

Research Article

Synergistic Effect and Antiquorum Sensing Activity of *Nymphaea tetragona* (Water Lily) Extract

Md. Akil Hossain,¹ Ji-Yong Park,^{1,2} Jin-Yoon Kim,¹ Joo-Won Suh,³ and Seung-Chun Park¹

¹ Laboratory of Clinical Pharmacokinetics and Pharmacodynamics, Department of Pharmacology, College of Veterinary Medicine, Kyungpook National University, Daegu 702-701, Republic of Korea

² Institute of Clean Bio, Daejeon 301-212, Republic of Korea

³ Division of Bioscience and Bioinformatics, Myongji University, Science Campus, Gyeonggi 449-728, Republic of Korea

Correspondence should be addressed to Seung-Chun Park; parksch@knu.ac.kr

Received 26 February 2014; Accepted 1 April 2014; Published 8 May 2014

Academic Editor: Kota V. Ramana

Copyright © 2014 Md. Akil Hossain et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Salmonellosis is a common and widely distributed food borne disease where *Salmonella typhimurium* is one of the most important etiologic agents. The purpose of this study was to investigate the antimicrobial activity of *Nymphaea tetragona* alone and in combination with antibiotics against *S. typhimurium*. It also aimed to assess the plant for quorum sensing inhibition (QSI) activity and to identify the bioactive compounds. The antibacterial activities of the extract were assessed using broth microdilution method. Disk agar diffusion method was employed to determine the QSI and bioactive compounds were identified by GC-MS analysis. Ethyl acetate fraction of *N. tetragona* extract (EFNTE) demonstrated good antimicrobial activity (MIC 781 µg/mL) against 4 strains out of 5. FIC index ranged from 0.375 to 1.031 between EFNTE/tylosin and 0.515 to 1.250 between EFNTE/streptomycin against *S. typhimurium*. Among all extracts, EFNTE and butanol fraction more significantly inhibited pigment production of *C. violaceum*. Polyphenols were identified as major compound of EFNTE and butanol fraction. These results indicate that combination among *N. tetragona* extract and antibiotics could be useful to combat drug-resistance *Salmonella* infections and polyphenols are promising new components from *N. tetragona* that warrant further investigation as a candidate anti-*Salmonella* agent and quorum sensing inhibitor.

1. Introduction

Salmonella species are the leading cause of bacterial gastroenteritis in humans and animals all over the world [1, 2]. Food animals and water are the most important reservoirs of the bacteria [2] where the outbreaks of *Salmonella* infections have increasingly been associated with processed foods [3, 4]. There are 1300 million cases of gastroenteritis, 16 million cases of typhoid fever, and 3 million cases of deaths worldwide each year due to *Salmonella* infections [5]. *Salmonella typhimurium* is one of the most common serovars associated with clinically reported salmonellosis in humans in most parts of the world, accounting for at least 15% of infections [2, 6].

S. typhimurium infects a wide range of animal hosts, including poultry, cattle, and pigs, and is termed ubiquitous which usually causes a self-limited gastroenteritis in

humans [7]. The use of antibiotics is a major strategy and they are commonly used therapeutically and prophylactically to treat *S. typhimurium* infections in human and animal. However, increased antimicrobial resistance is exacerbating impact on public health worldwide, which leads to increased morbidity, mortality, and treatment costs [8, 9]. Scientific studies showed that tylosin has low or no inhibitory effects on experimentally inoculated *S. typhimurium* in pigs [10, 11]. It was also reported that *S. typhimurium* illustrated resistance to ampicillin, chloramphenicol, streptomycin, sulphonamides, and tetracycline in 1980s, in the UK and later distributed extensively through Europe and North America. Additional resistance had attained to trimethoprim-sulfamethoxazole, ciprofloxacin, and extended-spectrum cephalosporins within the 1990s [12]. Development of alternative antibacterial therapies is necessary to overcome this outbreak. Approximately 80% of the world's inhabitants rely on traditional medicine for

their primary health care and plants also play an important role in the health care system [13]. The possible therapeutic use of *Nymphaeaceae* may be a good alternative of traditional antibacterial.

The Nymphaeaceae also called water lilies have a broad range of flower colors and are living on the banks of lakes and rivers, distributed in tropical areas around the world [14–16]. A number of species of *Nymphaea* in Nepal, India, and China are thought to act as functional drug plants [17]. Many bioactive and pharmacologically important compounds have been obtained from *Nymphaea* species and used in medicine and pharmacy [18]. Flower extract of *N. nouchali* which possess compounds with high antibacterial and cytotoxic properties [19, 20] and ethyl acetate extract of *N. nouchali* leaf extract have antibacterial activity against a wide range of strains [21]. *Nymphaea lotus* extract was reported to have bioactive compounds such as tannins, flavonoids, alkaloids, anthraquinones, saponins, cardiac glycosides, and phenolics where methicillin and vancomycin resistant *S. aureus*, *S. pyogenes*, and *E. coli* were highly susceptible to *N. lotus* [13, 22, 23]. The pygmy water lily, *Nymphaea tetragona* (Ait.) Georgi (Nymphaeaceae), is one of the widely distributed plants, ranging globally from Asia-temperate, Asia-tropical, Europe, and northern America [24]. It has ethnomedical uses as the rhizome is used to cure acute diarrhea and dysentery by tribal herbal practitioners in Indian region [24].

Although there are many reports in the ethnomedical values of *Nymphaea*, information on their antibacterial efficacy is scarce and with low scientific caliber for further commercial use. Hence, it is important to determine the antibacterial activity very clearly and to identify the antibacterial active compounds of this plant. Furthermore, the potentials of water lily in combating antimicrobial resistance alone and in combination with antibiotics and the inhibition of quorum sensing controlled virulent factors of microbial pathogens were not explored previously. Thus, the current study was designated to evaluate the antibacterial activity of *Nymphaea tetragona* extract alone and in combination with commercial antibiotics against *Salmonella typhimurium*. Quorum sensing inhibition activity of the extract against biomonitor strain *Chromobacterium violaceum* was also aimed at assessing in this study. Finally, a GC-MS analysis was performed to identify and quantify the major compounds of the *N. tetragona*.

