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Neurovascular pathways to neurodegeneration in Alzheimer's disease and other disorders

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Abstract

The neurovascular unit (NVU) comprises brain endothelial cells, pericytes or vascular smooth muscle cells, glia and neurons. The NVU controls blood–brain barrier (BBB) permeability and cerebral blood flow, and maintains the chemical composition of the neuronal ‘milieu’, which is required for proper functioning of neuronal circuits. Recent evidence indicates that BBB dysfunction is associated with the accumulation of several vasculotoxic and neurotoxic molecules within brain parenchyma, a reduction in cerebral blood flow, and hypoxia. Together, these vascular-derived insults might initiate and/or contribute to neuronal degeneration. This article examines mechanisms of BBB dysfunction in neurodegenerative disorders, notably Alzheimer's disease, and highlights therapeutic opportunities relating to these neurovascular deficits.

Neurons depend on blood vessels for their oxygen and nutrient supplies, and for the removal of carbon dioxide and other potentially toxic metabolites from the brain's interstitial fluid (ISF). The importance of the circulatory system to the human brain is highlighted by the fact that although the brain comprises ~2% of total body mass, it receives up to 20% of cardiac output and is responsible for ~20% and ~25% of the body's oxygen consumption and glucose consumption, respectively¹. To underline this point, when cerebral blood flow (CBF) stops, brain functions end within seconds and damage to neurons occurs within minutes².

Neurodegenerative disorders such as Alzheimer's disease and amyotrophic lateral sclerosis (ALS) are associated with microvascular dysfunction and/or degeneration in the brain, neurovascular disintegration, defective blood–brain barrier (BBB) function and/or vascular factors^{1,3–12}. Microvascular deficits diminish CBF and, consequently, the brain's supply of oxygen, energy substrates and nutrients. Moreover, such deficits impair the clearance of neurotoxic molecules that accumulate and/or are deposited in the ISF, non-neuronal cells and neurons. Recent evidence suggests that vascular dysfunction leads to neuronal dysfunction and neurodegeneration, and that it might contribute to the development of proteinaceous brain and cerebrovascular ‘storage’ disorders. Such disorders include cerebral

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β -amyloidosis and cerebral amyloid angiopathy (CAA), which are caused by accumulation of the peptide amyloid- β in the brain and the vessel wall, respectively, and are features of Alzheimer's disease¹.

In this Review, I will discuss neurovascular pathways to neurodegeneration, placing a focus on Alzheimer's disease because more is known about neurovascular dysfunction in this disease than in other neurodegenerative disorders. The article first examines transport mechanisms for molecules to cross the BBB, before exploring the processes that are involved in BBB breakdown at the molecular and cellular levels, and the consequences of BBB breakdown, hypoperfusion, and hypoxia and endothelial metabolic dysfunction for neuronal function. Next, the article reviews evidence for neurovascular changes during normal ageing and neurovascular BBB dysfunction in various neurodegenerative diseases, including evidence suggesting that vascular defects precede neuronal changes. Finally, the article considers specific mechanisms that are associated with BBB dysfunction in Alzheimer's disease and ALS, and therapeutic opportunities relating to these neurovascular deficits.

The neurovascular unit

The neurovascular unit (NVU) comprises vascular cells (that is, endothelium, pericytes and vascular smooth muscle cells (VSMCs)), glial cells (that is, astrocytes, microglia and oligodendroglia) and neurons^{1,2,13} (FIG. 1). In the NVU, the endothelial cells together form a highly specialized membrane around blood vessels. This membrane underlies the BBB and limits the entry of plasma components, red blood cells (RBCs) and leukocytes into the brain. The BBB also regulates the delivery into the CNS of circulating energy metabolites and essential nutrients that are required for proper neuronal and synaptic function. Non-neuronal cells and neurons act in concert to control BBB permeability and CBF. Vascular cells and glia are primarily responsible for maintenance of the constant 'chemical' composition of the ISF, and the BBB and the blood–spinal cord barrier (BSCB) work together with pericytes to prevent various potentially neurotoxic and vasculotoxic macromolecules in the blood from entering the CNS, and to promote clearance of these substances from the CNS¹.

Transport across the blood–brain barrier

The endothelial cells that form the BBB are connected by tight and adherens junctions, and it is the tight junctions that confer the low paracellular permeability of the BBB¹. Small lipophilic molecules, oxygen and carbon dioxide diffuse freely across the endothelial cells, and hence the BBB, but normal brain endothelium lacks fenestrae and has limited vesicular transport.

The high number of mitochondria in endothelial cells reflects a high energy demand for active ATP-dependent transport, conferred by transporters such as the sodium pump ((Na^+ + K^+) ATPase) and the ATP-binding cassette (ABC) efflux transporters. Sodium influx and potassium efflux across the abluminal side of the BBB is controlled by (Na^+ + K^+) ATPase (FIG. 2). Changes in sodium and potassium levels in the ISF influence the

generation of action potentials in neurons and thus directly affect neuronal and synaptic functions^{1,12}.

Brain endothelial cells express transporters that facilitate the transport of nutrients down their concentration gradients, as described in detail elsewhere (FIG. 2). Glucose transporter 1 (GLUT1; also known as solute carrier family 2, facilitated glucose transporter member 1 (SLC2A1)) — the BBB-specific glucose transporter — is of special importance because glucose is a key energy source for the brain. Monocarboxylate transporter 1 (MCT1), which transports lactate, and the L1 and γ + amino acid transporters are expressed at the luminal and abluminal membranes. Sodium-dependent excitatory amino acid transporter 1 (EAAT1), EAAT2 and EAAT3 are expressed at the abluminal side of the BBB¹⁵ and enable removal of glutamate, an excitatory neurotransmitter, from the brain (FIG. 2). Glutamate clearance at the BBB is essential for protecting neurons from overstimulation of glutamergic receptors, which is neurotoxic¹⁶. ABC transporters limit the penetration of many drugs into the brain¹⁷. For example, multidrug resistance protein 1 (ABCB1; also known as ATP-binding cassette subfamily B member 1) controls the rapid removal of ingested toxic lipophilic metabolites¹⁷ (FIG. 2). Some ABC transporters also mediate the efflux of nutrients from the endothelium into the ISF. For example, solute carrier organic anion transporter family member 1C1 (OATP1C1) transports thyroid hormones into the brain. MCT8 mediates influx of thyroid hormones from blood into the endothelium¹⁸ (FIG. 2).

The transport of circulating peptides across the BBB into the brain is restricted or slow compared with the transport of nutrients¹⁹. Carrier-mediated transport of neuroactive peptides controls their low levels in the ISF^{20–24} (FIG. 2). Some proteins, including transferrin, insulin, insulin-like growth factor 1 (IGF1), leptin^{25–27} and activated protein C (APC)²⁸, cross the BBB by receptor-mediated transcytosis (FIG. 2).

Circumventricular organs

Several small neuronal structures that surround brain ventricles lack the BBB and sense chemical changes in blood or the cerebrospinal fluid (CSF) directly. These brain areas are known as circumventricular organs (CVOs). CVOs have important roles in multiple endocrine and autonomic functions, including the control of feeding behaviour as well as regulation of water and salt metabolism²⁹. For example, the subfornical organ is one of the CVOs that are capable of sensing extracellular sodium using astrocyte derived lactate as a signal for local neurons to initiate neural, hormonal and behavioural responses underlying sodium homeostasis³⁰. Excessive sodium accumulation is detrimental, and increases in plasma sodium above a narrow range are incompatible with life, leading to cerebral oedema (swelling), seizures and death²⁹.

Vascular-mediated pathophysiology

The key pathways of vascular dysfunction that are linked to neurodegenerative diseases include BBB breakdown, hypoperfusion–hypoxia and endothelial metabolic dysfunction (FIG. 3). This section examines processes that are involved in BBB breakdown at the molecular and cellular levels, and explores the consequences of all three pathways for neuronal function and viability.

Blood–brain barrier breakdown

Disruption to tight and adherens junctions, an increase in bulk-flow fluid transcytosis, and/or enzymatic degradation of the capillary basement membrane cause physical breakdown of the BBB. The levels of many tight junction proteins, their adaptor molecules and adherens junction proteins decrease in Alzheimer's disease and other diseases that cause dementia^{1,9}, ALS³¹, multiple sclerosis³² and various animal models of neurological disease^{8,33}. These decreases might be partly explained by the fact that vascular-associated matrix metalloproteinase (MMP) activity rises in many neurodegenerative disorders and after ischaemic CNS injury^{34,35}; tight junction proteins and basement membrane extracellular matrix proteins are substrates for these enzymes³⁴. Lowered expression of messenger RNAs that encode several key tight junction proteins, however, has also been reported in some neurodegenerative disorders, such as ALS³¹.

Endothelial cell–pericyte interactions are crucial for the formation^{36,37} and maintenance of the BBB^{33,38}. Pericyte deficiency can lead to a reduction in expression of certain tight junction proteins, including occludin, claudin 5 and ZO1³³, and to an increase in bulk-flow transcytosis across the BBB, causing BBB breakdown³⁸. Both processes can lead to extravasation of multiple small and large circulating macromolecules (up to 500 kDa) into the brain parenchyma^{33,38}. Moreover, in mice, an age-dependent progressive loss of pericytes can lead to BBB disruption and microvascular degeneration and, subsequently, neuronal dysfunction, cognitive decline and neurodegenerative changes³³. In their lysosomes, pericytes concentrate and degrade multiple circulating exogenous³⁹ and endogenous proteins amplify BBB breakdown in cases of pericyte deficiency.

BBB breakdown typically leads to an accumulation of various molecules in the brain. The build up of serum proteins such as immunoglobulins and albumin can cause brain oedema and suppression of capillary blood flow whereas high concentrations of thrombin lead to neurotoxicity and memory impairment⁴⁰, and accelerate vascular damage and BBB disruption⁴¹. The accumulation of plasmin (derived from circulating plasminogen) can catalyze the degradation of neuronal laminin and, hence, promote neuronal injury⁴², and high fibrin levels accelerate neurovascular damage⁶. Finally, an increase in the number of RBCs causes deposition of haemoglobin-derived neurotoxic products including iron, which generates neurotoxic reactive oxygen species (ROS)^{8,43} (FIG. 3a). In addition to protein-mediated vasogenic oedema, local tissue ischaemia–hypoxia depletes ATP stores, causing (Na⁺+K⁺) ATPase pumps and Na⁺-dependent ion channels to stop working and, consequently, the endothelium and astrocytes to swell (known as cytotoxic oedema)⁴⁴. Upregulation of aquaporin 4 water channels in response to ischaemia facilitates the development of cytotoxic oedema in astrocytes⁴⁵.

