

# FOXE1 Association with Differentiated Thyroid Cancer and Its Progression

Marissa Penna-Martinez,<sup>1</sup> Friederike Epp,<sup>1</sup> Heinrich Kahles,<sup>1</sup> Elizabeth Ramos-Lopez,<sup>1</sup> Nora Hinsch,<sup>2</sup> Martin-Leo Hansmann,<sup>2</sup> Ivan Selkinski,<sup>3</sup> Frank Grünwald,<sup>3</sup> Katharina Holzer,<sup>4</sup> Wolf O. Bechstein,<sup>4</sup> Stefan Zeuzem,<sup>1</sup> Christian Vorländer,<sup>5</sup> and Klaus Badenhop<sup>1</sup>

**Background:** Single nucleotide polymorphisms (SNPs) near thyroid transcription factor genes (*FOXE1* rs965513/*NKX2-1* rs944289) have been shown to be associated with differentiated thyroid cancer (DTC) in Caucasoid populations. We investigated the role of those SNPs in German patients with DTC and also extended our analysis to tumor stages and lymphocytic infiltration of the tumors (ITL).

**Methods:** Patients with DTC ( $n=243$ ; papillary, PTC; follicular, FTC) and healthy controls (HC;  $n=270$ ) were analyzed for the rs965513 and rs944289 SNPs.

**Results:** The case-control analysis for rs965513 SNP showed that the genotypes “AA,” “AG,” and minor allele “A” were more frequent in patients with DTC than in HC (pronounced in PTC  $p_{\text{genotype}}=0.000084$ ,  $p_{\text{allele}}=0.006$  than FTC  $p_{\text{genotype}}=0.29$  and  $p_{\text{allele}}=0.06$ ). Furthermore, subgroup analysis of the DTC patients stratified for primary tumor stage (T1–T2, T3–T4), the absence or presence of regional lymph node metastases (N0, N1), for distant metastases (M0, M1), as well as for ITL, showed an association of rs965513 with stages T1–T2, T1–T3, N1, and absence of ITL. The *NKX2-1* SNP rs944289, however, was not associated with DTC.

**Conclusion:** Our results confirm that the *FOXE1* rs965513 SNP confers an increased risk for DTC in the German population, particularly allele “A” and the genotypes “AA” and “AG” for PTC. This increased risk was also observed in advanced tumor stages and absence of ITL, which may reflect the course of a more aggressive disease. The *NKX2-1* rs944289 SNP, however, appears to play a secondary role in the development of DTC in the German population.

## Introduction

THE MAJORITY OF MALIGNANT THYROID TUMORS derive from follicular epithelial cells. The differentiated thyroid cancers (DTC) are papillary (PTC; ~85% of cases) and follicular (FTC; ~10% of cases) subtypes (1), and their etiology is still not well characterized. The transformation of normal into malignant tissue is a multifactorial process involving genetic and environmental factors such as radiation exposure (2). Some environmental factors such as hepatitis C infection or late and multiple pregnancies have been suggested as risk factors, but they have not been confirmed (3,4). Moreover, genetic components—for example proto-oncogenes, which encode receptor tyrosine kinases (*RET*) and small GTPases (*RAS*)—may contribute to thyroid cancer formation in familial syndromes (5). However, none of these factors is thyroid specific, and therefore the identification of additional

candidate genes is important for a better understanding of the pathogenesis of thyroid cancer. In this context, key genes involved in thyroid organogenesis, for example the thyroid transcription factors (TTFs), have been identified as genetic susceptibility factors of various thyroid conditions. This illustrates the pivotal function of these genes in the development of thyroid diseases (6). Recently, three genome wide association (GWA) studies demonstrated associations of single nucleotide polymorphisms (SNPs) located near the *TTF-1* (also known as thyroid-specific enhancer-binding protein: *NKX2-1*, rs944289) and *TTF-2* (also known as forkhead box protein: *FOXE1*, rs965513) loci with an increased risk of sporadic thyroid carcinoma in Icelandic, British, and Japanese populations (7–9). Radiation-related PTC of Belarusian patients showed the strongest association only with the *FOXE1* gene (10). In order to elucidate the role of TTFs as susceptibility genes in DTC of the German

<sup>1</sup>Department of Internal Medicine I, Division of Endocrinology, Diabetes and Metabolism, <sup>2</sup>Senckenberg Institute for Pathology, and Departments of <sup>3</sup>Nuclear Medicine and <sup>4</sup>Surgery, Goethe-University Hospital Frankfurt, Frankfurt am Main, Germany.

