

RESEARCH ARTICLE

Open Access

Bioactive Terpenoids and Flavonoids from *Daucus littoralis* Smith subsp. *hyrcanicus* Rech.f, an Endemic Species of Iran

Fatemeh Yousefbeyk^{1†}, Ahmad Reza Gohari^{1†}, Zeinabsadat Hashemighahderijani^{2†}, Sayed Nasser Ostad^{2†}, Mohamad Hossein Salehi Sourmaghi², Mohsen Amini³, Fereshteh Golfakhrabadi¹, Hossein Jamalifar⁴ and Gholamreza Amin^{1*†}

A correction to this article has been published: <http://www.darujps.com/content/22/1/33>

Abstract

Background: *Daucus littoralis* Smith subsp. *hyrcanicus* Rech.f. (Apiaceae) is an endemic species in northern parts of Iran where it is commonly named Caspian carrot. The fruits have been used as condiment.

Methods: In a series of *in vitro* assays, antioxidant (DPPH and FRAP assays), cytotoxic and antimicrobial activities of different extracts of roots and fruits were evaluated for the first time. The separation and purification of the compounds were carried out on the most potent extracts using various chromatographic methods and identified by spectroscopic data (¹H and ¹³C NMR).

Results: The results showed that among the extracts only fruit methanol extract (FME) has significant antioxidant activity (IC₅₀ = 145.93 μg.ml⁻¹ in DPPH assay and 358 ± 0.02 mmol FeII/g dry extract in FRAP assay). The radical scavenging activity of FME at 400 μg.ml⁻¹ was comparable with α-tocopherol (40 μg.ml⁻¹) and with BHA (100 μg.ml⁻¹) (*p* > 0.05). FME did not show any toxicity against cancerous and normal cell lines. Fruit ethyl acetate extract (FEE) had cytotoxic activity against breast carcinoma and hepatocellular carcinoma cells (IC₅₀ 168.4 and 185 μg.ml⁻¹, respectively), while it did not possess antioxidant activity in comparison with α-tocopherol and BHA as standard compounds. Ethyl acetate and methanol extract of fruits showed antimicrobial activity against *Staphylococcus aureus* (MIC: 3.75 mg.ml⁻¹) and *Candida albicans* (MIC: 15.6 and 7.8 mg.ml⁻¹, respectively). Four terpenoids were isolated from FEE including: β-sitosterol (1), stigmasterol (2), caryophyllene oxide (3), β-amyrin (4). Also, three flavonoids namely quercetin 3-O-β-glucoside (5), quercetin 3-O-β-galactoside (6) and luteolin (7) were isolated from FME.

Conclusion: This study showed that FEE and FME of *D. littoralis* Smith subsp. *hyrcanicus* Rech.f. had the highest biological activities which may be correlated with *in vitro* cytotoxic, antimicrobial and antioxidant activities of terpenoids and flavonoids components of the extracts.

Keywords: *Daucus littoralis* Smith subsp. *hyrcanicus* Rech.f, Antioxidant, Cytotoxic activity, Antimicrobial

Background

Daucus littoralis Smith subsp. *hyrcanicus* Rech.f. (Umbelliferae or Apiaceae) is an endemic species which is distributed in north of Iran (Mazandaran and Guilan provinces). It is an annual or perennial herb growing up to 3 to 10 cm high on the sandy dunes of Caspian Sea coasts where the

fruits have been used as condiment by the rural population [1]. The fruits of the related species, *D. carota*, have been used in Traditional Chinese Medicine (TCM) as a remedy for the treatment of ancylostomiasis, dropsy, chronic kidney diseases and bladder afflictions [2]. A wide range of pharmacologic properties such as antibacterial, antifungal, anthelmintic, hepatoprotective and cytotoxic activities have been reported on *D. carota* [2]. Phytochemical studies indicated the presence of sesquiterpenes, chromones, flavonoids, coumarins and anthocyanins from *D. carota* [2], sesquiterpene lactone [3] and phenylpropanoid

* Correspondence: amin@tums.ac.ir

†Equal contributors

¹Department of Pharmacognosy, Faculty of Pharmacy and Medicinal Plants Research Centre, Tehran University of Medical Sciences, Tehran 14155-6451, Iran
Full list of author information is available at the end of the article

triesters [4] from *D. glaber* (Forssk.) Thell. Recently we reported the composition and antimicrobial activity of the essential oil from leaves and stems, fruits, flowers and roots of *D. littoralis* Smith subsp. *hyrcanicus* Rech.f. [5]. No data on the phytochemistry and biological activity have been published for this species. In this study, we investigated the antioxidant, antimicrobial and cytotoxic activities of different extracts from roots and fruits of this plant. Also the isolation and structure elucidation of active compounds from most active extracts are reported.

