RESEARCH PAPER

Genetic signature of differential sensitivity to stevioside in the Italian population

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Abstract The demand for *diet* products is continuously increasing, together with that for *natural* food ingredients. Stevioside and other steviol glycosides extracted from the leaves of the plant *Stevia rebaudiana* Bertoni are the first natural high-potency sweeteners to be approved for consumption in the United States and the European Union. However, the sweetness of these compounds is generally accompanied by aversive sensations, such as bitter and offtastes, which may constitute a limit to their consumption. Moreover, consumers' differences in sensitivity to high-potency sweeteners are well known, as well as difficulties in characterizing their aftertaste. Recently, TAS2R4 and TAS2R14 have been identified as the receptors that mediate the bitter off-taste of steviol glycosides in vitro. In

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the present study, we demonstrate that *TAS2R4* gene polymorphism rs2234001 and *TAS2R14* gene polymorphism rs3741843 are functional for stevioside bitterness perception.

Keywords Stevioside · Bitter aftertaste · Genetic polymorphisms · hTAS2R4 · hTAS2R14 · hTAS2R38

Introduction

Stevioside (Fig. 1) is the most abundant compound of a group of structurally related high-potency sweeteners, the steviol glycosides, which are secondary metabolites extracted from the leaves of the *Stevia rebaudiana* Bertoni. This plant is native to Paraguay where its leaves have a long history of use as sweetener and to treat several diseases (Kinghorn 2002; Yadav and Guleria 2012; Vega-Gálvez et al. 2012). All these compounds share a common aglycone, known as steviol (*ent*-13-hydroxykaur-16-en-18-oic acid), but differ in the number and types of sugar residues.

All steviol glycosides are high-potency sweeteners, and stevioside has been reported to have a relative sweetness, compared to sucrose, between 210 and 300, depending on the protocol used (Crammer and Ikan 1987; Kinghorn and Soejarto 1986). A detailed study of concentration/response functions for several sweeteners, including stevioside, was presented by DuBois et al. (1991). The demand for this kind of sweeteners in the production of zero and reduced-calorie food products is continuously expanding as a response to increasing health awareness. Such awareness stems from a growing body of evidence showing that overweight and obesity strongly contribute to a large proportion of non-communicable diseases.



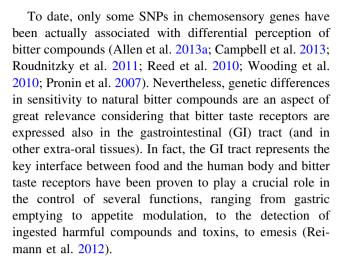
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Fig. 1 Chemical structure of stevioside

As for other high-potency sweeteners (DuBois and Prakash 2012), the sweet taste of steviol glycosides presents bitter and off-tastes (Kinghorn and Soejarto 1986; Prakash et al. 2008). Moreover, consumers' differences in sweeteners sensitivity and acceptance, as well as the difficulties in characterizing their aftertaste, have been reported along with evidences suggesting that most of variation in sensitivity to chemical stimuli may have a genetic basis (Simons et al. 2008).

The recent introduction (2008 in USA, 2011 in EU) of purified steviol glycosides (at least 95 %) as the first natural high-potency sweeteners on the market opens up questions on the genetic basis of stevioside taste perception and its worldwide distribution. Particularly, understanding the differential ability to sense stevioside bitter aftertaste of various human populations has implications on its worldwide use as a natural sweetener and, consequently, on the diffusion of related healthy reduced-calorie food products. Bitter taste is detected by a set of 25 taste 2 receptors (TAS2R) (Meyerhof et al. 2010), and individual differences in the ability to taste substances like phenylthiocarbamide have been known since a long time (Fox 1932). Single nucleotide polymorphisms (SNPs) in the TAS2R38 gene have been identified as the key determinants of this capability, as well as for that of tasting the related compound 6-n-propylthiouracil (PROP) (Bufe et al. 2005; Kim et al. 2003; Duffy et al. 2004). In particular, PROP status (i.e., taster or non-taster) has been proposed by some authors to have a broad effect on other oral sensations and therefore has been considered a marker for food preferences and diet selection (Tepper 2008). However, this issue is highly debated as other studies showed that PROP responsiveness had no influence on food preferences (Baranowski et al. 2011).

