clayton, E. H., and Genin, G. M., 2012, "Quantitative Imaging Methods for the Development and Validation of Brain Biomechanics Models," *Annu. Rev. Biomed. Eng.*, 14, pp. 369–396.
- [12] LaPlaca, M. C., Simon, C. M., Prado, G. R., and Cullen, D. K., 2007, "CNS Injury Biomechanics and Experimental Models," *Prog. Brain Res.*, 161, pp. 13–26.
- [13] King, A. I., 2000, "Fundamentals of Impact Biomechanics: Part I—Biomechanics of the Head, Neck, and Thorax," *Annu. Rev. Biomed. Eng.*, 2, pp. 55–81.
- [14] Leung, L. Y., VandeVord, P. J., Dal Cengio, A. L., Bir, C., Yang, K. H., and King, A. I., 2008, "Blast Related Neurotrauma: A Review of Cellular Injury," *Mol. Cell Biomech.*, 5(3), pp. 155–168.
- [15] Gennarelli, T. A., Spielman, G. M., Langfitt, T. W., Gildenberg, P. L., Harrington, T., Jane, J. A., Marshall, L. F., Miller, J. D., and Pitts, L. H., 1982, "Influence of the Type of Intracranial Lesion on Outcome From Severe Head Injury," *J. Neurosurg.*, 56(1), pp. 26–32.
- [16] Orman, J. A., Geyer, D., Jones, J., Schneider, E. B., Grafman, J., Pugh, M. J., and Dubose, J., 2012, "Epidemiology of Moderate-to-Severe Penetrating Versus Closed Traumatic Brain Injury in the Iraq and Afghanistan Wars," *J. Trauma Acute Care Surg.*, 73(6 Suppl. 5), pp. S496–502.
- [17] Andriessen, T. M., Horn, J., Franschman, G., van der Naalt, J., Haitsma, I., Jacobs, B., Steyerberg, E. W., and Vos, P. E., 2011, "Epidemiology, Severity Classification, and Outcome of Moderate and Severe Traumatic Brain Injury: A Prospective Multicenter Study," *J. Neurotrauma*, 28(10), pp. 2019–2031.

- [18] Morrison, B. 3rd, Meaney, D. F., Margulies, S. S., and McIntosh, T. K., 2000, "Dynamic Mechanical Stretch of Organotypic Brain Slice Cultures Induces Differential Genomic Expression: Relationship to Mechanical Parameters," *ASME J. Biomech. Eng.*, **122**(3), pp. 224–230.
- [19] O'Dell, D. M., Raghupathi, R., Crino, P. B., Eberwine, J. H., and McIntosh, T. K., 2000, "Traumatic Brain Injury Alters the Molecular Fingerprint of TUNEL-Positive Cortical Neurons in vivo: A Single-Cell Analysis," *J. Neurosci.*, **20**(13), pp. 4821–4828.
- [20] Davis, J. E., Eberwine, J. H., Hinkle, D. A., Marciano, P. G., Meaney, D. F., and McIntosh, T. K., 2004, "Methodological Considerations Regarding Single-Cell Gene Expression Profiling for Brain Injury," *Neurochem. Res.*, **29**(6), pp. 1113–1121.
- [21] Rafaels, K., Bass, C. R., Salzar, R. S., Panzer, M. B., Woods, W., Feldman, S., Cummings, T., and Capehart, B., 2011, "Survival Risk Assessment for Primary Blast Exposures to the Head," *J. Neurotrauma*, **28**(11), pp. 2319–2328.
- [22] Bass, C. R., Panzer, M. B., Rafaels, K. A., Wood, G., Shridharani, J., and Capehart, B., 2012, "Brain Injuries From Blast," *Ann. Biomed. Eng.*, **40**(1), pp. 185–202.
- [23] Effgen, G. B., Hue, C. D., Vogel, E. 3rd, Panzer, M. B., Meaney, D. F., Bass, C. R., and Morrison, B., 3rd, 2012, "A Multiscale Approach to Blast Neurotrauma Modeling: Part II: Methodology for Inducing Blast Injury to in vitro Models," *Front. Neurol.*, **3**, pp. 1–10.
- [24] Sosin, D. M., Sniezek, J. E., and Waxweiler, R. J., 1995, "Trends in Death Associated With Traumatic Brain Injury, 1979 Through 1992. Success and Failure," *JAMA*, **273**(22), pp. 1778–1780.
- [25] Adekoya, N., Thurman, D. J., White, D. D., and Webb, K. W., 2002, "Surveillance for Traumatic Brain Injury Deaths—United States, 1989–1998," *MMWR Surveill. Summ.*, **51**(10), pp. 1–14.
- [26] National Highway Traffic Safety Administration, 1999, "Federal Motor Vehicle Safety Standards and Regulations," U.S. Government Printing Office, Washington, DC.
- [27] Fabbri, A., Servadei, F., Marchesini, G., Negro, A., and Vandelli, A., 2010, "The Changing Face of Mild Head Injury: Temporal Trends and Patterns in Adolescents and Adults From 1997 to 2008," *Injury*, **41**(9), pp. 913–917.
- [28] Helmick, K. M., Warden, D. L., and Ryan, L. M., 2005, "War Neurotrauma: the Defense and Veterans Brain Injury Center (DVBIC) Experience at Walter Reed Army Medical Center (WRAMC)," *J. Neurotrauma*, **22**(10), p. 1178.
- [29] Okie, S., 2005, "Traumatic Brain Injury in the War Zone," *N. Engl. J. Med.*, **352**(20), pp. 2043–2047.
- [30] Wilk, J. E., Thomas, J. L., McGurk, D. M., Riviere, L. A., Castro, C. A., and Hoge, C. W., 2010, "Mild Traumatic Brain Injury (Concussion) During Combat: Lack of Association of Blast Mechanism With Persistent Postconcussive Symptoms," *J. Head Trauma Rehabil.*, **25**(1), pp. 9–14.
- [31] Wojcik, B. E., Stein, C. R., Bagg, K., Humphrey, R. J., and Orosco, J., 2010, "Traumatic Brain Injury Hospitalizations of U.S. Army Soldiers Deployed to Afghanistan and Iraq," *Am. J. Prev. Med.*, **38**(1 Suppl.), pp. S108–S116.
- [32] Owens, B. D., Kragh, J. F. Jr., Wenke, J. C., Macaitis, J., Wade, C. E., and Holcomb, J. B., 2008, "Combat Wounds in Operation Iraqi Freedom and Operation Enduring Freedom," *J. Trauma*, **64**(2), pp. 295–299.
- [33] Rafaels, K. A., Bass, C. R., Panzer, M. B., Salzar, R. S., Woods, W. A., Feldman, S. H., Walilko, T., Kent, R. W., Capehart, B. P., Foster, J. B., Derkunt, B., and Toman, A., 2012, "Brain Injury Risk From Primary Blast," *J. Trauma Acute Care Surg.*, **73**(4), pp. 895–901.
- [34] White, C. S., Richmond, D. R., Fletcher, E. R., Jones, R. K., and Damon, E. G., 1971, *The Biodynamics of Airblast*, N. D. Agency, ed., Washington, DC.
- [35] Gennarelli, T. A., 1993, "Mechanisms of Brain Injury," *J. Emerg. Med.*, **11**(Suppl. 1), pp. 5–11.
- [36] Armonda, R. A., Bell, R. S., Vo, A. H., Ling, G., DeGraba, T. J., Crandall, B., Ecklund, J., and Campbell, W. W., 2006, "Wartime Traumatic Cerebral Vasospasm: Recent Review of Combat Casualties," *Neurosurgery*, **59**(6), pp. 1215–1225, discussion p. 1225.
- [37] Ling, G., Bandak, F., Armonda, R., Grant, G., and Ecklund, J., 2009, "Explosive Blast Neurotrauma," *J. Neurotrauma*, **26**(6), pp. 815–825.
- [38] Gennarelli, T. A., Thibault, L. E., Adams, J. H., Graham, D. I., Thompson, C. J., and Marcincin, R. P., 1982, "Diffuse Axonal Injury and Traumatic Coma in the Primate," *Ann. Neurol.*, **12**(6), pp. 564–574.
- [39] Farkas, O., and Povlishock, J. T., 2007, "Cellular and Subcellular Change Evoked by Diffuse Traumatic Brain Injury: a Complex Web of Change Extending Far Beyond Focal Damage," *Prog. Brain Res.*, **161**, pp. 43–59.