<sup>5</sup>Department of Surgery, Bürgerhospital Frankfurt, Frankfurt am Main, Germany.

population, in the present case-control study, we investigated the SNPs near *FOXE1* and *NKX2-1* genes (rs965513 and rs944289 respectively). In addition, we explored whether the tumor stage and an immune-cell infiltration in the thyroid gland modified the risk of DTC.

## Subjects and Methods

### Subjects

In total, 243 patients (160 females, 83 males) with a pathologically confirmed diagnosis of DTC (196 papillary: 130 females and 66 males; 47 follicular: 30 females and 17 males) and known tumor node metastasis (TNM) stage were recruited from the Department of Nuclear Medicine at the University Hospital Frankfurt am Main, Germany. Healthy controls (HC;  $n=270$ , 113 females, 157 males) were volunteer blood donors from the staff personnel or medical students from the University Hospital Frankfurt am Main, Germany. All participants were of Caucasian (mostly German) origin and inhabitants of the area surrounding Frankfurt am Main, Germany. The median age of the patients with DTC and HC was 55 years (range 14–92 years) and 40 years (range 23–101 years) respectively. The study protocol was approved by the Ethics Committee of the University Hospital Frankfurt am Main, and informed consent was obtained from all participants.

### SNP selection

In total, two SNPs (rs965513 and rs944289) previously associated with thyroid cancer susceptibility in the Icelandic population were investigated (7). While the rs965513 variant (A/G; Chromosome 9) is located ~59 kb centromeric to the *FOXE1* gene, the rs944289 variant (C/T; Chromosome 14) is situated 337 kb telomeric to the *NKX2-1* gene. The SNP positions are given according to the National Center for Biotechnology Information (NCBI; www.ncbi.nlm.nih.gov).

### Genotyping

Genomic DNA from whole blood containing ethylenediaminetetraacetic acid was isolated by salting out procedure (11) and subjected to real time polymerase chain reaction (PCR). For the genotype analysis, the PCR reaction was carried out in 25  $\mu$ L containing 3  $\mu$ L (60 ng) DNA template, 6.25  $\mu$ L ABsoluteTMQPCR mix (Abgene®; Thermo Fischer Scientific, Schwerte, Germany), 15.45  $\mu$ L H<sub>2</sub>O, and 0.3  $\mu$ L Taqman assay (*FOXE1* rs965513/C\_1593670\_20, or *NKX2-1* rs944289/C\_1444137\_10), and analyzed in an ABI 7300 PCR system (Applied Biosystems, Darmstadt, Germany). The conditions were denaturation at 95°C for 10 min followed by 50 cycles at 92°C for 50 s and 60°C for 1 min. In order to confirm the accuracy of the method, random samples of the *FOXE1* rs965513 and the *NKX2-1* rs944289 SNPs were genotyped twice with a concordance of 100%.

### Lymphocytic infiltration and thyroid antibodies

Fifty-five tissues from patients with DTC were examined for lymphocyte infiltration using the alkaline phosphatase anti-alkaline phosphatase (APPAP) method (12). Additionally, thyroglobulin (TG) and thyroid peroxidase (TPO) antibodies (Abs) were measured using an enzyme-linked immunosorbent assay (Phadia, Freiburg, Germany). Concentrations >100 UI/mL for thyroid Abs were defined as positive.

### Statistical analysis

Deviation from Hardy–Weinberg equilibrium and differences in genotype and allele distributions between groups were evaluated by chi-square test (BiAS software, package 9.08; Epsilon, Weinheim, Germany). The odds ratio (OR) and its confidence interval [CI] were estimated by unconditional logistic regression as a measure of the associations between genotypes or alleles and thyroid cancer risk. In order to elucidate the role of TTFs as susceptibility genes in DTC of the German population, the comparison of the genotype/allele frequencies between cases and controls were analyzed for each cancer group separately (DTC/PTC/FTC vs. HC respectively, DTC<sub>female</sub>/PTC<sub>female</sub>/FTC<sub>female</sub> vs. HC<sub>female</sub> respectively, and DTC<sub>male</sub>/PTC<sub>male</sub>/FTC<sub>male</sub> vs. HC<sub>male</sub> respectively). The observed  $p$ -values were corrected by multiplication with the number of genotypes<sup>( $\times 3$ )</sup>/alleles<sup>( $\times 2$ )</sup> tested. Furthermore, to address whether a certain risk genotype/allele influences the progression of DTC, the patients were classified in four groups with regard to the extent of the primary tumor (T), regional lymph node metastases (N), distant metastatic lesions (M), and thyroid lymphocytic infiltration (ITL). The comparisons were performed within each of those groups (see Table 2) using the Mantel–Haenszel procedure. With a significance at  $p_{\text{global}} \leq 0.05$ , a Multiple Mantel–Haenszel comparison was applied and corrected  $p$ -values ( $p_c$ ) were calculated using the Bonferroni method (13). The assumed significance level was  $\leq 0.05$ . A power calculation for each group was performed assuming an allele frequency of 34.1% (rs965513 A) and 57.3% (rs944289 T) (7) and a type I error rate of 5%. On the basis of these assumptions, we estimate that we have more than 80% power to detect an allelic odds ratio of 1.44 (DTC), 1.47 (PTC), and 1.9 (FTC) for disease susceptibility in the case/control data set by using the program Power and Sample Size Calculations v3.0 (14).