Hypoperfusion and hypoxia

CBF is regulated by local neuronal activity and metabolism, known as neurovascular coupling⁴⁶. The pial and intracerebral arteries control the local increase in CBF that occurs during brain activation, which is termed 'functional hyperaemia'. Neurovascular coupling requires intact pial circulation, and for VSMCs and pericytes to respond normally to vasoactive stimuli^{33,46,47}. In addition to VSMC-mediated constriction and vasodilation of

cerebral arteries, recent studies have shown that pericytes modulate brain capillary diameter through constriction of the vessel wall⁴⁷, which obstructs capillary flow during ischaemia⁴⁸. Astrocytes regulate the contractility of intracerebral arteries^{49,50}.

Progressive CBF reductions have increasingly serious consequences for neurons (FIG. 3b). Briefly, mild hypoperfusion — termed oligoemia — affects protein synthesis, which is required for the synaptic plasticity mediating learning and memory⁴⁶. Moderate to severe CBF reductions and hypoxia affect ATP synthesis, diminishing ($\text{Na}^+\text{+K}^+$) ATPase activity and the ability of neurons to generate action potentials⁹. In addition, such reductions can lower or increase pH, and alter electrolyte balances and water gradients, leading to the development of oedema and white matter lesions, and the accumulation of glutamate and proteinaceous toxins (for example, amyloid- β and hyperphosphorylated tau) in the brain. A reduction of greater than 80% in CBF results in neuronal death².

The effect of CBF reductions has been extensively studied at the molecular and cellular levels in relation to Alzheimer's disease. Reduced CBF and/or CBF dysregulation occurs in elderly individuals at high risk of Alzheimer's disease before cognitive decline, brain atrophy and amyloid- β accumulation^{10,46,51–54}. In animal models, hypoperfusion can induce or amplify Alzheimer's disease-like neuronal dysfunction and/or neuropathological changes. For example, bilateral carotid occlusion in rats causes memory impairment, neuronal dysfunction, synaptic changes and amyloid- β oligomerization⁵⁵, leading to accumulation of neurotoxic amyloid- β oligomers⁵⁶. In a mouse model of Alzheimer's disease, oligoemia increases neuronal amyloid- β levels and neuronal tau phosphorylation at an epitope that is associated with Alzheimer's disease-type paired helical filaments⁵⁷. In rodents, ischaemia leads to the accumulation of hyperphosphorylated tau in neurons and the formation of filaments that resemble those present in human neurodegenerative tauopathies and Alzheimer's disease⁵⁸. Mice expressing amyloid- β precursor protein (APP) and transforming growth factor β 1 (TGF β 1) develop deficient neurovascular coupling, cholinergic denervation, enhanced cerebral and cerebrovascular amyloid- β deposition, and age-dependent cognitive decline⁵⁹.

Recent studies have shown that ischaemia–hypoxia influences amyloidogenic APP processing through mechanisms that increase the activity of two key enzymes that are necessary for amyloid- β production; that is, β -secretase and γ -secretase^{60–63}. Hypoxia-inducible factor 1 α (HIF1 α) mediates transcriptional increase in β -secretase expression⁶¹. Hypoxia also promotes phosphorylation of tau through the mitogen-activated protein kinase (MAPK; also known as extracellular signal-regulated kinase (ERK)) pathway⁶⁴, downregulates neprilysin — an amyloid- β -degrading enzyme⁶⁵ — and leads to alterations in the expression of vascular-specific genes, including a reduction in the expression of the homeobox protein MOX2 gene mesenchyme homeobox 2 (*MEOX2*) in brain endothelial cells⁵ and an increase in the expression of the myocardin gene (*MYOCD*) in VSMCs⁶⁶. In patients with Alzheimer's disease and in models of this disorder, these changes cause vessel regression, hypoperfusion and amyloid- β accumulation resulting from the loss of the key amyloid- β clearance lipoprotein receptor (see below). In addition, hypoxia facilitates alternative splicing of *Eaat2* mRNA in Alzheimer's disease transgenic mice before amyloid- β deposition⁶⁷ and suppresses glutamate reuptake by astrocytes independently of amyloid

formation⁶⁸, resulting in glutamate-mediated neuronal injury that is independent of amyloid- β .

In response to hypoxia, mitochondria release ROS that mediate oxidative damage to the vascular endothelium and to the selective population of neurons that has high metabolic activity. Such damage has been suggested to occur before neuronal degeneration and amyloid- β deposition in Alzheimer's disease^{69,70}. Although the exact triggers of hypoxia-mediated neurodegeneration and the role of HIF1 α in neurodegeneration versus preconditioning-mediated neuroprotection remain topics of debate, mitochondria-generated ROS seem to have a primary role in the regulation of the HIF1 α -mediated transcriptional switch that can activate an array of responses, ranging from mechanisms that increase cell survival and adaptation to mechanisms inducing cell cycle arrest and death⁷¹. Whether inhibition of hypoxia-mediated pathogenic pathways will delay onset and/or control progression in neurodegenerative conditions such as Alzheimer's disease remains to be determined.

When comparing the contributions of BBB break-down and hypoperfusion to neuronal injury, it is interesting to consider *Meox2*^{+/-} mice. Such animals have normal pericyte coverage and an intact BBB but a substantial perfusion deficit⁵ that is comparable to that found in pericyte-deficient mice that develop BBB breakdown³³. Notably, however, *Meox2*^{+/-} mice show less pronounced neurodegenerative changes than pericyte-deficient mice, indicating that chronic hypoperfusion–hypoxia alone can cause neuronal injury, but not to the same extent as hypoperfusion–hypoxia combined with BBB breakdown.

Endothelial neurotoxic and inflammatory factors

Alterations in cerebrovascular metabolic functions can lead to the secretion of multiple neurotoxic and inflammatory factors^{72,73}. For example, brain microvessels that have been isolated from individuals with Alzheimer's disease (but not from neurologically normal age-matched and young individuals) and brain microvessels that have been treated with inflammatory proteins release neurotoxic factors that kill neurons^{74,75}. These factors include thrombin, the levels of which increase with the onset of Alzheimer's disease⁷⁶. Thrombin can injure neurons directly⁴⁰ and indirectly by activating microglia and astrocytes⁷³. Compared with those from age-matched controls, brain microvessels from individuals with Alzheimer's disease secrete increased levels of multiple inflammatory mediators, such as nitric oxide, cytokines (for example, tumour necrosis factor (TNF), TGF β 1, interleukin-1 β (IL-1 β) and IL-6), chemokines (for example, CC-chemokine ligand 2 (CCL2; also known as monocyte chemoattractant protein 1 (MCP1)) and IL-8), prostaglandins, MMPs and leukocyte adhesion molecules⁷³. Endothelium-derived neurotoxic and inflammatory factors together provide a molecular link between vascular metabolic dysfunction, neuronal injury and inflammation in Alzheimer's disease and, possibly, in other neurodegenerative disorders.

Neurovascular changes

This section examines evidence for neurovascular changes during normal ageing and for neurovascular and/or BBB dysfunction in various neurodegenerative diseases, as well as the possibility that vascular defects can precede neuronal changes.

Age-associated neurovascular changes

Normal ageing diminishes brain circulatory functions, including a detectable decay of CBF in the limbic and association cortices that has been suggested to underlie age-related cognitive changes⁷⁷. Alterations in the cerebral microvasculature, but not changes in neural activity, have been shown to lead to age-dependent reductions in functional hyperaemia in the visual system in cats⁷⁸ and in the sensorimotor cortex in pericyte-deficient mice³³. Importantly, a recent longitudinal CBF study in neurologically normal individuals revealed that people bearing the apolipoprotein E (APOE) $\epsilon 4$ allele — the major genetic risk factor for late-onset Alzheimer's disease^{79–81} — showed greater regional CBF decline in brain regions that are particularly vulnerable to pathological changes in Alzheimer's disease than did people without this allele⁸².

A meta-analysis of BBB permeability in 1,953 individuals showed that neurologically healthy humans had an age-dependent increase in vascular permeability⁸³. Moreover, patients with vascular or Alzheimer's disease-type dementia and leukoaraiosis — a small-vessel disease of the cerebral white matter — had an even greater age-dependent increase in vascular permeability⁸³. Interestingly, an increase in BBB permeability in brain areas with normal white matter in patients with leukoaraiosis has been suggested to play a causal part in disease and the development of lacunar strokes⁸⁴. Age-related changes in the permeability of the blood–CSF barrier and the choroid plexus have been reported in sheep⁸⁵.

Vascular pathology

Patients with Alzheimer's disease or other dementia-causing diseases frequently show focal changes in brain microcirculation. These changes include the appearance of string vessels (collapsed and acellular membrane tubes), a reduction in capillary density, a rise in endothelial pinocytosis, a decrease in mitochondrial content, accumulation of collagen and perlecan in the basement membrane, loss of tight junctions and/or adherens junctions^{3–6,9,46,86}, and BBB breakdown with leakage of blood-borne molecules^{4,6,7,9}. The time course of these vascular alterations and how they relate to dementia and Alzheimer's disease pathology remain unclear, as no protocol that allows the development of the diverse brain vascular pathology to be scored, and hence to be tracked with ageing, has so far been developed and widely validated⁸⁷. Interestingly, a recent study involving 500 individuals who died between the ages of 69 and 103 years showed that small-vessel disease, infarcts and the presence of more than one vascular pathological change were associated with Alzheimer's disease-type pathological lesions and dementia in people aged 75 years of age⁸⁷. These associations were, however, less pronounced in individuals aged 95 years of age, mainly because of a marked ageing-related reduction in Alzheimer's disease neuropathology relative to a moderate but insignificant ageing-related reduction in vascular pathology⁸⁷.

Accumulation of amyloid- β and amyloid deposition in pial and intracerebral arteries results in CAA, which is present in over 80% of Alzheimer's disease cases⁸⁸. Inpatients who have Alzheimer's disease with established CAA in small arteries and arterioles, the VSMC layer frequently shows atrophy, which causes a rupture of the vessel wall and intracerebral bleeding in about 30% of these patients^{89,90}. These intracerebral bleedings contribute to, and aggravate, dementia. Patients with hereditary cerebral β -amyloidosis and CAA of the Dutch, Iowa, Arctic, Flemish, Italian or Piedmont L34V type have accelerated VSMC degeneration resulting in haemorrhagic strokes and dementia⁹¹. Duplication of the gene encoding APP causes early-onset Alzheimer's disease dementia with CAA and intracerebral haemorrhage⁹².

Early studies of serum immunoglobulin leakage reported that patients with ALS had BSCB break-down and BBB breakdown in the motor cortex⁹³. Microhaemorrhages and BSCB breakdown have been shown in the spinal cord of transgenic mice expressing mutant variants of human superoxide dismutase 1 (SOD1), which in mice cause an ALS-like disease^{8,94,95}. In mice with ALS-like disease and in patients with ALS, BSCB breakdown has been shown to occur before motor neuron degeneration or brain atrophy^{8,11,95}.

BBB breakdown in the substantia nigra and the striatum has been detected in murine models of Parkinson's disease that are induced by administration of 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP)⁹⁶⁻⁹⁸. However, the temporal relationship between BBB breakdown and neurodegeneration in Parkinson's disease is currently unknown. Notably, the prevalence of CAA and vascular lesions increases in Parkinson's disease^{99,100}. Vascular lesions in the striatum and lacunar infarcts can cause vascular parkinsonism syndrome¹⁰¹. A recent study reported BBB breakdown in a rat model of Huntington's disease that is induced with the toxin 3-nitropropionic acid¹⁰².