- [40] Park, E., Eisen, R., Kinio, A., and Baker, A. J., 2013, "Electrophysiological White Matter Dysfunction and Association With Neurobehavioral Deficits Following Low-Level Primary Blast Trauma," *Neurobiol. Dis.*, **52**, pp. 150–159.
- [41] Garman, R. H., Jenkins, L. W., Switzer, R. C. 3rd, Bauman, R. A., Tong, L. C., Swauger, P. V., Parks, S. A., Ritzel, D. V., Dixon, C. E., Clark, R. S., Bayir, H., Kagan, V., Jackson, E. K., and Kochanek, P. M., 2011, "Blast Exposure in Rats With Body Shielding Is Characterized Primarily by Diffuse Axonal Injury," *J. Neurotrauma*, **28**(6), pp. 947–959.
- [42] Mac Donald, C. L., Johnson, A. M., Cooper, D., Nelson, E. C., Werner, N. J., Shimony, J. S., Snyder, A. Z., Raichle, M. E., Witherow, J. R., Fang, R., Flaherty, S. F., and Brody, D. L., 2011, "Detection of Blast-Related Traumatic Brain Injury in U.S. Military Personnel," *N. Engl. J. Med.*, **364**(22), pp. 2091–2100.
- [43] McGowan, J. C., McCormack, T. M., Grossman, R. I., Mendonca, R., Chen, X. H., Berlin, J. A., Meaney, D. F., Xu, B. N., Cecil, K. M., McIntosh, T. K., and Smith, D. H., 1999, "Diffuse Axonal Pathology Detected With Magnetization Transfer Imaging Following Brain Injury in the Pig," *Magn. Reson. Med.*, **41**(4), pp. 727–733.
- [44] Kimura, H., Meaney, D. F., McGowan, J. C., Grossman, R. I., Lenkinski, R. E., Ross, D. T., McIntosh, T. K., Gennarelli, T. A., and Smith, D. H., 1996, "Magnetization Transfer Imaging of Diffuse Axonal Injury Following Experimental Brain Injury in the Pig: Characterization by Magnetization Transfer Ratio With Histopathologic Correlation," *J. Comput. Assist. Tomogr.*, **20**(4), pp. 540–546.
- [45] Adams, J. H., Graham, D. I., Murray, L. S., and Scott, G., 1982, "Diffuse Axonal Injury due to Nonmissile Head Injury in Humans: An Analysis of 45 Cases," *Ann. Neurol.*, **12**(6), pp. 557–563.
- [46] Yoganandan, N., Gennarelli, T. A., Zhang, J., Pintar, F. A., Takhounts, E., and Ridella, S. A., 2009, "Association of Contact Loading in Diffuse Axonal Injuries From Motor Vehicle Crashes," *J. Trauma*, **66**(2), pp. 309–315.
- [47] Skandsen, T., Kvistad, K. A., Solheim, O., Strand, I. H., Folvik, M., and Vik, A., 2010, "Prevalence and Impact of Diffuse Axonal Injury in Patients With Moderate and Severe Head Injury: A Cohort Study of Early Magnetic Resonance Imaging Findings and 1-Year Outcome," *J. Neurosurg.*, **113**(3), pp. 556–563.
- [48] Capehart, B., and Bass, D., 2012, "Review: Managing Posttraumatic Stress Disorder in Combat Veterans With Comorbid Traumatic Brain Injury," *J. Rehabil. Res. Dev.*, **49**(5), pp. 789–812.
- [49] Sigurdsson, S., Aspelund, T., Forsberg, L., Fredriksson, J., Kjartansson, O., Oskarsdottir, B., Jonsson, P. V., Eiriksdottir, G., Harris, T. B., Zijdenbos, A., van Buchem, M. A., Launer, L. J., and Gudnason, V., 2012, "Brain Tissue Volumes in the General Population of the Elderly: The AGES-Reykjavik Study," *Neuroimage*, **59**(4), pp. 3862–3870.
- [50] Sharp, D. J., Beckmann, C. F., Greenwood, R., Kinnunen, K. M., Bonnelle, V., De Boissezon, X., Powell, J. H., Counsell, S. J., Patel, M. C., and Leech, R., 2011, "Default Mode Network Functional and Structural Connectivity After Traumatic Brain Injury," *Brain*, **134**(Pt 8), pp. 2233–2247.
- [51] Margulies, S. S., and Meaney, D. F., 1994, "Physical Properties of Brain Tissue," *Handbook of Biomaterial Properties*, J. Black, and G. Hastings, eds., Chapman and Hall, pp. 70–80, London, UK.
- [52] McElhane, J. H., Hilyard, J. F., and Roberts, V. L., 1976, *Handbook of Human Tolerance*, JARI, Tokyo, Japan.
- [53] Maikos, J. T., Elias, R. A., and Shreiber, D. I., 2008, "Mechanical Properties of Dura Mater From the Rat Brain and Spinal Cord," *J. Neurotrauma*, **25**(1), pp. 38–51.
- [54] Zarzur, E., 1996, "Mechanical Properties of the Human Lumbar Dura Mater," *Arq. Neuropsiquiatr.*, **54**(3), pp. 455–460.
- [55] Dunn, M. G., and Silver, F. H., 1983, "Viscoelastic Behavior of Human Connective Tissues: Relative Contribution of Viscous and Elastic Components," *Connect Tissue Res.*, **12**(1), pp. 59–70.
- [56] van Noort, R., Martin, T. R., Black, M. M., Barker, A. T., and Montero, C. G., 1981, "The Mechanical Properties of Human Dura Mater and the Effects of Storage Media," *Clin. Phys. Physiol. Meas.*, **2**(3), pp. 197–203.
- [57] van Noort, R., Black, M. M., Martin, T. R., and Meaney, S., 1981, "A Study of the Uniaxial Mechanical Properties of Human Dura Mater Preserved in Glycerol," *Biomaterials*, **2**(1), pp. 41–45.
- [58] Zhang, L., Bae, J., Hardy, W. N., Monson, K. L., Manley, G. T., Goldsmith, W., Yang, K. H., and King, A. I., 2002, "Computational Study of the Contribution of the Vasculature on the Dynamic Response of the Brain," *Stapp Car Crash J.*, **46**, pp. 145–164.
- [59] Delye, H., Goffin, J., Verschuere, P., Vander Sloten, J., Van der Perre, G., Alaerts, H., Verpoest, I., and Berckmans, D., 2006, "Biomechanical Properties of the Superior Sagittal Sinus-Bridging Vein Complex," *Stapp Car Crash J.*, **50**, pp. 625–636.
- [60] Lee, M. C., and Haut, R. C., 1989, "Insensitivity of Tensile Failure Properties of Human Bridging Veins to Strain Rate: Implications in Biomechanics of Subdural Hematoma," *J. Biomech.*, **22**(6–7), pp. 537–542.
- [61] Lowenhielm, P., 1974, "Dynamic Properties of the Parasagittal Bridging Veins," *Z. Rechtsmed.*, **74**(1), pp. 55–62.
- [62] Monson, K. L., Goldsmith, W., Barbaro, N. M., and Manley, G. T., 2003, "Axial Mechanical Properties of Fresh Human Cerebral Blood Vessels," *ASME J. Biomech. Eng.*, **125**(2), pp. 288–294.
- [63] Monson, K. L., Goldsmith, W., Barbaro, N. M., and Manley, G. T., 2005, "Significance of Source and Size in the Mechanical Response of Human Cerebral Blood Vessels," *J. Biomech.*, **38**(4), pp. 737–744.
- [64] Metz, H., McElhane, J., and Ommaya, A. K., 1970, "A Comparison of the Elasticity of Live, Dead, and Fixed Brain Tissue," *J. Biomech.*, **3**(4), pp. 453–458.
- [65] Galford, J. E., and McElhane, J. H., 1970, "A Viscoelastic Study of Scalp, Brain, and Dura," *J. Biomech.*, **3**(2), pp. 211–221.
- [66] Chatelin, S., Constantinesco, A., and Willinger, R., 2010, "Fifty Years of Brain Tissue Mechanical Testing: From in vitro to in vivo Investigations," *Biorheology*, **47**(5–6), pp. 255–276.