## Results

In our study, patients with DTC ( $n=243$ ) had 81% papillary and 19% follicular subtypes, and there were more females (66%) than males (34%). All genotypes were in Hardy–Weinberg equilibrium ( $p > 0.05$ ) for the two investigated SNPs.

### Genotype and allele analysis and DTC

The case-controls analysis for *FOXE1* rs965513 SNP showed that the genotypes “AA” and “AG” (17.3% vs. 15.2% and 57.2% vs. 40.7% respectively) were more frequent in patients with DTC than in HC, while the genotype “GG” was less frequent (25.5% vs. 44.1%;  $p_c=0.00014$ ). Additionally, the allele “A” was observed more often in DTC patients than in HC (45.9% vs. 35.6%; OR 1.54 [CI 1.20–1.97],  $p_c=0.0019$ ). These differences in the genotype and allele distribution between patients and controls was more pronounced in the DTC histological PTC subtype ( $p_{c \text{ genotype}}=0.000084$  and  $p_{c \text{ allele}}=0.006$  respectively) than FTC ( $p_{c \text{ genotype}}=0.29$  and  $p_{c \text{ allele}}=0.06$  respectively). Subsequently, a sex-stratified analysis revealed differences for the *FOXE1* rs965513 genotypes in female as well as in male DTC/PTC patients compared with their corresponding HC group ( $p_c=0.01/p_c=0.01$  and  $p_c=0.03/p_c=0.007$  respectively). This result was not confirmed in FTC ( $p_c=0.33$  and  $p_c=0.55$  respectively). Finally, the comparison of the allele

TABLE 1. DISTRIBUTION OF *FOXE1* rs965513 AND *NKX2-1* rs944289 GENE SINGLE NUCLEOTIDE POLYMORPHISMS IN PATIENTS WITH DIFFERENTIATED THYROID CANCER, PAPILLARY THYROID CANCER, AND FOLLICULAR THYROID CANCER COMPARED WITH HEALTHY CONTROLS