Methods

General procedures

^1H and ^{13}C -NMR spectra was acquired using a Bruker Avance TM500 DRX (500 MHz for ^1H and 125 MHz for ^{13}C) spectrometer with tetramethylsilane as an internal standard, and chemical shifts are given in δ (ppm). Column chromatography was performed using silica gel (70–230, 230–400 mesh) (Merck, Germany) and Sephadex LH₂₀ (Fluka, Switzerland). Silica gel 60 F254 precoated plates (Merck, Germany) were used for TLC. The spots were detected by spraying anisaldehyde- H_2SO_4 (Sigma-Aldrich Chemie, Germany) reagent followed by heating. HPLC separations were carried out on a Knauer system (Smart line system, Germany) connected to a photodiode array detector. All the solvents, standards and reagents were obtained from Merck (Germany).

Plant material

The plant was collected from Bandar-e-Anzaly sea coast, province of Guilan, north of Iran, during the fruiting stage in June 2012. A voucher specimen of plant (6734-TEH) was deposited in Herbarium of Department of Pharmacognosy, Faculty of Pharmacy, Tehran University of Medical Sciences, Tehran, Iran.

Extraction and Isolation

The roots and fruits of plant (1 kg, each) were powdered and extracted successively with ethyl acetate, methanol and methanol–water (1:1), at room temperature. The fruit ethyl acetate extract (FEE) (88 g) was subjected to silica gel column chromatography (CC) with CHCl_3 : AcOEt (9: 1) as eluent to give ten fractions (A–J). The fraction H (5 g) was submitted to silica gel CC with hexane: AcOEt (8: 2) to obtain 20 fractions (Ha– Ht). The fractions Hm and Hr result compounds 1 and 2 (5 and 5.3 mg). The fraction Hc (300 mg) was subjected to CC with hexane: AcOEt (9: 1) to give three fractions (Hc₁–Hc₃). Compound 3 (5.9 mg) was obtained from the fraction Hc₃ (25 mg) by silica gel CC and hexane: CHCl_3 : AcOEt (18: 1: 1) as mobile phase.

The fraction Hd (500 mg) was chromatographed on Sephadex LH₂₀ with CHCl_3 : MeOH (3:7) to obtain nine fractions (Hd₁–Hd₉). Fraction Hd₉ (30 mg) was subjected

to normal phase semi-HPLC a Eurospher column (250 × 18 mm i.d.) and a PDA detector (λ : 210 nm). The initial eluted ratio was adjusted with 95: 5 (hexane: AcOEt) and delivered to the column for 20 min (flow-rate: 3 ml. min⁻¹). Then the eluted ratio was changed to 85:15 (hexane: AcOEt) until 50 min. The program was continued with the same ratio of solvents for next 20 min (70 min after starting point). The compound 4 (5.5 mg) was purified with this method.

The FME was dissolved in distilled water and after filtration; the aqueous solution was extracted with petroleum ether three times. The water soluble phase was evaporated to dryness and then extracted with n-BuOH for three times. The butanolic extract (8 g) was subjected to column chromatography on Sephadex LH₂₀ with MeOH to obtain 7 fractions (B₁–B₇). Fraction B₆ was submitted to reversed phase semi- HPLC including a Eurospher (column 250 × 20 mm i.d.) and a PDA detector (λ : 310 nm). Mobile phase including 40:60 (H₂O: MeOH) was delivered at flow rate 3 ml.min⁻¹ to give compounds 5 and 6 (5.5 and 5 mg respectively).

For purification of fraction B₇, a gradient reversed phase semi-HPLC was used with the same column, flow rate and detection condition. The eluted ratio was adjusted with 65:35 (H₂O: MeOH) as starting ratio and delivered to the column for 50 min and then was changed to 9:11 (H₂O: MeOH) until 70 min. Chromatography was continued with the same ratio of mobile phase for next 100 min (170 min after starting point) to give compound 7 (6.5 mg).