2. Materials and Methods

2.1. Bacterial Strains and Culture Medium. *Salmonella typhimurium* QC strain KTCC2515 and clinical isolates ST171, ST482, ST688, and ST21A were used for experiments in this study (Table 1) which were collected from different farms in Republic of Korea. Bacterial strains were suspended in Mueller Hinton broth (MHB, Difco, USA) and then incubated at 37°C with 200 rpm for 20 h. Mueller Hinton agar (MHA, Difco) was used for the agar diffusion method.

2.2. Plant Extraction and Fractionation. *N. tetragona* powder of body and root mixture was purchased from Chamsamgol

Lotus Farm (Chungju, Republic of Korea). 100 g of the powdered material was boiled with 1000 mL of 50% methanol in a 2000 mL three-neck round bottom boiling flask (Schott Duran, NY, USA) at 100°C setting temperature on nonasbestos surface for 3 h when the % Brix and absorbance of the extract became the highest. The supernatant of *N. tetragona* 50% methanol extract (NTME) was collected by filtration (70 mm, Advantec; Toyo Roshi Kaisha Ltd., Tokyo, Japan) and solid particles retained on the filter were discarded. The solvent was then removed under reduced pressure in Buchi Rotavapor R-114 (BUCHI Labortechnik AG in Flawil, Switzerland) at 10 rpm and Eyela CCA-1111 (Tokyo Rikakikai Co. Ltd., made in China) and solidified by freeze-drying prior to use. The yield of extract was 10.71%.

10 g of lyophilized *N. tetragona* extract was suspended in 50 mL of water and fractionated with equivalent amount of dichloromethane, ethyl acetate, and butanol correspondingly by separating funnel. The solvent fractions were solidified and the distribution of extract was 0.43% in dichloromethane, 4.01% in ethyl acetate, and 46.74% in butanol. 47.36% was obtained as residue in water and 1.46% was process loss.

2.3. Antibacterial Resistance Testing. The disk-agar diffusion method validated by the Clinical and Laboratory Standards Institute [25] was used to verify the resistance pattern of four *S. typhimurium* clinical isolates against different commercial antibiotics. *S. typhimurium* KTCC 2515 was used as quality control strain in this experiment. Antibiotic sensitivity was considered according to the zone diameter interpretative standards of CLSI [26].

2.4. Determination of MIC and MBC. MIC was determined by the standard broth microdilution method according to the CLSI guidelines [27] in MHB using $\sim 5 \times 10^5$ CFU/mL of inoculums concentration. The MHB was supplemented with serial dilutions ranging from 24.4–25000, 4.9–2500, 12.2–6250, and 4.9–25000 $\mu\text{g/mL}$, respectively, for NTME, DFNTE, EFNTE, and BFNTE in 100 μL volumes in 96-well plates. Bacterial suspensions were adjusted to 0.1 OD at 600 nm, diluted 1/100, and dispensed in 100 μL aliquots to all the wells, including drug-free controls. Initial CFU/mL of the bacterial suspensions was determined by plating 10-fold dilutions on MHA plates. After overnight incubation at 37°C, inoculums from each well were diluted 10-fold and plated on MHA plates to determine the CFU/mL of each well. MIC was determined by comparing the final CFU with initial CFU. The lowest concentration of the extract inhibiting the increase of CFU was considered as MIC. 100 μL drug dilutions from wells of 96-well plates were cultured on MHA plates to determine the existing bacterial number. MBC was considered as the lowest concentration which can eradicate 99.9% bacteria [28].

2.5. Killing Rate of *Salmonella typhimurium*. Killing rate was evaluated as previously described by Tia Dubuisson et al., [28] with some modification. 0.25 \times MIC, 0.5 \times MIC, 1 \times MIC, and 2 \times MIC of EFNTE were prepared in 10 mL of MHB. Four-hour old *S. typhimurium* (KTCC 2515) cultures were adjusted to 0.1 OD in 600 nm, diluted to make suitable

TABLE 1: List of *Salmonella* strains used in this study along with their sources and sensitivities against commercial antibiotics.

Antimicrobials	Concentration ($\mu\text{g}/\text{disk}$)	KCTC2515	Isolates			
			ST171 (chicken)	ST482 (pig)	ST688 (cattle)	ST21A (pig)
Chloramphenicol	30	S	S	S	R	S
Ciprofloxacin	5	S	S	S	S	S
Erythromycin	15	R	R	R	R	R
Gentamycin	10	S	R	S	R	R
Kanamycin	30	S	S	R	R	I
Norfloxacin	10	S	S	S	S	S
Streptomycin	10	I	R	R	R	R
Tetracycline	30	S	R	R	R	S

Antibiotic sensitivity is considered according to the zone diameter interpretative standards of CLSI [26]. R: resistant; I: intermediate, and S: sensitive.

concentration which will be 10^6 CFU/mL after inoculating to each tube. Incubation of tubes was done at 200 rpm on a shaker incubator at 37°C . In 0, 1, 2, 3, 4, 6, 8, 12, and 24 hours, the CFU of the cultures was determined by culturing $20 \mu\text{L}$ aliquots of 10-fold dilutions on MHA plates. Plates were incubated at 37°C overnight before counting CFU. The mean \log_{10} CFU/mL for each extract was plotted against different times.

2.6. *Nymphaea tetragona* Extract and Antimicrobial Combination Activities In Vitro. Combination effects of EFNTE with ampicillin, marbofloxacin, norfloxacin, Streptomycin, trimethoprim, and tylosin (Sigma-Aldrich Co. St. Louis, MO, USA) against *S. typhimurium* were screened out, since these antimicrobials have reported for becoming resistant to *S. typhimurium* [10–12]. Previously described double disk agar diffusion method [29] was used with slight modification. A zone diameter of individual antimicrobial was determined by CLSI guideline [25].