- [67] Cheng, S., Clarke, E. C., and Bilston, L. E., 2008, "Rheological Properties of the Tissues of the Central Nervous System: A Review," *Med. Eng. Phys.*, **30**(10), pp. 1318–1337.
- [68] Pervin, F., and Chen, W. W., 2009, "Dynamic Mechanical Response of Bovine Gray Matter and White Matter Brain Tissues Under Compression," *J. Biomech.*, **42**(6), pp. 731–735.
- [69] Urbanczyk, C., Palmeri, M., Rouze, N., Kloppenborg, N., and Bass, C. R., 2012, "Acoustic Radiation Force Impulse Imaging Improves Ultrasound Resolution in Neural Tissue: Effects of Temperature and Confinement on Brain Material Property Characterization," *J. Neurotrauma*, **22**(10), pp. A200–A201.
- [70] Lippert, S. A., Rang, E. M., and Grimm, M. J., 2004, "The High Frequency Properties of Brain Tissue," *Biorheology*, **41**(6), pp. 681–691.

- [71] Weaver, J. B., Pattison, A. J., McGarry, M. D., Perreard, I. M., Swienkowski, J. G., Eskey, C. J., Lollis, S. S., and Paulsen, K. D., 2012, "Brain Mechanical Property Measurement Using MRE With Intrinsic Activation," *Phys. Med. Biol.*, **57**(22), pp. 7275–7287.
- [72] Clayton, E. H., Genin, G. M., and Bayly, P. V., 2012, "Transmission, Attenuation and Reflection of Shear Waves in the Human Brain," *J. R. Soc. Interface*, **9**(76), pp. 2899–2910.
- [73] Boulet, T., Kelso, M. L., and Othman, S. F., 2011, "Microscopic Magnetic Resonance Elastography of Traumatic Brain Injury Model," *J. Neurosci. Meth.*, **201**(2), pp. 296–306.
- [74] Zhang, J., Green, M. A., Sinkus, R., and Bilston, L. E., 2011, "Viscoelastic Properties of Human Cerebellum Using Magnetic Resonance Elastography," *J. Biomech.*, **44**(10), pp. 1909–1913.
- [75] Hamhaber, U., Klatt, D., Papazoglou, S., Hollmann, M., Stadler, J., Sack, I., Bernarding, J., and Braun, J., 2010, "In vivo Magnetic Resonance Elastography of Human Brain at 7 T and 1.5 T," *J. Magn. Reson. Imaging*, **32**(3), pp. 577–583.
- [76] Elkin, B. S., Ilankovan, A. I., and Morrison, B., 3rd, 2011, "A Detailed Viscoelastic Characterization of the P17 and Adult Rat Brain," *J. Neurotrauma*, **28**(11), pp. 2235–2244.
- [77] Elkin, B. S., Ilankova, A., and Morrison, B., 2011, "Dynamic, Regional Mechanical Properties of the Porcine Brain: Indentation in the Coronal Plane," *ASME J. Biomech. Eng.*, **133**(7), p. 071009.
- [78] Finan, J. D., Elkin, B. S., Pearson, E. M., Kalbian, I. L., and Morrison, B., 3rd, 2012, "Viscoelastic Properties of the Rat Brain in the Sagittal Plane: Effects of Anatomical Structure and Age," *Ann. Biomed. Eng.*, **40**(1), pp. 70–78.
- [79] Feng, Y., Okamoto, R. J., Namani, R., Genin, G. M., and Bayly, P. V., 2013, "Measurements of Mechanical Anisotropy in Brain Tissue and Implications for Transversely Isotropic Material Models of White Matter," *J. Mech. Behav. Biomed. Mater.*, **23**, pp. 117–132.
- [80] Velardi, F., Fraternali, F., and Angelillo, M., 2006, "Anisotropic Constitutive Equations and Experimental Tensile Behavior of Brain Tissue," *Biomech. Model. Mechanobiol.*, **5**(1), pp. 53–61.
- [81] Arbogast, K. B., and Margulies, S. S., 1998, "Material Characterization of the Brainstem From Oscillatory Shear Tests," *J. Biomech.*, **31**(9), pp. 801–807.
- [82] Elkin, B. S., Ilankovan, A., and Morrison, B., 3rd, 2010, "Age-Dependent Regional Mechanical Properties of the Rat Hippocampus and Cortex," *ASME J. Biomech. Eng.*, **132**(1), p. 011010.
- [83] Sarninoranont, M., Lee, S. J., Hong, Y., King, M. A., Subhash, G., Kwon, J., and Moore, D. F., 2012, "High-Strain-Rate Brain Injury Model Using Submerged Acute Rat Brain Tissue Slices," *J. Neurotrauma*, **29**(2), pp. 418–429.
- [84] Zhang, J., Yoganandan, N., Pintar, F. A., Guan, Y., Shender, B., Paskoff, G., and Laud, P., 2011, "Effects of Tissue Preservation Temperature on High Strain-Rate Material Properties of Brain," *J. Biomech.*, **44**(3), pp. 391–396.
- [85] Zhang, J., Song, B., Pintar, F. A., Yoganandan, N., Chen, W., and Gennarelli, T. A., 2008, "How to Test Brain and Brain Simulant at Ballistic and Blast Strain Rates," *Biomed. Sci. Instrum.*, **44**, pp. 129–134.
- [86] Bain, A. C., Shreiber, D. I., and Meaney, D. F., 2003, "Modeling of Microstructural Kinematics During Simple Elongation of Central Nervous System Tissue," *ASME J. Biomech. Eng.*, **125**(6), pp. 798–804.
- [87] Meaney, D. F., 2003, "Relationship Between Structural Modeling and Hyperelastic Material Behavior: Application to CNS White Matter," *Biomech. Model. Mechanobiol.*, **1**(4), pp. 279–293.
- [88] Karami, G., Grundman, N., Abolfathi, N., Naik, A., and Ziejewski, M., 2009, "A Micromechanical Hyperelastic Modeling of Brain White Matter Under Large Deformation," *J. Mech. Behav. Biomed. Mater.*, **2**(3), pp. 243–254.
- [89] Abolfathi, N., Naik, A., Sotudeh Chafi, M., Karami, G., and Ziejewski, M., 2009, "A Micromechanical Procedure for Modelling the Anisotropic Mechanical Properties of Brain White Matter," *Comput. Meth. Biomech. Biomed. Eng.*, **12**(3), pp. 249–262.
- [90] Pan, Y., Shreiber, D. I., and Pelegri, A. A., 2011, "A Transition Model for Finite Element Simulation of Kinematics of Central Nervous System White Matter," *IEEE Trans. Biomed. Eng.*, **58**(12), pp. 3443–3446.
- [91] Bernick, K. B., Prevost, T. P., Suresh, S., and Socrate, S., 2011, "Biomechanics of Single Cortical Neurons," *Acta Biomater.*, **7**(3), pp. 1210–1219.
- [92] Mustata, M., Ritchie, K., and McNally, H. A., 2010, "Neuronal Elasticity as Measured by Atomic Force Microscopy," *J. Neurosci. Meth.*, **186**(1), pp. 35–41.
- [93] Ma, J., Liu, B. F., Xu, Q. Y., and Cui, F. Z., 2005, "AFM Study of Hippocampal Cells Cultured on Silicon Wafers With Nano-Scale Surface Topograph," *Colloids Surf. B Biointerfaces*, **44**(2–3), pp. 152–157.
- [94] Parpura, V., Haydon, P. G., and Henderson, E., 1993, "Three-Dimensional Imaging of Living Neurons and Glia With the Atomic Force Microscope," *J. Cell Sci.*, **104**(Pt. 2), pp. 427–432.
- [95] Miller, W. J., Leventhal, I., Scarsella, D., Haydon, P. G., Janmey, P., and Meaney, D. F., 2009, "Mechanically Induced Reactive Gliosis Causes ATP-Mediated Alterations in Astrocyte Stiffness," *J. Neurotrauma*, **26**(5), pp. 789–797.
- [96] Haydon, P. G., Lartius, R., Parpura, V., and Marchese-Ragona, S. P., 1996, "Membrane Deformation of Living Glial Cells Using Atomic Force Microscopy," *J. Microsc.*, **182**(Pt. 2), pp. 114–120.
- [97] Holbourn, A. H. S., 1943, "Mechanics of Head Injuries," *Lancet*, **242**(6267), pp. 438–441.