Antimicrobial activity of extracts

Antimicrobial activity of different extracts were tested against a Gram-positive (*Staphylococcus aureus* ATCC 6538), two Gram-negatives (*Escherichia coli* ATCC 8739 and *Pseudomonas aeruginosa* ATCC 9027) and a fungal strain (*Candida albicans* ATCC 1023). Minimum inhibitory concentration (MIC) of the extracts was determined by broth micro dilution method using 96 U-shaped wells plates [6]. A stock solution of 300 mg.ml⁻¹ from each extract was prepared in DMSO. Then two-fold serial dilution of the stock solution of each extract (100 μl) was prepared by using Mueller Hinton Broth (MHB) and Sabourad Dextrose Broth (SDB) (100 μl , each) in ten wells. The stock microbial suspension with twofold test inoculum was prepared in MHB and SDB from a 24-h old culture. Then aliquot of 100 μl of twofold test strain inoculum was added to each well to reach the final inoculum size of 5×10^5 cfu.ml⁻¹ [7]. The minimum bactericidal concentration (MBC) was determined by quantitative subculture of 100 μl from each clear well onto Mueller Hinton Agar (MHA) and Sabourad Dextrose Agar (SDA) plates. Plates were incubated at 37°C and 20–25°C for bacterial and fungal strains, for 48 h, respectively. The MBC

is defined as the lowest concentration of extracts that results in more than 99.9% killing of the bacteria being tested [7].

Antioxidant activity

DPPH radical-scavenging activity assay

The antioxidant activity of extracts were measured by the DPPH (2, 2'-diphenyl-1-picrylhydrazyl) free radical scavenging method based on an established protocol [8]. Sample solutions (1 ml) in methanol at different concentration were added to DPPH methanol solution (2 ml, 40 $\mu\text{g}\cdot\text{ml}^{-1}$). The mixtures were incubated at room temperature for 30 min and the absorbance was measured at 517 nm. Vitamin E and butyl hydroxyanisole (BHA) were used as positive controls. IC_{50} values (indicate the concentration of the test samples providing 50% radical scavenging) were calculated from graph-plotted scavenging percentage against extract concentration.

Ferric reducing antioxidant potential (FRAP scavenging) assay:

The FRAP assay was done according to the method described by Benzie and Strain [9,10]. Briefly, the FRAP reagent contained 5 ml of a (10 $\text{mmol}\cdot\text{l}^{-1}$) TPTZ (2, 4, 6-tripyridyl- s- triazine) solution in 40 $\text{mmol}\cdot\text{l}^{-1}$ HCl plus 5 ml of (20 $\text{mmol}\cdot\text{l}^{-1}$) FeCl_3 and 50 ml of (0.3 $\text{mmol}\cdot\text{l}^{-1}$) acetate buffer, pH 3.6 and was prepared freshly. Aliquots of extract (50 μl) were mixed with FRAP reagent (1.5 ml), incubated at 37°C, for 10 min, and then the absorbance was measured at 593 nm. For construction of calibration curve, five concentrations of $\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$ (125, 250, 500, 750, 1000 $\text{mmol}\cdot\text{l}^{-1}$) were used. The antioxidant activities were expressed as the concentration of antioxidants having a reducing ability equivalent for 1 $\text{mmol}\cdot\text{l}^{-1}$ FeSO_4 [11].

Measurement of total phenolic contents

Total phenolics were determined colorimetrically by the Folin-Ciocalteu method as described by Miliauskas, et al. [12]. The prepared extracts (1 ml) were mixed with 5 ml of Folin-Ciocalteu reagent (previously diluted tenfold with distilled water) and allowed to stand at room temperature for 10 min. A 4 ml sodium bicarbonate solution (75 $\text{g}\cdot\text{l}^{-1}$) was added to the mixture. After 30 min at room temperature, absorbance was measured at 765 nm using a UV spectrophotometer (Pharmacia Biotech). Total phenolics were quantified by calibration curve obtained from measuring the absorbance of a known concentration of gallic acid (GA) standard (20–200 $\text{mg}\cdot\text{l}^{-1}$). The concentrations are expressed as milligrams of gallic acid equivalents (GA) per g dry extract [11].