Recently, hTAS2R4 and hTAS2R14 have been identified as the receptors that specifically mediate the bitter off-taste of steviol glycosides in vitro (Hellfritsch et al. 2012), opening the way to investigate whether variation at the genes encoding for these receptors could explain individual differences in aftertaste perception.



That being so, in this work, we set to investigate putative associations between stevioside bitter aftertaste and SNPs on genes known to be involved in bitterness perception (i.e., *TAS2R4*, *TAS2R14* and *TAS2R38*), in a panel of Italian subjects. Furthermore, we explore their worldwide patterns of diversity, looking for signatures of ongoing selection at these loci that could explain potential differences in the ability to sense stevioside bitter aftertaste in different human populations.

Subjects and methods

Study design and sampled population

A total of 86 healthy adult Italian donors (48 females and 38 males) were recruited. Subjects (average age of 47 ± 14 years) were not following a prescribed diet or using drugs that might interfere with taste perception, and none of them had food allergies. In addition, individuals who smoked more than ten cigarettes per week were excluded.

Experimental protocol

A training session regarding taste perception, taste qualities and how to rate perception using the Labeled Magnitude Scale (LMS, Green et al. 1993) was performed. Volunteers were asked to refrain from eating and drinking for at least 3 h before the beginning of the session and to rinse their mouth with room temperature deionized water prior to the first and between each sample. Weight (in kg) and height (in m) were collected, in order to calculate the body mass index (BMI). For the perception test, 10 mL of a solution of stevioside 1.26 mM (Nastevia, Stevia Italia s.r.l., Italy) was used. The PROP-taster status was assessed using cotton swabs dipped in 50 mM 6-n-propylthiouracil solution (Sigma Aldrich S.r.l.) (Caremoli 2011), modifying the



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Table 1 Details of the studied SNPs in *TAS2R4*, *TAS2R14* and *TAS2R38* genes

Chr	Gene	dbSNP	cSNP	Allele	Type of mutation	Residue change
7	TAS2R4	rs2234001	G286-C	G	Missense	V [Val]
				C		L [Leu]
12	TAS2R14	rs11610105	G11088981-A	G	Silent	\
				A		
		rs3741843	G375-A	G	Cds synon	R [Arg]
				A		R [Arg]
		rs7138535	A114-T	A	Cds synon	G [Gly]
				T		G [Gly]
7	TAS2R38	rs713598	G229-C	G	Missense	A [Ala]
				C		P [Pro]
		rs10246939	A970-G	A	Missense	I [Ile]
				G		V [Val]
		rs1726866	T869-C	T	Missense	V [Val]
				C		A [Ala]

protocol for filter paper discs described in Zhao et al. (2003). Subjects were informed that they may receive stimuli eliciting more than one taste quality. They were asked to held the stevioside solution in the mouth for 5 s and the cotton PROP swab for 10 s. For both stimuli (i.e., stevioside and PROP), participants were allowed to choose among different tastes such as bitter, sweet or tasteless, as well as to rate their perceived intensity separately on multiple LMSs, one for each taste quality.

DNA collection, extraction, genotyping and quality control

DNA was collected using Oragene saliva collection kits and extracted according to the manufacturer's protocol (Genotek Inc., Kanata, Ontario, Canada). A list of genotyped SNPs in TAS2R14 (chromosome 12), TAS2R4 and TAS2R38 (chromosome 7), which were selected from the literature and by surveying lists of genetic variants contained in different databases, is shown in Table 1. Genotypes for these variants were determined using Sequenom MassARRAY technology (Sequenom, San Diego, CA, USA). Genotype calls were analyzed by using SEQUE-NOM Typer 4.0 software, and the individual spectrograms were checked in order to evaluate the presence of calling errors. None of them showed significant deviations from Hardy-Weinberg equilibrium (Online Resource 1). We also assessed their minor allele frequency to be above 5 % and their call rate to be above 95 %.