Several studies have established disruption of BBB with a loss of tight junction proteins during neuroinflammatory conditions such as multiple sclerosis and its murine model, experimental allergic encephalitis. Such disruption facilitates leukocyte infiltration, leading to oligodendrocyte death, axonal damage, demyelination and lesion development³².

Functional changes in the vasculature

In individuals with Alzheimer's disease, GLUT1 expression at the BBB decreases¹⁰³, suggesting a shortage in necessary metabolic substrates. Studies using 18F-fluorodeoxyglucose (FDG) positron emission tomography (PET) have identified reductions in glucose uptake in asymptomatic individuals with a high risk of dementia^{104,105}. Several studies have suggested that reduced glucose uptake across the BBB, as seen by FDG PET, precedes brain atrophy¹⁰⁴⁻¹⁰⁸.

Amyloid- β constricts cerebral arteries¹⁰⁹. In a mouse model of Alzheimer's disease, impairment of endothelium-dependent regulation of neocortical microcirculation^{110,111} occurs before amyloid- β accumulation. Recent studies have shown that CD36, a scavenger receptor that binds amyloid- β , is essential for the vascular oxidative stress and diminished functional hyperaemia that occurs in response to amyloid- β exposure¹¹². Neuroimaging studies in patients with Alzheimer's disease have shown that neurovascular uncoupling

occurs before neurodegenerative changes^{10,51–53}. Moreover, cognitively normal APOE $\epsilon 4$ carriers at risk of Alzheimer's disease show impaired CBF responses to brain activation in the absence of neurodegenerative changes or amyloid- β accumulation⁵⁴. Recently, patients with Alzheimer's disease as well as mouse models of this disease with high cerebrovascular levels of serum response factor (SRF) and MYOCD, the two transcription factors that control VSMC differentiation, have been shown to develop a hypercontractile arterial phenotype resulting in brain hypoperfusion, diminished functional hyperaemia and CAA^{66,113}. More work is needed to establish the exact role of SRF and MYOCD in the vascular dysfunction that results in the Alzheimer's disease phenotype and CAA.

PET studies with ¹¹C-verapamil, an ABCB1 substrate, have indicated that the function of ABCB1, which removes multiple drugs and toxins from the brain, decreases with ageing¹¹⁴ and is particularly compromised in the midbrain of patients with Parkinson's disease, progressive supranuclear palsy or multiple system atrophy¹¹⁵. More work is needed to establish the exact roles of ABC BBB transporters in neurodegeneration and whether their failure precedes the loss of dopaminergic neurons that occurs in Parkinson's disease.

In mice with ALS-like disease and in patients with ALS, hypoperfusion and/or dysregulated CBF have been shown to occur before motor neuron degeneration or brain atrophy^{8,116}. Reduced regional CBF in basal ganglia and reduced blood volume have been reported in pre-symptomatic gene-tested individuals at risk for Huntington's disease¹¹⁷. Patients with Huntington's disease display a reduction in vasomotor activity in the cerebral anterior artery during motor activation¹¹⁸.

Vascular and neuronal common growth factors

Blood vessels and neurons share common growth factors and molecular pathways that regulate their development and maintenance^{119,120}. Angioneurins are growth factors that exert both vasculotrophic and neurotrophic activities¹²¹. The best studied angioneurin is vascular endothelial growth factor (VEGF). VEGF regulates vessel formation, axonal growth and neuronal survival¹²⁰. Ephrins, semaphorins, slits and netrins are axon guidance factors that also regulate the development of the vascular system¹²¹. During embryonic development of the neural tube, blood vessels and choroid plexus secrete IGF2 into the CSF, which regulates the proliferation of neuronal progenitor cells¹²². Genetic and pharmacological manipulations of angioneurin activity yielded various vascular and cerebral phenotypes¹²¹. Given the dual nature of angioneurin action, these studies have not been able to address whether neuronal dysfunction results from a primary insult to neurons and/or whether it is secondary to vascular dysfunction.

Increased levels of VEGF, a hypoxia-inducible angiogenic factor, were found in the walls of intraparenchymal vessels, perivascular deposits, astrocytes and intrathecal space of patients with Alzheimer's disease, and were consistent with the chronic cerebral hypoperfusion and hypoxia that were observed in these individuals⁷³. In addition to VEGF, brain microvessels in Alzheimer's disease release several molecules that can influence angiogenesis, including IL-1 β , IL-6, IL-8, TNF, TGF β , MCP1, thrombin, angiopoietin 2, α V β 3 and α V β 5 integrins, and HIF1 α ⁷³. However, evidence for increased vascularity in Alzheimer's disease is lacking. On the contrary, several studies have reported that focal vascular regression and

diminished microvascular density occur in Alzheimer's disease^{4,5,73} and in Alzheimer's disease transgenic mice¹²³. The reason for this discrepancy is not clear. The anti-angiogenic activity of amyloid- β , which accumulates in the brains of individuals with Alzheimer's disease and Alzheimer's disease models, may contribute to hypo-vascularity¹²³. Conversely, genome-wide transcriptional profiling of brain endothelial cells from patients with Alzheimer's disease revealed that extremely low expression of vascular-restricted MEOX2 mediates aberrant angiogenic responses to VEGF and hypoxia, leading to capillary death⁵. This finding raises the interesting question of whether capillary degeneration in Alzheimer's disease results from unsuccessful vascular repair and/or remodelling. Moreover, mice that lack one *Meox2* allele have been shown to develop a primary cerebral endothelial hypoplasia with chronic brain hypoperfusion⁵, resulting in secondary neurodegenerative changes³³.

Does vascular dysfunction cause neuronal dysfunction?

In summary, the evidence that is discussed above clearly indicates that vascular dysfunction is tightly linked to neuronal dysfunction. There are many examples to illustrate that primary vascular deficits lead to secondary neurodegeneration, including CADASIL (cerebral autosomal dominant arteriopathy with subcortical infarcts), an hereditary small-vessel brain disease resulting in multiple small ischaemic lesions, neurodegeneration and dementia¹²⁴; mutations in *SLC2A1* that cause dysfunction of the BBB-specific GLUT1 transporter in humans resulting in seizures; cognitive impairment and microcephaly¹²⁵; microcephaly and epileptiform discharges in mice with genetic deletion of a single *Slc2a1* allele¹²⁶; and neurodegeneration mediated by a single *Meox2* homeobox gene deletion restricted to the vascular system³³. Patients with hereditary cerebral β -amyloidosis and CAA of the Dutch, Iowa, Arctic, Flemish, Italian or Piedmont L34V type provide another example showing that primary vascular dysfunction — which in this case is caused by deposition of vasculotropic amyloid- β mutants in the arterial vessel wall — leads to dementia and intracerebral bleeding. Moreover, as reviewed in the previous sections, recent evidence suggests that BBB dysfunction and/or breakdown, and CBF reductions and/or dysregulation may occur in sporadic Alzheimer's disease and experimental models of this disease before cognitive decline, amyloid- β deposition and brain atrophy. In patients with ALS and in some experimental models of ALS, CBF dysregulation, BSCB breakdown and spinal cord hypoperfusion have been reported to occur before motor neuron cell death. Whether neurological changes follow or precede vascular dysfunction in Parkinson's disease, Huntington's disease and multiple sclerosis remains less clear. However, there is little doubt that vascular injury mediates, amplifies and/or lowers the threshold for neuronal dysfunction and loss in several neurological disorders.

Disease-specific considerations

This section examines how amyloid- β levels are kept low in the brain, a process in which the BBB has a central role, and how faulty BBB-mediated clearance mechanisms go awry in Alzheimer's disease. On the basis of this evidence and the findings discussed elsewhere in the Review, a new hypothesis for the pathogenesis of Alzheimer's disease that incorporates

the vascular evidence is presented. ALS-specific disease mechanisms relating to the BBB are then examined.

Alzheimer's disease

Amyloid- β clearance from the brain by the BBB is the best studied example of clearance of a proteinaceous toxin from the CNS. Multiple pathways regulate brain amyloid- β levels, including its production and clearance (FIG. 4). Recent studies^{127–129} have confirmed earlier findings in multiple rodent and non-human primate models demonstrating that peripheral amyloid- β is an important precursor of brain amyloid- β ^{130–136}. Moreover, peripheral amyloid- β sequestering agents such as soluble LRP1¹³⁷, anti-amyloid- β antibodies^{138–140}, gelsolin and the ganglioside GM1¹⁴¹, or systemic expression of neprilysin have been shown to reduce the amyloid burden in Alzheimer's disease mice by eliminating contributions of the peripheral amyloid- β pool to the total brain pool of this peptide.

The receptor for advanced glycation end products (RAGE) mediates amyloid- β transport in brain and the propagation of its toxicity. RAGE expression in brain endothelium provides a mechanism for influx of amyloid- β ^{144,145} and amyloid- β -laden monocytes¹⁴⁶ across the BBB, as shown in Alzheimer's disease models (FIG. 4). The amyloid- β -rich environment in Alzheimer's disease and models of this disorder increases RAGE expression at the BBB and in neurons^{147,148}, amplifying amyloid- β -mediated pathogenic responses. Blockade of amyloid- β -RAGE signaling in Alzheimer's disease is a promising strategy to control self-propagation of amyloid- β -mediated injury.

Several studies in animal models of Alzheimer's disease and, more recently, in patients with this disorder¹⁴⁹ have shown that diminished amyloid- β clearance occurs in brain tissue in this disease. LRP1 plays an important part in the three-step serial clearance of this peptide from brain and the rest of the body¹⁵⁰ (FIG. 4). In step one, LRP1 in brain endothelium binds brain-derived amyloid- β at the abluminal side of the BBB, initiating its clearance to blood, as shown in many animal models^{151–156} and BBB models *in vitro*^{151,157,158}. The vasculotropic mutants of amyloid- β that have low binding affinity for LRP1 are poorly cleared from the brain or CSF^{151,159,160}. APOE4, but not APOE3 or APOE2, blocks LRP1-mediated amyloid- β clearance from the brain and, hence, promotes its retention¹⁶¹, whereas clusterin (also known as apolipoprotein J (APOJ)) mediates amyloid- β clearance across the BBB via LRP2¹⁵³. APOE and clusterin influence amyloid- β aggregation^{162,163}. Reduced LRP1 levels in brain microvessels, perhaps in addition to altered levels of ABCB1, are associated with amyloid- β cerebrovascular and brain accumulation during ageing in rodents, non-human primates, humans, Alzheimer's disease mice and patients with Alzheimer's disease^{66,151,152,164–166}. Moreover, recent work has shown that brain LRP1 is oxidized in Alzheimer's disease¹⁶⁷, and may contribute to amyloid- β retention in brain because the oxidized form cannot bind and/or transport amyloid- β ¹³⁷. LRP1 also mediates the removal of amyloid- β from the choroid plexus¹⁶⁸.

