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Concise total synthesis of (±)-actinophyllic acid

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Abstract

A concise total synthesis of the complex indole alkaloid (\pm) -actinophyllic acid was accomplished by a sequence of reactions requiring only 10 steps from readily-available, known starting materials. The approach featured a Lewis acid-catalyzed cascade of reactions involving stabilized carbocations that delivered the tetracyclic core of the natural product in a single chemical operation. Optimal conversion of this key intermediate into (\pm) -actinophyllic acid required judicious selection of a protecting group strategy.

Keywords

Total Synthesis; Cascade Reaction; Iminium Ion; Mannich-type Reaction

1. Introduction

It has been estimated that as many as 600,000 people suffer annually from thrombotic diseases in the United States.¹ The mortality rate associated with thrombosis is exceedingly high, necessitating an improvement in the current standard of care. One risk factor for thrombosis is unregulated fibrinolysis, or the breakdown of fibrin clots by plasmin. Fibrinolysis is in part regulated by carboxypeptidase U (CPU), alternatively called activated thrombin-activatable fibrinolysis inhibitor (TAFIa), which catalyzes the cleavage of *C*-terminal arginine and lysine residues from fibrin and fibrin degradation products.² These basic residues are essential for the activation of plasmin via the formation of a ternary complex between fibrin, plasminogen, and tissue-type plasminogen activator (tPA). Because plasmin promotes the breakdown of fibrin, CPU acts as an endogenous inhibitor of the fibrinolysis process.³ Accordingly, CPU has attracted considerable attention from the scientific community as a potential target for the treatment of thrombosis,^{4–7} although there is currently no CPU inhibitor in the clinic.⁸ In 2005, Carroll and co-workers reported the

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Supplementary Data

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Experimental procedures and characterization data for selected compounds and x-ray crystallographic data for compounds 2,³⁷ 24,³⁸ and 28.³⁹ Supplementary data related to this article can be found at http://dx.doi.....

isolation of the indole alkaloid actinophyllic acid (1) from the leaves of *Alstonia actinophylla*, and using a CPU/hippuricase coupled enzyme assay they discovered that 1 was a potent CPU inhibitor (IC₅₀ = 0.84μ M) (Scheme 1).⁹

The key structural features of the bridged architecture of **1**, which is unique among known natural products, comprise a pyrrolidine ring, a cyclic hemiketal, a carboxylic acid appended to a quaternary carbon atom, an indol-3-ylmethanamine moiety, and five contiguous stereocenters. Indeed, the compact and highly functionalized hexacyclic framework embodied in **1** has prompted several total synthesis campaigns. Overman and co-workers completed the first total syntheses of (\pm) - and (-)-**1** through the elegant application of the venerable aza-Cope/Mannich sequence.¹⁰ Importantly, the synthesis of (-)-**1** allowed for the determination of the absolute configuration of the natural product via analysis of its methyl ester derivatives through electronic circular dichroism (ECD).¹¹ Other reports from the groups of Wood,¹² Taniguchi,¹³ Maldonado,¹⁴ and Coldham¹⁵ have revealed a diverse array of approaches toward **1**.

Our plan for the synthesis of (\pm) -actinophyllic acid (1) is outlined retrosynthetically in Scheme 1 and commences with the oxidation of the neopentyl alcohol in 2, which would result from deprotection of the primary alcohols in 3 and subsequent hemiketalization. The pyrrolidine ring in 3 would be installed via deprotection of the nitrogen atom in 4, followed by reductive alkylation of N(2) and cyclization at C(19) by intramolecular enolate alkylation. We envisioned that the tetracyclic core of the natural product embodied in 4 would arise from the union of electrophile 6 and π -nucleophile 7. In particular, we reasoned that capture of the stabilized carbocation generated upon Lewis-acid initiated ionization of 6 with the π -nucleophile 7 via a vinylogous Mannich-like reaction¹⁶ would produce the *N*acyliminium ion 5 that would then undergo attack by the 3-position of the indole in a Pictet-Spengler type reaction.¹⁷

It is notable that the pivotal reaction cascade proposed for the transformation of **6** and **7** into the tetracyclic intermediate **4** was inspired by chemistry developed during efforts directed toward our syntheses of the welwitindolinone alkaloids.¹⁸ Indeed, we had developed this reaction into a general method to couple stabilized carbocations derived from heteroaromatic carbinols with various π -nucleophiles to deliver β -heteroaryl propionates as exemplified by the preparation of **10** by the TMSOTf-promoted reaction of **8** and **9** (Equation 1).¹⁹



(1)

We thus sought to further demonstrate the utility of this powerful transformation by applying it to the total synthesis of (\pm) -actinophyllic acid (1), and selected aspects of our work in the area and a total synthesis of (\pm) -1 were recently disclosed.²⁰ Herein, we provide a full account of those studies.

2. Results and Discussion

In order to probe the feasibility of the key cascade sequence, we sought an efficient route to the requisite starting materials. Accordingly, carboxylate-directed deprotonation at the 2-position of indole (**11**) produced a dianion that underwent reaction with 1,3-dibenzyloxyacetone²¹ to afford an alkoxide intermediate that was acetylated with acetic anhydride to deliver acetate **12** in 83% yield.²² The structure of compound **12** was confirmed through single crystal X-ray analysis. Deprotonation of the known hydroazepinone **13**, which was prepared in a single step from the photolysis of *N*-vinylpyrrolidinone,²³ followed by reaction with several alkyl chloroformates gave carbamates **14–16**, which were then converted into their corresponding triethylsilyl enol ethers **17–19**.

With the desired electrophile precursor 12 and the π -nucleophiles 17–19 in hand, we explored the pivotal cascade sequence, beginning with conditions similar to those shown in Equation 1. However, upon treatment of a mixture of 12 and 17 with TMSOTf in CH₂Cl₂ at -78 °C, the formation of the desired tetracycle 20 was not observed, and the α , β -unsaturated ketone 21 and tetracycle 22 were isolated in 15% and 18% yields, respectively (Scheme 3). The ratio of 21:22 was found to be temperature dependent with tricycle 21 being the sole product (isolated in 63% yield) when the temperature was maintained at -78 °C. We found that compound 21 was converted completely to tetracycle 22 upon resubjection to the reaction conditions at room temperature. Although this transformation did not provide the desired bridged compound 20, the observation that the two key C-C bonds had been successfully formed encouraged us to pursue the reaction.

Based on this unexpected result, we reasoned that **21** and **22** were formed by degradation of the initially formed product **20** via two potential pathways (Scheme 4). In one scenario, β -elimination of the carbamate group from the silyl enol ether **20** would give the α , β -unsaturated ketone **21** (Path A), which upon cyclization and dehydration would produce tetracycle **22**. Alternatively, **20** could suffer a gramine-type fragmentation to give intermediate **23** (Path B).²⁴ Loss of the silyl group would then furnish **21**, which may then cyclize to **22**.

These plausible degradation pathways suggested that the initially formed tetracycle **20** was unstable to the acidic reaction conditions, so we queried whether adding a suitable base might suppress any deleterious side reactions. In an initial experiment, we found that the presence of Hünig's base precluded any reaction, presumably due to coordination of TMSOTf with the basic nitrogen atom of the amine. In order to prevent such coordination, we employed 2,6-di-*tert*-butylpyridine as the base, but upon the usual aqueous workup, we again isolated a mixture of **21** and **22**. We eventually discovered that quenching the reaction at -78 °C with NaOMe in MeOH, followed by an aqueous work-up, avoided the formation of any fragmentation products. In the event, treatment of a mixture of indole **12** and the π -nucleophiles **17–19** with TMSOTf in the presence of 2,6-di*tert*-butylpyridine, followed by addition of NaOMe in MeOH at -78 °C delivered the silyl enol ethers **20**, **24**, and **25** in 53–80% yields (Equation 2). The structure of methyl carbamate **24** was confirmed by single crystal X-ray analysis (Figure 1). To our dismay, we found that **20**, **24**, and **25** were rather

unstable to further synthetic manipulations, a fact that severely limited their utility as intermediates en route to (\pm) -actinophyllic acid (1).



(2)

Given that the enol ether moieties of **20**, **24**, and **25** are poised to undergo facile β elimination (Scheme 4, Path A), we hypothesized that the corresponding ketones might be more stable. Recognizing that *N*-Alloc groups can be easily removed under mild conditions, we prepared the ketone **26** in 92% yield by treating **25**, which was formed *in situ* by the reaction of **12** with **19** in the presence of TMSOTf and 2,6-di-*tert*-butylpyridine, with TBAF (Scheme 5). Palladium-catalyzed removal of the *N*-Alloc group²⁵ gave the secondary amine **27**, but this intermediate proved to be rather labile and difficult to isolate in pure form. Although we did not fully characterize any of the side-products derived from **27**, ¹H NMR spectra of product mixtures suggested that fragmentation pathways similar to those outlined in Scheme 4 were operative.

Two possible strategies to increase the stability of intermediates derived from **26** were thus considered: We could either reduce the ketone, a tactic with some precedent,^{13,26} or place an electron withdrawing group on the nitrogen atom of the indole ring. Because we had encountered some difficulties in preliminary attempts to introduce an electron withdrawing group on the indole nitrogen atom of **26**, we elected instead to reduce the ketone function in **26** with lithium borohydride and obtained **28** as a single diastereomer in 82% yield (Scheme 5). The structure of **28** was unambiguously assigned via single crystal X-ray analysis (Figure 2). Alcohol **28** underwent facile palladium-catalyzed Alloc cleavage using phenylsilane as the π -allyl cation scavenger to yield the amino alcohol **29** in 73% yield.²⁵

With **29** in hand, we turned our attention to forming the pyrrolidine ring, thereby establishing the pentacyclic core of actinophyllic acid (**1**). In an exploratory experiment, the secondary amine group in **29** was reductively alkylated with chloroacetaldehyde, but several attempts to oxidize the alcohol group in **30** and induce a base-catalyzed ring closure of the resultant ketone to give **31**, even under the conditions described by Taniguchi for a similar conversion,¹³ were unavailing; the tetracyclic alcohol **34** was obtained in 41% yield instead (Scheme 6). Isolation of the primary alcohol **34** may be rationalized by the formation of an aziridinium ion such as **32** that underwent subsequent fragmentation (*cf.*, Scheme 4) and nucleophilic ringopening with water to give **33**, which could then provide **34** via sequential cyclization and dehydration.²⁷

At this juncture, the main issue to be confronted was whether **26** or a compound derived therefrom was a viable intermediate in the synthesis of (\pm) -actinophyllic acid (1). Because

step-count was a secondary consideration in establishing proof-of-principle, we developed a strategy that relied upon installing an electron withdrawing group on the indole nitrogen atom to mitigate fragmentation via a gramine-type pathway. Toward this end, protection of the hydroxyl group in **28** as its trimethylsilyl ether, followed by treatment with neat Bocanhydride in the presence of a catalytic amount of DMAP at 100 °C and then TBAFmediated deprotection of the alcohol furnished the *N*-protected indole **35** in 88% overall yield (Scheme 7). Utilizing the previously developed conditions for palladium-catalyzed deprotection and reductive amination, **36** was obtained in 56% overall yield. Although oxidation of **36** with IBX gave mixtures of the desired ketone **37** together with side products that appeared to arise from fragmentation, oxidation of **36** under Swern conditions cleanly delivered **37** in 86% yield. Treatment of **37** with sodium *tert*-butoxide resulted in intramolecular enolate alkylation and formation of the pyrrolidine ring, giving the pentacycle **38** in 94% yield.