Statistical analyses

Allelic and genotypic association analyses were performed with PLINK v. 1.07 (Purcell et al. 2007) using Fisher's

exact method, and the significance threshold was adjusted according to the Bonferroni multiple testing correction (adjusted P = P value \times number of individual tests). Linkage disequilibrium (LD) patterns at the TAS2R38 gene were explored by means of the solid spine approach implemented in the Haploview package (Barrett et al. 2005). The PHASE software (Stephens et al. 2001) was then used to statistically infer TAS2R38 haplotypes within the identified high-LD blocks and to reconstruct haplotype pairs for each subject. For association analyses between individual SNPs and bitterness perception, analysis of covariance (ANCOVA) was performed using STATISTI-CA v. 6.0 (Stat-Soft Inc, Tulsa, OK), considering age, sex and BMI as covariates. Analysis of molecular variance (AMOVA) was carried out with Arlequin v. 3.5 (Excoffier et al. 2007). Exploration of worldwide variation patterns for the examined SNPs was performed using 1000 Genomes Project PHASE 1, HapMap and Human Genome Diversity Project datasets. Functional annotations and SIFT (http://sift.jcvi.org/) predictions (Kumar et al. 2009) for each SNP were retrieved from the Ensembl database (http://ensembl.org).

Results

Assessing taste phenotypes

Volunteers involved in the study were classified, after the perception test, in different tasting categories for both PROP and stevioside. Regarding the PROP status, the distribution of PROP sensitivity showed the classical bimodal curve (Online Resource 2), with 18 individuals (20.9 %) being classified as non-tasters and 68 (79.1 %) as



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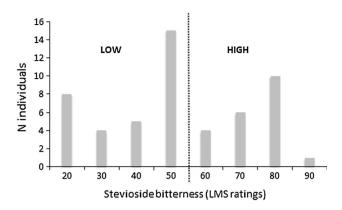


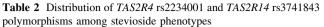
Fig. 2 Distribution of stevioside sensitivity in the Italian sample

tasters. In order to test whether the adopted deviation from the method described in Zhao et al. (2003) could affect PROP phenotype assessment, we calculated the overall genotype—phenotype concordance (90.7 %), thus inferring the correctness of our cotton swab PROP test. After stevioside tasting, 11 individuals (13 %) were able to perceive only its bitter taste. On the contrary, 22 individuals (26 %) perceived only its sweet taste, whereas the majority of samples constituted by 53 individuals (61 %) identified a sweet taste followed by a bitter after-taste.

The 53 bitter/sweet-tasters were subsequently distinguished into 32 "bitter-low" (LMS scores up to 50) and 21 "bitter-high" (LMS from 60 to 100) tasters (Fig. 2) according to LMS scale scores.

TAS2R4 SNP regulates the ability to perceive the bitter taste of stevioside

The TAS2R4 rs2234001 (C/G) turned out to be associated with stevioside bitter status. In fact, the "bitter" and "sweet" phenotype groups showed statistically significant differences for both genotypic (adjusted P = 0.002, Fisher's exact test) and allelic frequencies (adjusted P = 0.039, Fisher's exact test). In particular, the GG genotype and the G allele were more frequent in bitter-tasters (n = 11), whereas genotype CC and the C allele were more frequent in sweet-tasters (n = 22). More specifically, 68.18 % of bitter-tasters carried the G allele, whereas 76.19 % of sweet-tasters carried the C allele at this locus (Table 2). The same segregation was observed between the sweet and the "bitter-low" tasters (n = 32), with the genotype GG (adjusted P = 0.008, Fisher's exact test) and the G (adjusted P = 0.013, Fisher's exact test) allele being more frequent in the "bitter-low" group. Analyses conducted with the SIFT software showed that rs2234001 caused an amino acidic substitution at residue 96, resulting in a valine-leucine change, without altering the secondary structure of the protein. However, this SNP was found to be