Disks were placed separately with a distance equal to the sum of the zone radii for each disk tested separately. The interface of inhibition zones was observed after incubation. Generally, synergism shows an improved zone, indifference shows no change, and antagonism shows a reduced zone compared to the zones of individual test [29]. Further test of synergism was done only for tylosin and streptomycin with the extract fraction, as those 2 antibiotics showed synergism in disk diffusion test.

Combination interactions of the extract fraction with tylosin and streptomycin were determined in 96-well plates by previously described chequerboard microdilution method with slight modifications [30]. Antibiotic was vertically and the extract was horizontally diluted to get a matrix of different combinations of 2 antibacterials. Similar dilutions of individual drugs and the drug-free medium control were included in each test plate. Plates were incubated at 35°C for 16 to 20 hours after the addition of $\sim 5 \times 10^5$ CFU/mL of inoculums. From the MIC of the drugs alone and in combination, we calculated the fractional inhibitory concentration (FIC) and the FIC index (FICI). FIC is the MIC in combination divided by the MIC of the individual drug and FICI is the sum of

the FICs of individual drugs. An FICI of ≤ 0.5 is considered synergistic, an FICI of 4.0 is considered antagonistic, and an FICI of 0.5–4 is considered to indicate no interaction [31].

2.7. Quorum Sensing Inhibition. Quorum sensing inhibition of NTME and its solvent fractions were verified in accordance with the method described by Alvarez et al. [32]. *C. violaceum* CV12472 was employed to find out the pigment inhibition of extracts for attaining a qualitative screening. $100 \mu\text{L}$ of fresh culture diluted as 2.5×10^6 CFU/mL was poured on media for the preparation of LB agar plates. $60 \mu\text{L}$ of each extract with specific concentrations was applied to saturate the sterile paper disks (8 mm). Normal saline ($60 \mu\text{L}$) was used as negative control and purified furanone ($100 \mu\text{g}$) was applied as positive control whereas tetracycline ($10 \mu\text{g}$) was employed to compare antibacterial and anti-quorum sensing activity. Inhibition of pigment production around the disc was checked after 18–24 h incubation at 30°C . The sensitivity to different agents was classified by the diameter of the inhibition zones as follows: “not sensitive” for diameter less than 8 mm, “sensitive” for diameter between 9 and 14 mm, “very sensitive” for diameter between 15 and 19 mm, and “extremely sensitive” for diameter larger than 20 mm [33, 34].

2.8. Gas Liquid Chromatography Coupled Mass Spectrophotometric (GC-MS) Analysis. GC-MS analysis of DFNTE, EFNTE, and BFNTE was performed by “Center for Scientific Instruments” of Kyungpook National University and carried out using a HP 6890 Plus GC gas chromatograph with a (MSD)—HP 5973 MSD mass selective detector (Hewlett-Packard). Samples were diluted 1:1000 (v:v) with HPLC grade dichloromethane. Aliquots of the sample ($1 \mu\text{L}$) were injected into an HP-5 column. The GC oven temperature was set at 50°C for 4 min, increased to 280°C at a rate of $4^\circ\text{C}/\text{min}$, and held at the final temperature for 2 min. Velocity of the He carrier gas (99.99%) was $0.7 \text{ mL}/\text{min}$. Quantitative analysis was performed using the area normalization method.

2.9. Statistical Analysis. The mean values and the standard deviation were calculated from the data obtained from triplicate trials. Analysis of variance (ANOVA) was used to verify

TABLE 2: Antimicrobial activity of NTME and its solvent fractions against five strains of *Salmonella typhimurium*.

	KCTC 2515	ST171	ST482	ST688	ST21A
MIC ($\mu\text{g/mL}$)					
NOR ^a	0.063	0.063	0.125	0.063	0.125
NTME	6250	6250	6250	6250	6250
DFNTE	>2500	>2500	>2500	>2500	>2500
EFNTE	781	781	>781	781	781
BFNTE	6250	6250	6250	6250	6250
MBC ($\mu\text{g/mL}$)					
NOR ^a	1	1	1	1	1
NTME	12500	12500	25000	12500	12500
DFNTE	>2500	>2500	>2500	>2500	>2500
EFNTE	1562	1562	1562	1562	1562
BFNTE	25000	25000	25000	25000	25000

NOR: norfloxacin, NTME: *N. tetragona* 50% methanol extract, EFNTE: ethyl acetate fraction of *N. tetragona* 50% methanol extract, DFNTE: dichloromethane fraction of *N. tetragona* 50% methanol extract, BFNTE: butanol fraction of *N. tetragona* 50% methanol extract. ^aPositive control.

differences and *F*-test was applied for the determination of statistical significance between groups.

3. Results

3.1. MIC and MBC of *Nymphaea tetragona* 50% Methanol Extract. The antimicrobial activity of NTME and its solvent fractions were confirmed by determining the minimum inhibitory concentration (MIC) and minimum bactericidal concentration (MBC) against different strains of *S. typhimurium*. The results indicated that the EFNTE possessed the strongest antibacterial activity among all fractions. MBC of EFNTE against all tested *S. typhimurium* strains was $\leq 1562 \mu\text{g/mL}$, whereas the MIC values were within $781 \mu\text{g/mL}$ for all strains except one clinical isolate (Table 2).

3.2. Killing Rate of *Salmonella typhimurium*. Time-kill curves of *S. typhimurium* after treatment with EFNTE are demonstrated in Figure 1. The EFNTE of both the $1 \times \text{MIC}$ and $2 \times \text{MIC}$ concentrations showed complete inhibition up to 8 hours and started their log phase from this time point. At 24 hours, the growth level was 2-fold lower in $1 \times \text{MIC}$ and 3-fold lower in $2 \times \text{MIC}$ of EFNTE than the growth control. At $0.25 \times \text{MIC}$ and $0.5 \times \text{MIC}$, bacteria reached log phase after 1 hour and stationary phase after 8 hours whereas the control started stationary phase after the 4th hour. None of the tested concentrations showed complete killing effect within 24 h.