Cell cultures and cytotoxicity assay

Three cancerous cell lines HT29 (colon carcinoma), HepG2 (hepatocellular carcinoma), MCF7 (breast ductal carcinoma) and a normal cell line NIH-3T3 (Swiss mouse embryo fibroblast) were purchased from the Pasteur Institute, Tehran, Iran. The cells were maintained in RPMI 1640, supplemented with 10% fetal bovine serum, 0.28 $\text{units}\cdot\text{ml}^{-1}$ insulin, 100 $\mu\text{g}\cdot\text{ml}^{-1}$ streptomycin, 100 $\text{units}\cdot\text{ml}^{-1}$ penicillin, and 0.3 $\text{mg}\cdot\text{ml}^{-1}$ glutamine. The cells were grown at 37°C in a humidified atmosphere of 5% CO_2 . The cytotoxicity of different extracts was assayed using the MTT cytotoxicity assay. The cells (1×10^4) were plated in 100 μl of medium/well in 96-well plates (NUNC, Denmark). After 48 hours incubation at 37°C, in 5% CO_2 , and a humidified atmosphere, the different extracts were added to the cells of different concentrations (800, 400, 200, 100, 50, 25, 12.5 and 6, 25 $\mu\text{g}\cdot\text{ml}^{-1}$). Methotrexate (positive control) and extracts were incubated at 37°C, in 5% CO_2 , humidified atmosphere, for 48 hours. After 48 hours, 25 μl of 5 $\text{mg}\cdot\text{ml}^{-1}$ MTT (dissolved in PBS) was added per well. After three hours of incubation, the MTT solution was removed and the cells were washed twice with 100 μl of PBS. One hundred and fifty microliters of DMSO was added per well, to solubilize the formazan crystals. The optical densities of the wells were then measured at 570 nm (690 nm reference wavelength). By referring to the control (medium with DMSO), the cell survival was assessed [13]. The median growth inhibitory concentration (IC_{50} values) was obtained from the IC_{50} of dose response curve in the Sigma Plot 12 software. Each data is the mean value of three independent experiments and presented as mean \pm SD.

Results and discussion

In the present study extracts from roots and fruits of *D. littoralis* Smith subsp. *hyrcanicus* Rech.f. were investigated for bioactivity, the first time. Among the tested extracts, fruits methanol extract (FME) and fruit ethyl acetate extract (FEE) showed highest bioactive properties. As shown in Table 1, FME had the highest content of total phenol (99.1 ± 0.08 mg gallic acid equivalent/g dry extract) and the highest antioxidant activity in the DPPH assay ($\text{IC}_{50} = 145.93$ $\mu\text{g}\cdot\text{ml}^{-1}$) and in FRAP assay (358 ± 0.02 mmol FeII/g dry extract). Radical scavenging activity of FME at 400 $\mu\text{g}\cdot\text{ml}^{-1}$ was comparable with α -tocopherol (40 $\mu\text{g}\cdot\text{ml}^{-1}$) and BHA (100 $\mu\text{g}\cdot\text{ml}^{-1}$) ($p > 0.05$). Other extracts did not have any antioxidant activity in comparison with α -tocopherol and BHA. Other extracts did not have any significant antioxidant activities.

The results of antimicrobial assays are shown in Table 2. Among the extracts, only FME exhibited antimicrobial activity against all four microorganisms. FME showed better antimicrobial activity against *S. aureus*

Table 1 Antioxidant activity and total phenolic content of different extracts from fruits and roots of *D. littoralis* subsp. *hyrcanicus*

	DPPH ($\mu\text{g}\cdot\text{ml}^{-1}$)	FRAP (mmol FeII/g dry extract)	Total phenol contents (mg GAE/g dry extract)
FEE	789.74	44.6 ± 0.2	25.13 ± 0.06
FME	145.93	358 ± 0.02	99.1 ± 0.08
FMWE	172.3	306 ± 0.08	41.35 ± 0.04
REE	>1000	78 ± 0.01	36.06 ± 0.01
RME	467.2	258 ± 0.30	32.12 ± 0.03
RMWE	269.75	214 ± 0.20	27.79 ± 0.01
Vitamin E	14.12	313 ± 0.01	-
BHA	7.8	880 ± 0.06	-

Key to extracts employed: FEE: Fruits Ethyl acetate Extract; FME: Fruits Methanol Extract; FMWE: Fruits Methanol–water Extract; REE: Roots Ethyl acetate Extract; RME: Roots Methanol Extract; RMWE: Roots Methanol–water Extract.

(MIC: 3.75 and MBC: 7.5 $\text{mg}\cdot\text{ml}^{-1}$), whereas it showed weak activity towards Gram negative bacteria.

Antiproliferative activity was determined in HepG2, MCF7, HT-29, and NIH-3T3 cells, shown in Table 3. Only FEE and root ethyl acetate extract (REE) showed toxicity on cancerous cell lines. FEE showed higher cytotoxicity on HepG2 and MCF7 (IC_{50} 185.01 ± 2.1 and 168 ± 1.5 $\mu\text{g}\cdot\text{ml}^{-1}$, respectively) than REE.