Group	n	Genotype [n (frequency)]				p	p <sub>c</sub>	Allele [n (frequency)]				OR [CI]	p	p <sub>c</sub>	
		AA	AG	GG	GA			A	G	A	G				
<b><i>FOXE1</i> rs965513</b>															
All															
HC	270	41 (15.2%)	110 (40.7%)	119 (44.1%)			192 (35.6%)	348 (64.4%)							
DTC	243	42 (17.3%)	139 (57.2%)	62 (25.5%)	<b>0.000046</b>	<b>0.000014</b>	223 (45.9%)	263 (54.1%)				1.54 [1.20–1.97]	0.65 [0.51–0.84]	<b>0.00096</b>	<b>0.0019</b>
PTC	196	30 (15.3%)	118 (60.2%)	48 (24.5%)	<b>0.000028</b>	<b>0.000084</b>	178 (45.4%)	214 (54.6%)				1.51 [1.16–1.97]	0.66 [0.51–0.87]	<b>0.003</b>	<b>0.006</b>
FTC	47	12 (25.5%)	21 (44.7%)	14 (29.8%)	0.097	0.29	45 (47.9%)	49 (52.1%)				1.66 [1.07–2.59]	0.60 [0.39–0.93]	<b>0.03</b>	0.06
Females															
HC	113	21 (18.6%)	43 (38.1%)	49 (43.4%)			85 (37.6%)	141 (62.4%)							
DTC	160	34 (21.3%)	87 (54.4%)	39 (24.4%)	<b>0.0034</b>	<b>0.01</b>	155 (48.4%)	165 (51.6%)				1.56 [1.10–2.21]	0.64 [0.45–0.91]	<b>0.01</b>	<b>0.03</b>
PTC	130	25 (19.2%)	73 (56.2%)	32 (24.6%)	<b>0.0051</b>	<b>0.01</b>	123 (47.3%)	137 (52.7%)				1.49 [1.04–2.14]	0.67 [0.47–0.97]	<b>0.04</b>	0.08
FTC	30	9 (30.0%)	14 (46.7%)	7 (23.3%)	0.11	0.33	32 (53.3%)	28 (46.7%)				1.90 [1.07–3.37]	0.53 [0.30–0.94]	<b>0.04</b>	0.08
Males															
HC	157	20 (12.7%)	67 (42.7%)	70 (44.6%)			107 (34.1%)	207 (65.9%)							
DTC	83	8 (9.60%)	52 (62.7%)	23 (27.7%)	<b>0.01</b>	<b>0.03</b>	68 (41.0%)	98 (59.0%)				1.34 [0.91–1.98]	0.74 [0.51–1.10]	0.16	0.32
PTC	66	5 (7.60%)	45 (68.2%)	16 (24.2%)	<b>0.0023</b>	<b>0.007</b>	55 (41.7%)	77 (58.3%)				1.38 [0.91–2.10]	0.72 [0.48–1.10]	0.16	0.32
FTC	17	3 (17.6%)	7 (41.2%)	7 (41.2%)	0.85	2.55	13 (38.2%)	21 (61.8%)				1.20 [0.58–2.49]	0.84 [0.40–1.73]	0.77	1.54
<b><i>NKX2-1</i> rs944289</b>															
All															
HC	270	47 (17.4%)	128 (47.4%)	95 (35.2%)			222 (41.1%)	318 (58.9%)							
DTC	243	42 (17.3%)	109 (44.9%)	92 (37.9%)	0.8	2.4	193 (39.7%)	293 (60.3%)				0.94 [0.73–1.21]	1.06 [0.83–1.36]	0.69	1.4
PTC	196	34 (17.3%)	93 (47.4%)	69 (35.2%)	1.0	3.0	161 (41.1%)	231 (58.9%)				1.00 [0.77–1.30]	1.00 [0.77–1.30]	0.95	1.9
FTC	47	8 (17.0%)	16 (34.0%)	23 (48.9%)	0.16	0.48	32 (34.0%)	62 (66.1%)				0.74 [0.47–1.17]	1.35 [0.85–2.14]	0.24	0.48
Females															
HC	113	21 (18.6%)	56 (49.6%)	36 (31.9%)			98 (43.4%)	128 (56.6%)							
DTC	160	30 (18.8%)	70 (43.8%)	60 (37.5%)	0.58	1.74	130 (40.6%)	190 (59.4%)				0.89 [0.63–1.26]	1.12 [0.79–1.58]	0.58	1.16
PTC	130	25 (19.2%)	59 (45.4%)	46 (35.4%)	0.79	2.37	109 (41.9%)	151 (58.1%)				0.94 [0.66–1.35]	1.06 [0.74–1.52]	0.81	1.62
FTC	30	5 (16.7%)	11 (36.7%)	14 (46.7%)	0.30	0.9	21 (35.0%)	39 (65.0%)				0.70 [0.39–1.27]	1.42 [0.79–2.57]	0.31	0.62
Males															
HC	157	26 (16.6%)	72 (45.9%)	59 (37.6%)			124 (39.5%)	190 (60.5%)							
DTC	83	12 (14.5%)	39 (47.0%)	32 (38.6%)	0.91	2.73	63 (38.0%)	103 (62.0%)				0.94 [0.64–1.38]	1.07 [0.72–1.57]	0.82	1.64
PTC	66	9 (13.6%)	34 (51.5%)	23 (34.8%)	0.72	2.16	52 (39.4%)	80 (60.6%)				1.00 [0.66–1.51]	1.00 [0.66–1.52]	0.93	1.86
FTC	17	3 (17.6%)	5 (29.4%)	9 (52.9%)	0.39	1.17	11 (32.4%)	23 (67.6%)				0.73 [0.35–1.56]	1.37 [0.64–2.90]	0.53	1.06

*p*-Values <0.05 are in bold type.

CI, confidence interval; DTC, differentiated thyroid cancer; FTC, follicular thyroid cancer; HC, healthy controls; OR, odds ratio; p<sub>c</sub>, corrected *p*-values; PTC, papillary thyroid cancer.

distribution between patients and HC reached significant differences only in the DTC/PTC/FTC females but not in DTC/PTC/FTC males ( $p_c=0.03/p_c \text{ trend}=0.08/p_c \text{ trend}=0.08$  and  $p_c=0.32/p_c=0.32/p_c=1.54$  respectively). In contrast, the genotypes and alleles of the *NKX2-1* rs944289 SNP showed no associations with DTC (Table 1). In order to detect a possible contribution of the *NKX2-1* rs944289 SNP to DTC susceptibility, we analyzed its genotype combinations with *FOXE1* rs965513 (presented in Supplementary Table S1; Supplementary Data are available online at [www.liebert.com/thy](http://www.liebert.com/thy)), revealing nine different combinations. DTC patients carry the genotype combination “GG-rs965513/CT-rs944289” less often than HC (DTC,  $p_c=0.0027$ ; PTC,  $p_c=0.02$ ; FTC,  $p_c=0.18$ ). There were also significant differences between patients and controls in the *FOXE1* rs965513 carrying the *NKX2-1* rs944289 “CT” genotype, but not the *NKX2-1* rs944289 “TT” or “CC” genotypes (Supplementary Table S2).