rs2234001	Bitter	Bitter/sweet	Sweet	
Genotype	%	%	%	
CC	0	19.22	57.1	
CG	63.64	50	38.1	
GG	36.36	30.78	4.8	
Allele				
C	31.82	44.23	76.19	
G	68.18	55.77	23.81	
rs3741843	Bitte	er-low	Bitter-high	
Genotype	%		%	
AA	93.5	55	42.85	
GA	6.45		57.15	
GG	0		0	
Allele				
A	96.7	7	28.57	
G	3.23		71.43	

in strong LD ($r^2 \ge 0.9$) in European populations with two other non-synonymous SNPs (rs2227264 and rs2233998), both predicted by SIFT to alter the function of TAS2R5 and TAS2R4 proteins, respectively.

TAS2R4 SNP does not predict variations in stevioside bitterness perception

We also tested whether TAS2R4 rs2234001 (C/G) could predict stevioside bitterness or sweetness. Differently than TAS2R14 rs3741843, this SNP did not show variation among different levels of bitter and sweet perception (ANCOVA, P = 0.601 and P = 0.623, respectively).

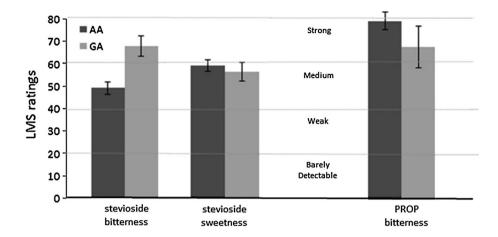
TAS2R14 SNP predicts variations in stevioside bitterness perception

We found evidence that TAS2R14 rs3741843 (A/G) has a significant impact on bitterness perception. Through comparing the "bitter-low" and "bitter-high" groups, the allelic frequency of this SNP was found to be significantly different (adjusted P=0.002; Fisher's exact test). In particular, the G allele was more frequent in the "bitter-high" group (n=21) compared to the "bitter-low" one (n=32). Genotypic analyses confirmed this statistically significant different distribution, with genotype AA being more frequent in the "bitter-low" group (adjusted $P \leq 0.001$, Fisher's exact test). The same tests were repeated by removing the LMS modal group (LMS = 50), which is suspected to introduce a confounding effect, and obtained results confirmed our previous estimation



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Fig. 3 Effect of the *TAS2R14* rs3741843 on bitterness and sweetness of stevioside and on bitterness of PROP



(P = 0.003 and P = 0.001, Fisher's exact test, respectively).

In addition, to test whether arbitrary classifications in taste phenotypes were plausible, an ANCOVA was performed on the subset of individuals who were able to perceive stevioside bitterness (n = 64), considering age, sex and BMI as covariates. As shown in Fig. 3, homozygote individuals with the AA genotype of TAS2R14 rs3741843 reported less bitterness from stevioside than heterozygote ones (P = 0.002). In contrast, there was no evidence that this allele predicts stevioside sweetness (ANCOVA, P = 0.621). We also found minimal evidence that stevioside bitterness is predictive of PROP bitterness (ANCOVA P = 0.081).

TAS2R38 SNPs predict variations in PROP bitterness but not in stevioside bitterness perception