3.3. In Vitro Synergy with Commercial Antimicrobials. To investigate whether there is any synergy between EFNTE and commercial antibiotics, we examined six antibiotics with EFNTE by disc diffusion method. EFNTE exhibited synergism only with tylosin and streptomycin and has additive/indifferent effects with other antibiotics. There was no antagonistic effect observed in EFNTE with those antibiotics (Figure 2).

To reconfirm the synergistic activity of EFNTE with tylosin and streptomycin, we further performed checkerboard microdilution assay. The results of the checkerboard

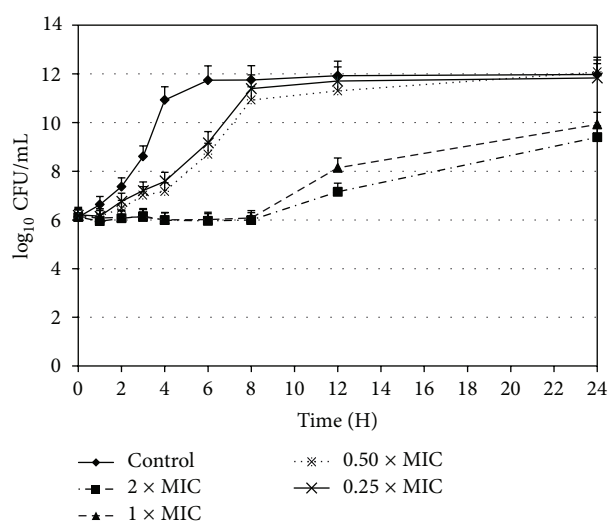


FIGURE 1: Time-kill curves of *S. typhimurium* KCTC 2515 after treatment with EFNTE at $0.25 \times \text{MIC}$ (x), $0.50 \times \text{MIC}$ (x), $1 \times \text{MIC}$ (▲), and $2 \times \text{MIC}$ (■). Ca-MHB was used as a control (◆). The results are presented as mean \pm standard deviations ($n = 3$), and the coefficients of variation for all concentrations are $\leq 5\%$.

analysis are summarized in Tables 3 and 4 and Figure 3. An increased sensitivity against tylosin was observed in combination with EFNTE. The corresponding FICs were ≤ 0.5 in tested strain, demonstrating a synergistic effect. However, EFNTE in combination with streptomycin showed additive and indifferent instead of synergistic interaction.

3.4. Quorum Sensing Inhibition of NTME. The inhibition diameter of *C. violaceum* pigment in presence of NTME and solvent fractions of NTME is presented in Table 5. *C. violaceum* showed sensitivity to all fractions including the crude extract. Although the crude extract and all fractions of extract significantly prevented the pigment, ethyl acetate and butanol fractions showed highest inhibition among all.

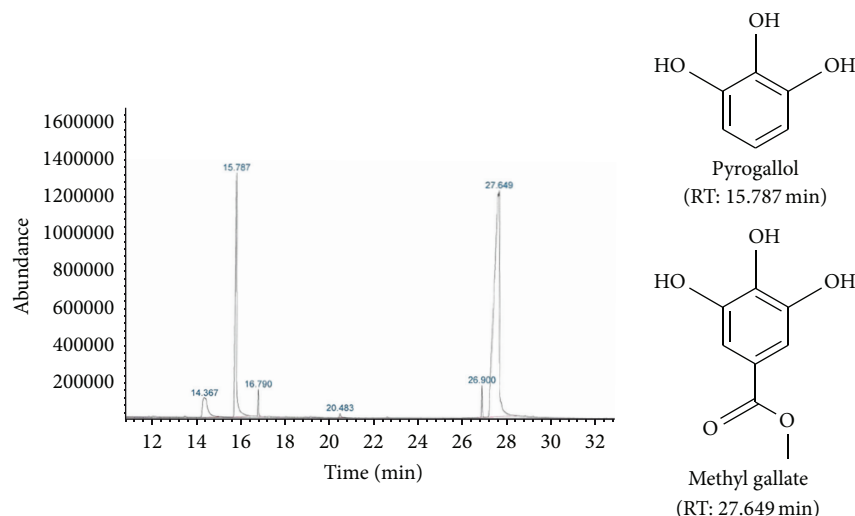


FIGURE 4: Gas chromatography coupled with mass spectroscopy (GC-MS) chromatogram of EFNTE and chemical structure of major compounds.

TABLE 3: Fractional inhibitory concentration (FIC) and FIC index (FICI) of combination between tylosin and EFNTE.

Tylosin		EFNTE		FICI	Interpretation
Con ^c (μg/mL)	FIC of drug A	Con ^c (μg/mL)	FIC of drug B	(FIC of A + FIC of B)	
1024	MIC of A	0	MIC of A	MIC of A	MIC of A
1024	1	6	0.0077	1.008	IND
1024	1	12	0.0154	1.015	IND
1024	1	24	0.0307	1.031	IND
512	0.5	49	0.063	0.563	ADI
256	0.25	98	0.125	0.375	SYN
256	0.25	195	0.25	0.499	SYN
128	0.125	391	0.5	0.626	ADI
64	0.0625	391	0.5	0.563	ADI
0	MIC of B	781	MIC of B	MIC of B	MIC of B

EFNTE: ethyl acetate fraction of *N. tetragona* 50% methanol extract; Con^c: concentration; drug A: tylosin; drug B: EFNTE; SYN: synergistic effect; ADI: additive effect; IND: indifferent effect; MIC: minimum inhibitory concentration. The FICI was interpreted as follows: synergistic effect ($0 < \text{FICI} \leq 0.5$), additive effect ($0.5 < \text{FICI} \leq 1$), and indifferent effect ($1 < \text{FICI} \leq 4$).

TABLE 4: Fractional inhibitory concentration (FIC) and FIC index (FICI) of combination between streptomycin and EFNTE.