In step two, circulating soluble LRP1 binds more than 70% of plasma amyloid- β in neurologically normal humans¹³⁷. In patients with Alzheimer's disease or mild cognitive impairment (MCI), and in Alzheimer's disease mice, amyloid- β binding to soluble LRP1 is compromised due to oxidative changes^{137,169}, resulting in elevated plasma levels of free

amyloid- β isoforms comprising 40 or 42 amino acids (amyloid- β_{1-40} and amyloid- β_{1-42}). These peptides can then re-enter the brain, as has been shown in a mouse model of Alzheimer's disease¹³⁷. Rapid systemic removal of amyloid- β by the liver is also mediated by LRP1 and comprises step three of the clearance process¹⁷⁰.

In brain, amyloid- β is enzymatically degraded by neprilysin¹⁷¹, insulin-degrading enzyme¹⁷², tissue plasminogen activator¹⁷³ and MMPs^{173,174} in various cell types, including endothelial cells, pericytes, astrocytes, neurons and microglia. Cellular clearance of this peptide by astrocytes and VSMCs is mediated by LRP1 and/or another lipoprotein receptor^{66,175}. Clearance of amyloid- β aggregates by microglia has an important role in amyloid- β -directed immunotherapy¹⁷⁶ and reduction of the amyloid load in brain¹⁷⁷. Passive ISF-CSF bulk flow and subsequent clearance through the CSF might contribute to 10–15% of total amyloid- β removal^{152,153,178}. In the injured human brain, increasing soluble amyloid- β concentrations in the ISF correlated with improvements in neurological status, suggesting that neuronal activity might regulate extracellular amyloid- β levels¹⁷⁹.

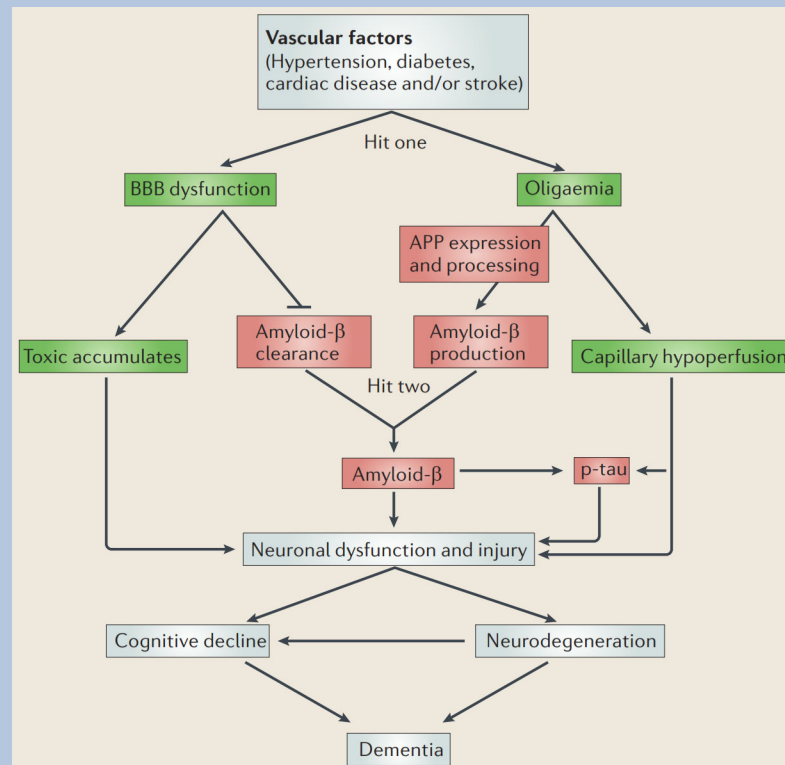
The role of BBB dysfunction in amyloid- β accumulation, as discussed above, underlies the contribution of vascular dysfunction to Alzheimer's disease (see FIG. 5 for a model of vascular damage in Alzheimer's disease). The amyloid hypothesis for the pathogenesis of Alzheimer's disease maintains that this peptide initiates a cascade of events leading to neuronal injury and loss and, eventually, dementia^{180,181}. Here, I present an alternative hypothesis — the two-hit vascular hypothesis of Alzheimer's disease — that incorporates the vascular contribution to this disease, as discussed in this Review (BOX 1). This hypothesis states that primary damage to brain microcirculation (hit one) initiates a non-amyloidogenic pathway of vascular-mediated neuronal dysfunction and injury, which is mediated by BBB dysfunction and is associated with leakage and secretion of multiple neurotoxic molecules and/or diminished brain capillary flow that causes multiple focal ischaemic or hypoxic micro-injuries. BBB dysfunction also leads to impairment of amyloid- β clearance, and oligoemia leads to increased amyloid- β generation. Both processes contribute to accumulation of amyloid- β species in the brain (hit two), where these peptides exert vasculotoxic and neurotoxic effects. According to the two-hit vascular hypothesis of Alzheimer's disease, tau pathology develops secondary to vascular and/or amyloid- β injury.

Box 1

The two-hit vascular hypothesis for Alzheimer's disease

Substantial overlap exists among risk factors for cerebrovascular disorder and Alzheimer's disease^{9,88,205}. For example, midlife diabetes^{10,206}, hypertension²⁰⁷ and obesity²⁰⁸ are vascular risk factors that predispose individuals to Alzheimer's disease and vascular dementia. It is now widely recognized that most cases of Alzheimer's disease have mixed vascular pathology and small-vessel disease^{88,209}. Moreover, brain hypoperfusion-hypoxia⁵³, silent infarcts²¹⁰, the presence of one or more infarctions²¹¹, stroke episodes and transient ischaemic or hypoxic attacks all increase the risk of Alzheimer's disease. In this disorder, although the molecular and cellular events for each step in the disease process and for each risk factor are not absolutely clear, vascular factors might all converge on a common final disease pathway, involving brain

microvascular dysfunction and/or degeneration, as well as amyloid- β and tau pathology. According to the two-hit vascular hypothesis of Alzheimer's disease, vascular risk factors (hit one) lead to blood-brain barrier (BBB) dysfunction and a reduction in cerebral blood flow (oligaemia), initiating a cascade of events that precedes dementia. In the non-amyloid- β pathway (see the figure, shown in green boxes), toxic accumulates and capillary hypoperfusion induce early neuronal dysfunction. Vascular injury also reduces amyloid- β clearance at the BBB and increases production of this peptide from the amyloid- β precursor protein (APP), leading to amyloid- β accumulation (the amyloid- β pathway; see the figure, shown in red boxes). The increase in amyloid- β (hit two) amplifies neuronal dysfunction, accelerates neurodegeneration and dementia, and contributes to disease self-propagation. Amyloid- β and/or hypoperfusion can induce hyperphosphorylation of tau (p-tau), leading to neurofibrillary tangle formation.



Box 1.
The two-hit vascular hypothesis for Alzheimer's disease

Amyotrophic lateral sclerosis

The cause of sporadic ALS, a fatal adult-onset motor neuron neurodegenerative disease, is not known¹⁸². In a relatively small number of patients with inherited SOD1 mutations, the disease is caused by toxic properties of mutant SOD1¹⁸³. Mutations in the genes encoding ataxin 2 and TAR DNA-binding protein 43 (TDP43) that cause these proteins to aggregate have been associated with ALS^{182,184}. Some studies have suggested that abnormal SOD1 species accumulate in sporadic ALS¹⁸⁵. Interestingly, studies in ALS transgenic mice

expressing a mutant version of human *SOD1* in neurons, and in non-neuronal cells neighbouring these neurons, have shown that deletion of this gene from neurons does not influence disease progression¹⁸⁶, whereas deletion of this gene from microglia¹⁸⁶ or astrocytes¹⁸⁷ substantially increases an animal's lifespan. According to an emerging hypothesis of ALS that is based on studies in *SOD1* mutant mice, the toxicity that is derived from non-neuronal neighbouring cells, particularly microglia and astrocytes, contributes to disease progression and motor neuron degeneration^{186–190}, whereas BBB dysfunction might be critical for disease initiation^{8,43,94,95}. More work is needed to determine whether this concept of disease initiation and progression may also apply to cases of sporadic ALS.

Human data support a role for angiogenic factors and vessels in the pathogenesis of ALS. For example, the presence of *VEGF* variations (which were identified in large meta-analysis studies) has been linked to ALS¹⁹¹. Angiogenin is another pro-angiogenic gene that is implicated in ALS because heterozygous missense mutations in angiogenin cause familial and sporadic ALS¹⁹². Moreover, mice with a mutation that eliminates hypoxia-responsive induction of the *Vegf* gene (*Vegf*^{δ/δ} mice) develop late-onset motor neuron degeneration¹⁹³. Spinal cord ischaemia worsens motor neuron degeneration and functional outcome in *Vegf*^{δ/δ} mice, whereas the absence of hypoxic induction of VEGF in mice that develop motor neuron disease from expression of ALS-linked mutant SOD1G93A results in substantially reduced survival¹⁹¹.

Therapeutic opportunities

Many investigators believe that primary neuronal dysfunction resulting from an intrinsic neuronal disorder is the key underlying event in human neurodegenerative diseases. Thus, most therapeutic efforts for neurodegenerative diseases have so far been directed at the development of so-called 'single-target, single-action' agents to target neuronal cells directly and reverse neuronal dysfunction and/or protect neurons from injurious insults. However, most preclinical and clinical studies have shown that such drugs are unable to cure or control human neurological disorders^{2,181,183,194,195}. For example, although pathological overstimulation of glutamatergic NMDA receptors (NMDARs) has been shown to lead to neuronal injury and death in several disorders, including stroke, Alzheimer's disease, ALS and Huntington's disease¹⁶, NMDAR antagonists have failed to show a therapeutic benefit in the above-mentioned human neurological disorders.

Recently, my colleagues and I coined the term vasculo-neuronal-inflammatory triad¹⁹⁵ to indicate that vascular damage, neuronal injury and/or neurodegeneration, and neuroinflammation comprise a common pathological triad that occurs in multiple neurological disorders. In line with this idea, it is conceivable that 'multiple-target, multiple-action' agents (that is, drugs that have more than one target and thus have more than one action) will have a better chance of controlling the complex disease mechanisms that mediate neurodegeneration than agents that have only one target (for example, neurons). According to the vasculo-neuronal-inflammatory triad model, in addition to neurons, brain endothelium, VSMCs, pericytes, astrocytes and activated microglia are all important therapeutic targets.

Here, I will briefly discuss a few therapeutic strategies based on the vasculo-neuronal-inflammatory triad model. VEGF and other angiogenic factors may have multiple targets, and thus multiple actions, in the CNS¹²⁰. For example, preclinical studies have shown that treatment of *SOD1^{G93A}* rats with intracerebroventricular VEGF¹⁹⁶ or intramuscular administration of a VEGF-expressing lentiviral vector that is transported retrogradely to motor neurons in *SOD1^{G93A}* mice¹⁹⁷ reduced pathology and extended survival, probably by promoting angiogenesis and increasing the blood flow through the spinal cord as well as through direct neuronal protective effects of VEGF on motor neurons. On the basis of these and other studies, a phase I–II clinical trial has been initiated to evaluate the safety of intracerebroventricular infusion of VEGF in patients with ALS¹⁹⁸. Treatment with angiogenin also slowed down disease progression in a mouse model of ALS¹⁹⁹.