As expected, PROP-tasters and non-tasters differed significantly for TAS2R38 alleles and haplotypes. In particular, TAS2R38 rs10246939 (T/C), rs1726866 (T/C) and rs713598 (G/C) were more frequent in PROP-tasters (n = 68) compared to PROP-non-tasters (n = 18) (adjusted P < 0.001, adjusted P < 0.001, adjusted P < 0.001, Fisher's exact tests). In the same way, perception of PROP bitterness varied with TAS2R38 haplotypes. The prolinealanine-valine (PAV) homozygotes (n = 15) reported significantly more bitterness than the heterozygotes (n = 42) (ANCOVA, P < 0.001) or the AVI (alanine, valine, and isoleucine) homozygotes (n = 24) (ANCOVA, P < 0.001). Moreover, bitterness reported by heterozygotes was similar to that reported by PAV homozygotes (ANCOVA, P = 0.345). Rare haplotypes were excluded from the analyses because they are known to have intermediate phenotypes that differ from both each other and the common haplotypes. In this study, we observed both AAI (3) and AAI/AVI (2) individuals. No associations were instead found between PROP haplotypes and stevioside bitterness (ANCOVA, P = 0.945) or sweetness (ANCOVA, P = 0.812).

Comparison between stevioside bitterness and sweetness perception

Great variability in both stevioside bitterness and sweetness perception was found. Figure 4 shows this distribution of variation, with stevioside bitterness and sweetness ranging from 20 to 80 on a LMS. We therefore tried to test whether a covariation between them existed. By plotting the bitterness and sweetness of stevioside simultaneously, we did not observe a covariation between them $(R^2 = 0.007, P = 0.776)$, with bitterness showing a decreasing trend toward the modal class and sweetness a decreasing trend followed by an increase after the modal class.

Population genetics of *TAS2R4* rs2234001 and *TAS2R14* rs3741843 SNPs

Alleles worldwide distribution

To better understand the differences observed for TAS2R4 and TAS2R14 SNPs, we examined their worldwide distribution. Figure 5 shows a map of allele frequencies of the two studied SNPs, based on 1000 Genomes Project data. Hardy–Weinberg test (Online Resource 3) and Heterozygosity test (Online Resource 4) were performed, confirming that all the examined populations are in Hardy–Weinberg equilibrium and no excess of heterozygosity was found (P > 0.05).

AMOVA

In order to identify the causes underlying the different distributions of rs3741843 and rs2234001, AMOVA based on the 1000 Genomes Project data was performed. As



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Fig. 4 Scatter plot showing sweetness and bitterness intensities of stevioside. Histograms shown along the axes illustrate the total amount of variation found across individuals

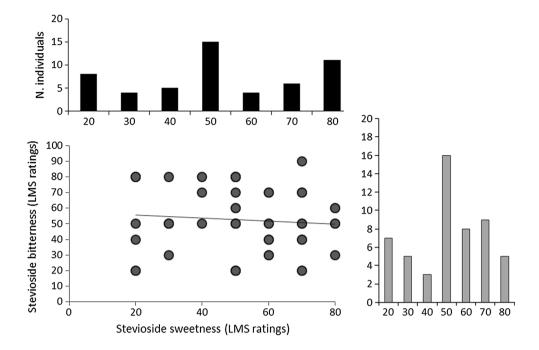




Fig. 5 Worldwide distribution of the studied TAS2R4 rs2234001 and TAS2R14 rs3741843 SNPs

shown in Table 3, 19.62 % of variation at rs3741843 was due to differences among population groups (P=0.002), 1.32 % was due to differences among populations within groups (P<0.001), and 79.06 % was accounted by differences among individuals within populations (P<0.001). rs2234001 showed a similar pattern, with 11.75 % of variation which was attributed to differences among groups (P<0.001), with the remaining 88.25 % accounting only for differences observable within populations (P=0.006).

Genetic differentiation analyses

Genome-wide pairwise Wright's F_{ST} was calculated as a measure of genetic differentiation on 1000 Genomes populations and continental groups, and a set of relevant percentiles was extracted (Online Resource 5). F_{ST} obtained for rs3741843 and rs2234001 fell within the top percentiles (0.95) of most of the continent-wide distributions and turned out to be outlier values also for some population-level comparison.