Toward completing the total synthesis of (\pm) -actinophyllic acid (1), our initial plan was to remove the 1,3-diol and indole protecting groups in **38** by hydrogenolysis in the presence of protic acid to give **2**, but these conditions led exclusively to the cyclic carbamate **39** (Scheme 8). Although **39** was converted into **2** upon standing in wet methanol, we developed a facile two-step, one-pot procedure whereby the Boc group was first removed by treating **38** with HCl in methanol at 65 °C. The solution was simply cooled, Pd/C (10%) was added, and the mixture was stirred under an atmosphere of hydrogen to furnish the hexacyclic alcohol **2** as its hydrochloride salt in 86% yield. The structure of **2** was confirmed through single crystal X-ray analysis (Figure 3).

The oxidation of the neopentyl alcohol **2** to give (\pm)-actinophyllic acid (**1**) proved surprisingly difficult. The insolubility of **2** precluded use of many standard reaction conditions, and numerous attempts to oxidize **2** led to poor material balances and low yields of **1**.^{28,29} We surmised that oxidation of the indole ring and/or ketal opening and oxidation of both primary alcohols were complicating factors, but we were not able to isolate products that support either of these hypotheses. After considerable experimentation, we discovered that oxidation of **2** with TPAP/NMO³⁰ or PDC in DMF³¹ produced (\pm)-actinophyllic acid (**1**), albeit in irreproducible yields. Finally, we found that oxidation of **2** with IBX in DMSO at 50 °C produced an intermediate aldehyde, which upon treatment with *N*hydroxysuccinimide and further oxidation yielded an activated ester that was readily hydrolyzed under basic conditions to deliver (\pm)-actinophyllic acid (**1**) in 31% yield (Equation 3).³² The spectral data for **1** matched that of a synthetic sample graciously provided by Professor Larry E. Overman.



This first generation synthesis of (\pm) -actinophyllic acid (1) was completed in 14 steps and 5% overall yield from known materials. However, the sequence suffers from far too many protection/deprotection and refunctionalization steps; avoiding so many "unproductive" steps was clearly desirable in order to enhance the overall efficiency of the synthesis. After considering various possibilities, we decided to revisit the tactic of placing an electron withdrawing group on the indole nitrogen atom of 26. This seemed a risky undertaking because 26 had previously proven difficult to handle (vide supra), and the forcing conditions required to introduce the Boc group onto a precursor of 35 did not augur well for such a protective move. However, despite our reservations, we discovered that treating 26 with Boc-anhydride and DMAP in toluene at room temperature delivered the desired N-protected compound 40 in 83% yield (Scheme 9). Under these milder reaction conditions no products resulting from fragmentation of 26 were observed in the ¹H NMR spectra of the crude reaction mixture. The subsequent Alloc deprotection was optimized through the use of N,Ndimethylbarbituric acid as the π -allyl cation scavenger.³³ and the amine **41** was obtained in 92% yield. Reductive amination of 41 with chloroacetaldehyde, followed by base-induced cyclization as before furnished pentacycle 38 in 83% yield over the two steps. This improved second generation synthesis delivered (\pm) -1 in only 10 total steps and 10% overall yield from known materials.

3. Summary

Inspired by previous work in our laboratories, a novel cascade of reactions involving stabilized carbocations and π -nucleophiles was developed to deliver the tetracyclic core of (\pm) -actinophyllic acid (1) in a single chemical operation from readily-available starting materials. Although several synthetic intermediates exhibited a proclivity to suffer a troublesome fragmentation, consideration of possible degradation pathways enabled us to solve the underlying problems and complete the synthesis of (\pm) -actinophyllic acid (1) via two different routes. The first of these required 14 steps and proceeded in 5% overall yield from readily-available starting materials, whereas the second generation approach proceeded in 10% overall yield and required only 10 steps from the same starting materials. Toward identifying novel, biologically-active compounds, several intermediates were diverted to prepare analogs for biological evaluation,³⁴ and a number of tetracyclic derivatives of **28** were found to exhibit significant anticancer activity as reported elsewhere.²⁰ Notably. these compounds are not readily accessible by known synthetic pathways, including those that have been reported for the synthesis of actinophyllic acid. Accordingly, this discovery of promising anticancer leads underscores the value, which is not universally acknowledged, of developing alternative entries to complex natural products as a strategy to generate compounds for biological screening that would not be otherwise accessible. Details of our biological studies and other applications of cascade reactions to address problems in alkaloid synthesis will be reported in due course.

4. Experimental

4.1. General

Tetrahydrofuran (THF) and toluene were passed through two columns of activated neutral alumina prior to use.³⁵ Dichloromethane (CH₂Cl₂), acetic anhydride (Ac₂O), and allyl chloroformate (Alloc-Cl) were freshly distilled over CaH₂. Trimethylsilyl trifluoromethanesulfonate (TMSOTf) was distilled over P₂O₅. All other reagents and solvents were reagent grade and were used as received unless otherwise noted. Reactions were performed under a nitrogen or argon atmosphere, and moisture sensitive reactions were performed with flame- or oven-dried glassware. Reaction temperatures are reported as the temperatures of the bath surrounding the vessel.

Proton nuclear magnetic resonance (¹H NMR) and carbon nuclear magnetic resonance (¹³C NMR) spectra were acquired in CDCl₃ unless otherwise noted. Chemical shifts are reported in parts per million (ppm, δ), downfield from tetramethylsilane (TMS, $\delta = 0.00$ ppm) and are referenced to residual solvent (CDCl₃, $\delta = 7.26$ ppm (¹H) and 77.16 ppm (¹³C)). Coupling constants (*J*) are reported in hertz (Hz) and the resonance multiplicity abbreviations used are: s, singlet; d, doublet; t, triplet; q, quartet; dt, doublet of triplets; dd, doublet of doublets; ddd, doublet of doublet of doublets; m, multiplet; comp, overlapping multiplets of magnetically non-equivalent protons. The abbreviations br and app stand for broad and apparent, respectively. Infrared (IR) spectra were obtained as thin films on sodium chloride plates. Thin-layer chromatography (TLC) was performed on EMD 60 F254 glass-backed pre-coated silica gel plates. Flash chromatography was performed using Silicycle[®] SiliaFlash F60[®] (40–63 µm) silica gel eluting with the solvents indicated according to the procedure of Still.³⁶ Reverse phase chromatography was performed on a CombiFlash[®] Companion instrument utilizing a RediSep[®] R_f 43 g C18 Reverse Phase column.

4.2. Selected Experimental Procedures

1,3-Bis(benzyloxy)-2-(1H-indol-2-yl)propan-2-yl acetate (12)—A solution of n-BuLi (6.8 mL of a 2.17 M solution in hexanes, 14.8 mmol) was added dropwise to a solution of indole (11) (1.6 g, 13.5 mmol) in dry THF (39 mL) at -78 °C. The reaction was stirred for 30 min, and dried CO_2 gas (passed through a short column of anhydrous $CaSO_4$) was bubbled through the reaction for 15 min. The reaction was stirred for an additional 15 min at -78 °C, the bath was removed, and the reaction was stirred for 1 h at room temperature. Excess CO₂ was removed via freeze-pump-thaw (3 cycles), and then the reaction was evacuated for 30 sec and sparged with Ar for 20 min. The solution was cooled to -78 °C, whereupon t-BuLi (10.0 mL of a 1.47 M solution in pentane, 14.8 mmol) was added dropwise. The reaction was stirred for 80 min and transferred via cannula to a solution of 1,3-dibenzyloxyacetone (4.0 g, 14.8 mmol) in dry THF (39 mL) at -78 °C. The reaction was stirred for 2.5 h, acetic anhydride (3.8 mL, 40.5 mmol) was added dropwise, and the reaction was allowed to warm to room temperature over 7 h. Saturated aqueous NaHCO₃ (50 mL) was added, and the reaction was stirred for 30 min, whereupon saturated brine (80 mL) and Et₂O (200 mL) were added. The layers were separated, and the aqueous layer was extracted with Et_2O (2 × 100 mL). The combined organic layers were dried (MgSO₄), filtered and

concentrated under reduced pressure. The resultant yellow oil was purified by flash column chromatography eluting with hexanes/EtOAc (90 : $10 \rightarrow 85 : 15 \rightarrow 80 : 20 \rightarrow 75 : 25 \rightarrow 70 : 20$) containing 1% Et₃N to give 5.0 g (85%) of **12** as a pale yellow solid: mp 78–79 °C; ¹H NMR (400 MHz, CD₃CN) & 9.55 (br s, 1 H), 7.58 (d, *J* = 8.0 Hz, 1 H), 7.45 (d, *J* = 8.0 Hz, 1 H), 7.39–731 (comp, 10 H), 7.21-7.17 (m, 1 H), 7.11-7.07 (m, 1 H), 6.46 (dd, *J* = 2.4, 0.8 Hz, 1 H), 4.58 (s, 4 H), 4.21 (s, 4 H), 2.08 (s, 3 H); ¹³C NMR (100 MHz, CD₃CN) & 169.8, 138.6, 137.3, 136.2, 128.6, 128.0, 127.9, 122.0, 120.5, 119.8, 111.4, 99.9, 80.7, 73.4, 70.7, 21.5; IR (neat) 3416, 2864, 1741, 1454, 1367, 1237, 1103, 1018, 738, 698 cm⁻¹; mass spectrum (CI) *m/z* 430.2010 [C₂₇H₂₈NO₄ (M+1) requires 430.2018].

General procedure for the acylation of hydroazepinone 13—A solution of *n*-BuLi (1.2 equiv of a 2.5 M solution in hexanes) was added dropwise to a solution of **13** (1 equiv) in THF (0.11 M) at -78 °C. After stirring for 0.5 h, the chloroformate (1.3 equiv) was added dropwise, and the reaction was stirred for 1.5 h. The bath was removed, and the reaction was stirred for 1.5 h at room temperature. The reaction was concentrated and partitioned between a mixture of saturated aqueous NaHCO₃, saturated brine and CH₂Cl₂. The layers were separated, and the aqueous layer was extracted twice with CH₂Cl₂. The combined organic layers were washed with saturated brine, dried (Na₂SO₄), filtered, and concentrated under reduced pressure. The crude product was purified by flash column chromatography.

Benzyl 5-oxo-2,3,4,5-tetrahydro-1*H***-azepine-1-carboxylate (14)**—Purification: hexanes/EtOAc (4 : 1 \rightarrow 3 : 2) to give 1.94 g (84%) of **14** as a colorless amorphous solid: ¹H NMR (400 MHz, CD₂Cl₂) δ 7.47 (d, *J* = 10.6 Hz, 1 H), 7.41-7.34 (comp, 5 H), 5.24 (s, 2 H), 5.19 (d, *J* = 10.6 Hz, 1 H), 3.97-3.94 (m, 2 H), 2.62 (app t, *J* = 6.5 Hz, 2 H), 2.06-2.00 (m, 2 H); ¹³C NMR (100 MHz, CD₂Cl₂) δ 200.7, 135.8, 129.0, 128.9, 128.6, 110.1, 110.0, 69.4, 48.5, 43.5, 22.9; IR (neat) 1722, 1613, 1315, 1250, 1221, 1112 cm⁻¹; mass spectrum (CI) *m/z* 246.1131 [C₁₄H₁₆NO₃ (M+1) requires 246.1130].