- [98] Gurdjian, E. S., and Lissner, H. R., 1961, "Photoelastic Confirmation of the Presence of Shear Strains at the Craniospinal Junction in Closed Head Injury," *J. Neurosurg.*, **18**, pp. 58–60.
- [99] Margulies, S. S., Thibault, L. E., and Gennarelli, T. A., 1990, "Physical Model Simulations of Brain Injury in the Primate," *J. Biomech.*, **23**(8), pp. 823–836.
- [100] Bayly, P. V., Ji, S., Song, S. K., Okamoto, R. J., Massouros, P., and Genin, G. M., 2004, "Measurement of Strain in Physical Models of Brain Injury: A Method Based on HARP Analysis of Tagged Magnetic Resonance Images (MRI)," *ASME J. Biomech. Eng.*, **126**(4), pp. 523–528.
- [101] Ivarsson, J., Viano, D. C., and Lovsund, P., 2002, "Influence of the Lateral Ventricles and Irregular Skull Base on Brain Kinematics due to Sagittal Plane Head Rotation," *ASME J. Biomech. Eng.*, **124**(4), pp. 422–431.
- [102] Ivarsson, J., Viano, D. C., Lovsund, P., and Aldman, B., 2000, "Strain Relief From the Cerebral Ventricles During Head Impact: Experimental Studies on Natural Protection of the Brain," *J. Biomech.*, **33**(2), pp. 181–189.
- [103] Meaney, D. F., Ross, D. T., Winkelstein, B. A., Brasko, J., Goldstein, D., Bilston, L. B., Thibault, L. E., and Gennarelli, T. A., 1994, "Modification of the Cortical Impact Model to Produce Axonal Injury in the Rat Cerebral Cortex," *J. Neurotrauma*, **11**(5), pp. 599–612.
- [104] Thibault, L. E., Meaney, D. F., Anderson, B. J., and Marmarou, A., 1992, "Biomechanical Aspects of a Fluid Percussion Model of Brain Injury," *J. Neurotrauma*, **9**(4), pp. 311–322.
- [105] Meaney, D. F., Smith, D. H., Shreiber, D. I., Bain, A. C., Miller, R. T., Ross, D. T., and Gennarelli, T. A., 1995, "Biomechanical Analysis of Experimental Diffuse Axonal Injury," *J. Neurotrauma*, **12**(4), pp. 689–694.
- [106] Mediavilla Varas, J., Philippens, M., Meijer, S. R., van den Berg, A. C., Sibma, P. C., van Bree, J. L., and de Vries, D. V., 2011, "Physics of IED Blast Shock Tube Simulations for mTBI Research," *Front. Neurol.*, **2**, pp. 1–14.
- [107] Zhu, F., Wagner, C., Dal Cengio Leonardi, A., Jin, X., Vandevord, P., Chou, C., Yang, K. H., and King, A. I., 2012, "Using a Gel/Plastic Surrogate to Study the Biomechanical Response of the Head Under Air Shock Loading: A Combined Experimental and Numerical Investigation," *Biomech. Model. Mechanobiol.*, **11**(3–4), pp. 341–353.
- [108] Alley, M. D., Schimzize, B. R., and Son, S. F., 2011, "Experimental Modeling of Explosive Blast-Related Traumatic Brain Injuries," *Neuroimage*, **54**(Suppl. 1), pp. S45–S54.
- [109] Zhang, J., Yoganandan, N., Pintar, F. A., Gennarelli, T. A., and Shender, B. S., 2009, "A Finite Element Study of Blast Traumatic Brain Injury—Biomed 2009," *Biomed. Sci. Instrum.*, **45**, pp. 119–124.
- [110] Hardy, W. N., Foster, C. D., Mason, M. J., Yang, K. H., King, A. I., and Tashman, S., 2001, "Investigation of Head Injury Mechanisms Using Neutral Density Technology and High-Speed Biplanar X-ray," *Stapp Car Crash J.*, **45**, pp. 337–368.
- [111] Wing, I. D., Merkle, A. C., Armiger, R. S., Carkhuff, B. G., and Roberts, J. C., 2013, "Development of a Miniaturized Position Sensing System for Measuring Brain Motion During Impact—Biomed 2013," *Biomed. Sci. Instrum.*, **49**, pp. 281–288.
- [112] Hardy, W. N., Mason, M. J., Foster, C. D., Shah, C. S., Kopacz, J. M., Yang, K. H., King, A. I., Bishop, J., Bey, M., Anderst, W., and Tashman, S., 2007, "A Study of the Response of the Human Cadaver Head to Impact," *Stapp Car Crash J.*, **51**, pp. 17–80.
- [113] Zhang, L., Yang, K. H., Dwarampudi, R., Omori, K., Li, T., Chang, K., Hardy, W. N., Khalil, T. B., and King, A. I., 2001, "Recent Advances in Brain Injury Research: A New Human Head Model Development and Validation," *Stapp Car Crash J.*, **45**, pp. 369–394.
- [114] Parnaik, Y., Beillas, P., Demetropoulos, C. K., Hardy, W. N., Yang, K. H., and King, A. I., 2004, "The Influence of Surrogate Blood Vessels on the Impact Response of a Physical Model of the Brain," *Stapp Car Crash J.*, **48**, pp. 259–277.
- [115] Kleiven, S., and Hardy, W. N., 2002, "Correlation of an FE Model of the Human Head With Local Brain Motion—Consequences for Injury Prediction," *Stapp Car Crash J.*, **46**, pp. 123–144.
- [116] Zou, H., Schmiedeler, J. P., and Hardy, W. N., 2007, "Separating Brain Motion Into Rigid Body Displacement and Deformation Under Low-Severity Impacts," *J. Biomech.*, **40**(6), pp. 1183–1191.
- [117] Bayly, P. V., Cohen, T. S., Leister, E. P., Ajo, D., Leuthardt, E. C., and Genin, G. M., 2005, "Deformation of the Human Brain Induced by Mild Acceleration," *J. Neurotrauma*, **22**(8), pp. 845–856.
- [118] Sabet, A. A., Christoforou, E., Zatlín, B., Genin, G. M., and Bayly, P. V., 2008, "Deformation of the Human Brain Induced by Mild Angular Head Acceleration," *J. Biomech.*, **41**(2), pp. 307–315.
- [119] Voo, K., Kumaresan, S., Pintar, F. A., Yoganandan, N., and Sances, A., Jr., 1996, "Finite-Element Models of the Human Head," *Med. Biol. Eng. Comput.*, **34**(5), pp. 375–381.
- [120] King, A. I., Ruan, J. S., Zhou, C., Hardy, W. N., and Khalil, T. B., 1995, "Recent Advances in Biomechanics of Brain Injury Research: a Review," *J. Neurotrauma*, **12**(4), pp. 651–658.
- [121] Mao, H., Zhang, L., Jiang, B., Genthikatti, V., Jin, X., Zhu, F., Makwana, R., Gill, A., Jandir, G., Singh, A., and Yang, K., 2013, "Development of a Finite Element Human Head Model Validated With Forty Nine Loading Cases From Experimental and Real World Impacts," *ASME J. Biomech. Eng.*, **135**(11), p. 111002.
- [122] Holst, H. V., and Li, X., 2013, "The Dynamic Triple Peak Impact Factor in Traumatic Brain Injury Influences Native Protein Structures in Gray and White Matter as Measured With Computational Simulation," *Neurol. Res.*, **35**(8), pp. 782–789.
- [123] Pintar, F. A., Philippens, M. M., Zhang, J., and Yoganandan, N., 2013, "Methodology to Determine Skull Bone and Brain Responses From Ballistic Helmet-to-Head Contact Loading Using Experiments and Finite Element Analysis," *Med. Eng. Phys.*, **135**(11), pp. 1682–1687.



- [124] Lillie, E. M., Urban, J. E., Lynch, S. K., Whitlow, C. T., and Stitzel, J. D., 2013, "Evaluation of the Extent and Distribution of Diffuse—Axonal Injury From Real World Motor Vehicle Crashes Biomed 2013," *Biomed. Sci. Instrum.*, **49**, pp. 297–304.
- [125] Watanabe, R., Katsuhara, T., Miyazaki, H., Kitagawa, Y., and Yasuki, T., 2012, "Research of the Relationship of Pedestrian Injury to Collision Speed, Car-Type, Impact Location and Pedestrian Sizes Using Human FE Model (THUMS Version 4)," *Stapp Car Crash J.*, **56**, pp. 269–321.