The FEE and FME were used for isolation and purification of main components with different chromatography methods. From FEE four terpenoids including β -sitosterol (1), stigmasterol (2) caryophyllene oxide (3) and β -amyrin (4) were isolated. Three flavonoids including: quercetin 3-O- β -glucoside (5), quercetin 3-O- β -galactoside (6) and luteolin (7) were isolated from FME (Figure 1). Compounds 1–7 were identified by comparison of their

spectroscopic data ($^1\text{H-NMR}$, $^{13}\text{C-NMR}$) with those in the literature and authentic compounds from our laboratory [14-16]. Spectroscopic data of compounds 1-7 are provided in Additional file 1.

β -Amyrin is a pentacyclic triterpene with anti-inflammatory, antimicrobial, antifungal, antiviral and cytotoxic properties [17]. β -Amyrin isolated from MeOH extracts of *Byrsonima crassifolia* showed moderate antimicrobial activity against *S. aureus* and *C. albicans* (MIC: 0.5 and 1.02 $\text{mg}\cdot\text{ml}^{-1}$, respectively) [18]. The better activity of FEE against *S. aureus* and *C. albicans* seems to be due to the presence of β -amyrin. β -Sitosterol and stigmasterol are two of the most prevalent phytoestrogens in the plant kingdom [19]. *In vivo* investigation showed that oral consumption of stigmasterol inhibits the absorption of sterols and cholesterol from the intestinal tract and suppress the biosynthesis of cholesterol and bile acids in rats [19]. β -Sitosterol modulates the production of inflammatory cytokines, and reduces prostate enlargement [20]. It showed cytotoxic activity against colon carcinoma (COLO 320 DM), breast cancer and Bowes cell lines [21]. β -Caryophyllene oxide has shown cytotoxic activity against HepG2, AGS (human lung cancer cells), HeLa (human cervical adenocarcinoma cells), SNU-1 (human gastric cancer cell) and SNU-16 (human stomach cancer), with IC_{50} values of 3.95, 12.6, 13.55, 16.79, and 27.39 μM , respectively [22]. The cytotoxic activity of FEE against MCF7 and HepG2 seems to be due to the presence of β -sitosterol, β -amyrin and β -caryophyllene oxide.

In general, phenolic compounds possess antibacterial and antifungal properties [23]. Among them, flavonoids are well known for their antibacterial, antifungal, antiviral, antioxidant, anti-inflammatory activities [24]. The antioxidant activity of flavonoids is due to their capability as radical scavengers [25]. Many medicinal plants containing flavonoids have been reported for their antibacterial activity [23]. The high amount of total phenols

Table 2 Minimum inhibitory concentration (MIC) and minimum bactericidal concentration (MBC) of different extract of fruits and roots of *D. littoralis* subsp. *hyrcanicus*

Microorganisms	<i>Staphylococcus aureus</i>		<i>Escherichia coli</i>		<i>Pseudomonas aeruginosa</i>		<i>Candida albicans</i>	
	MIC	MBC	MIC	MBC	MIC	MBC	MIC	MBC
FEE	3.7	7.5	-	-	-	-	15.6	15.6
FME	3.7	7.5	>100	>100	62.5	>100	7.8	15.6
FMWE	-	-	-	-	-	-	-	-
REE	-	-	-	-	-	-	-	-
RME	>100	>100	-	-	-	-	-	-
RMWE	>100	>100	-	-	-	-	-	-

Note: MIC and MBC were determined by broth micro dilution method and expressed in $\text{mg}\cdot\text{ml}^{-1}$ (W/V); Key to extracts employed: FEE: Fruits Ethyl acetate Extract; FME: Fruits Methanol Extract; FMWE: Fruits Methanol–water Extract; REE: Roots Ethyl acetate Extract; RME: Roots Methanol Extract; RMWE: Roots Methanol–water Extract.