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Table 3 AMOVA performed on *TAS2R14* rs3741843 and *TAS2R4* rs2234001 SNPs

	rs3741843	rs2234001		
Among groups ^a	19.62 %	11.75 %		
Among population with groups	1.32 %	0		
Within populations ^b	79.06 %	88.25 %		
F_{ct}	0.19619 (<i>P</i> value 0.0019)	0.11754 (<i>P</i> value <0.001)		
F_{sc}	0.01644 (<i>P</i> value <0.001)	0		
F_{st}	0.20940 (<i>P</i> value <0.001)	0.1164 (P value 0.00587)		

^a Africans, Americans, Asian, Europeans

Admixture analysis

The admixed nature of the American 1000 Genomes populations allowed us to test whether the studied SNPs could be differentially selected according to the ancestral components of these populations. The American, African and European genomic components of the 1000 Genomes American populations were extracted from the literature (1000 Genomes Project Consortium et al. 2012; Skotte et al. 2013) and used to infer the expected frequencies of the two SNPs, given the observed admixture. The African and European frequencies of the two SNPs were obtained from the relevant populations, while American frequencies were extracted from the Mexican one (MXL), after correcting for admixture. Such frequencies were then combined, according to their admixture proportions, to generate the expected frequencies of each of the American populations. Table 4 reports the observed and expected frequencies hence calculated, along with the observed excess. The difference between observed and expected frequencies is remarkable for rs3741843, where Colombians (CLM) show a defect of 22 % and Puertoricans (PUR) show an excess of 61 % on the expected frequencies.

Table 4 Ancestral proportions in American 1000 Genomes populations and observed/ expected frequencies of *TAS2R4* rs2234001 and *TAS2R14* rs3741843 SNPs

Population	Ancestral proportions			rs2234001		rs3741843			
	%African	%NatAm	%European	Obs.	Exp.	%Excess	Obs.	Exp.	%Excess
ASW	0.79	0.02	0.19	0.39	0.37	4.65	0.38	0.40	-6.06
CLM	0.07	0.24	0.69	0.54	0.51	5.98	0.11	0.14	-21.75
MXL	0.02	0.60	0.38	0.60	0.60	0.00	0.11	0.11	0.00
PUR	0.13	0.13	0.74	0.55	0.48	14.08	0.26	0.16	61.41
AFR	-	-	_	0.34	-	-	0.48	-	-
EUR	-	-	_	0.47	-	-	0.12	-	-
NATAM (MXL) ^a	-	_	_	0.69	-	-	0.08	-	-

^a % of Native Americans in the Mexican population

Discussion

To our knowledge, this study represents the first attempt to explore genotype–phenotype association of stevioside bitter taste perception, as well its relationships with PROP phenotypes/genotypes. In fact, a previous study by Allen et al. (2013b) tested only for relationships between other steviol glycosides, RebA and RebD, and *TAS2R* SNPs. For this purpose, we have assayed three SNPs on *TAS2R14* (rs11610105, rs3741843 and rs7138535), one on *TAS2R4* (rs2234001) and three on *TAS2R38* (rs10246939, rs1726866 and rs713598) genes for individual associations with stevioside perception phenotypes and for their relationships with PROP genotypes/phenotypes on a sample of 86 Italian subjects.

We confirmed the associations between PROP variants and PROP-taster status at both allelic and haplotypic levels. Surprisingly, compared to other published data reporting a correlation between PROP bitterness and intensity of different tastants, including sucrose, citric acid, sodium chloride and quinine (Hayes et al. 2008), the sweetness of aspartame (Duffy et al. 2006) and Acesulfame K (Allen et al. 2013a), as well as sweet foods, such as marshmallows and iced cakes (Lanier et al. 2005), we did not find any correlation between PROP bitterness and either stevioside sweetness or stevioside bitterness.