Methyl 5-oxo-2,3,4,5-tetrahydro-1*H*-azepine-1-carboxylate (15)—Purification: hexanes/EtOAc (4 : 1 →1 : 1) to give 1.27 g (79%) of 15 as a pale yellow oil: ¹H NMR (400 MHz, CD₂Cl₂) δ 7.44 (d, *J* = 10.6 Hz, 1 H), 5.18 (d, *J* = 10.6 Hz, 1 H), 3.94-3.92 (m, 2 H), 3.83 (s, 3 H), 2.62 (app t, *J* = 6.5 Hz, 2 H), 2.05-2.00 (m, 2 H); IR (neat) 2956, 1729, 1614, 1441, 1351, 1319, 1256, 1116, 943, 767 cm⁻¹; mass spectrum (CI) *m*/*z* 170.0815 [C₈H₁₂NO₃ (M+1) requires 170.0817].

Allyl 5-oxo-2,3,4,5-tetrahydro-1*H*-azepine-1-carboxylate (16)—Purification: hexanes/EtOAc (7 : 3 \rightarrow 8 : 4 \rightarrow 1 : 1) to give 3.7 g (84%) of 16 as a colorless solid: ¹H NMR (400 MHz) δ 7.46 (d, *J* = 10.6 Hz, 1 H), 5.99-5.89 (m, 1 H), 5.37-5.23 (comp, 3 H), 4.70 (app dt, *J* = 1.4, 5.8 Hz, 2 H), 3.96-3.93 (comp, 2 H), 2.65 (app t, *J* = 6.5 Hz, 2 H), 2.07-2.01 (comp, 2 H); ¹³C NMR (100 MHz) δ 200.9, 166.0, 137.2, 131.4, 119.3, 109.9, 68.0, 48.1, 42.7, 22.7; IR (neat) 3492, 3088, 2945, 1730, 1614, 1434, 1114, 943, 766 cm⁻¹; mass spectrum (CI) *m*/*z* 196.0974 [C₁₀H₁₃NO₃ (M+1) requires 196.0974].

Allyl 5-((triisopropylsilyl)oxy)-2,3-dihydro-1H-azepine-1-carboxylate (19)— Triisopropylsilyl chloride (7.2 g, 8.0 mL, 37.4 mmol) was added to a solution of azepine 16

(6.0 g, 30.9 mmol) in THF (330 mL) at -78 °C and stirred for 12 min. Sodium hexamethyldisilazide (20.0 mL of a 2.04 M solution in THF, 40.1 mmol) was added to the mixture dropwise, and the solution was stirred at -78 °C for 1.5 h. Saturated NaHCO₃ (100 mL) was added, the bath was removed, and stirring was continued for 15 min. Once at room temperature, the reaction was partitioned between a mixture of saturated aqueous NaHCO₃ (100 mL), saturated brine (150 mL), and Et₂O (200 mL). The layers were separated, and the aqueous layer was extracted with Et₂O (2×200 mL). The combined organic layers were washed with saturated brine (300 mL), dried (Na₂SO₄), filtered and concentrated under reduced pressure. The resultant yellow oil was purified by flash column chromatography eluting with hexanes/EtOAc (92 : $8 \rightarrow 93$: 7) containing 1% Et₃N to give 11.3 g (92%) of **16** as a pale yellow oil: ¹H NMR (400 MHz, CD₃CN) δ 6.84 (d, J = 8.9 Hz, 1 H), 6.03-5.93 (m, 1 H), 5.36-5.19 (comp, 3 H), 5.06 (d, J = 9.9 Hz, 1 H), 4.64 (app dt, J = 1.4, 5.5 Hz, 2H), 3.74 (app t, J = 4.1 Hz, 2 H), 2.35 (app q, J = 5.5 Hz, 2 H), 1.23-1.14 (comp, 3 H), 1.10-1.09 (comp, 18 H); ¹³C NMR (125 MHz, CD₃CN) 153.9, 148.9, 133.7, 128.6, 118.2, 110.4, 109.6, 67.6, 46.2, 28.6, 18.5, 13.3; IR (neat) 3060, 3030, 2944, 2866, 1691, 1461, 1411, 1366, 1233, 1199, 1106, 885, 743.6, 697.9 cm⁻¹; mass spectrum (CI) m/z 351.2225 [C₁₉H₃₃NO₃Si (M+1) requires 351.2230].

General procedure for the formation of silyl enol ethers 20, 24, and 25—

Trimethylsilyl triflate (1.1 equiv) was added dropwise to a solution of **12** (1.0 equiv), **17–19** (1.1 equiv) and 2,6-di-*tert*-butylpyridine (2.0 equiv) in CH_2Cl_2 (0.1 M in **12**) at -78 °C under argon. The reaction was stirred for 3.5 h, and sodium methoxide (14.5 equiv of a 3.20 M solution in dry methanol) was added. After 1 h, saturated aqueous NaHCO₃ was added, the dry-ice bath was removed, and the reaction was stirred for 1 h. Additional portions of saturated aqueous NaHCO₃ and saturated brine were added, and the reaction was extracted three-times with CH_2Cl_2 . The combined organic layers were washed with saturated brine, dried (Na₂SO₄), filtered and concentrated. The crude material was purified by flash column chromatography.

Benzyl (1S/R,5S/R)-6,6-bis((benzyloxy)methyl)-12-

((triisopropyIsilyI)oxy)-1,3,4,5,6,7-hexahydro-2H-1,5-ethenoazocino[4,3-

b]indole-2-carboxylate (20)—Purification: pentane/Et₂O (4 : 1) containing 1% Et₃N to give 33 mg (61%) of 20 as a colorless amorphous solid: ¹H NMR (600 MHz, CD₂Cl₂, as a mixture of rotamers) δ 9.17 (s, 0.5 H), 9.16 (s, 0.5 H), 7.56 (d, J = 7.8 Hz, 0.5 H), 7.49 (d, J = 7.4 Hz, 1 H), 7.43 (app t, J = 7.4 Hz, 2 H), 7.40-7.27 (comp, 13.5 H), 7.13-7.10 (m, 1 H), 7.08-7.03 (m, 1 H), 5.92 (d, J = 8.6 Hz, 0.5 H), 5.81 (d, J = 8.4 Hz, 0.5 H), 5.76 (dd, J = 2.4, 8.6 Hz, 0.5 H), 5.68 (dd, J = 2.4, 8.4 Hz, 0.5 H), 5.25 (d, J = 12.5 Hz, 0.5 H), 5.20 (d, J = 12.5 Hz, 0.5 H), 5.18 (d, J = 12.5 Hz, 0.5 H), 5.09 (d, J = 12.5 Hz, 0.5 H), 4.69-4.64 (comp, 2 H), 4.60-4.59 (d, J = 11.9 Hz, 1 H), 4.49-4.47 (dd, J = 3.5, 11.8 Hz, 1 H), 4.11-3.97 (m, 1 H), 3.98-3.96 (m, 1 H), 3.89-3.86 (m, 1 H), 3.25-3.20 (app dt, J = 2.6, 14.0 Hz, 0.5 H), 2.86-2.84 (comp 1H), 2.13-2.05 (m, 1 H), 1.82-1.78 (m, 1 H), 1.26-1.15 (comp, 3 H), 1.10-1.06 (comp, 18 H); ¹³C NMR (150 MHz, CD₂Cl₂, as a mixture of rotamers) δ 159.0, 158.7, 115.7, 155.5, 139.5, 139.4, 139.0, 138.52, 138.50, 138.0, 137.9, 135.0, 134.9, 128.9, 128.8, 128.7, 128.6, 128.2, 128.1, 128.1, 128.0, 128.0, 127.9, 127.8, 126.9, 126.7,

121.9, 119.8, 119.7, 118.3, 117.9, 111.1, 111.0, 109.7, 109.57, 109.55, 109.5, 74.2, 73.8, 72.5, 72.4, 72.2, 72.1, 67.2, 67.1, 60.6, 45.6, 45.4, 45.0, 41.5, 41.3, 27.5, 27.0, 18.3, 13.0; IR (neat) 2943, 2866, 1691, 1460, 1419, 1199, 1106, 884, 744, 697 cm⁻¹; mass spectrum (CI) m/z 771.4195 [C₄₈H₅₉N₂O₅Si (M+1) requires 771.4188].

(1S/R,5S/R)-Methyl 6,6-bis((benzyloxy)methyl)-12-((triisopropylsilyl)oxy)-3,4,5,6-tetrahydro-1*H*-1,5-ethenoazocino[4,3-

b]indole-2(7*H*)-carboxylate (24)—Purification: hexanes/EtOAc ($9: 1 \rightarrow 7: 3$) containing 1% Et₃N to give 324 mg (53%) of **24** a colorless amorphous solid: ¹H NMR (600 MHz, CD₂Cl₂, as a mixture of rotamers) δ 9.16-9.13 (comp, 1H), 7.52 (d, J = 30.7 Hz, 0.5 H), 7.47 (d, J = 30.7 Hz, 0.5 H), 7.37-7.25 (comp, 11 H), 7.10 (app dt, J = 2.9, 7.2 Hz, 1 H), 7.05 (app q, J = 6.9, 1 H), 5.85 (d, J = 8.5 Hz, 0.5 H), 5.72 (dd, J = 2.3, 8.6 Hz, 0.5 H), 5.67 (app s, 1 H), 4.65 (dd, J = 3.7, 11.9 Hz, 2 H), 4.58 (d, J = 11.9 Hz, 1 H), 4.46 (d, J = 11.8 Hz, 1 H), 4.01 (app dt, J = 3.7, 14.7 Hz, 0.5 H), 3.95 (d, J = 8.7 Hz, 1 H), 3.86 (app dd, J = 3.5, 12.0 Hz, 1 H), 3.88-3.84 (comp, 0.5 H), 3.79 (app dd, J = 2.3, 8.6 Hz, 1 H), 3.75 (s, 1.5 H), 3.68 (app t, 7.7 Hz, 1 H), 3.64 (s, 1.5 H), 3.25 (app t, *J* = 12.8 Hz, 1 H), 2.82 (d, *J* = 2.1 Hz, 1 H), 2.10-2.03 (comp, 0.5 H), 2.06-2.00 (comp, 0.5 H), 1.77-1.76 (comp, 0.5 H), 1.75-1.74 (comp, 0.5 H), 1.25-1.17 (comp, 3 H), 1.10-1.04 (comp, 18 H); ¹³C NMR (150 MHz, CD₂Cl₂ as a mixture of rotamers) δ 158.8, 158.6, 156.40, 156.36, 139.4, 139.0, 138.5, 135.0, 134.9, 128.9, 128.6, 128.2, 128.1, 128.0, 127.8, 126.9, 126.8, 121.9, 119.7, 118.3, 117.9, 111.1, 111.0, 110.0, 109.7, 109.6, 109.5, 74.2, 73.8, 72.43, 72.40, 72.2, 72.1, 52.7, 52.6, 45.5, 45.4, 45.3, 45.02, 45.00, 41.5, 41.2, 27.4, 26.8, 18.3, 13.0, 12.9; IR (neat) 3030, 2945, 2866, 1692, 1460, 1401, 1367, 1233, 1198, 1106, 1073, 885, 744, 698 cm⁻¹; mass spectrum (CI) *m/z* 694.3805 [C₄₂H₅₄N₂O₅Si (M+1) requires 694.3802].