IGF1 delivery has been shown to promote amyloid- β vascular clearance and to improve learning and memory in a mouse model of Alzheimer's disease²⁰⁰. Local intracerebral implantation of VEGF-secreting cells in a mouse model of Alzheimer's disease has been shown to enhance vascular repair, reduce amyloid burden and improve learning and memory²⁰¹. In contrast to VEGF, which can increase BBB permeability, TGF β , hepatocyte growth factor and fibroblast growth factor 2 promote BBB integrity by upregulating the expression of endothelial junction proteins¹²¹ in a similar way to APC⁴³. However, VEGF and most growth factors do not cross the BBB, so the development of delivery strategies such as Trojan horses is required for their systemic use²⁵.

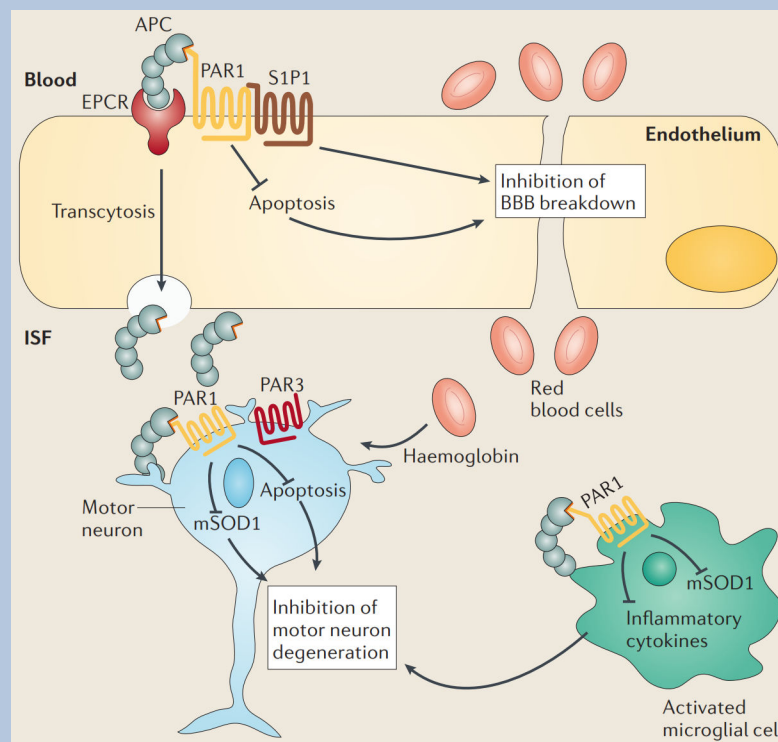
A recent experimental approach with APC provides an example of a neurovascular medicine that has been shown to favorably regulate multiple pathways in non-neuronal cells and neurons, resulting in vasculo-protection, stabilization of the BBB, neuroprotection and anti-inflammation in several acute and chronic models of the CNS disorders¹⁹⁵ (BOX 2).

Box 2

A model of multiple-target, multiple-action neurovascular medicine

Activated protein C (APC) is an endogenous plasma protease with antithrombotic, cytoprotective and anti-inflammatory activities in the CNS¹⁹⁵. APC and recombinant analogues of this protease that have been engineered to have reduced or no anticoagulant activity have excellent therapeutic potential as disease-modifying therapies for neuropathologies that are driven by the vasculo-neuronal-inflammatory triad and, hence, involve vascular damage with blood–brain barrier (BBB) or blood–spinal cord barrier (BSCB) breakdown, neuronal injury and/or neurodegeneration, and neuroinflammation. Although much remains to be learned with respect to the biology of APC, APC-mediated signaling within the injured neurovascular unit (NVU) has beneficial effects in experimental models of amyotrophic lateral sclerosis (ALS)⁴³, multiple sclerosis²¹², multiple models of stroke, spinal cord injury and brain trauma¹⁹⁵. Here, drawing on data from a transgenic *SOD1^{G93A}* mouse model of ALS, the multiple effects of APC on different cell types within the NVU in the spinal cord are shown⁴³. APC protects brain endothelium from divergent inducers of apoptosis by first binding to the endothelial protein C receptor (EPCR). Activation of this receptor is required for activation of proteinase-activated receptor 1 (PAR1), which in turn induces cell protective signaling in

the endothelium. PAR1 also cross-activates sphingosine 1 phosphate receptor 1 (S1P1), which enhances the integrity of endothelial barrier and inhibits BSCB breakdown⁹⁵. APC's action contributes to vascular repair of the NVU by ameliorating red blood cell extravasation, causing microhaemorrhages and inducing accumulation of neurotoxic reactive oxygen species in the spinal cord. EPCR also mediates transcytosis of APC across the BBB²⁸ and BSCB⁴³. Once in the spinal cord interstitial fluid (ISF), APC reaches motor neurons and exerts its direct neuronal protective activity by blocking the apoptotic pathways that are induced by divergent injurious stimuli, and by downregulating mutant superoxide dismutase 1 (mSOD1) expression. The effect of APC on neurons is mediated by PAR1 and PAR3 receptors. APC exerts its anti-inflammatory activity by activating PAR1 in microglia, which suppresses the activation of these cells and, therefore, inhibits the expression of inflammatory cytokines. PAR1 activation by APC also inhibits mSOD1 expression in microglia.



Box 2.
A model of multiple-target, multiple-action neurovascular medicine

The recognition of amyloid- β clearance pathways (FIG. 4), as discussed above, opens exciting new therapeutic opportunities for Alzheimer's disease. Amyloid- β clearance pathways are promising therapeutic targets for the future development of neurovascular medicines because it has been shown both in animal models of Alzheimer's disease¹ and in patients with sporadic Alzheimer's disease¹⁴⁹ that faulty clearance from brain and across the BBB primarily determines amyloid- β retention in brain, causing the formation of neurotoxic amyloid- β oligomers⁵⁶ and the promotion of brain and cerebrovascular amyloidosis³. The

targeting of clearance mechanisms might also be beneficial in other diseases; for example, the clearance of extracellular mutant SOD1 in familial ALS, the prion protein in prion disorders and α -synuclein in Parkinson's disease might all prove beneficial. However, the clearance mechanisms for these proteins in these diseases are not yet understood.

Conclusions and perspectives

Currently, no effective disease-modifying drugs are available to treat the major neurodegenerative disorders^{202–204}. This fact leads to a question: are we close to solving the mystery of neurodegeneration? The probable answer is yes, because the field has recently begun to recognize that, first, damage to neuronal cells is not the sole contributor to disease initiation and progression, and that second, correcting disease pathways in vascular and glial cells may offer an array of new approaches to control neuronal degeneration that do not involve targeting neurons directly. These realizations constitute an important shift in paradigm that should bring us closer to a cure for neurodegenerative diseases. Here, I raise some issues concerning the existing models of neurodegeneration and the new neurovascular paradigm.

The discovery of genetic abnormalities and associations by linkage analysis or DNA sequencing has broadened our understanding of neurodegeneration. However, insufficient effort has been made to link genetic findings with disease biology. Another concern for neurodegenerative research is how we should interpret findings from animal models²⁰². Genetically engineered models of human neurodegenerative disorders in *Drosophila melanogaster* and *Caenorhabditis elegans* have been useful for dissecting basic disease mechanisms and screening compounds. However, in addition to having much simpler nervous systems, insects and avascular species do not have cerebrovascular and immune systems that are comparable to humans and, therefore, are unlikely to replicate the complex disease pathology that is found in people.

For most neurodegenerative disorders, early steps in the disease processes remain unclear, and biomarkers for these stages have yet to be identified. Thus, it is difficult to predict whether mammalian models expressing human genes and proteins that we know are implicated in the intermediate or later stages of disease pathophysiology, such as amyloid- β or tau in Alzheimer's disease^{7,181}, will help us to discover therapies for the early stages of disease and for disease prevention, because the exact role of these pathological accumulations during disease onset remains uncertain. According to the two-hit vascular hypothesis of Alzheimer's disease, incorporating vascular factors that are associated with Alzheimer's disease into current models of this disease may more faithfully replicate dementia events in humans. Alternatively, by focusing on the comorbidities and the initial cellular and molecular mechanisms underlying early neurovascular dysfunction that are associated with Alzheimer's disease, new models of dementia and neurodegeneration may be developed that do not require the genetic manipulation of amyloid- β or tau expression.

The proposed neurovascular triad model of neurodegenerative diseases challenges the traditional neurocentric view of such disorders. At the same time, this model raises a set of new important issues that require further study. For example, the molecular basis of the

neurovascular link with neurodegenerative disorders is poorly understood, in terms of the adhesion molecules that keep the physical association of various cell types together, the molecular crosstalk between different cell types (including endothelial cells, pericytes and astrocytes) and how these cellular interactions influence neuronal activity. Addressing these issues promises to create new opportunities not only to better understand the molecular basis of the neurovascular link with neurodegeneration but also to develop novel neurovascular-based medicines.

The construction of a human BBB molecular atlas will be an important step towards understanding the role of the BBB and neurovascular interactions in health and disease. Achievement of this goal will require identifying new BBB transporters by using genomic and proteomic tools, and by cloning some of the transporters that are already known. Better knowledge of transporters at the human BBB will help us to better understand their potential as therapeutic targets for disease.

Development of higher-resolution imaging methods to evaluate BBB integrity, key transporters' functions and CBF responses in the microregions of interest (for example, in the entorhinal region of the hippocampus) will help us to understand how BBB dysfunction correlates with cognitive outcomes and neurodegenerative processes in MCI, Alzheimer's disease and related disorders.

The question persists: are we missing important therapeutic targets by studying the nervous system in isolation from the influence of the vascular system? The probable answer is yes. However, the current exciting and novel research that is based on the neurovascular model has already begun to change the way that we think about neurodegeneration, and will continue to provide further insights in the future, leading to the development of new neurovascular therapies.