*Genotype and allele analysis of FOXE1 rs965513 SNP in DTC according to tumor stages*

The patients were grouped according to the extent of the primary tumor (T1–T2, T3–T4), the absence or presence of regional lymph node metastases (N0, N1), and the absence or presence of distant metastatic lesions (M0, M1). The genotypes “AA,” “AG,” or allele “A” were more frequent both in patients with T1–T2 as well as those with T3–T4 in comparison to controls (both  $p_c \text{ genotype}$  and  $p_c \text{ allele}=0.02$ ;  $p_c \text{ genotype}=0.06$  and  $p_c \text{ allele}=0.03$  respectively). Also, the genotypes “AG” or allele “A” were only found more frequently in the group of patients with N1 compared to HC (both  $p_c \text{ genotype}$  and  $p_c \text{ allele}=0.03$ ). In contrast, no association of the rs965513 SNP with M (M0:  $p_c \text{ allele}=0.18$ ; M1:  $p_c \text{ allele}=0.12$ ) was observed. The patient groups with advanced stages T3–T4, N1, or M1 had higher odds ratios in the presence of allele “A” (Table 2). However, the frequency of allele “A” in the advanced stage groups in comparison with the groups with a low/nonadvanced stage did not reach the level of significance (T3–T4: 47% vs. T1–T2: 45.1%; N1: 46.8% vs. N0: 43.7%; and M1: 52.8% vs. M0: 46.3%;  $p_c \text{ allele}=2.16$ ,  $p_c \text{ allele}=1.77$ ,  $p_c \text{ allele}=1.59$  respectively).

*Thyroid antibodies in DTC patients in relation to the FOXE1 rs965513 genotypes*

In 121 DTC patients, thyroid antibodies (TPO-Ab/TG-Ab) levels were available. Among them, nine (7.4%) were positive (pos) and 112 (92.6%) negative (neg) for TPO-Ab and/or TG-Ab titers. A comparison of the rs965513 SNP in patients with TPO-Ab/TG-Ab<sup>pos</sup> and TPO-Ab/TG-Ab<sup>neg</sup> was performed and showed no differences between the groups ( $p_{\text{genotypes}}=0.79$  and  $p_{\text{allele}}=0.92$ ). A subsequent analysis in which the median values of TPO-Ab and TG-Ab in DTC patients were compared revealed no differences between the various genotypes of the rs965513 SNP (TPO-Ab AA: 1 IU/mL, AG: 2 IU/mL, GG 3 IU/mL,  $p_{\text{global}}=0.21$ ; TG-Ab AA: 10 IU/mL, AG: 14 IU/mL, GG: 13 IU/mL,  $p_{\text{global}}=0.37$ ).

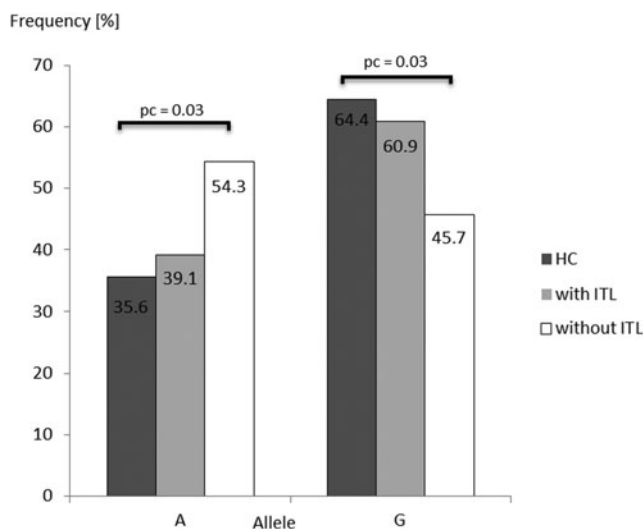
*Lymphocytic infiltration in DTC in relation to FOXE1 rs965513 SNP*

Histopathology reports were available for 55 patients with DTC (33 females, 22 males; median age 47 years). Among