Streptomycin		EFNTE		FICI	Interpretation
Con ^c (μg/mL)	FIC of drug A	Con ^c (μg/mL)	FIC of drug B	(FIC of A + FIC of B)	
32	MIC of A	0	MIC of A	MIC of A	MIC of A
32	1	6	0.0077	1.008	IND
16	0.5	12	0.0154	0.515	ADI
16	0.5	24	0.0307	0.531	ADI
16	0.5	49	0.063	0.563	ADI
16	0.5	98	0.125	0.625	ADI
16	0.5	195	0.25	0.75	ADI
16	0.5	391	0.5	1	IND
8	0.25	781	1	1.25	IND
4	0.125	781	1	1.125	IND
2	0.0625	781	1	1.063	IND
0	MIC of B	781	MIC of B	MIC of B	MIC of B

EFNTE: ethyl acetate fraction of *N. tetragona* 50% methanol extract; Con^c: concentration; drug A: streptomycin; drug B: EFNTE; ADI: additive effect; IND: indifferent effect; MIC: minimum inhibitory concentration. The FICI was interpreted as follows: synergistic effect ($0 < \text{FICI} \leq 0.5$), additive effect ($0.5 < \text{FICI} \leq 1$), and indifferent effect ($1 < \text{FICI} \leq 4$).

TABLE 5: Quorum sensing inhibition activity (as pigment inhibition zone diameters) of *Nymphaea tetragona* 50% methanol extract and solvent fractions of the crude extract against *C. violaceum*.

Solvent fraction/antibiotic	Concentration ($\mu\text{g}/\text{disk}$)	Pigment inhibition diameter (mm)
		Mean \pm SD
Tetracycline	10	16.33 \pm 0.58*
Furanone	100	16.67 \pm 0.58 ^a
Normal Saline	—	ND
NTME	600	13.67 \pm 1.15 ^c
DFNTE	600	13.33 \pm 0.58 ^c
EFNTE	600	14.67 \pm 1.15 ^b
BFNTE	600	14.33 \pm 1.53 ^b

NTME: *N. tetragona* 50% methanol extract, DFNTE: dichloromethane fraction of *N. tetragona* 50% methanol extract, EFNTE: ethyl acetate fraction of *N. tetragona* 50% methanol extract, BFNTE: butanol fraction of *N. tetragona* 50% methanol extract. *Diameter of clear zone. ND: no activity detected. Data shown represent the mean \pm SD of three replicates. Different alphabets indicate significant difference.

used in the treatment of infectious diseases of human and animals [45]. Thus, to combat infectious diseases associated with resistant pathogens, development of alternative antimicrobial drugs is urgent [46, 47]. The *in vitro* activity of EFNTE against resistance strains of *Salmonella typhimurium* (Table 1) reflects that the plant could be a good candidate as a source of active phytoconstituents to minimize the development of bacterial resistance and to ensure clinical cure of bacterial infection.

In the present study, MIC results of EFNTE exhibited antibacterial activity against all strains of *S. typhimurium* tested which have been shown to be resistant in one to six out of eight antibiotics (Table 1). The time-kill assay also exposed that the extract fraction effectively inhibited the growth of *S. typhimurium*. Utilities of EFNTE are again explored by the combination interactions with commercial antibiotics where it possessed synergistic effect with tylosin and additive and/or indifferent effects with other tested antibiotics. Quorum sensing inhibition activity of NTME and solvent fractions of the extract demonstrated varied level of effects statistically wherever EFNTE and BFNTE showed the greatest activity among all fractions and crude extract. Furthermore, the GC-MS analysis was performed to investigate possible components from the NTME for its antibacterial potential. This GC-MS analysis efficiently confirmed the existence of some major phenolic compounds (methyl gallate and pyrogallol) along with several other minor constituents.

MIC and MBC of NTME and its solvent fractions were studied against 5 strains of *S. typhimurium*. The results in Table 2 indicated that the EFNTE possessed the strongest antibacterial activity among all the fractions and crude extract. Considering the antibacterial activity, EFNTE was selected for further investigation. Extract having MIC values less than 8 mg/mL was believed as active crude extract [48]. Again, it was suggested to avoid the MIC greater than 1 mg/mL for crude extract and 0.1 mg/mL for isolated compounds. 0.1 mg/mL and 0.01 mg/mL MICs correspondingly for crude extract and isolated compounds would be very interesting activity [49]. Including all, MIC values less than 1 mg/mL were considered as good activity in the current study.

Time-kill curves have been used to determine whether the effects of EFNTE are either bacteriostatic or bactericidal

and are useful for the evaluation of the pharmacodynamic characteristics of new antimicrobial agents [50]. According to our results, EFNTE shows bacteriostatic activity against the tested bacteria, since a reduction $\geq 99.9\%$ of the inoculums was not observed compared to the growth control (Figure 1). At its 1 \times MIC and 2 \times MIC, EFNTE was able to inhibit *S. typhimurium* in the first 8 h of incubation. Then, 1 \times MIC and 2 \times MIC of EFNTE treated groups started their log phase. At the end of the incubation period (24 h), 3-fold reduced inoculums concentration in 2 \times MIC of EFNTE and 2-fold lower inoculums concentration in 1 \times MIC of EFNTE were achieved in contrast to control, indicating that EFNTE displays a bacteriostatic effect.

It is always suggested to treat bacterial infections with a combination of antimicrobial agents for the prevention of drug resistance development and to improve efficacy. Drug combinations having synergistic interactions are generally considered as more effective and, therefore, preferable [28]. Incidentally, EFNTE is proved to have synergistic effects with tylosin against *S. typhimurium*. The EFNTE also has additive and indifferent effects with streptomycin and other antibiotics tested against the same bacterial strain and has no antagonistic interactions with those antibiotics. Excellent *in vitro* activity combined with synergistic effects with other antibacterial drugs underscores the potential utility of EFNTE for the treatment of *Salmonella* associated infections.