Allyl (1S/R,5S/R)-6,6-bis((benzyloxy)methyl)-12-

((triisopropylsilyl)oxy)-1,3,4,5,6,7-hexahydro-2H-1,5-ethenoazocino[4,3**b]indole-2-carboxylate (25)**—Purification: hexanes/EtOAc ($9: 1 \rightarrow 7: 3$) containing 1% Et₃N to give 915 mg (80%) of 25 of an amorphous, colorless solid: ¹H NMR (600 MHz, CD₃CN, as a mixture of rotamers) 9.49-9.47 (m, 1 H), 7.41 (d, J = 7.9 Hz, 1 H), 7.38-7.21 (comp, 11 H), 7.11-710 (m, 1 H), 7.02 (dd, J = 7.6, 15.1 Hz, 1 H), 6.08-6.00 (m, 0.5 H),5.95-5.88 (m, 0.5 H), 5.81 (d, J = 8.6 Hz, 0.5 H), 5.74 (d, J = 8.4 Hz, 0.5 H), 5.67 (dd, J = 2.3, 8.3 Hz, 1 H), 5.43-5.20 (m, 1 H), 5.40 (dd, J = 1.5, 17.3 Hz, 0.5 H), 5.13 (d, J = 10.5 Hz, 0.5 H), 4.63 (d, J = 12.0 Hz, 1 H), 4.58 (d, J = 12.0 Hz, 1 H), 4.64-4.58 (m, 1 H), 4.50-4.58 (m, 1 H), 4.53 (d, J = 12.3 Hz, 1 H), 4.45 (d, J = 12.3 Hz, 1 H), 4.04 (d, J = 9.0 Hz, 1 H), 3.93 (app dt, J = 3.1, 13.9 Hz, 0.5 H), 3.88 (app dt, J = 3.1, 13.9 Hz, 0.5 H), 3.73 (d, J = 9.2 Hz, 1 H), 3.71 (d, J = 9.8 Hz, 1 H), 3.64 (d, J = 8.9 Hz, 1 H), 3.20 (dt, J = 2.3, 12.8, Hz, 0.5 H), 3.11 (dt, J = 2.3, 12.8, Hz, 0.5 H), 2.70 (dd, J = 2.8, 5.4 Hz, 1 H), 1.97-1.91 (m, 1 H), 1.87-1.79 (m, 1 H), 1.19-1.12 (comp, 3 H), 1.04-1.01 (comp, 18 H); ¹³C NMR (150 MHz, CD₃CN, as a mixture of rotamers) 159.51, 159.46, 156.0, 155.9, 139.9, 139.8, 139.5, 139.4, 135.84, 135.81, 134.8, 129.4, 129.2, 128.8, 128.6, 128.4, 127.24, 127.22, 122.4, 120.2, 117.14, 117.05 112.0, 111.9, 110.4, 110.1, 109.5, 74.2, 74.1, 73.2, 71.9, 71.8, 66.5, 66.3, 46.3, 46.0, 45.72, 45.68, 42.0, 41.8, 27.9, 27.3, 18.6, 18.5, 13.3; IR (neat) 3030, 2943, 2866, 1691, 1461, 1411, 1366, 1233, 1199, 1106, 885, 744 cm⁻¹; mass spectrum (CI) m/z720.3954 [C₄₄H₅₆N₂O₅Si (M+1) requires 720.3959].

Allyl (1S/R,5S/R)-6,6-bis((benzyloxy)methyl)-12-oxo-1,3,4,5,6,7hexahydro-2H-1,5-ethanoazocino[4,3-b]indole-2-carboxylate (26)—A solution of TMSOTf (0.9 g, 0.7 mL, 3.7 mmol) in CH₂Cl₂ (1.5 mL) was added dropwise to a solution of 19 (1.3 g, 3.6 mmol), 12 (1.5 g, 3.5 mmol), and 2,6-di(tert-butyl) pyridine (1.4 g, 1.3 mL, 7.0 mmol) in dry CH₂Cl₂ (13.5 mL) at -78 °C. The reaction was stirred for 3 h at -78 °C, whereupon a solution of TBAF·3H₂O (5.5 g, 17.5 mmol) in CH₂Cl₂ (27.0 mL) was added via cannula. The bath was removed, and the reaction was stirred at room temperature for 1 h. The reaction was quenched with saturated aqueous NaHCO₃ (30 mL), and the layers were separated. The aqueous layer was extracted with CH_2Cl_2 (2 × 30 mL), and the combined organic layers were dried (Na₂SO₄), filtered, and concentrated under reduced pressure. The resultant yellow oil was purified by flash column chromatography eluting with hexanes/ EtOAc (8 : 2 \rightarrow 7 : 3 \rightarrow 6 : 4) containing 1% Et₃N to give 1.8 g (92%) of **26** as an amorphous yellow solid: ¹H NMR (600 MHz, CD₃CN, as a mixture of rotamers) § 9.60 (s, 1 H), 7.45 (d, J = 7.81 Hz, 1 H), 7.36-7.23 (comp 9 H), 7.38 (d, J = 8.1 Hz, 1 H), 7.17-7.15 (m, 1 H), 7.16-7.12 (m, 1 H), 7.07 (app t, J = 7.9 Hz, 1 H), 6.10-5.89 (m, 1 H), 5.81-5.76 (m, 1 H), 5.40-5.15 (comp, 2 H), 4.66-4.54 (comp, 2 H), 4.62 (d, *J* = 11.0 Hz, 1 H), 4.57 (d, *J* = 11.0 Hz, 1 H), 4.38 (d, J = 11.9 Hz, 1 H), 4.31 (d, J = 11.9 Hz, 1 H), 3.98-3.80 (m, 1 H), 3.96 (d, J = 9.3 Hz, 1 H), 3.75 (d, J = 9.3 Hz, 1 H), 3.70 (d, J = 9.3 Hz, 1 H), 3.63 (d, J = 9.3 Hz, 1 H), 3.07 (dd, J = 2.6, 5.9 Hz, 1 H), 3.04 (app dd, J = 4.4, 18.8 Hz, 1 H), 3.03-2.87 (m, 1 H), 2.98-2.84 (m, 1 H), 2.13-2.01 (m, 1 H), 1.80-1.70 (m, 1 H); ¹³C NMR (150 MHz, CD₃CN, as a mixture of rotamers) 211.9, 155.6, 139.13, 139.09, 136.4, 134.6, 129.4, 129.2, 128.9, 128.8, 128.7, 128.5, 127.4, 123.0, 120.5, 117.3, 112.0, 110.4, 75.9, 74.2, 74.0, 73.0, 66.6, 58.1, 50.4, 49.6, 46.1, 45.7, 40.5, 28.8, 28.4; IR (neat) 3031, 2863, 1694, 1414, 1344, 1305, 1228, 1103, 1207, 983, 941, 861, 744, 699 cm⁻¹; mass spectrum (CI) m/z 564.2625 [C₃₅H₃₆N₂O₅ (M+1) requires 564.2725].

Allyl (1S/R,5S/R,12R/S)-6,6-bis((benzyloxy)methyl)-12-hydroxy-1,3,4,5,6,7hexahydro-2H-5,1-ethanoazocino[4,3-b]indole-2-carboxylate (28)-Lithium borohydride (4 mg, 0.18 mmol) was added to a solution of 26 (20 mg, 0.04 mmol) in MeOH (0.4 mL) at 0 °C. The reaction was stirred for 18 h at 0 °C and then partitioned between saturated brine (3 mL) and CH₂Cl₂ (3 mL). The layers were separated, and the aqueous layer was extracted (2 × 3 mL). The combined organic layers were dried (Na₂SO₄), filtered, and concentrated under reduced pressure. The resultant yellow solid was purified by flash column chromatography, eluting with hexanes/EtOAc (1:1) containing 1% Et₃N to give 16 mg (82%) of **28** as a colorless solid: mp 167–169 °C; ¹H NMR (500 MHz, DMSO-*d*₆, 120 °C) δ 10.23 (br s, 1 H), 7.40 (d, J = 8.0 Hz, 1 H), 7.36-7.31 (comp, 5 H), 7.29-7.21 (comp, 6 H), 7.07-7.04 (m, 1 H), 7.00-6.97 (m, 1 H), 6.05-5.97 (m, 1 H), 5.56 (d, *J* = 6.5 Hz, 1 H), 5.34 (d, J = 17.5 Hz, 1 H), 5.22 (d, J = 10.0 Hz, 1 H), 4.68-4.59 (comp, 4 H), 4.48 (dd, J = 10.0 Hz, 1 H), 4.68-4.59 (comp, 4 H), 4.48 (dd, J = 10.0 Hz, 1 H), 4.68-4.59 (comp, 4 H), 4.48 (dd, J = 10.0 Hz, 1 H), 4.68-4.59 (comp, 4 H), 4.48 (dd, J = 10.0 Hz, 1 H), 4.68-4.59 (comp, 4 H), 4.48 (dd, J = 10.0 Hz, 1 H), 4.68-4.59 (comp, 4 H), 4.48 (dd, J = 10.0 Hz, 1 H), 4.68-4.59 (comp, 4 H), 4.48 (dd, J = 10.0 Hz, 1 H), 4.68-4.59 (comp, 4 H), 4.48 (dd, J = 10.0 Hz, 1 H), 4.68-4.59 (comp, 4 H), 4.48 (dd, J = 10.0 Hz, 1 H), 4.68-4.59 (comp, 4 H), 4.48 (dd, J = 10.0 Hz, 1 H), 4.68-4.59 (comp, 4 H), 4.48 (dd, J = 10.0 Hz, 1 H), 4.68-4.59 (comp, 4 H), 4.48 (dd, J = 10.0 Hz, 1 H), 4.68-4.59 (comp, 4 H), 4.48 (dd, J = 10.0 Hz, 1 H), 4.68-4.59 (comp, 4 H), 4.48 (dd, J = 10.0 Hz, 1 H), 4.68-4.59 (comp, 4 H), 4.48 (dd, J = 10.0 Hz, 1 H), 4.68-4.59 (comp, 4 H), 4.48 (dd, J = 10.0 Hz, 1 H), 4.68-4.59 (comp, 4 H), 4.48 (dd, J = 10.0 Hz, 1 H), 4.68-4.59 (comp, 4 H), 4.48 (dd, J = 10.0 Hz, 1 H), 4.68-4.59 (comp, 4 H), 4.48 (dd, J = 10.0 Hz, 1 Hz), 4.58 (dd, J = 10.0 Hz), 4.58 (dd, J = 114.5, 13.0 Hz, 2 H), 4.41 (br s, 1 H), 4.29 (d, J = 9.0 Hz, 1 H), 4.26-4.23 (m, 1 H), 4.08 (d, J = 10.0 Hz, 1 H), 4.02 (d, J = 10.0 Hz, 1 H), 3.76-3.73 (comp, 2 H), 2.97-2.91 (m, 1 H), 2.68-2.63 (comp, 2 H), 1.93 (app dt, *J* = 14.5, 3.5 Hz, 1 H), 1.85 (dd, *J* = 13.5, 9.5 Hz, 1 H), 1.76-1.68 (m, 1 H); ¹³C NMR (125 MHz, DMSO-*d*₆, 120 °C) δ 154.0, 138.9, 138.2, 138.0, 134.9, 133.3, 127.5, 127.4, 126.74, 126.7, 126.6, 126.4, 125.5, 120.2, 118.2, 116.0, 115.97, 110.8, 110.5, 73.3, 73.2, 72.4, 72.2, 71.8, 64.4, 46.7, 46.4, 45.4, 39.9, 39.8, 29.4; IR (neat)

3408, 2943, 1680, 1454, 1413, 1104, 739 cm⁻¹; mass spectrum (ESI) m/z 566.2823 [C₃₅H₃₉N₂O₅ (M+1) requires 566.2853].