Stevioside perception showed great variability among the candidate subjects, with the majority of the individuals perceiving both a sweet and a bitter after-taste and some individuals being not able to perceive its bitterness or sweetness. Moreover, also bitterness intensity strongly varied across subjects. Investigating polymorphisms on *TAS214* and *TAS2R4* genes, which have been recently identified as the loci encoding for receptors that mediate the bitter off-taste of steviol glycosides in vitro (Hellfritsch et al. 2012), our results suggest that two SNPs (rs2234001 and rs3741843) contribute to the perceived bitterness of stevioside in humans. After phenotype identification, we showed that *TAS2R4* rs2234001 varies among people who were able to perceive or not the bitter taste of stevioside. Specifically, our results demonstrate that the G allele of



b YRI, LWK, ASW, CLM, MXL, PUR, CHB, JPT, CHS, CEU, FIN, GBR, IBS, TSI

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this locus was associated with capability to perceive bitter after-taste of stevioside, while the C allele was more abundant in subjects who could not perceive it. At the genotype level, the recessive homozygote status GG was associated with the ability to perceive the bitter taste of stevioside, while the dominant CC homozygote status was associated with the inability.

In addition, when comparing strong and low bitter stevioside perceptions, one SNP located on the *TAS2R14* gene (rs3741843) showed significant frequency differences. In particular, its G allele was associated with a stronger bitter perception. In order to confirm this, we tried to verify whether perception of stevioside bitterness varied with the genotype and, as a matter of fact, the AA genotype turned out to be strongly correlated with lower values of bitter perception. Regarding PROP status, we did not find any relationships with neither stevioside perception nor the *TAS2R4/14* studied SNPs, although minimal evidence that the rs3741843 AA genotype is predictive of PROP bitterness was observed.

While variation at rs3741843 locus was not linked with any protein modification, we found that not only rs2234001 is a non-synonymous (albeit not deleterious) SNP, but that it is also in strong LD ($r^2 > 0.9$) with two other deleterious mutations in CEU (Northern Europeans from Utah). Both such polymorphisms (rs2227264 and rs2233998) were found to be associated with eating behavior (Dotson et al. 2010), while only rs2227264 has been proved to be related to the perceived bitterness of the espresso coffee (Hayes et al. 2011), hence representing putative functional candidates for the reported genotype–phenotype association.

Patterns of worldwide variation of two SNPs related to stevioside perception, together with comparison of their pairwise population $F_{\rm st}$ values with empirical genome wide distributions, highlighted great variability levels within human populations. In particular, rs2234001 showed a significant differentiation between African-Asian groups (P=0.05), but not between Asian-European ones (P=0.10), while rs3741843 was highly differentiated between African-Asian (P=0.025) and African-European (P=0.05) groups (Table S5). Furthermore, considering the 10 kbp haplotypes centered on the rs3741843 locus, the modal haplotype class broadly differed between African and European populations (Online Resource 6).

The strong differentiation between African and European groups for this latter SNP allowed us to detect a 61 % excess of the African variant in Puertoricans, after accounting for their history of genetic admixture. The enrichment of the African component in Puertoricans may suggest the presence of some natural or social selective pressure acting on such variant after its arrival in their genic pool some 200 ya (Kidd et al. 2012). Such selective pressure could perhaps be related with cultural/dietary

shifts caused by the introduction of African traditions at the time of their arrival in Puerto Rico. While these results might be affected by our choice of using the corrected Mexicans as proxy for the ancestral American components, we believe such choice to be conservative given that only two observed–expected pairs showed an excess higher than 20 %.

In conclusion, we found genotype–phenotype associations between stevioside perception and SNPs at the *TAS2R4* and *TAS2R14* genes, which are significantly differentiated worldwide. This finding and the strong LD of one of these variants with deleterious mutations suggest a role played by natural selection in shaping the current patterns of variation observed in different human groups.

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Ethical standards Davide Risso, Gabriella Morini, Luca Pagani, Andrea Quagliariello, Cristina Giuliani, Sara De Fanti, Marco Sazzini, Donata Luiselli and Sergio Tofanelli declare that they have no conflict of interest. All procedures followed were in accordance with the ethical standards of the Ethics Committee of the University of Bologna and with the Helsinki Declaration of 1975, as revised in 2000. Informed consent was obtained from all subjects for being included in the study.

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