In this study, NTME and its solvent fractions also possessed quorum sensing inhibition activity. EFNTE and BFNTE are considered as very sensitive or sensitive among them, whereas the crude extract and the rest of the fractions are sensitive according to the previous report [33]. Every fraction of extract may have many major and minor compounds with QS-inhibition activity which is reflected in the result. Furthermore, many compounds having antimicrobial activity were identified by the GC-MS analysis of DFNTE, EFNTE, and BFNTE. Methyl gallate, pyrogallol, and some hydrocarbons (hexacosan, heptacosan, octacosan, etc.) are important in response to desired type of effects and the proportion of those compounds in extracts. Pyrogallol is reported to have quorum sensing inhibition activity [36, 37] and antimicrobial activity [38] that is present in both EFNTE (20.61%) and BFNTE (22.09%). Methyl gallate is identified

TABLE 6: Major compound list of three solvent fractions according to their contribution in respective solvent and compounds were listed followed by their elution order including reported activity.

RT	% area	Compound	Activity	Reference
Dichloromethane fraction of <i>Nymphaea tetragona</i> 50% methanol extract				
35.46	3.18	Heneicosan	—	
37.12	5.44	Tetracosan	—	
38.71	7.9	Pentacosan	—	
40.24	11.18	Hexacosan	Antimicrobial effects	[35]
41.71	10.57	Heptacosan	Antifungal activity	[35]
43.14	8.19	Octacosan	Antimicrobial effects	[35]
Ethyl acetate fraction of <i>Nymphaea tetragona</i> 50% methanol extract				
14.37	5.9	6,8-Dimethylbenzocyclooctene	—	
15.79	20.61	Pyrogallol	Antibacterial, QS inhibition, and potent tyrosinase inhibitor.	[36–39]
16.79	1.31	1,4-Cyclohexanedicarboxylic acid, 2,5-dioxo-diethyl ester	Anticancer.	[40]
26.90	1.39	1-[7'-Methylbenzofuran-2'-carbonyl]-3-ethylazulene	Derivatives have antibacterial, antifungal, anti-inflammatory, analgesic, antidepressant, anticonvulsant, antitumor, anti-HIV, antidiabetic, antitubercular activity.	[41, 42]
27.65	70.44	Methyl gallate	Antimicrobial	[43]
Butanol fraction of <i>Nymphaea tetragona</i> 50% methanol extract				
12.86	57.61	2-Hydrazinoquinoline	—	
15.8	22.09	Pyrogallol	Antibacterial, QS inhibition, and potent tyrosinase inhibitor.	[36–39]
27.39	12.77	Methyl gallate	Antibacterial	[43, 44]

from EFNTE (70.44%) and BFNTE (12.77%) which is an antibacterial agent [43]. The scenario is now clear that methyl gallate and pyrogallol were contributing the vital role of the activity of EFNTE where they occupy about 91% of the total. The similar level of QS inhibition activity of EFNTE and BFNTE may be because of the presence of pyrogallol almost in equivalent amount in both fractions. The combined activity of those hydrocarbons or minor compounds present in DFNTE may possess the anti-QS activity.

This is the first study describing the combination interaction with commercial antibiotics and quorum sensing inhibition activity of *N. tetragona* extract. This study is also reporting the existence of methyl gallate and pyrogallol for the first time from this species. Together with all the promising *in vitro* assay findings, we believe that *N. tetragona* 50% methanol extract is expected to become a novel antimicrobial treatment for *Salmonella* infection of animals and human. Further study is needed to explore the mechanism of quorum sensing inhibition, antibacterial, and synergistic activity of the extract. *In vivo* activity and cytotoxic effects of the extract are also necessary to expose.

Abbreviations

NTME: *Nymphaea tetragona* 50% methanol extract
 DFNTE: Dichloromethane fraction of *Nymphaea tetragona* 50% methanol extract
 EFNTE: Ethyl acetate fraction of *Nymphaea tetragona* 50% methanol extract
 BFNTE: Butanol fraction of *Nymphaea tetragona* 50% methanol extract
 GC-MS: Gas chromatography-mass spectrometry.

Conflict of Interests

None of the authors have any conflict of interests to declare.

Acknowledgments

This research was supported in part by the Next Generation BioGreen 21 Program (no. PJ009007), Republic of Korea, and in part by the Ministry of Trade, Industry and Energy (MOTIE) and Korea Institute for Advancement of Technology (KIAT) through the Promoting Regional specialized Industry.