(1S/R,5S/R,12R/S)-6,6-bis((Benzyloxy)methyl)-2,3,4,5,6,7-hexahydro-1*H*-5,1ethanoazocino[4,3-*b*]indol-12-ol (29)—A solution of

tetrakis(triphenylphosphine)palladium(0) (92 mg, 0.08 mmol) in CH₂Cl₂ (2.0 mL) was added dropwise to a solution of alcohol 28 (0.9 g, 1.59 mmol) and PhSiH₃ (688 mg, 0.8 mL, 6.36 mmol) in CH₂Cl₂ (7.4 mL) at room temperature, and the reaction was stirred for 0.5 h. Saturated aqueous NaHCO3 (20 mL) was added, and the layers were separated. The aqueous layer was extracted with CH_2Cl_2 (2 × 25 mL). The combined organic layers were dried (Na_2SO_4) , filtered, and concentrated under reduced pressure. The resultant brown gum was purified by flash column chromatography eluting with CH₂Cl₂/MeOH (75:25) containing 1% Et₃N to give 0.56 g (73%) of **29** as a tan amorphous solid foam:¹H NMR (600 MHz, CD₂Cl₂) δ 9.40 (br s, 1 H), 7.65 (d, J = 7.8 Hz, 1 H), 7.42-7.22 (comp, 11 H), 7.13 (ddd, J = 7.2, 7.2, 1.2 Hz, 1 H), 7.09 (ddd, J = 8.4, 8.4, 1.2 Hz, 1 H), 4.96 (d, J = 6.0 Hz, 1 H), 4.71-4.60 (comp, 3 H), 4.52-4.48 (m, 1 H), 4.09-4.00 (comp, 3 H), 3.93 (d, J = 9.0 Hz, 1 H), 3.71 (d, J = 9.0 Hz, 1 H), 3.05 (ddd, J = 14.4, 8.4, 6.6 Hz, 1 H), 2.87 (dd, J = 13.8, 4.8 Hz, 1 H), 2.66-2.55 (comp, 2 H), 2.07 (dd, J = 13.8, 9.0 Hz, 1 H), 2.01-1.85 (comp, 2 H); ${}^{13}C$ NMR (150 Hz, CD₂Cl₂) & 140.0, 138.42, 138.38, 135.5, 128.97, 128.94, 128.7, 128.3, 128.2, 128.1, 126.5, 122.2, 119.9, 118.5, 111.0, 110.3, 74.7, 74.2, 74.0, 72.8, 64.5, 47.8, 46.5, 41.8, 40.6, 31.0, 29.0; mass spectrum m/z (ESI) 483.2642 [C₃₁H₃₅N₂O₃ (M+1) requires 483.2642].

Allyl (1S/R,5S/R,12R/S)-6,6-bis((benzyloxy)methyl)-12-

((trimethylsilyl)oxy)-1,3,4,5,6,7-hexahydro-2H-5,1-ethanoazocino[4,3blindole-2-carboxylate—TMSCI (2.2 mL, 1.8 g, 17.4 mmol) was added to a solution of 28 (3.3 g, 5.8 mmol) and imidazole (2.0 g, 29.0 mmol) in anhydrous DMF (62 mL), and the reaction was stirred 2 h. The mixture was partitioned between saturated aqueous NaHCO₃ (100 mL) and Et₂O (150 mL), and the layers were separated. The aqueous layer was extracted with Et₂O (2×100 mL), and the combined organic layers were washed with brine $(1 \times 100 \text{ mL})$. The organic layer was dried (MgSO₄), filtered, and concentrated under reduced pressure. The resultant clear oil was purified by flash column chromatography eluting with hexanes/EtOAc (9:1) containing 1% Et₃N to provide 3.6 g (97%) of the trimethylsilyl protected alcohol as a colorless amorphous solid foam: mp 115-116 °C (colorless crystals from *i*-PrOH/H₂O); ¹H NMR (500 MHz, DMSO-*d*6, 120 °C) & 10.23 (br s, 1 H), 7.39 (d, J = 8.0 Hz, 1H), 7.32-7.29 (comp, 5 H), 7.27-7.22 (comp, 6 H), 7.05 (t, J = 8.0 Hz, 1 H), 6.97 (t, J = 8.0 Hz, 1 H), 6.04-5.97 (m, 1 H), 5.51 (d, J = 6.5 Hz, 1 H), 5.33 (d, J = 17.0 Hz, 1 H), 5.21 (d, J = 10.5 Hz, 1 H), 4.67-4.58 (comp, 4 H), 4.48 (d, J = 12.5 Hz, 1 H), 4.45 (d, J = 12.5 Hz, 1 H), 4.32-4.27 (comp, 2 H), 3.98 (d, J = 9.5 Hz, 1 H), 3.92 (d, J = 9.5 Hz, 1 H), 3.75-3.71 (comp, 2 H), 2.93-2.85 (m, 1 H), 2.67-2.61 (comp, 2 H), 1.92 (ddd, J

= 14.5, 3.5, 3.5 Hz, 1 H), 1.86 (dd, *J* = 13.5, 9.0 Hz, 1 H), 1.75-1.67 (m, 1 H), 0.05 (s, 9 H); ¹³C NMR (125 MHz, DMSO-*d6*, 120 °C) δ 154.0, 138.8, 138.1, 138.0, 134.9, 133.2, 127.4, 127.3, 126.6, 126.6, 126.5, 126.4, 125.3, 120.2, 118.1, 116.0, 116.0 110.6, 110.5, 74.8, 72.5, 72.3, 72.1, 71.5, 64.4, 46.7, 46.2, 45.5, 40.5 39.7, 29.1, -0.8; IR (neat) 3030,

2951, 2871, 1693, 1454, 1412, 1251, 1072, 873, 842, 744, 698 cm⁻¹; mass spectrum (ESI) *m*/*z* 661.3066 [C₃₈H₄₆N₂NaO₅Si (M+1) requires 661.3068].

2-Allyl 7-(*tert*-butyl) (1S/R,5S/R,12R/S)-6,6-bis((benzyloxy)methyl)-12-((trimethylsilyl)oxy)-3,4,5,6-tetrahydro-1*H*-5,1-ethanoazocino[4,3-*b*]indole-2,7-

dicarboxylate—DMAP (0.2 g, 1.64 mmol) was added to a solution of the trimethylsilyl protected alcohol (3.5 g, 5.5 mmol) from the previous step in Boc₂O (16 mL) at 100 °C, and the reaction was stirred for 30 min at 100 °C. The mixture was partitioned between H₂O (100 mL) and Et₂O (150 mL), and the layers were separated. The aqueous layer was extracted with Et₂O (2×150 mL), and the combined organic layers were dried (MgSO₄), filtered, and concentrated under reduced pressure. The resultant brown oil was purified by flash column chromatography eluting with hexanes/EtOAc $(1: 0 \rightarrow 95: 5 \rightarrow 9: 1)$ to provide 3.8 g (93%) of the Boc protected indole as a colorless amorphous solid foam: mp 139-140 °C (colorless crystals from i-PrOH/H₂O); ¹H NMR (400 MHz, as a mixture of rotamers) δ 7.72 (dd, J = 8.4, 2.4 Hz, 1 H), 7.45 (d, J = 7.6 Hz, 0.5 H), 7.36-7.27 (comp, 4 H), 7.24-7.19 (comp, 2.5 H), 7.16-7.08 (comp, 4 H), 7.00-6.96 (comp, 2 H), 6.06-5.96 (m, 0.5 H), 5.93-5.84 (m, 0.5 H), 5.66 (d, *J* = 6.8 Hz, 0.5 H), 5.54 (d, *J* = 6.8 Hz, 0.5 H), 5.37 (dd, J = 17.2, 1.6 Hz, 0.5 H), 5.26-5.20 (m, 1 H), 5.13 (dd, J = 10.4, 1.2, 0.5 H), 4.77-4.72 (comp, 1.5 H), 4.67-4.49 (comp, 3.5 H), 4.45-4.41 (comp, 2 H), 4.35-4.32 (m, 1 H), 4.28-4.15 (comp, 2 H), 3.80 (dd, J = 14.4, 6.4 Hz, 0.5 H), 3.58 (dd, J = 14.4, 6.4 Hz, 0.5 H), 3.58 (d, J = 10.0 Hz, 1 H), 2.94-2.87 (comp, 1.5 H), 2.83-2.76 (m, 0.5 H), 2.69-2.57 (m, 1 H), 2.08-1.99 (m, 1 H), 1.91-1.65 (comp, 2 H), 1.50 (s, 9 H); 0.01 (s, 9 H); ¹³C NMR (100 MHz, as a mixture of rotamers) & 155.4, 155.3, 151.0, 139.5, 138.9, 138.8, 138.2, 138.1, 136.3, 136.2, 133.31, 133.28, 127.2, 127.1, 126.9, 126.8, 126.7, 126.4, 126.3, 124.1, 121.74, 121.71, 121.3, 121.1, 118.0, 117.6, 117.4, 116.9, 114.1, 114.0, 83.7, 83.6, 78.0, 77.8, 77.2, 75.5, 75.4, 72.5, 72.4, 72.3, 72.0, 65.9, 65.8, 52.7, 52.6, 49.5, 47.0, 46.8, 41.1, 41.0, 39.8, 38.8, 31.5, 30.7, 28.0, 27.4, -0.08; IR (neat) 2952, 1739, 1696, 1456, 1316, 1250, 1112, 845, 737 cm⁻¹; mass spectrum (ESI) m/z 739.3765 [C₄₃H₅₅N₂O₇Si (M+1) requires 739.3773].

2-Allyl 7-(*tert*-butyl) (1S/R,5S/R,12R/S)-6,6-bis((benzyloxy)methyl)-12hydroxy-3,4,5,6-tetrahydro-1*H*-5,1-ethanoazocino[4,3-*b*]indole-2,7-

dicarboxylate (35)—TBAF·3H₂O (289 mg, 0.915 mmol) was added to a solution of the Boc protected indole (135 mg, 0.183 mmol) from the previous step in CH₂Cl₂ (1.8 mL), and the reaction was stirred for 2 h. The mixture was partitioned between saturated aqueous NaHCO₃ (10 mL) and CH₂Cl₂ (10 mL), and the layers were separated. The aqueous layer was extracted with CH₂Cl₂ (2×10 mL), and the combined organic layers were dried (Na₂SO₄), filtered, and concentrated under reduced pressure. The resultant yellow gum was purified by flash column chromatography eluting with hexanes/EtOAc (70 : 30) containing 1% Et₃N to provide 118 mg (97%) of **35** as a colorless amorphous solid foam: mp 182–183 °C (colorless crystals from *i*-PrOH/H₂O); ¹H NMR (400 MHz, as a mixture of rotamers) δ 7.75 (dd, *J* = 8.4, 4.0 Hz, 1 H), 7.50 (d, *J* = 8.4 Hz, 0.5 H), 7.39 (d, *J* = 7.6 Hz, 0.5 H), 7.31-7.12 (comp, 12 H), 6.09-5.99 (m, 0.5 H), 5.95-5.87 (m, 0.5 H), 5.74 (d, *J* = 6.4 Hz, 0.5 H), 5.43-5.15 (comp, 2 H), 4.73-4.70 (m, 1 H), 4.63-4.25 (comp, 9 H), 4.12 (d, *J* = 9.2 Hz, 0.5 H), 4.05 (d, *J* = 9.2 Hz, 0.5 H), 3.88-3.66 (comp, 2 H), 3.09-2.95 (m, 1 H), 2.84 (br s, 1 H), 2.61-2.48 (m, 1 H), 2.14-2.05 (m, 1 H), 1.97-1.73