- [126] Patton, D. A., McIntosh, A. S., and Kleiven, S., 2013, "The Biomechanical Determinants of Concussion: Finite Element Simulations to Investigate Brain Tissue Deformations During Sporting Impacts to the Unprotected Head," *J. Appl. Biomech* (accepted).
- [127] Jazi, M. S., Rezaei, A., Karami, G., Azarmi, F., and Ziejewski, M., 2013, "A Computational Study of Influence of Helmet Padding Materials on the Human Brain Under Ballistic Impacts," *Comput. Meth. Biomech. Biomed. Eng. Jan. 3* (ePub ahead of print).
- [128] Roberts, J. C., Harrigan, T. P., Ward, E. E., Taylor, T. M., Annett, M. S., and Merkle, A. C., 2012, "Human Head-Neck Computational Model for Assessing Blast Injury," *J. Biomech.*, **45**(16), pp. 2899–2906.
- [129] Wright, R. M., Post, A., Hoshizaki, B., and Ramesh, K. T., 2013, "A Multi-scale Computational Approach to Estimating Axonal Damage Under Inertial Loading of the Head," *J. Neurotrauma*, **30**(2), pp. 102–118.
- [130] Lamy, M., Baumgartner, D., Willinger, R., Yoganandan, N., and Stemper, B. D., 2011, "Study of Mild Traumatic Brain Injuries Using Experiments and Finite Element Modeling," *Ann. Adv. Automot. Med.*, **55**, pp. 125–135.
- [131] Cloots, R. J., van Dommelen, J. A., and Geers, M. G., 2012, "A Tissue-Level Anisotropic Criterion for Brain Injury Based on Microstructural Axonal Deformation," *J. Mech. Behav. Biomed. Mater.*, **5**(1), pp. 41–52.
- [132] Chatelin, S., Deck, C., Renard, F., Kremer, S., Heinrich, C., Armspach, J. P., and Willinger, R., 2011, "Computation of Axonal Elongation in Head Trauma Finite Element Simulation," *J. Mech. Behav. Biomed. Mater.*, **4**(8), pp. 1905–1919.
- [133] Kimpara, H., and Iwamoto, M., 2012, "Mild Traumatic Brain Injury Predictors Based on Angular Accelerations During Impacts," *Ann. Biomed. Eng.*, **40**(1), pp. 114–126.
- [134] McAllister, T. W., Ford, J. C., Ji, S., Beckwith, J. G., Flashman, L. A., Paulsen, K., and Greenwald, R. M., 2012, "Maximum Principal Strain and Strain Rate Associated With Concussion Diagnosis Correlates With Changes in Corpus Callosum White Matter Indices," *Ann. Biomed. Eng.*, **40**(1), pp. 127–140.
- [135] Ganpule, S., Gu, L., Alai, A., and Chandra, N., 2012, "Role of Helmet in the Mechanics of Shock Wave Propagation Under Blast Loading Conditions," *Comput. Meth. Biomech. Biomed. Eng.*, **15**(11), pp. 1233–1244.
- [136] Wright, R. M., and Ramesh, K. T., 2012, "An Axonal Strain Injury Criterion for Traumatic Brain Injury," *Biomech. Model. Mechanobiol.*, **11**(1–2), pp. 245–260.
- [137] Yan, W., and Pangestu, O. D., 2011, "A Modified Human Head Model for the Study of Impact Head Injury," *Comput. Meth. Biomech. Biomed. Eng.*, **14**(12), pp. 1049–1057.
- [138] Nyein, M. K., Jason, A. M., Yu, L., Pita, C. M., Joannopoulos, J. D., Moore, D. F., and Radovitzky, R. A., 2010, "In Silico Investigation of Intracranial Blast Mitigation With Relevance to Military Traumatic Brain Injury," *Proc. Natl. Acad. Sci. USA*, **107**(48), pp. 20703–20708.
- [139] Colgan, N. C., Gilchrist, M. D., and Curran, K. M., 2010, "Applying DTI White Matter Orientations to Finite Element Head Models to Examine Diffuse TBI Under High Rotational Accelerations," *Prog. Biophys. Mol. Biol.*, **103**(2–3), pp. 304–309.
- [140] Lockhart, P., Cronin, D., Williams, K., and Ouellet, S., 2011, "Investigation of Head Response to Blast Loading," *J. Trauma*, **70**(2), pp. E29–36.
- [141] Cheng, S., and Bilston, L. E., 2010, "Computational Model of the Cerebral Ventricles in Hydrocephalus," *ASME J. Biomech. Eng.*, **132**(5), p. 054501.
- [142] Fijalkowski, R. J., Yoganandan, N., Zhang, J., and Pintar, F. A., 2009, "A Finite Element Model of Region-Specific Response for Mild Diffuse Brain Injury," *Stapp Car Crash J.*, **53**, pp. 193–213.
- [143] Chafi, M. S., Karami, G., and Ziejewski, M., 2010, "Biomechanical Assessment of Brain Dynamic Responses due to Blast Pressure Waves," *Ann. Biomed. Eng.*, **38**(2), pp. 490–504.
- [144] Moore, D. F., Jerusalem, A., Nyein, M., Noels, L., Jaffee, M. S., and Radovitzky, R. A., 2009, "Computational Biology—Modeling of Primary Blast Effects on the Central Nervous System," *Neuroimage*, **47**(Suppl. 2), pp. T10–T20.
- [145] Cloots, R. J., Gervaise, H. M., van Dommelen, J. A., and Geers, M. G., 2008, "Biomechanics of Traumatic Brain Injury: Influences of the Morphologic Heterogeneities of the Cerebral Cortex," *Ann. Biomed. Eng.*, **36**(7), pp. 1203–1215.
- [146] Fredriksson, R., Zhang, L., Bostrom, O., and Yang, K., 2007, "Influence of Impact Speed on Head and Brain Injury Outcome in Vulnerable Road User Impacts to the Car Hood," *Stapp Car Crash J.*, **51**, pp. 155–167.
- [147] Kleiven, S., 2007, "Predictors for Traumatic Brain Injuries Evaluated Through Accident Reconstructions," *Stapp Car Crash J.*, **51**, pp. 81–114.
- [148] Ishikawa, R., Kato, K., Kubo, M., Uzuka, T., and Takahashi, H., 2006, "Finite Element Analysis and Experimental Study on Mechanism of Brain Injury Using Brain Model," *Conf. Proc. IEEE Eng. Med. Biol. Soc.*, **1**, pp. 1327–1330.
- [149] Viano, D. C., Casson, I. R., and Pellman, E. J., 2007, "Concussion in Professional Football: Biomechanics of the Struck Player—part 14," *Neurosurgery*, **61**(2), pp. 313–327, discussion pp. 327–318.
- [150] Takahashi, T., Kato, K., Ishikawa, R., Watanabe, T., Kubo, M., Uzuka, T., Fujii, Y., and Takahashi, H., 2007, "3-D Finite Element Analysis and Experimental Study on Brain Injury Mechanism," *Conf. Proc. IEEE Eng. Med. Biol. Soc.*, pp. 3613–3616.
- [151] Zhang, L., Yang, K. H., and King, A. I., 2004, "A Proposed Injury Threshold for Mild Traumatic Brain Injury," *ASME J. Biomech. Eng.*, **126**(2), pp. 226–236.
- [152] Viano, D. C., Casson, I. R., Pellman, E. J., Zhang, L., King, A. I., and Yang, K. H., 2005, "Concussion in Professional Football: Brain Responses by Finite Element Analysis: Part 9," *Neurosurgery*, **57**(5), pp. 891–916, discussion pp. 891–916.
- [153] Sanchez-Molina, D., Velazquez-Ameijide, J., Arregui-Dalmases, C., Crandall, J. R., and Untaroiu, C. D., 2012, "Minimization of Analytical Injury Metrics for Head Impact Injuries," *Traffic Inj. Prev.*, **13**(3), pp. 278–285.
- [154] Duma, S. M., and Rowson, S., 2009, "Every Newton Hertz: A Macro to Micro Approach to Investigating Brain Injury," *Conf. Proc. IEEE Eng. Med. Biol. Soc.*, pp. 1123–1126.