TABLE 2. DISTRIBUTION OF *FOXE1* rs965513 SINGLE NUCLEOTIDE POLYMORPHISMS IN PATIENTS WITH DIFFERENTIATED THYROID CARCINOMA ACCORDING TO THE EXTENT OF PRIMARY TUMOR AND THE ABSENCE OR PRESENCE OF REGIONAL LYMPH NODE METASTASES AND DISTANT METASTATIC LESIONS

Group	n	Genotype [n (frequency)]			p	p <sub>c</sub>	Allele [n (frequency)]			OR [CI]	p	p <sub>c</sub>
		AA	AG	GG			A	G	A			
<i>FOXE1</i> rs965513												
HC	270	41 (15.2%)	110 (40.7%)	119 (44.1%)		192 (35.6%)	348 (64.4%)					
T1–T2	144	23 (16.0%)	84 (58.3%)	37 (25.7%)	<b>0.0075</b>	130 (45.1%)	158 (54.9%)	1.49 [1.11–2.00]	0.67 [0.50–0.90]	<b>0.0071</b>	<b>0.02</b>	
T3–T4	66	13 (19.7%)	36 (54.5%)	17 (25.8%)	<b>0.02</b>	62 (47.0%)	70 (53.0%)	1.61 [1.09–2.36]	0.62 [0.42–0.92]	<b>0.01</b>	<b>0.03</b>	
T1–T2 vs. T3–T4					0.70					0.72	2.16	
HC	270	41 (15.2%)	110 (40.7%)	119 (44.1%)		192 (35.6%)	348 (64.4%)					
N0	71	12 (16.9%)	38 (53.5%)	21 (29.6%)	0.09	62 (43.7%)	80 (56.3%)	1.40 [0.96–2.04]	0.71 [0.49–1.04]	0.08	0.24	
N1	78	10 (12.8%)	53 (67.9%)	15 (19.2%)	<b>0.01</b>	73 (46.8%)	83 (53.2%)	1.59 [1.11–2.29]	0.63 [0.44–0.90]	<b>0.01</b>	<b>0.03</b>	
N0 vs. N1					0.54					0.59	1.77	
HC	270	41 (15.2%)	110 (40.7%)	119 (44.1%)		192 (35.6%)	348 (64.4%)					
M0	40	8 (20.0%)	21 (52.5%)	11 (27.5%)	0.08	37 (46.3%)	43 (53.8%)	1.56 [0.97–2.50]	0.64 [0.40–1.03]	0.06	0.18	
M1	18	4 (22.2%)	11 (61.1%)	3 (16.7%)	<b>0.05</b>	19 (52.8%)	17 (47.2%)	2.03 [1.03–3.99]	0.49 [0.25–0.97]	<b>0.04</b>	0.12	
M0 vs. M1					0.49					0.52	1.56	

p-Values <0.05 are in bold type. Group T:  $p_{\text{global/genotype}}=0.0060$ ,  $p_{\text{global/allele}}=0.0059$ ; group N:  $p_{\text{global/genotype}}=0.02$ ,  $p_{\text{global/allele}}=0.04$ ; group M:  $p_{\text{global/genotype}}=0.04$ ,  $p_{\text{global/allele}}=0.03$ .



**FIG. 1.** Comparison of allelic (A;G) distribution for *FOXE1* rs965513 SNP in DTC patients with or without thyroid lymphocytic infiltration (ITL) and healthy controls (HC). *p*-Values <0.05 are shown. SNP, single-nucleotide polymorphism; DTC, differentiated thyroid cancer; *p<sub>c</sub>*, corrected *p*-value.

them, 58.2% showed evidence for a lymphocytic infiltrate. Among the 32 patients with lymphocytic infiltrates within the thyroid gland, 31% suffered from an autoimmune thyroid disease (nine Hashimoto's thyroiditis and one Graves' disease), and 6% were only positive for thyroid antibodies. Whether the lymphocyte infiltrate interacts with the allele risk of DTC was addressed by dividing into two groups—(i) absence or (ii) presence of lymphocytic infiltration—and comparing both groups with HC and among themselves ( $p_{\text{global}}^{\text{allele}}=0.04$ ). Only DTC patients without lymphocytic infiltrate had the “A” allele significantly more often and the “G” allele less often compared to HC (54.3% vs. 35.6%, OR 2.16 [CI 1.18–3.96]; 45.7% vs. 64.4%, OR 0.46 [CI 0.25–0.85];  $p=0.01$ ,  $p_c=0.03$ , respectively; Fig. 1). In contrast, patients with lymphocytic infiltration showed no differences in the allele distribution (“A” allele 39.1% vs. 35.6%, OR 1.16 [CI 0.68–1.98]; “G” allele, 60.9% vs. 64.4%, OR 0.86 [CI 0.51–1.41],  $p=0.58$ ,  $p_c=1.74$ ). Furthermore, no difference in allele distribution was observed between patients with lymphocytic infiltration and those without ( $p=0.11$ ,  $p_c=0.33$ ).