Table 3 Cytotoxic activity of different extracts of fruits and roots *D. littoralis* subsp. *hyrcanicus* using MTT assay

	HepG2	MCF7	HT-29	NIH-3T3
FEE	185.01 ± 2.1	168.41 ± 1.5	412.8 ± 1.3	149.48 ± 1.1
FME	- ^a	-	-	-
FMWE	-	-	-	-
REE	219.58 ± 1.1	279.68 ± 2.6	351.26 ± 3.2	155.34 ± 1.3
RME	935.34 ± 2.4	-	-	-
RMWE	-	-	-	-
Methotrexate	-	0.16 ± 0.09	0.23 ± 0.02	0.24 ± 0.01

Notes: Results are expressed as IC₅₀ values (µg.m.l⁻¹), ^a inactive (IC₅₀ > 1000); Key to cell Lines employed: HepG2(Hepatocellular carcinoma); MCF7 (breast carcinoma); HT29 (colon carcinoma); NIH-3T3 (Swiss embryo fibroblast). Key to extracts employed: FEE: Fruts Ethyl acetate Extract; FME: Fruits Methanol Extract; FMWE: Fruits Methanol–water Extract; REE: Roots Ethyl acetate Extract; RME: Roots Methanol Extract; RMWE: Roots Methanol–water Extract.

and in particular quercetin 3-O-β-glucoside, quercetin 3-O-β-galactoside and luteolin are responsible for better antibacterial activity of FME.

Conclusion

In this study, a screening of different extracts of *D. littoralis* Smith subsp. *hyrcanicus* Rech.f. was carried out for the first time. The presence of β-sitosterol, β-amyrin and β-caryophyllene oxide explained the cytotoxic activity of FEE in breast carcinoma and hepatocellular carcinoma cell lines. The high amount of phenolic compounds and flavonoids was responsible for the antioxidant and antimicrobial activity of FME. Based on these observations, FEE and FME can be good candidates for further *in vivo* biological studies and phytochemical investigations.

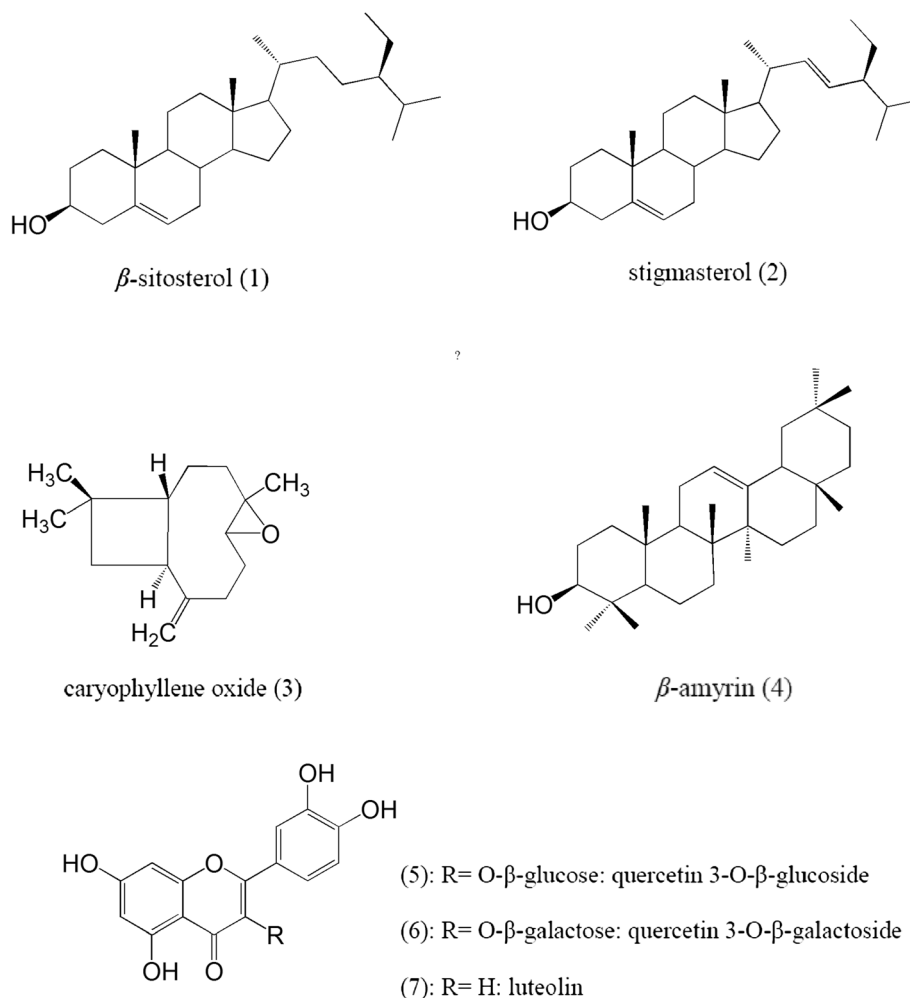


Figure 1 Structures of compounds 1-7 isolated form *D. littoralis* Smith subsp. *hyrcanicus* Rech.f including β-sitosterol (1), stigmasterol (2), caryophyllene oxide (3), β-amyrin (4), quercetin 3-O-β-glucoside (5), quercetin 3-O-β-galactoside (6) and luteolin (7).