- [155] Rigby, P., and Chan, P., 2009, "Evaluation of the Biodeficiency of FMVSS No. 218 Injury Criteria," *Traffic Inj. Prev.*, **10**(2), pp. 170–177.
- [156] Danelson, K. A., Geer, C. P., Stitzel, J. D., Slice, D. E., and Takhounts, E. G., 2008, "Age and Gender Based Biomechanical Shape and Size Analysis of the Pediatric Brain," *Stapp Car Crash J.*, **52**, pp. 59–81.
- [157] Takhounts, E. G., Ridella, S. A., Hasija, V., Tannous, R. E., Campbell, J. Q., Malone, D., Danelson, K., Stitzel, J., Rowson, S., and Duma, S., 2008, "Investigation of Traumatic Brain Injuries Using the Next Generation of Simulated Injury Monitor (SIMon) Finite Element Head Model," *Stapp Car Crash J.*, **52**, pp. 1–31.
- [158] Franklyn, M., Fildes, B., Zhang, L., Yang, K., and Sparke, L., 2005, "Analysis of Finite Element Models for Head Injury Investigation: Reconstruction of Four Real-World Impacts," *Stapp Car Crash J.*, **49**, pp. 1–32.
- [159] Takhounts, E. G., Eppinger, R. H., Campbell, J. Q., Tannous, R. E., Power, E. D., and Shook, L. S., 2003, "On the Development of the SIMon Finite Element Head Model," *Stapp Car Crash J.*, **47**, pp. 107–133.
- [160] Zhang, J., Yoganandan, N., Pintar, F. A., and Gennarelli, T. A., 2006, "Role of Translational and Rotational Accelerations on Brain Strain in Lateral Head Impact," *Biomed. Sci. Instrum.*, **42**, pp. 501–506.
- [161] Rezaei, A., Salimi Jazi, M., and Karami, G., 2013, "Computational Modeling of Human Head Under Blast in Confined and Open Spaces: Primary Blast Injury," *Int. J. Num. Method Biomed. Eng.*, Aug. 28 (ePub ahead of print).
- [162] Zhang, L., Makwana, R., and Sharma, S., 2013, "Brain Response to Primary Blast Wave Using Validated Finite Element Models of Human Head and Advanced Combat Helmet," *Front Neurol.*, **4**(8), pp. 1–12.
- [163] Sundaramurthy, A., Alai, A., Ganpule, S., Holmberg, A., Plougonven, E., and Chandra, N., 2012, "Blast-Induced Biomechanical Loading of the Rat: an Experimental and Anatomically Accurate Computational Blast Injury Model," *J. Neurotrauma*, **29**(13), pp. 2352–2364.
- [164] Panzer, M. B., Myers, B. S., Capehart, B. P., and Bass, C. R., 2012, "Development of a Finite Element Model for Blast Brain Injury and the Effects of CSF Cavitation," *Ann. Biomed. Eng.*, **40**(7), pp. 1530–1544.
- [165] Mao, H., Zhang, L., Yang, K. H., and King, A. I., 2006, "Application of a Finite Element Model of the Brain to Study Traumatic Brain Injury Mechanisms in the Rat," *Stapp Car Crash J.*, **50**, pp. 583–600.
- [166] Levchakov, A., Linder-Ganz, E., Raghupathi, R., Margulies, S. S., and Gefen, A., 2006, "Computational Studies of Strain Exposures in Neonate and Mature Rat Brains During Closed Head Impact," *J. Neurotrauma*, **23**(10), pp. 1570–1580.
- [167] Pena, A., Pickard, J. D., Stiller, D., Harris, N. G., and Schuhmann, M. U., 2005, "Brain Tissue Biomechanics in Cortical Contusion Injury: a Finite Element Analysis," *Acta Neurochir. Suppl.*, **95**, pp. 333–336.
- [168] Gefen, A., Gefen, N., Zhu, Q., Raghupathi, R., and Margulies, S. S., 2003, "Age-Dependent Changes in Material Properties of the Brain and Braincase of the Rat," *J. Neurotrauma*, **20**(11), pp. 1163–1177.
- [169] Zhu, F., Skelton, P., Chou, C. C., Mao, H., Yang, K. H., and King, A. I., 2013, "Biomechanical Responses of a Pig Head Under Blast Loading: a Computational Simulation," *Int. J. Num. Meth. Biomed. Eng.*, **29**(3), pp. 392–407.
- [170] Coats, B., Eucker, S. A., Sullivan, S., and Margulies, S. S., 2012, "Finite Element Model Predictions of Intracranial Hemorrhage From Non-Impact, Rapid Head Rotations in the Piglet," *Int. J. Dev. Neurosci.*, **30**(3), pp. 191–200.
- [171] Zhu, Q., Prange, M., and Margulies, S., 2006, "Predicting Unconsciousness From a Pediatric Brain Injury Threshold," *Dev. Neurosci.*, **28**(4–5), pp. 388–395.
- [172] Coats, B., and Margulies, S. S., 2006, "Material Properties of Porcine Parietal Cortex," *J. Biomech.*, **39**(13), pp. 2521–2525.
- [173] Margulies, S. S., and Thibault, K. L., 2000, "Infant Skull and Suture Properties: Measurements and Implications for Mechanisms of Pediatric Brain Injury," *ASME J. Biomech. Eng.*, **122**(4), pp. 364–371.
- [174] Mazumder, M. M., Miller, K., Bunt, S., Mostayed, A., Joldes, G., Day, R., Hart, R., and Wittek, A., 2013, "Mechanical Properties of the Brain-Skull Interface," *Acta Bioeng. Biomech.*, **15**(2), pp. 3–11.
- [175] Maitry, P., and Tekalur, S. A., 2011, "Finite Element Analysis of Ramming in *Ovis canadensis*," *ASME J. Biomech. Eng.*, **133**(2), p. 021009.
- [176] Anderson, R. W., Brown, C. J., Blumberg, P. C., McLean, A. J., and Jones, N. R., 2003, "Impact Mechanics and Axonal Injury in a Sheep Model," *J. Neurotrauma*, **20**(10), pp. 961–974.
- [177] Mao, H., Yang, K. H., King, A. I., and Yang, K., 2010, "Computational Neurotrauma—Design, Simulation, and Analysis of Controlled Cortical Impact Model," *Biomech. Model. Mechanobiol.*, **9**(6), pp. 763–772.

- [178] Li, Y., Zhang, L., Kallakuri, S., Zhou, R., and Cavanaugh, J. M., 2011, "Quantitative Relationship Between Axonal Injury and Mechanical Response in a Rodent Head Impact Acceleration Model," *J. Neurotrauma*, **28**(9), pp. 1767–1782.
- [179] Singh, A., Kallakuri, S., Chen, C., and Cavanaugh, J. M., 2009, "Structural and Functional Changes in Nerve Roots due to Tension at Various Strains and Strain Rates: an in-vivo Study," *J. Neurotrauma*, **26**(4), pp. 627–640.
- [180] Singh, A., Lu, Y., Chen, C., Kallakuri, S., and Cavanaugh, J. M., 2006, "A New Model of Traumatic Axonal Injury to Determine the Effects of Strain and Displacement Rates," *Stapp Car Crash J.*, **50**, pp. 601–623.
- [181] Bain, A. C., Raghupathi, R., and Meaney, D. F., 2001, "Dynamic Stretch Correlates to Both Morphological Abnormalities and Electrophysiological Impairment in a Model of Traumatic Axonal Injury," *J. Neurotrauma*, **18**(5), pp. 499–511.
- [182] Bain, A. C., and Meaney, D. F., 2000, "Tissue-Level Thresholds for Axonal Damage in an Experimental Model of Central Nervous System White Matter Injury," *ASME J. Biomech. Eng.*, **122**(6), pp. 615–622.
- [183] LaPlaca, M. C., Prado, G. R., Cullen, D. K., and Irons, H. R., 2006, "High Rate Shear Insult Delivered to Cortical Neurons Produces Heterogeneous Membrane Permeability Alterations," *Conf. Proc. IEEE Eng. Med. Biol. Soc.*, **1**, pp. 2384–2387.
- [184] Hallow, D. M., Seeger, R. A., Kamaev, P. P., Prado, G. R., LaPlaca, M. C., and Prausnitz, M. R., 2008, "Shear-Induced Intracellular Loading of Cells With Molecules by Controlled Microfluidics," *Biotechnol. Bioeng.*, **99**(4), pp. 846–854.