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References

1. Zlokovic BV. The blood-brain barrier in health and chronic neurodegenerative disorders. *Neuron*. 2008; 57:178–201. [PubMed: 18215617]
2. Moskowitz MA, Lo EH, Iadecola C. The science of stroke: mechanisms in search of treatments. *Neuron*. 2010; 67:181–198. Comprehensive review describing mechanisms of ischemic injury to the neurovascular unit. [PubMed: 20670828]
3. Zlokovic BV. Neurovascular mechanisms of Alzheimer's neurodegeneration. *Trends Neurosci*. 2005; 28:202–208. [PubMed: 15808355]
4. Brown WR, Thore CR. Review: cerebral microvascular pathology in ageing and neurodegeneration. *Neuropathol Appl Neurobiol*. 2011; 37:56–74. [PubMed: 20946471]
5. Wu Z, et al. Role of the *MEOX2* homeobox gene in neurovascular dysfunction in Alzheimer disease. *Nat Med*. 2005; 11:959–965. Important study demonstrating that low expression of *MEOX2* gene in brain endothelium leads to aberrant angiogenesis and vascular regression in Alzheimer's disease. [PubMed: 16116430]

6. Paul J, Strickland S, Melchor JP. Fibrin deposition accelerates neurovascular damage and neuroinflammation in mouse models of Alzheimer's disease. *J Exp Med.* 2007; 204:1999–2008. Important study showing the blood-brain barrier breakdown in models of Alzheimer's disease. [PubMed: 17664291]
7. Zipser BD, et al. Microvascular injury and blood-brain barrier leakage in Alzheimer's disease. *Neurobiol Aging.* 2007; 28:977–986. [PubMed: 16782234]
8. Zhong Z, et al. ALS-causing SOD1 mutants generate vascular changes prior to motor neuron degeneration. *Nat Neurosci.* 2008; 11:420–422. Important study demonstrating the blood-spinal cord barrier defects precede motor neuron degeneration in mice developing an ALS-like disease. [PubMed: 18344992]
9. Kalaria RN. Vascular basis for brain degeneration: faltering controls and risk factors for dementia. *Nutr Rev.* 2010; 68 (Suppl 2):S74–87. [PubMed: 21091952]
10. Knopman DS, Roberts R. Vascular risk factors: imaging and neuropathologic correlates. *J Alzheimers Dis.* 2010; 20:699–709. [PubMed: 20182020]
11. Miyazaki K, et al. Disruption of neurovascular unit prior to motor neuron degeneration in amyotrophic lateral sclerosis. *J Neurosci Res.* 2011; 89:718–728. [PubMed: 21337372]
12. Neuwelt EA, et al. Engaging neuroscience to advance translational research in brain barrier biology. *Nat Rev Neurosci.* 2011; 12:169–182. [PubMed: 21331083]
13. Guo S, Lo EH. Dysfunctional cell-cell signaling in the neurovascular unit as a paradigm for central nervous system disease. *Stroke.* 2009; 40:S4–7. [PubMed: 19064781]
14. Redzic Z. Molecular biology of the blood-brain and the blood-cerebrospinal fluid barriers: similarities and differences. *Fluids Barriers CNS.* 2011; 8:3. [PubMed: 21349151]
15. O'Kane RL, Martinez-Lopez I, DeJoseph MR, Vina JR, Hawkins RA. Na(+)-dependent glutamate transporters (EAAT1, EAAT2, and EAAT3) of the blood-brain barrier. A mechanism for glutamate removal. *J Biol Chem.* 1999; 274:31891–31895. [PubMed: 10542215]
16. Hardingham GE. Coupling of the NMDA receptor to neuroprotective and neurodestructive events. *Biochem Soc Trans.* 2009; 37:1147–1160. [PubMed: 19909238]
17. Elali A, Hermann DM. ATP-Binding Cassette Transporters and Their Roles in Protecting the Brain. *Neuroscientist.* 2011
18. Visser WE, Friesema EC, Visser TJ. Minireview: thyroid hormone transporters: the knowns and the unknowns. *Mol Endocrinol.* 2011; 25:1–14. [PubMed: 20660303]
19. Zlokovic BV, Begley DJ, Chain-Eliash DG. Blood-brain barrier permeability to leucine-enkephalin, D-alanine2-D-leucine5-enkephalin and their N-terminal amino acid (tyrosine). *Brain Res.* 1985; 336:125–132. [PubMed: 3891014]
20. Zlokovic BV, Lipovac MN, Begley DJ, Davson H, Rakic L. Transport of leucine-enkephalin across the blood-brain barrier in the perfused guinea pig brain. *J Neurochem.* 1987; 49:310–315. [PubMed: 3585338]
21. Zlokovic BV, Mackic JB, Djuricic B, Davson H. Kinetic analysis of leucine-enkephalin cellular uptake at the luminal side of the blood-brain barrier of an in situ perfused guinea-pig brain. *J Neurochem.* 1989; 53:1333–1340. [PubMed: 2795003]
22. Zlokovic BV, et al. Kinetics of arginine-vasopressin uptake at the blood-brain barrier. *Biochim Biophys Acta.* 1990; 1025:191–198. [PubMed: 2364078]
23. Zlokovic BV, Segal MB, Begley DJ, Davson H, Rakic L. Permeability of the blood-cerebrospinal fluid and blood-brain barriers to thyrotropin-releasing hormone. *Brain Res.* 1985; 358:191–199. [PubMed: 3935272]
24. Dogrukol-Ak D, et al. Isolation of peptide transport system-6 from brain endothelial cells: therapeutic effects with antisense inhibition in Alzheimer and stroke models. *J Cereb Blood Flow Metab.* 2009; 29:411–422. [PubMed: 19002200]
25. Pardridge WM. Blood-brain barrier delivery. *Drug Discov Today.* 2007; 12:54–61. [PubMed: 17198973]
26. Nishijima T, et al. Neuronal activity drives localized blood-brain-barrier transport of serum insulin-like growth factor-I into the CNS. *Neuron.* 2010; 67:834–846. [PubMed: 20826314]
27. Banks WA. Blood-brain barrier as a regulatory interface. *Forum Nutr.* 2010; 63:102–110. [PubMed: 19955778]

28. Deane R, et al. Endothelial protein C receptor-assisted transport of activated protein C across the mouse blood-brain barrier. *J Cereb Blood Flow Metab.* 2009; 29:25–33. [PubMed: 18841163]
29. Iadecola C. Astrocytes take center stage in salt sensing. *Neuron.* 2007; 54:3–5. [PubMed: 17408570]
30. Shimizu H, et al. Glial Nax channels control lactate signaling to neurons for brain $[Na^+]$ sensing. *Neuron.* 2007; 54:59–72. [PubMed: 17408578]
31. Henkel JS, Beers DR, Wen S, Bowser R, Appel SH. Decreased mRNA expression of tight junction proteins in lumbar spinal cords of patients with ALS. *Neurology.* 2009; 72:1614–1616. [PubMed: 19414730]
32. Alvarez JI, Cayrol R, Prat A. Disruption of central nervous system barriers in multiple sclerosis. *Biochim Biophys Acta.* 2011; 1812:252–264. [PubMed: 20619340]
33. Bell RD, et al. Pericytes control key neurovascular functions and neuronal phenotype in the adult brain and during brain aging. *Neuron.* 2010; 68:409–427. Important study demonstrating that loss of pericytes leads to blood-brain barrier breakdown and hypoperfusion resulting in secondary neurodegenerative changes. [PubMed: 21040844]
34. Rosenberg GA. Matrix metalloproteinases and their multiple roles in neurodegenerative diseases. *Lancet Neurol.* 2009; 8:205–216. [PubMed: 19161911]
35. Cheng T, et al. Activated protein C inhibits tissue plasminogen activator-induced brain hemorrhage. *Nat Med.* 2006; 12:1278–1285. [PubMed: 17072311]
36. Daneman R, Zhou L, Kebede AA, Barres BA. Pericytes are required for blood-brain barrier integrity during embryogenesis. *Nature.* 2010; 468:562–566. Important study describing that pericytes control the formation of blood-brain barrier during embryonic development. [PubMed: 20944625]
37. Li F, et al. Endothelial Smad4 maintains cerebrovascular integrity by activating N-cadherin through cooperation with Notch. *Dev Cell.* 2011; 20:291–302. Important study demonstrating that N-cadherin mediates pericyte-endothelial attachment in the cerebral blood vessels preventing microhemorrhages. [PubMed: 21397841]
38. Armulik A, et al. Pericytes regulate the blood-brain barrier. *Nature.* 2010; 468:557–561. Important study demonstrating the role of pericytes in the maintenance of the blood-brain barrier *in vivo* during adulthood. [PubMed: 20944627]
39. Broadwell RD, Salzman M. Expanding the definition of the blood-brain barrier to protein. *Proc Natl Acad Sci U S A.* 1981; 78:7820–7824. [PubMed: 6950422]
40. Mhatre M, et al. Thrombin, a mediator of neurotoxicity and memory impairment. *Neurobiol Aging.* 2004; 25:783–793. [PubMed: 15165703]
41. Chen B, Cheng Q, Yang K, Lyden PD. Thrombin mediates severe neurovascular injury during ischemia. *Stroke.* 2010; 41:2348–2352. [PubMed: 20705928]
42. Chen ZL, Strickland S. Neuronal death in the hippocampus is promoted by plasmin-catalyzed degradation of laminin. *Cell.* 1997; 91:917–925. [PubMed: 9428515]
43. Zhong Z, et al. Activated protein C therapy slows ALS-like disease in mice by transcriptionally inhibiting SOD1 in motor neurons and microglia cells. *J Clin Invest.* 2009; 119:3437–3449. Important study demonstrating that activated protein C prevents the blood-spinal cord barrier breakdown, suppresses microglia activation and protects motor neurons in ALS mice. [PubMed: 19841542]
44. Simard JM, Kent TA, Chen M, Tarasov KV, Gerzanich V. Brain oedema in focal ischaemia: molecular pathophysiology and theoretical implications. *Lancet Neurol.* 2007; 6:258–268. [PubMed: 17303532]
45. Hoshi A, Yamamoto T, Shimizu K, Sugiura Y, Ugawa Y. Chemical preconditioning-induced reactive astrocytosis contributes to the reduction of post-ischemic edema through aquaporin-4 downregulation. *Exp Neurol.* 2011; 227:89–95. [PubMed: 20887723]
46. Iadecola C. Neurovascular regulation in the normal brain and in Alzheimer's disease. *Nat Rev Neurosci.* 2004; 5:347–360. [PubMed: 15100718]
47. Peppiatt CM, Howarth C, Mobbs P, Attwell D. Bidirectional control of CNS capillary diameter by pericytes. *Nature.* 2006; 443:700–704. Important study demonstrating that pericytes control the diameter of brain capillaries in response to signals from neurons. [PubMed: 17036005]