References

- [1] T. J. Humphrey, "Public health aspects of *Salmonella* infections," in *Salmonella in Domestic Animals*, C. Wray and A. Wray, Eds., CABI, Wallingford, UK, 2000.
- [2] E. Scallan, R. M. Hoekstra, F. J. Angulo et al., "Foodborne illness acquired in the United States—major pathogens," *Emerging Infectious Diseases*, vol. 17, no. 1, pp. 7–15, 2011.
- [3] M. J. Sotir, G. Ewald, A. C. Kimura et al., "Outbreak of *Salmonella wandsworth* and *Typhimurium* infections in infants and toddlers traced to a commercial vegetable-coated snack food," *The Pediatric Infectious Disease Journal*, vol. 28, no. 12, pp. 1041–1046, 2009.
- [4] *Foodborne Outbreaks due to Confirmed Bacteria Etiologies*, Centers for Disease Control and Prevention, Atlanta, Ga, USA, 2007.
- [5] A. K. Bhunia, *Foodborne Microbial Pathogens: Mechanisms and Pathogenesis*, Food Science Text Series, Springer, New York, NY, USA, 2008.
- [6] G. A. Grassl, Y. Valdez, K. S. B. Bergstrom, B. A. Vallance, and B. B. Finlay, "Chronic enteric *Salmonella* infection in mice leads to severe and persistent intestinal fibrosis," *Gastroenterology*, vol. 134, no. 3, pp. 768–780, 2008.
- [7] M. E. Ohl and S. I. Miller, "*Salmonella*: a model for bacterial pathogenesis," *Annual Review of Medicine*, vol. 52, pp. 259–274, 2001.
- [8] A. E. Mather, S. W. J. Reid, D. J. Maskell et al., "Distinguishable epidemics of multidrug-resistant *Salmonella typhimurium* DT104 in different hosts," *Science*, vol. 341, no. 6153, pp. 1514–1517, 2013.
- [9] M.-H. Lee, H. A. Kwon, D.-Y. Kwon et al., "Antibacterial activity of medicinal herb extracts against *Salmonella*," *International Journal of Food Microbiology*, vol. 111, no. 3, pp. 270–275, 2006.
- [10] D. L. Baggesen, A. Wingstrand, B. Carstensen, B. Nielsen, and F. M. Aarestrup, "Effects of the antimicrobial growth promoter tylosin on subclinical infection of pigs with *Salmonella enterica* serotype typhimurium," *American Journal of Veterinary Research*, vol. 60, no. 10, pp. 1201–1206, 1999.
- [11] T. R. Shryock, R. A. Elliott, T. H. Bennett, R. P. Basson, and R. E. Bowen, "Effect of tylosin on an experimental *Salmonella* infection in pigs," *Journal of Swine Health and Production*, vol. 6, no. 5, pp. 211–216, 1998.
- [12] M. B. Zaidi, V. Leon, C. Canche et al., "Rapid and widespread dissemination of multidrug-resistant *bla_{CMY-2}* *Salmonella typhimurium* in Mexico," *Journal of Antimicrobial Chemotherapy*, vol. 60, no. 2, pp. 398–401, 2007.
- [13] O. J. Akinjogunla, C. S. Yah, N. O. Eghafona, and F. O. Ogbeumudia, "Antibacterial activity of leave extracts of *Nymphaea lotus* (*Nymphaeaceae*) on Methicillin resistant *Staphylococcus aureus* (MRSA) and Vancomycin resistant *Staphylococcus aureus* (VRSA) isolated from clinical samples," *Annals of Biological Research*, vol. 1, no. 2, pp. 174–184, 2010.
- [14] V. K. Agnihotri, H. N. ElSohly, S. I. Khan, T. J. Smillie, I. A. Khan, and L. A. Walker, "Antioxidant constituents of *Nymphaea caerulea* flowers," *Phytochemistry*, vol. 69, no. 10, pp. 2061–2066, 2008.
- [15] J. Zhao, T. Liu, L. Ma et al., "Antioxidant and preventive effects of extract from *Nymphaea candida* flower on *in vitro* immunological liver injury of rat primary hepatocyte cultures," *Evidence-Based Complementary and Alternative Medicine*, vol. 2011, Article ID 497673, 8 pages, 2011.
- [16] K. A. Beckett, *The Concise Encyclopedia of Garden Plants*, Orbis, London, UK, 1984.
- [17] J.-H. Cheng, S.-Y. Lee, Y.-Y. Lien, M.-S. Lee, and S.-C. Sheu, "Immunomodulating activity of *Nymphaea rubra* Roxb. extracts: activation of rat dendritic cells and improvement of the TH1 immune response," *International Journal of Molecular Sciences*, vol. 13, no. 9, pp. 10722–10735, 2012.
- [18] A. K. Siddhanta, K. H. Mody, B. K. Ramavat et al., "Bioactivity of marine organisms: part VIII—screening of some marine flora of western coast of India," *Indian Journal of Experimental Biology*, vol. 35, no. 6, pp. 638–643, 1997.
- [19] B. K. Dash, M. K. Sen, K. Alam et al., "Antibacterial activity of *Nymphaea nouchali* (Burm. f) flower," *Annals of Clinical Microbiology and Antimicrobials*, vol. 12, article 27, 2013.