- [185] LaPlaca, M. C., Cullen, D. K., McLoughlin, J. J., and Cargill, R. S., 2nd, 2005, "High Rate Shear Strain of Three-Dimensional Neural Cell Cultures: A New in vitro Traumatic Brain Injury Model," *J. Biomech.*, **38**(5), pp. 1093–1105.
- [186] Geddes, D. M., LaPlaca, M. C., and Cargill, R. S., 2nd, 2003, "Susceptibility of Hippocampal Neurons to Mechanically Induced Injury," *Exp. Neurol.*, **184**(1), pp. 420–427.
- [187] McKinney, J. S., Willoughby, K. A., Liang, S., and Ellis, E. F., 1996, "Stretch-Induced Injury of Cultured Neuronal, Glial, and Endothelial Cells. Effect of Polyethylene Glycol-Conjugated Superoxide Dismutase," *Stroke*, **27**(5), pp. 934–940.
- [188] Lusardi, T. A., Wolf, J. A., Putt, M. E., Smith, D. H., and Meaney, D. F., 2004, "Effect of Acute Calcium Influx After Mechanical Stretch Injury in vitro on the Viability of Hippocampal Neurons," *J. Neurotrauma*, **21**(1), pp. 61–72.
- [189] Spaethling, J. M., Geddes-Klein, D. M., Miller, W. J., von Reyn, C. R., Singh, P., Mesfin, M., Bernstein, S. J., and Meaney, D. F., 2007, "Linking Impact to Cellular and Molecular Sequelae of CNS Injury: Modeling in vivo Complexity With in vitro Simplicity," *Prog. Brain Res.*, **161**, pp. 27–39.
- [190] Ferrario, C. R., Ndukwe, B. O., Ren, J., Satin, L. S., and Goforth, P. B., 2013, "Stretch Injury Selectively Enhances Extrasynaptic, GluN2B-Containing NMDA Receptor Function in Cortical Neurons," *J. Neurophysiol.*, **110**(1), pp. 131–140.
- [191] Goforth, P. B., Ellis, E. F., and Satin, L. S., 2004, "Mechanical Injury Modulates AMPA Receptor Kinetics via an NMDA Receptor-Dependent Pathway," *J. Neurotrauma*, **21**(6), pp. 719–732.
- [192] Kao, C. Q., Goforth, P. B., Ellis, E. F., and Satin, L. S., 2004, "Potentiation of GABA(A) Currents After Mechanical Injury of Cortical Neurons," *J. Neurotrauma*, **21**(3), pp. 259–270.
- [193] Di, X., Goforth, P. B., Bullock, R., Ellis, E., and Satin, L., 2000, "Mechanical Injury Alters Volume Activated Ion Channels in Cortical Astrocytes," *Acta Neurochir. Suppl.*, **76**, pp. 379–383.
- [194] Goforth, P. B., Ellis, E. F., and Satin, L. S., 1999, "Enhancement of AMPA-Mediated Current After Traumatic Injury in Cortical Neurons," *J. Neurosci.*, **19**(17), pp. 7367–7374.
- [195] Singh, P., Doshi, S., Spaethling, J. M., Hockenberry, A. J., Patel, T. P., Geddes-Klein, D. M., Lynch, D. R., and Meaney, D. F., 2012, "N-methyl-D-aspartate Receptor Mechanosensitivity is Governed by C Terminus of NR2B Subunit," *J. Biol. Chem.*, **287**(6), pp. 4348–4359.
- [196] Kilinc, D., Gallo, G., and Barbee, K. A., 2009, "Mechanical Membrane Injury Induces Axonal Beading Through Localized Activation of Calpain," *Exp. Neurol.*, **219**(2), pp. 553–561.
- [197] Kilinc, D., Gallo, G., and Barbee, K. A., 2008, "Mechanically-Induced Membrane Poration Causes Axonal Beading and Localized Cytoskeletal Damage," *Exp. Neurol.*, **212**(2), pp. 422–430.
- [198] Barbee, K. A., 2005, "Mechanical Cell Injury," *Ann. NY Acad. Sci.*, **1066**, pp. 67–84.
- [199] Cullen, D. K., Vernekar, V. N., and LaPlaca, M. C., 2011, "Trauma-Induced Plasmalemma Disruptions in Three-Dimensional Neural Cultures Are Dependent on Strain Modality and Rate," *J. Neurotrauma*, **28**(11), pp. 2219–2233.
- [200] LaPlaca, M. C., Prado, G. R., Cullen, D., and Simon, C. M., 2009, "Plasma Membrane Damage as a Marker of Neuronal Injury," *Conf. Proc. IEEE Eng. Med. Biol. Soc.*, pp. 1113–1116.
- [201] Geddes, D. M., Cargill, R. S., 2nd, and LaPlaca, M. C., 2003, "Mechanical Stretch to Neurons Results in a Strain Rate and Magnitude-Dependent Increase in Plasma Membrane Permeability," *J. Neurotrauma*, **20**(10), pp. 1039–1049.
- [202] LaPlaca, M. C., and Thibault, L. E., 1998, "Dynamic Mechanical Deformation of Neurons Triggers an Acute Calcium Response and Cell Injury Involving the N-methyl-D-aspartate glutamate Receptor," *J. Neurosci. Res.*, **52**(2), pp. 220–229.
- [203] Hue, C. D., Cao, S., Haider, S. F., Vo, K. V., Effgen, G. B., Vogel, E. 3rd, Panzer, M. B., Bass, C. R., Meaney, D. F., and Morrison, B., 3rd, 2013, "Blood-Brain Barrier Dysfunction After Primary Blast Injury in vitro," *J. Neurotrauma*, **30**(19), pp. 1652–1663.
- [204] Panzer, M. B., Matthews, K. A., Yu, A. W., Morrison, B., 3rd, Meaney, D. F., and Bass, C. R., 2012, "A Multiscale Approach to Blast Neurotrauma Modeling: Part I—Development of Novel Test Devices for in vivo and in vitro Blast Injury Models," *Front. Neurol.*, **3**, pp. 1–11.
- [205] Arun, P., Abu-Taleb, R., Valiyaveetil, M., Wang, Y., Long, J. B., and Nambiar, M. P., 2012, "Transient Changes in Neuronal Cell Membrane Permeability After Blast Exposure," *Neuroreport*, **23**(6), pp. 342–346.
- [206] Skotak, M., Wang, F., and Chandra, N., 2012, "An in vitro Injury Model for SH-SY5Y Neuroblastoma Cells: effect of Strain and Strain Rate," *J. Neurosci. Meth.*, **205**(1), pp. 159–168.
- [207] Nienaber, M., Lee, J. S., Feng, R., and Lim, J. Y., 2011, "Impulsive Pressurization of Neuronal Cells for Traumatic Brain Injury Study," *J. Vis. Exp.*, **56**(e2723), pp. 1–4.
- [208] Alford, P. W., Dabiri, B. E., Goss, J. A., Hemphill, M. A., Brigham, M. D., and Parker, K. K., 2011, "Blast-Induced Phenotypic Switching in Cerebral Vasospasm," *Proc. Natl. Acad. Sci. USA*, **108**(31), pp. 12705–12710.
- [209] Arun, P., Spadaro, J., John, J., Gharavi, R. B., Bentley, T. B., and Nambiar, M. P., 2011, "Studies on Blast Traumatic Brain Injury Using in-vitro Model With Shock Tube," *Neuroreport*, **22**(8), pp. 379–384.
- [210] Nakagawa, A., Manley, G. T., Gean, A. D., Ohtani, K., Armonda, R., Tsukamoto, A., Yamamoto, H., Takayama, K., and Tominaga, T., 2011, "Mechanisms of Primary Blast-Induced Traumatic Brain Injury: Insights From Shock-Wave Research," *J. Neurotrauma*, **28**(6), pp. 1101–1119.
- [211] Sonden, A., Svensson, B., Roman, N., Ostmark, H., Brismar, B., Palmblad, J., and Kjellstrom, B. T., 2000, "Laser-Induced Shock Wave Endothelial Cell Injury," *Lasers Surg. Med.*, **26**(4), pp. 364–375.