48. Yemisci M, et al. Pericyte contraction induced by oxidative-nitrative stress impairs capillary reflow despite successful opening of an occluded cerebral artery. *Nat Med.* 2009; 15:1031–1037. [PubMed: 19718040]
49. Kuchibhotla KV, Lattarulo CR, Hyman BT, Bacskai BJ. Synchronous hyperactivity and intercellular calcium waves in astrocytes in Alzheimer mice. *Science.* 2009; 323:1211–1215. [PubMed: 19251629]
50. Takano T, Han X, Deane R, Zlokovic B, Nedergaard M. Two-photon imaging of astrocytic Ca²⁺ signaling and the microvasculature in experimental mice models of Alzheimer's disease. *Ann N Y Acad Sci.* 2007; 1097:40–50. [PubMed: 17413008]
51. Smith CD, et al. Altered brain activation in cognitively intact individuals at high risk for Alzheimer's disease. *Neurology.* 1999; 53:1391–1396. [PubMed: 10534240]
52. Bookheimer SY, et al. Patterns of brain activation in people at risk for Alzheimer's disease. *N Engl J Med.* 2000; 343:450–456. [PubMed: 10944562]
53. Ruitenberg A, et al. Cerebral hypoperfusion and clinical onset of dementia: the Rotterdam Study. *Ann Neurol.* 2005; 57:789–794. [PubMed: 15929050]
54. Sheline YI, et al. APOE4 allele disrupts resting state fMRI connectivity in the absence of amyloid plaques or decreased CSF Aβ₄₂. *J Neurosci.* 2010; 30:17035–17040. [PubMed: 21159973]
55. Wang X, et al. Cerebrovascular hypoperfusion induces spatial memory impairment, synaptic changes, and amyloid-beta oligomerization in rats. *J Alzheimers Dis.* 2010; 21:813–822. [PubMed: 20634588]
56. Walsh DM, et al. Naturally secreted oligomers of amyloid beta protein potently inhibit hippocampal long-term potentiation in vivo. *Nature.* 2002; 416:535–539. Important study showing that amyloid-β oligomers inhibit neuronal activity in the hippocampus. [PubMed: 11932745]
57. Koike MA, Green KN, Blurton-Jones M, Laferla FM. Oligemic hypoperfusion differentially affects tau and amyloid-β. *Am J Pathol.* 2010; 177:300–310. [PubMed: 20472896]
58. Gordon-Krajcer W, Kozniowska E, Lazarewicz JW, Ksiazek-Reding H. Differential changes in phosphorylation of tau at PHF-1 and 12E8 epitopes during brain ischemia and reperfusion in gerbils. *Neurochem Res.* 2007; 32:729–737. [PubMed: 17191139]
59. Ongali B, et al. Transgenic mice overexpressing APP and transforming growth factor-beta1 feature cognitive and vascular hallmarks of Alzheimer's disease. *Am J Pathol.* 2010; 177:3071–3080. [PubMed: 21088218]
60. Sun X, et al. Hypoxia facilitates Alzheimer's disease pathogenesis by up-regulating BACE1 gene expression. *Proc Natl Acad Sci U S A.* 2006; 103:18727–18732. [PubMed: 17121991]
61. Zhang X, et al. Hypoxia-inducible factor 1α (HIF-1α)-mediated hypoxia increases BACE1 expression and beta-amyloid generation. *J Biol Chem.* 2007; 282:10873–10880. [PubMed: 17303576]
62. Guglielmo M, et al. The up-regulation of BACE1 mediated by hypoxia and ischemic injury: role of oxidative stress and HIF1α. *J Neurochem.* 2009; 108:1045–1056. [PubMed: 19196431]
63. Li L, et al. Hypoxia increases Aβ generation by altering beta- and gamma-cleavage of APP. *Neurobiol Aging.* 2009; 30:1091–1098. [PubMed: 18063223]
64. Fang H, Zhang LF, Meng FT, Du X, Zhou JN. Acute hypoxia promote the phosphorylation of tau via ERK pathway. *Neurosci Lett.* 2010; 474:173–177. [PubMed: 20304032]
65. Wang Z, et al. Hypoxia-induced down-regulation of neprilysin by histone modification in mouse primary cortical and hippocampal neurons. *PLoS One.* 2011; 6:e19229. [PubMed: 21559427]
66. Bell RD, et al. SRF and myocardin regulate LRP-mediated amyloid-beta clearance in brain vascular cells. *Nat Cell Biol.* 2009; 11:143–153. Important study demonstrating that hypoxia leads to a failure of LRP-1-mediated amyloid-β clearance from brain arteries via elevated levels of myocardin and serum response factor. [PubMed: 19098903]
67. Munch C, et al. Chemical hypoxia facilitates alternative splicing of EAAT2 in presymptomatic APP23 transgenic mice. *Neurochem Res.* 2008; 33:1005–1010. [PubMed: 17999180]
68. Boycott HE, Dallas M, Boyle JP, Pearson HA, Peers C. Hypoxia suppresses astrocyte glutamate transport independently of amyloid formation. *Biochem Biophys Res Commun.* 2007; 364:100–104. [PubMed: 17927959]

69. Carvalho C, et al. Role of mitochondrial-mediated signaling pathways in Alzheimer disease and hypoxia. *J Bioenerg Biomembr.* 2009; 41:433–440. [PubMed: 19830532]
70. Fernandez-Checa JC, et al. Oxidative stress and altered mitochondrial function in neurodegenerative diseases: lessons from mouse models. *CNS Neurol Disord Drug Targets.* 2010; 9:439–454. [PubMed: 20522012]
71. Correia SC, et al. Mitochondria: the missing link between preconditioning and neuroprotection. *J Alzheimers Dis.* 2010; 20 (Suppl 2):S475–485. [PubMed: 20463394]
72. Glass CK, Saijo K, Winner B, Marchetto MC, Gage FH. Mechanisms underlying inflammation in neurodegeneration. *Cell.* 2010; 140:918–934. [PubMed: 20303880]
73. Grammas P. Neurovascular dysfunction, inflammation and endothelial activation: implications for the pathogenesis of Alzheimer's disease. *J Neuroinflammation.* 2011; 8:26. [PubMed: 21439035]
74. Grammas P, Moore P, Weigel PH. Microvessels from Alzheimer's disease brains kill neurons in vitro. *Am J Pathol.* 1999; 154:337–342. [PubMed: 10027392]
75. Moser KV, Stockl P, Humpel C. Cholinergic neurons degenerate when exposed to conditioned medium of primary rat brain capillary endothelial cells: counteraction by NGF, MK-801 and inflammation. *Exp Gerontol.* 2006; 41:609–618. [PubMed: 16701975]
76. Yin X, Wright J, Wall T, Grammas P. Brain endothelial cells synthesize neurotoxic thrombin in Alzheimer's disease. *Am J Pathol.* 2010; 176:1600–1606. [PubMed: 20150433]
77. Martin AJ, Friston KJ, Colebatch JG, Frackowiak RS. Decreases in regional cerebral blood flow with normal aging. *J Cereb Blood Flow Metab.* 1991; 11:684–689. [PubMed: 2050757]
78. Li B, Freeman RD. Neurometabolic coupling in the lateral geniculate nucleus changes with extended age. *J Neurophysiol.* 2010; 104:414–425. [PubMed: 20463197]
79. Bertram L, McQueen MB, Mullin K, Blacker D, Tanzi RE. Systematic meta-analyses of Alzheimer disease genetic association studies: the AlzGene database. *Nat Genet.* 2007; 39:17–23. [PubMed: 17192785]
80. Kim J, Basak JM, Holtzman DM. The role of apolipoprotein E in Alzheimer's disease. *Neuron.* 2009; 63:287–303. [PubMed: 19679070]
81. Verghese PB, Castellano JM, Holtzman DM. Apolipoprotein E in Alzheimer's disease and other neurological disorders. *Lancet Neurol.* 2011; 10:241–252. [PubMed: 21349439]
82. Thambisetty M, Beason-Held L, An Y, Kraut MA, Resnick SM. APOE epsilon4 genotype and longitudinal changes in cerebral blood flow in normal aging. *Arch Neurol.* 2010; 67:93–98. [PubMed: 20065135]
83. Farrall AJ, Wardlaw JM. Blood-brain barrier: ageing and microvascular disease--systematic review and meta-analysis. *Neurobiol Aging.* 2009; 30:337–352. [PubMed: 17869382]
84. Topkian R, Barrick TR, Howe FA, Markus HS. Blood-brain barrier permeability is increased in normal-appearing white matter in patients with lacunar stroke and leucoaraiosis. *J Neurol Neurosurg Psychiatry.* 2010; 81:192–197. [PubMed: 19710048]
85. Chen RL, et al. Age-related changes in choroid plexus and blood-cerebrospinal fluid barrier function in the sheep. *Exp Gerontol.* 2009; 44:289–296. [PubMed: 19133323]
86. Farkas E, Luiten PG. Cerebral microvascular pathology in aging and Alzheimer's disease. *Prog Neurobiol.* 2001; 64:575–611. [PubMed: 11311463]
87. Savva GM, et al. Age, neuropathology, and dementia. *N Engl J Med.* 2009; 360:2302–2309. [PubMed: 19474427]
88. Jellinger KA. Prevalence and impact of cerebrovascular lesions in Alzheimer and lewy body diseases. *Neurodegener Dis.* 2010; 7:112–115. [PubMed: 20173339]
89. Cordonnier C. Brain microbleeds: more evidence, but still a clinical dilemma. *Curr Opin Neurol.* 2011; 24:69–74. [PubMed: 21124218]
90. Viswanathan A, Greenberg SM. Cerebral amyloid angiopathy (CAA) in the elderly. *Ann Neurol.* 2011; 10:225–236. [PubMed: 22516]
91. Fossati S, et al. Differential activation of mitochondrial apoptotic pathways by vasculotropic amyloid-beta variants in cells composing the cerebral vessel walls. *FASEB J.* 2010; 24:229–241. [PubMed: 19770225]