- [20] M. A. A. Sikder, H. R. Jisha, Md. R. Kuddus, F. Rumi, M. A. Kaisar, and M. A. Rashid, "Evaluation of bioactivities of *Nymphaea nouchali* (Burm. f)—the national flower of Bangladesh," *Bangladesh Pharmaceutical Journal*, vol. 15, no. 1, pp. 1–5, 2012.
- [21] A. K. Kumar, S. babu, and K. Ammani, "Antimicrobial and phytochemical analysis of *Nymphaea nauchali* leaf extracts," *International Journal of Research and Reviews in Pharmacy and Applied Science*, vol. 2, no. 2, pp. 142–151, 2012.
- [22] O. J. Akinjogunla, A. A. Adegoke, I. P. Udokang, and B. C. Adebayo-Tayo, "Antimicrobial potential of *Nymphaea lotus* (Nymphaeaceae) against wound pathogens," *Journal of Medicinal Plant Research*, vol. 3, no. 3, pp. 138–141, 2009.
- [23] J. Yisa, "Phytochemical analysis and antimicrobial activity of *Scoparia dulcis* and *Nymphaea lotus*," *Australian Journal of Basic and Applied Sciences*, vol. 3, no. 4, pp. 3975–3979, 2009.
- [24] 2014, <http://www.plant-talk.org/nymphaea-tetragona-india.htm>.
- [25] Clinical and Laboratory Standards Institute, "Performance standards for antimicrobial disk susceptibility tests," CLSI Approved Standards M2-A7, Clinical and Laboratory Standards Institute, Wayne, Pa, USA, 2001.
- [26] Clinical and Laboratory Standards Institute, "Performance standard for antimicrobial susceptibility testing," CLSI Seventh Informational Supplement M100-S17, Clinical and Laboratory Standards Institute, Wayne, Pa, USA, 2007.
- [27] Clinical and Laboratory Standards Institute, "Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically," CLSI Approved Standards M7-A5, Clinical and Laboratory Standards Institute, Wayne, Pa, USA, 2000.
- [28] T. Dubuisson, E. Bogatcheva, M. Y. Krishnan et al., "In vitro antimicrobial activities of capuramycin analogues against non-tuberculous mycobacteria," *Journal of Antimicrobial Chemotherapy*, vol. 65, no. 12, Article ID dkq372, pp. 2590–2597, 2010.
- [29] G. M. Eliopoulos and R. C. Moellering, "Antimicrobial combinations," in *Antibiotics in Laboratory Medicine*, V. Lorian, Ed., pp. 330–396, Williams & Wilkins, Baltimore, Md, USA, 4th edition, 1996.
- [30] V. M. Reddy, L. Einck, and C. A. Nacy, "In vitro antimycobacterial activities of capuramycin analogues," *Antimicrobial Agents and Chemotherapy*, vol. 52, no. 2, pp. 719–721, 2008.
- [31] F. C. Odds, "Synergy, antagonism, and what the checkerboard puts between them," *Journal of Antimicrobial Chemotherapy*, vol. 52, no. 1, article 1, 2003.
- [32] M. V. Alvarez, M. R. Moreira, and A. Ponce, "Antiquorum sensing and antimicrobial activity of natural agents with potential use in food," *Journal of Food Safety*, vol. 32, no. 3, pp. 379–387, 2012.
- [33] A. G. Ponce, R. Fritz, C. Del Valle, and S. I. Roura, "Antimicrobial activity of essential oils on the native microflora of organic Swiss chard," *LWT—Food Science and Technology*, vol. 36, no. 7, pp. 679–684, 2003.
- [34] M. R. Moreira, A. G. Ponce, C. E. Del Valle, and S. I. Roura, "Inhibitory parameters of essential oils to reduce a foodborne pathogen," *LWT—Food Science and Technology*, vol. 38, no. 5, pp. 565–570, 2005.
- [35] C. Gaspar-Marques, M. F. Simões, M. L. Valdeira, and B. Rodríguez, "Terpenoids and phenolics from *Plectranthus strigosus*, bioactivity screening," *Natural Product Research*, vol. 22, no. 2, pp. 167–177, 2008.
- [36] N. Ni, G. Choudhary, M. Li, and B. Wang, "Pyrogallol and its analogs can antagonize bacterial quorum sensing in *Vibrio harveyi*," *Bioorganic & Medicinal Chemistry Letters*, vol. 18, no. 5, pp. 1567–1572, 2008.
- [37] T. Defoirdt, G. S. Pande, K. Baruah, and P. Bossier, "The apparent quorum-sensing inhibitory activity of pyrogallol is a side effect of peroxide production," *Antimicrobial Agents and Chemotherapy*, vol. 57, no. 6, pp. 2870–2873, 2013.
- [38] I. Kocaçalışkan, I. Talan, and I. Terzi, "Antimicrobial activity of catechol and pyrogallol as allelochemicals," *Zeitschrift für Naturforschung C: Journal of Biosciences*, vol. 61, no. 9–10, pp. 639–642, 2006.
- [39] J. Choi, S. J. Bae, Y. M. Ha et al., "A newly synthesized, potent tyrosinase inhibitor: 5-(6-Hydroxy-2-naphthyl)-1,2,3-benzenetriol," *Bioorganic & Medicinal Chemistry Letters*, vol. 20, no. 16, pp. 4882–4884, 2010.
- [40] W. Wichitnithad, U. Nimmanit, S. Wacharasindhu, and P. Rojsitthisak, "Synthesis, characterization and biological evaluation of succinate prodrugs of curcuminoids for colon cancer treatment," *Molecules*, vol. 16, no. 2, pp. 1888–1900, 2011.
- [41] H. A. Abdel-Aziz, A. A. I. Mekawey, and K. M. Dawood, "Convenient synthesis and antimicrobial evaluation of some novel 2-substituted-3-methylbenzofuran derivatives," *European Journal of Medicinal Chemistry*, vol. 44, no. 9, pp. 3637–3644, 2009.
- [42] M. Kamal, A. K. Shakya, and T. Jawaid, "Benzofurans: a new profile of biological activities," *International Journal of Medical and Pharmaceutical Sciences*, vol. 1, no. 3, pp. 1–15, 2011.
- [43] J.-G. Choi, O.-H. Kang, Y.-S. Lee et al., "Antibacterial activity of methyl gallate isolated from *Galla rhois* or *carvacrol* combined with nalidixic acid against nalidixic acid resistant bacteria," *Molecules*, vol. 14, no. 5, pp. 1773–1780, 2009.
- [44] J.-G. Choi, O.-H. Kang, Y.-S. Lee et al., "In vitro activity of methyl gallate isolated from *Galla Rhois* alone and in combination with ciprofloxacin against clinical isolates of *Salmonella*," *Journal of Microbiology and Biotechnology*, vol. 18, no. 11, pp. 1848–1852, 2008.
- [45] J. Davies and D. Davies, "Origins and evolution of antibiotic resistance," *Microbiology and Molecular Biology Reviews*, vol. 74, no. 3, pp. 417–433, 2010.
- [46] A. Berahou, A. Auhmani, N. Fdil, A. Benharref, M. Jana, and C. A. Gadhi, "Antibacterial activity of *Quercus ilex* bark's extracts," *Journal of Ethnopharmacology*, vol. 112, no. 3, pp. 426–429, 2007.
- [47] K. Salomão, P. R. S. Pereira, L. C. Campos et al., "Brazilian propolis: correlation between chemical composition and antimicrobial activity," *Evidence-Based Complementary and Alternative Medicine*, vol. 5, no. 3, pp. 317–324, 2008.
- [48] B. Ncube, J. F. Finnie, and J. van Staden, "In vitro antimicrobial synergism within plant extract combinations from three South African medicinal bulbs," *Journal of Ethnopharmacology*, vol. 139, no. 1, pp. 81–89, 2012.
- [49] J. L. Ríos and M. C. Recio, "Medicinal plants and antimicrobial activity," *Journal of Ethnopharmacology*, vol. 100, no. 1–2, pp. 80–84, 2005.
- [50] M. A. Pfaller, D. J. Sheehan, and J. H. Rex, "Determination of fungicidal activities against yeasts and molds: lessons learned from bactericidal testing and the need for standardization," *Clinical Microbiology Reviews*, vol. 17, no. 2, pp. 268–280, 2004.