- [212] Cater, H. L., Gitterman, D., Davis, S. M., Benham, C. D., Morrison, B., 3rd, and Sundstrom, L. E., 2007, "Stretch-Induced Injury in Organotypic Hippocampal Slice Cultures Reproduces in vivo Post-Traumatic Neurodegeneration: Role of Glutamate Receptors and Voltage-Dependent calcium Channels," *J. Neurochem.*, **101**(2), pp. 434–447.
- [213] Cater, H. L., Sundstrom, L. E., and Morrison, B., 3rd, 2006, "Temporal Development of Hippocampal Cell Death is Dependent on Tissue Strain but Not Strain Rate," *J. Biomech.*, **39**(15), pp. 2810–2818.
- [214] Elkin, B. S., and Morrison, B., 3rd, 2007, "Region-Specific Tolerance Criteria for the Living Brain," *Stapp Car Crash J.*, **51**, pp. 127–138.
- [215] Morrison, B., 3rd, Cater, H. L., Benham, C. D., and Sundstrom, L. E., 2006, "An in vitro Model of Traumatic Brain Injury Utilising Two-Dimensional Stretch of Organotypic Hippocampal Slice Cultures," *J. Neurosci. Meth.*, **150**(2), pp. 192–201.
- [216] Morrison, B., 3rd, Cater, H. L., Wang, C. C., Thomas, F. C., Hung, C. T., Atehsian, G. A., and Sundstrom, L. E., 2003, "A Tissue Level Tolerance Criterion for Living Brain Developed With an in vitro Model of Traumatic Mechanical Loading," *Stapp Car Crash J.*, **47**, pp. 93–105.
- [217] Morrison, B., 3rd, Meaney, D. F., and McIntosh, T. K., 1998, "Mechanical Characterization of an in vitro Device Designed to Quantitatively Injure Living Brain Tissue," *Ann. Biomed. Eng.*, **26**(3), pp. 381–390.
- [218] Yu, Z., Elkin, B. S., and Morrison, B., 2009, "Quantification of Functional Alterations After in vitro Traumatic Brain Injury," *Conf. Proc. IEEE Eng. Med. Biol. Soc.*, pp. 1135–1138.
- [219] Yu, Z., and Morrison, B., 3rd, 2010, "Experimental Mild Traumatic Brain Injury Induces Functional Alteration of the Developing Hippocampus," *J. Neurophysiol.*, **103**(1), pp. 499–510.
- [220] Mertz, H. J., Irwin, A. L., and Prasad, P., 2003, "Biomechanical and Scaling Bases for Frontal and Side Impact Injury Assessment Reference Values," *Stapp Car Crash J.*, **47**, pp. 155–188.
- [221] Bass, C. R., Rafaels, K. A., and Salzar, R. S., 2008, "Pulmonary Injury Risk Assessment for Short-Duration Blasts," *J. Trauma*, **65**(3), pp. 604–615.
- [222] Prado, G. R., Ross, J. D., DeWeerth, S. P., and LaPlaca, M. C., 2005, "Mechanical Trauma Induces Immediate Changes in Neuronal Network Activity," *J. Neural. Eng.*, **2**(4), pp. 148–158.
- [223] Yu, Z., Graudejus, O., Tsay, C., Lacour, S. P., Wagner, S., and Morrison, B., 3rd, 2009, "Monitoring Hippocampus Electrical Activity in vitro on an Elastically Deformable Microelectrode Array," *J. Neurotrauma*, **26**(7), pp. 1135–1145.
- [224] Duma, S. M., Manoogian, S. J., Bussone, W. R., Brolinson, P. G., Goforth, M. W., Donnenwerth, J. J., Greenwald, R. M., Chu, J. J., and Crisco, J. J., 2005, "Analysis of Real-Time Head Accelerations in Collegiate Football Players," *Clin. J. Sport Med.*, **15**(1), pp. 3–8.
- [225] Rowson, S., Beckwith, J. G., Chu, J. J., Leonard, D. S., Greenwald, R. M., and Duma, S. M., 2011, "A Six Degree of Freedom Head Acceleration Measurement Device for Use in Football," *J. Appl. Biomech.*, **27**(1), pp. 8–14.
- [226] Greenwald, R. M., Gwin, J. T., Chu, J. J., and Crisco, J. J., 2008, "Head Impact Severity Measures for Evaluating Mild Traumatic Brain Injury Risk Exposure," *Neurosurgery*, **62**(4), pp. 789–798, discussion p. 798.
- [227] Crisco, J. J., Wilcox, B. J., Beckwith, J. G., Chu, J. J., Duhaime, A. C., Rowson, S., Duma, S. M., Maerlender, A. C., McAllister, T. W., and Greenwald, R. M., 2011, "Head Impact Exposure in Collegiate Football Players," *J. Biomech.*, **44**(15), pp. 2673–2678.
- [228] Beckwith, J. G., Greenwald, R. M., Chu, J. J., Crisco, J. J., Rowson, S., Duma, S. M., Broglio, S. P., McAllister, T. W., Guskiewicz, K. M., Mihalik, J. P., Anderson, S., Schobel, B., Brolinson, P. G., and Collins, M. W., 2013, "Head

- Impact Exposure Sustained by Football Players on Days of Diagnosed Concussion," *Med. Sci. Sports Exerc.*, **45**(4), pp. 737–746.
- [229] Pellman, E. J., Viano, D. C., Tucker, A. M., and Casson, I. R., 2003, "Concussion in Professional Football: Location and Direction of Helmet Impacts—Part 2," *Neurosurgery*, **53**(6), pp. 1328–1340, discussion pp. 1340–1321.
- [230] Pellman, E. J., Viano, D. C., Tucker, A. M., Casson, I. R., and Waeckerle, J. F., 2003, "Concussion in Professional football: Reconstruction of game Impacts and Injuries," *Neurosurgery*, **53**(4), pp. 799–812, discussion pp. 812–794.
- [231] Casson, I. R., Viano, D. C., Powell, J. W., and Pellman, E. J., 2010, "Twelve years of national football league Concussion data," *Sports Health*, **2**(6), pp. 471–483.
- [232] Jadschke, R., Viano, D. C., Dau, N., King, A. I., and McCarthy J., 2013, "On the Accuracy of the Head Impact Telemetry (HIT) System Used in Football Helmets," *J. Biomech.*, **46**(13), pp. 2310–2315.
- [233] Allison, M. A., Kang, Y. S., Maltese, M. R., Bolte, J. H. 4th, Arbogast, K. B., 2013, "Validation of a Helmet-Based System to Measure Head Impact Biomechanics in Ice Hockey," *Med. Sci. Sports Exerc.*, **46**(1), pp. 115–123.
- [234] Chu, J. J., Beckwith, J. G., Leonard, D. S., Paye, C. M., and Greenwald, R. M., 2012, "Development of a Multimodal Blast Sensor for Measurement of Head Impact and Over-Pressurization Exposure," *Ann. Biomed. Eng.*, **40**(1), pp. 203–212.
- [235] Wu, N., Wang, W., Tian, Y., Zou, X., Maffeo, M., Niezrecki, C., Chen, J., and Wang, X., 2011, "Low-Cost Rapid Miniature Optical Pressure Sensors for Blast Wave Measurements," *Opt. Express*, **19**(11), pp. 10797–10804.
- [236] Cullen, D. K., Browne, K. D., Xu, Y., Adeb, S., Wolf, J. A., McCarron, R. M., Yang, S., Chavko, M., and Smith, D. H., 2011, "Blast-Induced Color Change in Photonic Crystals Corresponds With Brain Pathology," *J. Neurotrauma*, **28**(11), pp. 2307–2318.
- [237] Palm, E. J., Bass, C. R., Panzer, M. B., Shridharani, J., Salzar, R. S., Rafaels, K. A., Waliko, T., Weiss, G., Perritt, C., Haynes, N., and Masters, K., 2010, "Test Methodology for the Assessment of Blast Trauma Behind Military Helmets," Personal Armor Systems Symposium (PASS-2010), Quebec City, Canada.
- [238] Shridharani, J., Wood, G. W., Panzer, M. B., Matthews, K. A., Perritt, C., Masters, K., and Bass, C. R., 2012, "Blast Effects Behind Ballistic Protective Helmets," Personal Armor Systems Symposium Nuremberg, Germany.
- [239] Smith, D. H., Johnson, V. E., and Stewart W., 2013, "Chronic Neuropathologies of Single and Repetitive TBI: Substrates of Dementia?," *Nat. Rev. Neurol.*, **9**(4), pp. 211–221.