(comp, 2 H), 1.58 (s, 9 H); ¹³C NMR (100 MHz, as a mixture of rotamers) δ 155.2, 155.1, 151.4, 151.3, 137.8, 137.75, 137.66, 137.6, 137.4, 137.3, 136.4, 133.2, 133.1, 128.4, 128.2, 128.1, 128.0, 127.8, 127.5, 127.4, 127.3, 127.2, 126.8, 126.6, 124.5, 122.2, 122.1, 118.3, 117.9, 117.4, 117.1, 113.9, 84.2, 84.1, 77.2, 76.2, 75.4, 75.1, 74.8, 74.7, 73.54, 73.48, 72.9, 72.8, 66.1, 65.9, 53.7, 53.4, 49.5, 49.4, 47.1, 47.0, 40.7, 40.6, 39.2, 38.2, 31.4, 30.8, 28.0; IR (neat) 2941, 1739, 1693, 1456, 1316, 1112, 738 cm⁻¹; mass spectrum (ESI) *m/z* 667.3374 [C₄₀H₄₇N₂O₇ (M+1) requires 667.3378].

tert-Butyl (1S/R,5S/R,12R/S)-6,6-bis((benzyloxy)methyl)-12-hydroxy-1,2,3,4,5,6hexahydro-7H-5,1-ethanoazocino[4,3-b]indole-7-carboxylate—A solution of tetrakis-(triphenylphosphine)palladium(0) (10.4 mg, 0.009 mmol) in CH₂Cl₂ (0.1 mL) was added to a solution of **35** (118 mg, 0.177 mmol) and PhSiH₃ (0.44 mL, 383 mg, 3.54 mmol) in CH₂Cl₂ (1.7 mL), and the reaction was stirred for 0.5 h at room temperature. The mixture was partitioned between saturated aqueous NaHCO₃ (10 mL) and CH₂Cl₂ (10 mL), and the layers were separated. The aqueous layer was extracted with CH_2Cl_2 (2 × 10 mL), and the combined organic layers were dried (Na₂SO₄), filtered, and concentrated under reduced pressure. The resultant yellow oil was purified by flash column chromatography eluting with EtOAc/MeOH (85:15) containing 1% Et₃N to provide 75 mg (73%) of the secondary amine as a tan amorphous solid foam: ¹H NMR (400 MHz) δ 7.76 (d, J = 8.0 Hz, 1 H), 7.46 (d, *J* = 7.2 Hz, 1 H), 7.30-7.20 (comp, 10 H), 7.17-7.14 (comp, 2 H), 4.64-4.60 (comp, 2 H), 4.56 (s, 2 H), 4.51-4.46 (comp, 3 H), 4.42 (d, *J* = 12.4 Hz, 1 H), 4.05 (d, *J* = 9.2 Hz, 1 H), 3.74 (d, J = 9.2 Hz, 1 H), 2.86-2.78 (comp, 2 H), 2.58 (dd, J = 12.8, 5.2 Hz, 1 H), 2.36 (ddd, *J* = 12.8, 12.8, 3.6 Hz, 1 H), 2.13 (dd, *J* = 13.2, 9.2 Hz, 1 H), 1.97-1.90 (m, 1 H), 1.80-1.71 (m, 1 H), 1.58 (s, 9 H); ¹³C NMR (100 MHz) & 151.6, 138.0, 137.5, 137.3, 136.8, 128.3, 128.2, 127.9, 127.7, 127.2, 126.8, 124.1, 122.8, 121.8, 117.7, 114.0, 83.8, 77.2, 76.3, 75.6, 75.0, 73.5, 72.9, 53.4, 49.5, 47.5, 42.3, 41.7, 31.8, 28.0; IR (neat) 2931, 1736, 1456, 1316, 1135, 738 cm⁻¹; mass spectrum (ESI) m/z 583.3170 [C₃₆H₄₃N₂O₅ (M+1) requires 583.3166].

tert-Butyl (1S/R,5S/R,12R/S)-6,6-bis((benzyloxy)methyl)-2-(2-chloroethyl)-12hydr-oxy-1,2,3,4,5,6-hexahydro-7*H*-5,1-ethanoazocino[4,3-*b*]indole-7-

carboxylate (36)—Sodium triacetoxyborohydride (15 mg, 0.068 mmol) was added to a solution of the secondary amine (20 mg, 0.034 mmol) from the previous step and chloroacetaldehyde (11 µL of a 50% by weight solution in H₂O, 0.068 mmol) in 1,2-dichloroethane (0.4 mL) at 0 °C, and the reaction was stirred for 0.5 h at 0 °C. The mixture was partitioned between saturated aqueous NaHCO₃ (3 mL) and CH₂Cl₂ (3 mL), and the layers were separated. The aqueous layer was extracted with CH₂Cl₂ (2×3 mL), and the combined organic layers were dried (Na₂SO₄), filtered, and concentrated under reduced pressure. The resultant yellow oil was purified by flash column chromatography eluting with hexanes/EtOAc (60 : 40) to provide 17 mg (77%) of **36** as a yellow gum: ¹H NMR (400 MHz, CD₃CN) δ 7.69 (dd, *J* = 8.0, 0.4 Hz, 1 H), 7.48 (d, *J* = 8.0 Hz, 1 H), 7.28-7.12 (comp, 12 H), 4.48-4.28 (comp, 8 H), 4.20 (d, *J* = 9.6 Hz, 1 H), 3.69 (d, *J* = 6.4 Hz, 1 H), 3.64-3.59 (comp, 2 H), 3.10-3.04 (m, 1 H), 2.79-2.72 (m, 1 H), 2.70-2.62 (comp, 2 H), 2.38-2.23 (comp, 2 H), 2.15 (br s, 1 H), 1.89-1.85 (m, 1 H), 1.79-1.71 (m, 1 H), 1.56 (s, 9 H); ¹³C NMR (400 MHz, CD₃CN) δ 152.6, 139.3, 138.4, 137.3, 129.3, 129.2, 129.0, 128.7, 128.5,

128.2, 128.1, 124.9, 122.6, 122.4, 119.5, 114.6, 85.3, 77.9, 75.6, 73.6, 73.5, 60.0, 54.4, 54.3, 50.2, 49.4, 44.4, 44.3, 42.0, 32.2, 31.7, 28.2; IR (neat) 3450, 2934, 1737, 1456, 1315, 1142, 738 cm⁻¹; mass spectrum (ESI) *m*/*z* 645.3081 [C₃₈H_{46 35}ClN₂O₅ (M+1) requires 645.3090].

tert-Butyl (1S/R,5S/R)-6,6-bis((benzyloxy)methyl)-2-(2-chloroethyl)-12oxo-1,2,3,4,5,6-hexahydro-7*H*-1,5-ethanoazocino[4,3-*b*]indole-7-carboxylate

(37)—Anhydrous DMSO (56 µL, 62 mg, 0.792 mmol) was added to a solution of oxalyl chloride (35 µL, 50 mg, 0.397 mmol) in CH₂Cl₂ (1.1 mL) at -78 °C. The reaction was stirred for 10 min at -78 °C, whereupon a solution of 36 (128 mg, 0.198 mmol) in CH₂Cl₂ (1.1 mL) was added, and the reaction was stirred an additional 15 min at -78 °C. Et₃N (220 μ L, 160 mg, 1.58 mmol) was added, and the reaction was stirred for 15 min at -78 °C. The bath was removed, and the reaction was stirred for 1 h at room temperature. The mixture was partitioned between H₂O (15 mL) and CH₂Cl₂ (15 mL), and the layers were separated. The aqueous layer was extracted with CH_2Cl_2 (2 × 15 mL), and the combined organic layers were dried (Na_2SO_4) , filtered, and concentrated under reduced pressure. The resultant yellow oil was purified by flash column chromatography eluting with hexanes/EtOAc (8:2) to provide 109 mg (86%) of **37** as a yellow amorphous solid foam: ¹H NMR (400 MHz, CD_2Cl_2 δ 7.78 (d, J = 8.4 Hz, 1 H), 7.53 (d, J = 7.6 Hz, 1 H), 7.35-7.20 (comp, 12 H), 4.56-4.52 (comp, 2 H), 4.46-4.43 (comp, 2 H), 4.40 (d, *J* = 11.2 Hz, 1 H), 4.30 (d, *J* = 11.2 Hz, 1 H), 4.03 (d, J = 9.2 Hz, 1 H), 3.83-3.78 (comp, 2 H), 3.63-3.59 (comp, 2 H), 3.24 (dd, *J* = 5.6, 3.2 Hz, 1 H), 3.13 (ddd, *J* = 13.2, 6.8, 6.8 Hz, 1 H), 3.06-3.94 (comp, 2 H), 2.81-2.74 (m, 1 H), 2.66 (ddd, J = 12.8, 12.8, 2.8 Hz, 1 H), 2.51-2.45 (m, 1 H), 2.02-1.96 (m, 1 H), 1.84-1.76 (m, 1 H), 1.66 (s, 9 H); ¹³C NMR (100 MHz, CD₂Cl₂) δ 212.2, 151.9, 138.6, 138.3, 137.0, 136.9, 129.0, 128.7, 128.5, 128.3, 127.9, 127.8, 127.7, 124.5, 122.4, 118.9, 118.8, 114.4, 84.9, 76.2, 73.7, 73.6, 73.2, 63.0, 60.0, 53.0, 49.2, 49.0, 48.2, 43.3, 28.2, 28.0; IR (neat) 2978, 2934, 2859, 1739, 1699, 1455, 1312, 1145, 1097, 740, 699 cm⁻¹; mass spectrum (ESI) m/z 643.2931 [C₃₈H_{44 35}ClN₂O₅ (M+1) requires 643.2933].

tert-Butyl (1R/S,4R/S,7S/R,13*c*S)-8,8-bis((benzyloxy)-methyl)-14oxo-1,2,3,5,6,7,8,13*c*-octahydro-9*H*-1,7-methanopyrrolo[1',2':1,2]azocino[4,3tilin data 0, each survivate (22) Math ad 0, a fill a survivation of 1, (21) and (21) a

b]indole-9-carboxylate (38) Method A—Sodium *tert*-butoxide (81 mg, 0.847 mmol) was added to a solution of **37** (109 mg, 0.169 mmol) in THF/*t*-BuOH (4.2 mL/1.4 mL) at 0 °C. The bath was removed, and the reaction was stirred for 1 h at room temperature. The mixture was partitioned between saturated aqueous NH₄Cl (15 mL) and Et₂O (15 mL), and the layers were separated. The aqueous layer was extracted with Et₂O (2 × 15 mL), and the combined organic layers were dried (MgSO₄), filtered, and concentrated under reduced pressure. The resultant yellow oil was purified by flash column chromatography eluting with CH₂Cl₂/MeOH (95 : 5) to provide 96 mg (94%) of **38** as a pale yellow amorphous solid foam: ¹H NMR (400 MHz) δ 7.82 (d, *J* = 7.2 Hz, 1 H), 7.77 (dd, *J* = 7.6, 1.2 Hz, 1 H), 7.37-7.22 (comp, 10 H), 7.18-7.16 (comp, 2 H), 4.73 (d, *J* = 4.8 Hz, 1 H), 4.53 (d, *J* = 12.4 Hz, 1 H), 4.45- 4.39 (comp, 2 H), 4.37-4.32 (comp, 2 H), 4.14 (d, *J* = 9.6 Hz, 1 H), 3.95 (d, *J* = 5.2 Hz, 1 H), 3.78 (d, *J* = 9.6 Hz, 1 H), 3.67-3.60 (m, 1 H), 3.29-3.28 (m, 1 H), 3.23 (dd, *J* = 7.2, 5.2 Hz, 1 H), 3.19-3.11 (comp, 2 H), 2.67 (dd, *J* = 14.4, 3.2 Hz, 1 H), 2.45-2.29 (comp, 2 H), 2.01-1.93 (m, 1 H), 1.88-1.80 (m, 1 H), 1.64 (s, 9 H); ¹³C NMR (100 MHz) δ 214.2, 151.0, 137.8, 137.7, 137.2, 136.2, 128.4, 128.1, 127.9, 127.6, 127.4, 127.1, 127.0,

124.4, 122.3, 118.6, 115.0, 113.7, 84.3, 74.4, 73.0, 72.6, 72.5, 61.6, 58.0, 54.3, 49.2, 49.1, 47.5, 32.3, 27.8, 23.8; IR (neat) 2929, 2873, 1740, 1695, 1314, 1147, 742 cm⁻¹; mass spectrum (ESI) m/z 607.3169 [C₃₈H₄₃N₂O₅ (M+1) requires 607.3166].