Additional file

Additional file 1: Spectroscopic data of compounds 1-7 isolated from *D. littoralis* Smith subsp. *hyrcanicus* Rech.f.:

Competing interests

No conflict of interest has been declared.

Authors' contributions

FY performed plant preparation, extraction, isolation, identification of plant substances, evaluated antimicrobial activity of extracts, advised on antioxidant and total phenol content method and drafted the manuscript. AR-G advised on separation of plant substances and identification of compounds. ZH carried out antioxidant assays, total phenol content and cytotoxic activity of the extracts. SN-O advised on cytotoxic activity of extract by MTT assay. MH-SS conceived the study and edited the manuscript. MA advised on NMR techniques of isolated compounds. FG contributed in antimicrobial assay and edited the manuscript. HJ advised antimicrobial activity of extracts. GR-A did the botanical studies and identified scientific name of the Plant, conceived the study and edited the manuscript. All authors read and approved the final manuscript.

Acknowledgements

This investigation granted by research chancellor of Tehran University of Medical Sciences. The authors are grateful to Dr. Mohsen Amin (Faculty of Pharmacy, Tehran University of Medical Sciences) for proof reading and revising the manuscript. Also we acknowledge Mr Amir Yousefbeyk for his assistance in collecting plants.

Author details

¹Department of Pharmacognosy, Faculty of Pharmacy and Medicinal Plants Research Centre, Tehran University of Medical Sciences, Tehran 14155-6451, Iran.

²Department of Toxicology, Pharmacology and Nanotechnology Research Centre, Tehran University of Medical Sciences, Tehran 14155-6451, Iran.

³Department of Medicinal Chemistry, Faculty of Pharmacy, Tehran University of Medical Sciences, Tehran 14155-6451, Iran. ⁴Department of Drug and Food Control, Faculty of Pharmacy and Pharmaceutical Quality Assurance Research Centre, Tehran University of Medical Sciences, Tehran 14155-6451, Iran.

Received: 16 September 2013 Accepted: 18 November 2013

Published: 7 January 2014

References

1. Mozafarian V: *Flora of Iran. (Umbelliferae) volume 162*. Tehran: Research Institute of Forests and Rangelands; 2007.
2. Fu HW, Zhang L, Yi T, Chen RN, Wang X, Tian JK: **Two New guaiane-type sesquiterpene glycosides from the fruits of *Daucus carota* L.** *Chem Pharm Bull* 2010, **58**:125–128.
3. Sallam AA, Hitotsuyanagi Y, Mansour ESS, Ahmed AF, Gedara S, Fukaya H, Takeya K: **Sesquiterpene Lactones from *Daucus glaber*.** *Helv Chim Acta* 2010, **93**:48–57.
4. Sallam AA, Hitotsuyanagi Y, Mansour ESS, Ahmed AF, Gedara S, Fukaya H, Takeya K: **Phenylpropanoid triesters from *Daucus glaber*.** *Phytochem Lett* 2009, **2**:188–191.
5. Yousefbeyk F, Gohari AR, Salehi Sourmaghi MH, Amini M, Jamalifar H, Golfakhrabadi F, Ramezani N, Amin GH: **Chemical composition and antimicrobial activity of essential oils from different parts of *Daucus littoralis* Smith subsp. *hyrcanicus*.** *Rech J Essent Oil Bear.* in press.
6. NCCLS: *Methods for Dilution Antimicrobial Susceptibility Tests for Bacteria that Grow Aerobically. Approved Standard M7-A7*. Pennsylvania: Wayne; 2006.
7. Vazirian M, Taheri Kashani S, Shams Ardekani M, Khanavi M: **Antimicrobial activity of lemongrass (*Cymbopogon citratus* (DC) Stapf.) essential oil against food-borne pathogens added to cream filled cakes and pastries.** *J Essent Oil Res* 2010, **24**:579–582.
8. Yassa N, Razavi-Beni H, Hadjiakhoondi A: **Free radical scavenging and lipid peroxidation activity of the Shahani black grape.** *Pak J Biol Sci* 2008, **11**:1–4.
9. Benzie IF, Strain JJ: **The ferric reducing ability of plasma (FRAP) as a measure of "antioxidant power": the FRAP assay.** *Anal Biochem* 1996, **239**:70–76.