## Discussion

The development of the thyroid gland and its normal migration in the embryonic stage, as well as the maintenance of the differentiated state throughout life, depends on the interplay between three TTFs: *TTF-1/NKX2-1*, *TTF-2/FOXO1*, and paired box 8 (*PAX-8*) (15). That set of transcriptional factors is unique in the thyroid follicular cell type and regulates the transcription of genes such as thyrotropin receptor, thyroperoxidase, thyroglobulin, and sodium iodide symporter that are critical for thyroid hormone synthesis (16). Besides their role for normal thyroid physiology, they show an altered expression in thyroid tumors and are probably involved in thyroid oncogenesis (17,18). A GWA study, analyzing

Icelandic cases and cohorts of European descent, showed that two SNPs, situated on chromosomes 9q22.33 (near *FOXO1*: rs965513) and 14q13.3 (near *NKX2-1*: rs944289), are associated with both PTC and FTC (7). This finding has been confirmed in other populations (British and Japanese) (8,9) but not yet in the German or other Central European population. Also, whether certain genotypes/alleles of the rs965513 and rs944289 SNPs differentiate the risk for tumor progression is unknown so far. Lymphocytic infiltration within the thyroid gland is frequently observed in DTC, and its presence surrounding or within the tumor might be a useful marker predicting a favorable prognosis. It has been described that DTC patients with lymphocytic infiltration tend not to develop recurrent disease (19). Therefore, it is of interest whether certain alleles of the rs965513 and rs944289 SNPs are associated with such intrathyroidal immune reaction in DTC patients.

Our results demonstrate that solely the *FOXO1* rs965513 variant affects the risk for DTC, and this finding is more pronounced in PTC than in FTC. Remarkably, the association between *FOXO1* rs965513 SNP and DTC was found especially in individuals with extent of the primary tumor (T1–T2 vs. HC, T3–T4 vs. HC), affected lymph nodes (N1 vs. HC), and was detectable in patients without lymphocytic infiltrate (without ITL vs. HC). We therefore assume that patients with this *FOXO1* risk allele also carry a risk for more aggressive disease, but the data need to be confirmed with a larger cohort and additional BRAF status screening in thyroid tissues of DTC patients. These examinations have already been performed by Matsuse *et al.* (9). That work investigated the BRAF mutation status in PTC patients according to the *FOXO1* rs965513 in a Japanese cohort and found an association of rs965513 with BRAF<sup>V600E-pos</sup> but not with BRAF<sup>V600E-neg</sup> cases. Because there was no difference between the subgroups BRAF<sup>V600E-neg</sup> and BRAF<sup>V600E-pos</sup>, the authors concluded that a clear relationship between *FOXO1* rs965513 SNP and the BRAF<sup>V600E</sup> status does not exist. It is important to note that thyroid carcinomas in the Japanese population show a higher incidence of PTC and higher BRAF<sup>V600E-pos</sup> rate than in the European population (80% and 50% respectively) (9). Also, the role of the intrathyroidal immune cells in DTC needs to be studied with larger numbers and in more detail. In this regard, we found no association between the *FOXO1* rs965513 SNP and thyroid antibody status as well as lymphocytic infiltration in DTC patients. However, there was a limited number of patients who were either thyroid antibody positive or who had lymphocytic infiltrations. Future and sufficiently powered studies must address whether the *FOXO1* rs965513 SNP is involved if DTC develops in the background of autoimmune thyroid disease.

The association between *FOXO1* rs965513 and thyroid cancer in the present work confirms the results of other studies. The association with DTC (only PTC or both cancer subtypes PTC/FTC) is marked by the minor allele “A” of the *FOXO1* rs965513 SNP (7–10,20). In contrast, Denny *et al.* (6) found in a thyroid-related phenotype study only a trend ( $p=0.09$ ), attributed to the smaller number of patients ( $n=96$ ). Also, the lack of an association between FTC and the rs965513 SNP observed in the present study may be due to the smaller number of our patients, since the power to detect a suspected OR of 1.75 (7) is too low for this sample size. Here, only an OR of 1.9 can be detected with sufficient power