(1R/S,4R/S,7S/R,8R/S,13cS/R,16S/R)-8-(Hydroxymethyl)-2,3,5,6,7,8,9,13coctahydro-1*H*-1,7,8-(epimethanetriyloxy-methano) pyrrolo[1',2':

1,2]azocino[4,3-b]indol-16-ol hydrochloride (2)—A solution of 38 (1.0 g, 1.65 mmol) in a mixture of MeOH (23 mL) and 5 M HCl_(aq) (23 mL) was stirred at 65 °C for 1 h. The reaction was cooled to room temperature, palladium on carbon (250 mg of 10% loading) was added, and the reaction was stirred under a balloon of H₂ for 4 h. The reaction was filtered through a plug of cotton and concentrated under reduced pressure. The resultant yellow residue was purified by reverse phase chromatography eluting with MeOH/H₂O (0 : $100 \rightarrow 20$: 80 over 15 mins) containing 0.1% TFA to give 513 mg (86%) of 2 as a pale yellow amorphous solid foam: ¹H NMR (600 MHz, CD₃OD) δ 7.61 (d, J = 8.0 Hz, 1 H), 7.44 (d, J = 8.0 Hz, 1 H), 7.16 (ddd, J = 8.0, 7.0, 1.0 Hz, 1 H), 7.11 (ddd, J = 8.0, 7.0, 1.0 Hz, 1 H), 5.45 (d, *J* = 7.4 Hz, 1 H), 4.17 (d, *J* = 11.3 Hz, 1 H), 4.13 (d, *J* = 11.3 Hz, 1 H), 3.97 (d, J = 7.8 Hz, 1 H), 3.70 (ddd, J = 12.4, 12.4, 8.0 Hz, 1 H), 3.60-3.55 (m, 1 H), 3.56 (d, *J* = 7.8 Hz, 1 H), 3.21-3.14 (comp, 2 H), 2.95 (ddd, *J* = 13.7, 3.2, 3.2 Hz, 1 H), 2.74 (dddd, J = 12.8, 10.3, 8.0, 2.4 Hz, 1 H), 2.64-2.63 (m, 1 H), 2.53-2.46 (m, 1 H), 2.40-2.33 (m, 1 H), 2.23 (dddd, J = 12.2, 9.0, 3.0, 3.0 Hz, 1 H); ¹³C NMR (150 MHz, CD₃OD) δ 143.1, 137.2, 128.4, 123.3, 121.1, 118.0, 112.5, 108.4, 102.5, 75.3, 64.3, 62.8, 55.3, 53.4, 53.1, 50.5, 47.3, 26.2, 19.7; IR (neat) 3395, 3285, 1441, 1233, 1086, 997, 738, 721 cm⁻¹; mass spectrum (ESI) m/z 327.1704 [C19H23N2O3 (M+1) requires 327.1703].

(±)-Actinophyllic acid hydrochloride (1)—A solution of IBX (115 mg, 0.41 mmol) and alcohol 2 (30 mg, 0.08 mmol) in DMSO (0.33 mL) was stirred at 50 °C for 2 hours. Nhydroxysuccinimide (67 mg, 0.58 mmol) was added, and the reaction was stirred at 50 °C for 2 h. The reaction was cooled to room temperature, whereupon a solution of 1 M NaOH was added until the reaction was basic (~pH 10) according to pH paper. The reaction mixture was dry-loaded onto SiO₂ and purified by flash column chromatography eluting with $CHCl_3/MeOH/NH_4OH$ (6 : 3 : 0.5), followed by reverse phase chromatography eluting with MeOH/H₂O (0 : 100 \rightarrow 20 : 80 over 15 mins) to give 9.8 mg (31%) of **1** as a colorless amorphous solid: ¹H NMR (600 MHz, D₂O, internal reference to MeOH at 3.34 ppm) & 7.64 (d, J = 8.0 Hz, 1 H), 7.54 (d, J = 8.1 Hz, 1 H), 7.33-7.30 (m, 1 H), 7.25-7.22 (m, 1 H), 5.52 (d, J = 7.5 Hz, 1 H), 4.40 (d, J = 8.2 Hz, 1 H), 3.79 (d, J = 8.2 Hz, 1 H), 3.80-3.74 (m, 1 H), 3.64-3.60 (m, 1 H), 3.30-3.25 (comp, 2 H), 3.09-3.05 (comp, 2 H), 2.70-2.63 (m, 1 H), 2.61-2.54 (m, 1 H), 2.45-2.40 (m, 1 H), 2.37-2.33 (m, 1 H); ¹³C NMR (150 MHz, D₂O, internal reference to MeOH at 49.5 ppm) δ 173.8, 138.5, 136.1, 127.2, 123.7, 121.2, 117.8, 112.4, 107.7, 102.3, 74.6, 61.2, 58.3, 54.4, 51.8, 50.2, 46.2, 25.3, 18.7; IR (neat) 3321, 1589, 1460, 1372, 1108, 1035, 738 cm⁻¹; mass spectrum (ESI) m/z 341.1497 [C₁₉H₂₁N₂O₄ (M+1) requires 341.1496].

2-Allyl 7-(*tert*-butyl) (1S/R,5S/R)-6,6-bis((benzyloxy)methyl)-12-oxo-3,4,5,6tetrahydro-1*H*-1,5-ethanoazocino[4,3-*b*]indole-2,7-dicarboxylate (40)—4-

Dimethylaminopyridine (1.5 g, 12.3 mmol) was added to a solution of 26 (4.6 g, 8.2 mmol)

in a mixture of di-tert-butyl dicarbonate (23 mL, 22 g, 102.5 mmol) and toluene (18 mL), and the reaction was stirred for 3 h at room temperature. The reaction was partitioned between H_2O (100 mL) and Et_2O (100 mL). The layers were separated, and the aqueous layer was extracted with Et_2O (2 × 100 mL). The combined organic layers were dried $(MgSO_4)$, filtered, and concentrated under reduced pressure. The resultant yellow oil was purified by flash column chromatography eluting with hexanes/EtOAc (8 : 2) to give 4.6 g (83%) of 40 as a yellow amorphous solid: ¹H NMR (400 MHz, as a mixture of rotamers) δ 7.80 (d, J = 8.0 Hz, 1 H), 7.58 (d, J = 7.6 Hz, 0.5 H), 7.43 (d, J = 8.4 Hz, 0.5 H), 7.34-7.15 (comp, 12 H), 6.09-5.76 (comp, 2 H), 5.44-5.18 (comp, 2 H), 4.75-4.34 (comp, 7 H), 4.06-3.93 (comp, 1.5 H), 3.83-3.71 (comp, 2.5 H), 3.33-3.28 (m, 1 H), 3.17-3.03 (comp, 2 H), 2.95-2.78 (m, 1 H), 2.07-1.95 (m, 1 H), 1.90-1.75 (m, 1 H), 1.64 (s, 9 H); ¹³C NMR (100 MHz, as a mixture of rotamers) & 211.7, 211.5, 154.8, 154.5, 151.1, 137.7, 137.5, 137.3, 137.2, 136.3, 132.9, 132.8, 128.2, 128.0, 127.9, 127.6, 127.5, 127.4, 127.1, 127.0, 124.6, 122.3, 119.4, 119.2, 118.3, 117.7, 117.3, 114.1, 114.0, 84.4, 84.3, 75.5, 75.2, 73.2, 73.1, 73.0, 72.6, 66.3, 66.1, 63.3, 63.1, 48.8, 47.5, 46.7, 45.0, 39.7, 28.8, 28.3, 27.9; IR (neat) 2979, 2936, 2862, 1784, 1698, 1456, 1306, 1145, 1111, 767, 741, 700 cm⁻¹; mass spectrum (ESI) *m/z* 665.3215 [C₄₀H₄₅N₂O₇ (M+1) requires 665.3221].

tert-Butyl (1S/R,5S/S)-6,6-bis((benzyloxy)methyl)-12-oxo-1,2,3,4,5,6-

hexahydro-7H-1,5-ethanoazocino[4,3-b]indole-7-carboxylate (41)—A solution of Pd₂(dba)₃ (8.5 mg, 0.009 mmol) and 1,4-bis(diphenylphosphino)butane (dppb) (3.9 mg, 0.009 mmol) in THF (3.1 mL) was stirred for 5 min at room temperature, and the resultant solution was transferred to a solution of 40 (614 mg, 0.92 mmol) and N,Ndimethylbarbituric acid (NMDBA) (1.44 g, 9.24 mmol) in THF (6.2 mL). The reaction was stirred for 1 h at room temperature, whereupon Et₂O (20 mL) and saturated aqueous NaHCO₃ (30 mL) were slowly added. The layers were separated, and the aqueous layer was extracted with Et₂O (2×15 mL). The combined organic layers were dried (MgSO₄), filtered, and concentrated under reduced pressure. The resultant yellow oil was purified by flash column chromatography eluting with EtOAc containing 1% Et₃N to provide 495 mg (92%) of **41** as a pale yellow amorphous solid: ¹H NMR (400 MHz) δ 7.82 (d, J = 8.8 Hz, 1 H), 7.49 (d, J = 7.6 Hz, 1 H), 7.35-7.20 (comp, 12 H), 4.67-4.65 (m, 1 H), 4.60 (d, J = 8.8Hz, 1 H), 4.53 (d, J = 12.0 Hz, 1 H), 4.43 (d, J = 11.6 Hz, 1 H), 4.35 (d, J = 12.0 Hz, 1 H), 3.97 (d, J = 8.8 Hz, 1 H), 3.85- 3.81 (comp, 2 H), 3.37-3.34 (m, 1 H), 3.18 (dd, J = 2.4, 18.0 Hz, 1 H), 2.97 (dd, J = 4.4, 18.0 Hz, 1 H), 2.72-2.65 (comp, 2 H), 2.07-1.97 (comp, 2 H), 1.80-1.73 (m, 1 H), 1.64 (s, 9 H); ¹³C NMR (100 MHz) & 213.2, 151.7, 138.4, 138.0, 137.1, 136.9, 128.6, 128.4, 128.2, 128.1, 127.8, 127.6, 127.4, 124.6, 122.4, 120.6, 117.9, 114.5, 84.5, 75.9, 73.5, 73.4, 73.0, 62.9, 50.3, 49.2, 46.0, 41.4, 29.0, 28.3; IR (neat) 2920, 2857, 1737, 1698, 1456, 1311, 1143, 1098, 740, 699 cm⁻¹; mass spectrum (ESI) m/z 581.3012 [C₃₆H₄₁N₂O₅ (M+1) requires 581.3010].