10. Moradi-Afrapoli F, Asghari B, Saeidnia S, Ajani Y, Mirjani M, Malmir M, Dolatabadi Bazaz R, Hadjiakhoondi A, Salehi P, Hamburger M, Yassa N: **vitro α -glucosidase inhibitory activity of phenolic constituents from aerial parts of *Polygonum hyrcanicum*.** *Daru* 2012, **20**:37–41.
11. Sadati N, Khanavi M, Mahrokh A, Nabavi S, Sohrabipour J, Hadjiakhoondi A: **Comparison of Antioxidant Activity and Total Phenolic Contents of some Persian Gulf Marine Algae.** *J Med Plants* 2011, **10**:73–79.
12. Millauskas G, Venskutonis P, Beek T: **Screening of radical scavenging activity of some medicinal and aromatic plant extract.** *Food Chem* 2004, **85**:231–237.
13. Momtaz S, Lall N, Hussein A, Ostad S, Abdollahi M: **Investigation of the possible biological activities of a poisonous South African plant; *Hybanthus globosa* (Euphorbiaceae).** *Phacog Mag* 2010, **6**:34–41.
14. Goad LJ, Akihisa T: *Analysis of sterols*. London: Blackie Academic & Professional; 1997.
15. Gohari AR, Hadjiakhoondi A, Sadat-Ebrahimi S, Saeidnia S, Shafiee A: **Cytotoxic triterpenoids from *Satureja macrantha* C.A. Mey.** *Daru* 2005, **13**:177–181.
16. Agrawal P: *Carbon-13 NMR of Flavonoids*. New York: Elsevier; 1989.
17. Hernández Vázquez L, Palazon J, Navarro-Ocana A: *The pentacyclic triterpenes α , β amyryns: A review of sources and biological activities*. *Phytochemicals - A Global perspective of their role in nutrition and health*. InTech. Croatia: In Tech Europ; 2012.
18. Rivero-Cruz JF, Sánchez-Nieto S, Benítez G, Casimiro X, Ibarra-Alvarado C, Rojas-Molina A, Rivero-Cruz B: **Antibacterial compounds isolated from *Byrsonima crassifolia*.** *Rev Latinoamer Quím* 2009, **37**:155–163.
19. Saeidnia S, Perme P, Gohari A, Mashinchian-Moradi A: **Gracilariopsis persica from Persian Gulf Contains Bioactive Sterols.** *Iranian J Pharm Res* 2012, **11**:845–849.
20. Paniagua-Perez R, Madrigal-Bujaidar E, Reyes-Cadena S, Molina-Jaso D, Perez Gallaga J, Silva-Miranda A, Velazcoet O, Hernandezal N, Chamorro G: **Genotoxic and cytotoxic studies of beta-sitosterol and pteropodine in mouse.** *J Biomed Biotechnol* 2005, **3**:242–2473.
21. Baskar AA, Ignacimuthu S, Paulraj GM, Numair KSA: **Potential of β -Sitosterol in experimental colon cancer model - an In vitro and In vivo study.** *BMC Complement Altern Med* 2010, **10**:24–34.
22. Jun NJ, Mosaddik A, Moon JY, Jang KC, Lee DS, Ahn KS, Cho SK: **Cytotoxic activity of β -Caryophyllene oxide isolated from Jeju Guava (*Psidium cattleianum* Sabine) Leaf Rec.** *Nat Prod* 2011, **5**:242–246.
23. Martini N, Katerere D, Eloff J: **Biological activity of five antibacterial flavonoids from *Combretum erythrophyllum* (Combretaceae).** *J Ethnopharmacol* 2004, **93**:207–212.
24. Cushnie TT, Lamb AJ: **Antimicrobial activity of flavonoids.** *Int J Antimicrob Ag* 2005, **26**:343–356.
25. Saeidnia S, Abdollahi M: **Who plays dual role in cancerous and normal cells? Natural antioxidants or free radicals or cell environment.** *Int J Pharmacol* 2012, **8**:711–712.

doi:10.1186/2008-2231-22-12

Cite this article as: Yousefbeyk et al.: Bioactive Terpenoids and Flavonoids from *Daucus littoralis* Smith subsp. *hyrcanicus* Rech.f, an Endemic Species of Iran. *DARU Journal of Pharmaceutical Sciences* 2014 **22**:12.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