(80%). It is noteworthy that association studies on other SNPs near *FOXE* reinforce this region on chromosome 9 as a good candidate for thyroid cancer susceptibility: rs965513 is in strong pair-wise linkage disequilibrium with three other SNPs near *FOXE* (rs925489, rs7850258, and rs10759944) (6,10). These SNPs demonstrated a significant association with Chernobyl radiation-related PTC in Belarusian patients (10). Furthermore, another SNP located in the *FOXE1* promoter (rs1867277) and in the same LD region as rs965513 was associated with DTC in Spanish/Italian, Belarusian, and British cohorts (8,21,22). In this context, the risk allele “A” of the SNP rs1867277 is capable of augmenting *FOXE1* transcription by creating a binding site for leucine zipper upstream stimulatory factors 1 and 2 (USF1/ USF2) (22). Also, in three recent studies, both the rs1867277 and the *FOXE1* polyAla tract variants were associated with DTC cancer in Spanish, Australian, and Portuguese cohorts (20,21,23).

The rs944289 SNP is situated in a 249-kb LD-region. Neighboring genes of that region are *Breast Cancer Metastasis Suppressor 1-Like (BRMS1L)*, *MAP3K12 binding inhibitory protein 1 (MBIP)*, *surfactant associated 3 (SFTA3)*, and *NKX2-1*. Among these genes, *NKX2-1* is a suitable candidate gene for thyroid cancer risk because of its role in thyroid development (24), and its altered expression in thyroid tumors (18). It is interesting to note that whereas the *NKX2-1* rs944289 SNP is associated with DTC in Icelandic and Japanese populations, no associations were found in the Belarusian and our present study (7,9,10). However, the case-control comparison for the *NKX2-1* rs944289 SNP from Gudmundsson *et al.* (7) (192 cases and 37,196 controls;  $p=0.000000002$ ) and Takahashi *et al.* (10) (507 cases and 2766 controls;  $p=0.01$ ) reached different  $p$ -values based on asymmetric case/control group sizes. We therefore assume that the limited number of our study participants (243 cases and 270 controls) cannot solely explain the lack of association between DTC and the *NKX2-1* rs944289 SNP (expected for 80% power OR 1.44, reached OR 1.06). Furthermore, the only differences between cases and controls in our current study concerning this SNP are detected by the combination analysis of the two *FOXE1* and *NKX2-1* variants. We identified two genotype combinations: risk “AA-rs965513/CT-rs944289” and protective “GG-rs965513/CT-rs944289.” However, this finding has no impact on overall DTC risk. Moreover, analyses of this gene (addressing the A39V germline mutation) in several nonmedullary thyroid carcinoma families did not confirm *NKX2-1* as a risk gene (25). Finally, independent groups performed case-control studies of TTFs in thyroid cancer including the *NKX2-1* rs944289 SNP and provided contradictory results for an association (7,9,10). Gudmundsson *et al.* (7) found no association in the non-Icelandic individuals (cohort of Spanish origin) between *NKX2-1* and DTC. This may indicate a minor or secondary role of this variant in DTC susceptibility. These observations suggest that differences exist in the genetic profile of the TTFs depending on the population background. It is also conceivable that factors, which are related to rs944289 SNP, have a direct effect on the development of DTC and are more powerful in all population groups. In this direction, Jendzejewska *et al.* (26) investigated the mechanism of how the *NKX2-1* rs944289 SNP might predispose to PTC. They propose a dysregulation in the expression of the PTC suscepti-

bility candidate 3 (*PTCSC3*) located 3.2 kb downstream of rs944289. This long intergenic noncoding RNA gene acts as a tumor suppressor.

In summary, our findings are in agreement with earlier DTC studies that found an association with the *FOXE1* rs965513 SNP, indicating that this variant also confers DTC risk in the German population. Furthermore, we show for the first time that the association is also found in patients with advanced stages of DTC disease (T and N) and in the absence of lymphocytic infiltration in the thyroid gland. This observation may reflect more aggressive disease. In contrast, the *NKX2-1* rs944289 SNP appears to play a secondary role for DTC risk in Germans, and its effect is probably population specific.

The DTC risk gene *FOXE1* may serve to control thyroid growth both in early embryological stages as well as during adult life, and safeguard benign differentiation also under the influence of thyroid cell damage either by carcinogens or radiation effects.

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The authors declare that no competing financial interests exist.

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Address correspondence to:

*Klaus Badenhop, MD  
Department of Internal Medicine I  
Division of Endocrinology, Diabetes and Metabolism  
Goethe-University Hospital Frankfurt  
Theodor-Stern-Kai 7  
D-60590 Frankfurt am Main  
Germany*

*E-mail: badenhop@em.uni-frankfurt.de*