tert-Butyl (1R/S,4R/S,7S/R,13*c*S)-8,8-bis((benzyloxy)methyl)-14oxo-1,2,3,5,6,7,8,13*c*-octahydro- 9*H*-1,7-methanopyrrolo[1',2':1,2]azocino[4,3*b*]indole-9-carboxylate (38) Method B—Sodium triacetoxyborohydride (0.69 g, 3.27 mmol) was added to a mixture of 41 (0.95 g, 1.64 mmol) and chloroacetaldehyde (0.51 mL of a 50% by weight solution in H₂O, 3.27 mmol) in 1,2-dichloroethane (18 mL) at 0 °C, and

the reaction was stirred for 15 min at 0 °C. The reaction was partitioned between saturated aqueous NaHCO³ (20 mL) and CH₂Cl₂ (10 mL), and the layers were separated. The aqueous layer was extracted with CH₂Cl₂ (2 × 20 mL), and the combined organic layers were dried (Na₂SO₄), filtered, and concentrated under reduced pressure to give 1.16 g of yellow amorphous solid. The crude material was dissolved in a mixture of THF (38 mL) and *t*-BuOH (11.7 mL) and cooled to 0 °C. Sodium *tert*-butoxide (0.79 g, 8.20 mmol) was added at 0 °C, the bath was removed, and the reaction was stirred for 1.5 h at room temperature. The reaction was partitioned between saturated aqueous NH₄Cl (30 mL) and Et₂O (30 mL), and the layers were separated. The aqueous layer was extracted with Et₂O (2 × 30 mL), and the combined organic layers were dried (MgSO₄), filtered, and concentrated under reduced pressure. The resultant yellow oil was purified by flash column chromatography eluting with CH₂Cl₂/MeOH (95 : 5) to provide 820 mg (83% over the two steps) of **38** as a pale yellow amorphous solid. The spectra for **38** prepared via this procedure were identical to that prepared via the procedure listed as Method A above.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

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References

- 1. Beckman MG, Hooper WC, Critchley SE, Ortel TL. Am. J. Prev. Med. 2010; 38:S495–S501. [PubMed: 20331949]
- 2. Willemse JL, Hendriks DF. Clin. Chem. 2006; 52:30-36. [PubMed: 16299049]
- 3. Dubis J, Witkiewicz W. Adv. Clin. Exp. Med. 2010; 19:379-387.
- 4. (a) Barrow JC, Nantermet PG, Stauffer SR, Ngo PL, Steinbeiser MA, Mao SS, Carroll SS, Bailey C, Colussi D, Bosserman M, Burlein C, Cook JJ, Stiko G, Tiller PR, Miller-Stein CM, Rose M, McMasters DR, Vacca JP, Selnick HG. J. Med. Chem. 2003; 46:5294–5297. [PubMed: 14640538]
 (b) Nantermet PG, Barrow JC, Lindsley SR, Young M, Mao SS, Carroll S, Bailey C, Bosserman M, Colussi D, McMasters DR, Vacca JP, Selnick HG. Bioorg. Med. Chem. Lett. 2004; 14:2141–2145. [PubMed: 15080996]
- Bunnage ME, Blagg J, Steele J, Owen DR, Allerton C, McElroy AB, Miller D, Ringer T, Butcher K, Beaumont K, Evans K, Gray AJ, Holland SJ, Feeder N, Moore RS, Brown DG. J. Med. Chem. 2007; 50:6095–6103. [PubMed: 17990866]
- Islam I, Bryant J, May K, Mohan R, Yuan S, Kent L, Morser J, Zhao L, Vergona R, White K, Adler M, Whitlow M, Buckman BO. Bioorg. Med. Chem. Lett. 2007; 17:1349–1354. [PubMed: 17189688]
- 7. Bunnage ME, Owen DR. Curr. Opin. Drug Disc. Dev. 2008; 11:480-486.
- Willemse JL, Heylen E, Nesheim ME, Hendriks DF. J. Thromb. Haemost. 2009; 7:1962–1971. [PubMed: 19719827]
- 9. Carroll AR, Hyde E, Smith J, Quinn RJ, Guymer G, Forster PI. J. Org. Chem. 2005; 70:1096. [PubMed: 15675882]

- (a) Martin CL, Overman LE, Rohde JM. J. Am. Chem. Soc. 2008; 130:7568–7569. [PubMed: 18491907] (b) Martin CL, Overman LE, Rohde JM. J. Am. Chem. Soc. 2010; 132:4894–4906. [PubMed: 20218696]
- Taniguchi T, Martin CL, Monde K, Nakanishi K, Berova N, Overman LE. J. Nat. Prod. 2009; 39:806–810.
- 12. Vaswani RG, Day JJ, Wood JL. Org. Lett. 2009; 11:4532-4535. [PubMed: 19810762]
- 13. Zaimoku H, Taniguchi T, Ishibashi H. Org. Lett. 2012; 14:1656–1658. [PubMed: 22369137]
- 14. Galicia IZ, Maldonado LA. Tetrahedron Lett. 2013; 54:2180-2182.
- 15. Mortimer D, Whiting M, Harrity JPA, Jones S, Coldham I. Tetrahedron Lett. 2014; 55:1255–1257.
- For reviews of the vinylogous Mannich reaction, see: Martin SF. Acc. Chem. Res. 2002; 35:895–904. [PubMed: 12379142] Casiraghi G, Battistini L, Curti C, Rassu G, Zanardi F. Chem. Rev. 2011; 111:3076–3154. [PubMed: 21434642]
- 17. For a review of the Pictet-Spengler reaction, see: Stöckigt J, Antonchick AP, Wu F, Waldmann H. Angew. Chem. Int. Ed. 2011; 50:8538–8564.
- (a) Heidebrecht RW Jr, Gulledge B, Martin SF. Org. Lett. 2010; 12:2492–2495. [PubMed: 20446675] (b) Fu T–H, McElroy WT, Shamszad M, Heidebrecht RW Jr, Gulledge B, Martin SF. Org. Lett. 2012; 14:3834–3837. [PubMed: 22830424] (c) Fu T–H, McElroy WT, Shamszad M, Heidebrecht RW Jr, Gulledge B, Martin SF. Tetrahedron. 2013; 69:5588–5603. [PubMed: 23976796]
- 19. Fu T-H, Bonaparte A, Martin SF. Tetrahedron Lett. 2009; 50:3253-3257. [PubMed: 20161309]
- Granger BA, Jewett IT, Butler JD, Hua B, Knezevic CE, Parkinson EI, Hergenrother PJ, Martin SF. J. Am. Chem. Soc. 2013; 135:12984–12986. [PubMed: 23972114]
- 21. Choi M-H, Kim H-D. Arch. Pharm. Res. 2003; 26:990–996. [PubMed: 14723329]
- 22. Katritzky AR, Akutagawa K. Tetrahedron Lett. 1985; 26:5935–5938.
- (a) Buhr G. 1971 DE2013761, (b) Shizuka H, Ogiwara T, Morita T. Bull. Chem. Soc. Jpn. 1977; 50:2067–2073.
- For a review on gramine chemistry, see: Semenov BB, Granik VG. Pharm. Chem. J. 2004; 38:287– 310.
- 25. Minami I, Ohashi Y, Shimizu I, Tsuji J. Tetrahedron Lett. 1985; 26:2449-2452.
- 26. Martin CL, Nakamura S, Otte R, Overman LE. Org. Lett. 2011; 13:138. [PubMed: 21133399]
- Reactions of 1,2-aminochloroethanes with water are known to proceed through an aziridinium ion intermediate: Kerwin JF, Ullyot GE, Fuson RC, Zirkle CL. J. Am. Chem. Soc. 1974; 69:2961– 2965.
- 28. For a review on the oxidation of primary alcohols to carboxylic acids, see: Tojo G, Fernández M. Oxidation of Primary Alcohols to Carboxylic Acids: A Guide to Current Common Practices. 2007New YorkSpringer
- For attempted procedures, see: Jefford CW, Wang Y. J. Chem. Soc. Chem. Commun. 1988:634–635. Zhao M, Li J, Song Z, Desmond R, Tschaen DM, Grabowski EJJ, Reider PJ. Tetrahedron Lett. 1998; 39:5323–5326. Epp JB, Widlanski TS. J. Org. Chem. 1999; 64:293–295. [PubMed: 11674117] Ravi G, Lee K, Ji X–D, Kim HS, Soltysiak KA, Marquez VE, Jacobson KA. Bioorg. Med. Chem. Lett. 2001; 11:2295–2300. [PubMed: 11527718] Thottumkara AP, Bowsher MS, Vinod TK. Org. Lett. 2005; 7:2933–2936. [PubMed: 15987173] Grill JM, Ogle JW, Miller SA. J. Org. Chem. 2006; 71:9291–9296. [PubMed: 17137354] An G, Anh H, de Castro KA, Rhee H. Synthesis. 2010:477–485. Das R, Chakraborty D. Appl. Organometal. Chem. 2011; 25:437–442.
- 30. Schmidt A-KC, Stark CBW. Org. Lett. 2011; 13:4164–4167. [PubMed: 21793539]
- 31. Corey EJ, Schmidt G. Tetrahedron Lett. 1979; 5:399-402.
- 32. Mazitschek R, Mülbaier M, Giannis A. Angew. Chem. Int. Ed. 2002; 41:4059-4061.
- 33. Kunz H, März J. Angew. Chem. Int. Ed. 1988; 127:1375-1377.
- For some lead references on diverted total synthesis, see: Gaul C, Njardarson JT, Shan D, Dorn DC, Wu K–D, Tong WP, Huang X–Y, Moore MAS, Danishefsky SJ. J. Am. Chem. Soc. 2004; 126:11326–11337. [PubMed: 15355116] Wilson RM, Danishefsky SJ. J. Org. Chem. 2006; 71:8329–8351. [PubMed: 17064003] Szpilman AM, Carreira EM. Angew. Chem. Int. Ed. 2010; 49:9592–9628.

- Pangborn AB, Giardello MA, Grubbs RH, Rosen RK, Timmers FJ. Organometallics. 1996; 15:1518–1520.
- 36. Still WC, Kahn M, Mitra A. J. Org. Chem. 1978; 43:2923–2925.
- 37. CCDC 990247
- 38. CCDC 990248
- 39. CCDC 990249



Figure 1. X-ray structure of **24**; hydrogen atoms omitted for clarity.



Figure 2. X-ray structure of **28**; hydrogen atoms omitted for clarity.



Figure 3. X-ray structure of **2** with chloride ion (green); hydrogen atoms omitted for clarity.

OH,

0

OH



(±)-Actinophyllic acid (1)





Nin

N H

X

2





Scheme 1. Retrosynthetic analysis

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Scheme 2. Preparation of starting materials



Scheme 3. Failed cascade reaction

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Proposed fragmentation pathways







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Scheme 8. Synthesis of hexacyclic alcohol 2.



Scheme 9. Second generation synthesis

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