92. Rovelet-Lecrux A, et al. APP locus duplication causes autosomal dominant early-onset Alzheimer disease with cerebral amyloid angiopathy. *Nat Genet.* 2006; 38:24–26. [PubMed: 16369530]
93. Engelhardt JI, Appel SH. IgG reactivity in the spinal cord and motor cortex in amyotrophic lateral sclerosis. *Arch Neurol.* 1990; 47:1210–1216. [PubMed: 2122877]
94. Garbuzova-Davis S, et al. Evidence of compromised blood-spinal cord barrier in early and late symptomatic SOD1 mice modeling ALS. *PLoS One.* 2007; 2:e1205. [PubMed: 18030339]
95. Garbuzova-Davis S, et al. Amyotrophic lateral sclerosis: a neurovascular disease. *Brain Res.* 2011; 1398:113–125. [PubMed: 21632035]
96. Zhao C, Ling Z, Newman MB, Bhatia A, Carvey PM. TNF-alpha knockout and minocycline treatment attenuates blood-brain barrier leakage in MPTP-treated mice. *Neurobiol Dis.* 2007; 26:36–46. [PubMed: 17234424]
97. Chen X, Lan X, Roche I, Liu R, Geiger JD. Caffeine protects against MPTP-induced blood-brain barrier dysfunction in mouse striatum. *J Neurochem.* 2008; 107:1147–1157. [PubMed: 18808450]
98. Chao YX, He BP, Tay SS. Mesenchymal stem cell transplantation attenuates blood brain barrier damage and neuroinflammation and protects dopaminergic neurons against MPTP toxicity in the substantia nigra in a model of Parkinson's disease. *J Neuroimmunol.* 2009; 216:39–50. [PubMed: 19819031]
99. Elbaz A, Moisan F. Update in the epidemiology of Parkinson's disease. *Curr Opin Neurol.* 2008; 21:454–460. [PubMed: 18607207]
100. Bertrand E, et al. Amyloid angiopathy in idiopathic Parkinson's disease. Immunohistochemical and ultrastructural study. *Folia Neuropathol.* 2008; 46:255–270. [PubMed: 19169967]
101. Benamer HT, Grosset DG. Vascular parkinsonism: a clinical review. *Eur Neurol.* 2009; 61:11–15. [PubMed: 18948694]
102. Duran-Vilaregut J, et al. Blood-brain barrier disruption in the striatum of rats treated with 3-nitropropionic acid. *Neurotoxicology.* 2009; 30:136–143. [PubMed: 19026682]
103. Mooradian AD, Chung HC, Shah GN. GLUT-1 expression in the cerebra of patients with Alzheimer's disease. *Neurobiol Aging.* 1997; 18:469–474. [PubMed: 9390772]
104. Hunt A, et al. Reduced cerebral glucose metabolism in patients at risk for Alzheimer's disease. *Psychiatry Res.* 2007; 155:147–154. [PubMed: 17524628]
105. Herholz K. Cerebral glucose metabolism in preclinical and prodromal Alzheimer's disease. *Expert Rev Neurother.* 2010; 10:1667–1673. [PubMed: 20977325]
106. Mosconi L, et al. Hypometabolism exceeds atrophy in presymptomatic early-onset familial Alzheimer's disease. *J Nucl Med.* 2006; 47:1778–1786. [PubMed: 17079810]
107. Samuraki M, et al. Partial volume effect-corrected FDG PET and grey matter volume loss in patients with mild Alzheimer's disease. *Eur J Nucl Med Mol Imaging.* 2007; 34:1658–1669. [PubMed: 17520250]
108. Mosconi L, et al. Hippocampal hypometabolism predicts cognitive decline from normal aging. *Neurobiol Aging.* 2008; 29:676–692. [PubMed: 17222480]
109. Thomas T, Thomas G, McLendon C, Sutton T, Mullan M. beta-Amyloid-mediated vasoactivity and vascular endothelial damage. *Nature.* 1996; 380:168–171. Important study demonstrating that amyloid- β constricts blood vessels. [PubMed: 8600393]
110. Iadecola C, et al. SOD1 rescues cerebral endothelial dysfunction in mice overexpressing amyloid precursor protein. *Nat Neurosci.* 1999; 2:157–161. Important study showing dysregulation in the cerebral blood flow prior to amyloid- β deposition in Alzheimer's mice. [PubMed: 10195200]
111. Niwa K, et al. A β 1–40-related reduction in functional hyperemia in mouse neocortex during somatosensory activation. *Proc Natl Acad Sci U S A.* 2000; 97:9735–9740. [PubMed: 10944232]
112. Park L, et al. Scavenger receptor CD36 is essential for the cerebrovascular oxidative stress and neurovascular dysfunction induced by amyloid-beta. *Proc Natl Acad Sci U S A.* 2011; 108:5063–5068. [PubMed: 21383152]
113. Chow N, et al. Serum response factor and myocardin mediate arterial hypercontractility and cerebral blood flow dysregulation in Alzheimer's phenotype. *Proc Natl Acad Sci U S A.* 2007; 104:823–828. Important study demonstrating that elevated levels of myocardin and serum response factor lead to a hypercontractile phenotype of brain arteries in Alzheimer's disease. [PubMed: 17215356]

114. Bartels AL, et al. Blood-brain barrier P-glycoprotein function decreases in specific brain regions with aging: a possible role in progressive neurodegeneration. *Neurobiol Aging*. 2009; 30:1818–1824. [PubMed: 18358568]
115. Bartels AL, et al. Decreased blood-brain barrier P-glycoprotein function in the progression of Parkinson's disease, PSP and MSA. *J Neural Transm*. 2008; 115:1001–1009. [PubMed: 18265929]
116. Rule RR, Schuff N, Miller RG, Weiner MW. Gray matter perfusion correlates with disease severity in ALS. *Neurology*. 2010; 74:821–827. [PubMed: 20147656]
117. Harris GJ, et al. Reduced basal ganglia blood flow and volume in pre-symptomatic, gene-tested persons at-risk for Huntington's disease. *Brain*. 1999; 122 (Pt 9):1667–1678. [PubMed: 10468506]
118. Deckel AW, Duffy JD. Vasomotor hyporeactivity in the anterior cerebral artery during motor activation in Huntington's disease patients. *Brain Res*. 2000; 872:258–261. [PubMed: 10924705]
119. Greenberg DA, Jin K. From angiogenesis to neuropathology. *Nature*. 2005; 438:954–959. [PubMed: 16355213]
120. Ruiz de Almodovar C, Lambrechts D, Mazzone M, Carmeliet P. Role and therapeutic potential of VEGF in the nervous system. *Physiol Rev*. 2009; 89:607–648. [PubMed: 19342615]
121. Zacchigna S, Lambrechts D, Carmeliet P. Neurovascular signalling defects in neurodegeneration. *Nat Rev Neurosci*. 2008; 9:169–181. [PubMed: 18253131]
122. Lehtinen MK, et al. The cerebrospinal fluid provides a proliferative niche for neural progenitor cells. *Neuron*. 2011; 69:893–905. [PubMed: 21382550]
123. Paris D, et al. Impaired angiogenesis in a transgenic mouse model of cerebral amyloidosis. *Neurosci Lett*. 2004; 366:80–85. [PubMed: 15265595]
124. Chabriat H, Joutel A, Dichgans M, Tournier-Lasserre E, Boussier MG. Cadasil. *Lancet Neurol*. 2009; 8:643–653. [PubMed: 19539236]
125. Rotstein M, et al. Glut1 deficiency: inheritance pattern determined by haploinsufficiency. *Ann Neurol*. 2010; 68:955–958. [PubMed: 20687207]
126. Wang D, et al. A mouse model for Glut-1 haploinsufficiency. *Hum Mol Genet*. 2006; 15:1169–1179. [PubMed: 16497725]
127. Eisele YS, et al. Peripherally applied Abeta-containing inoculates induce cerebral beta-amyloidosis. *Science*. 2010; 330:980–982. Important study demonstrating that peripheral amyloid- β contributes to the development of cerebral β -amyloidosis in Alzheimer's mice. [PubMed: 20966215]
128. Sutcliffe JG, Hedlund PB, Thomas EA, Bloom FE, Hilbush BS. Peripheral reduction of beta-amyloid is sufficient to reduce brain beta-amyloid: Implications for Alzheimer's disease. *J Neurosci Res*. 2011; 89:808–814. [PubMed: 21374699]
129. Sagare AP, Winkler EA, Bell RD, Deane R, Zlokovic BV. From the liver to the blood-brain barrier: An interconnected system regulating brain amyloid-beta levels. *J Neurosci Res*. 2011; 89:967–968. [PubMed: 21544850]
130. Ujiie M, Dickstein DL, Carlow DA, Jefferies WA. Blood-brain barrier permeability precedes senile plaque formation in an Alzheimer disease model. *Microcirculation*. 2003; 10:463–470. [PubMed: 14745459]
131. Mackic JB, et al. Circulating amyloid-beta peptide crosses the blood-brain barrier in aged monkeys and contributes to Alzheimer's disease lesions. *Vascul Pharmacol*. 2002; 38:303–313. [PubMed: 12529925]
132. Mackic JB, et al. Cerebrovascular accumulation and increased blood-brain barrier permeability to circulating Alzheimer's amyloid beta peptide in aged squirrel monkey with cerebral amyloid angiopathy. *J Neurochem*. 1998; 70:210–215. [PubMed: 9422364]
133. Poduslo JF, Curran GL, Haggard JJ, Biere AL, Selkoe DJ. Permeability and residual plasma volume of human, Dutch variant, and rat amyloid beta-protein 1–40 at the blood-brain barrier. *Neurobiol Dis*. 1997; 4:27–34. [PubMed: 9258909]
134. Ghilardi JR, et al. Intra-arterial infusion of [125I]A beta 1–40 labels amyloid deposits in the aged primate brain in vivo. *Neuroreport*. 1996; 7:2607–2611. [PubMed: 8981432]

135. Zlokovic BV, et al. Blood-brain barrier transport of circulating Alzheimer's amyloid beta. *Biochem Biophys Res Commun.* 1993; 197:1034–1040. [PubMed: 8280117]
136. Martel CL, Mackic JB, McComb JG, Ghiso J, Zlokovic BV. Blood-brain barrier uptake of the 40 and 42 amino acid sequences of circulating Alzheimer's amyloid beta in guinea pigs. *Neurosci Lett.* 1996; 206:157–160. [PubMed: 8710175]
137. Sagare A, et al. Clearance of amyloid-beta by circulating lipoprotein receptors. *Nat Med.* 2007; 13:1029–1031. Important study showing that soluble LRP1 binds amyloid- β in the circulation preventing its reentry into the brain. [PubMed: 17694066]
138. DeMattos RB, Bales KR, Cummins DJ, Paul SM, Holtzman DM. Brain to plasma amyloid-beta efflux: a measure of brain amyloid burden in a mouse model of Alzheimer's disease. *Science.* 2002; 295:2264–2267. Important study showing that a circulating anti-amyloid- β antibody promotes efflux of this peptide from brain to blood. [PubMed: 11910111]
139. Sigurdsson EM, Scholtzova H, Mehta PD, Frangione B, Wisniewski T. Immunization with a nontoxic/nonfibrillar amyloid-beta homologous peptide reduces Alzheimer's disease-associated pathology in transgenic mice. *Am J Pathol.* 2001; 159:439–447. [PubMed: 11485902]
140. DeMattos RB, et al. Plaque-associated disruption of CSF and plasma amyloid-beta (A β) equilibrium in a mouse model of Alzheimer's disease. *J Neurochem.* 2002; 81:229–236. [PubMed: 12064470]
141. Matsuoka Y, et al. Novel therapeutic approach for the treatment of Alzheimer's disease by peripheral administration of agents with an affinity to beta-amyloid. *J Neurosci.* 2003; 23:29–33. [PubMed: 12514198]
142. Liu Y, et al. Expression of neprilysin in skeletal muscle reduces amyloid burden in a transgenic mouse model of Alzheimer disease. *Mol Ther.* 2009; 17:1381–1386. [PubMed: 19471248]
143. Liu Y, et al. Circulating neprilysin clears brain amyloid. *Mol Cell Neurosci.* 2010; 45:101–107. [PubMed: 20558294]
144. Deane R, et al. RAGE mediates amyloid-beta peptide transport across the blood-brain barrier and accumulation in brain. *Nat Med.* 2003; 9:907–913. Important study demonstrating that RAGE mediates influx of amyloid- β across the blood-brain barrier. [PubMed: 12808450]
145. Mackic JB, et al. Human blood-brain barrier receptors for Alzheimer's amyloid-beta 1–40. Asymmetrical binding, endocytosis, and transcytosis at the apical side of brain microvascular endothelial cell monolayer. *J Clin Invest.* 1998; 102:734–743. [PubMed: 9710442]
146. Giri R, et al. beta-amyloid-induced migration of monocytes across human brain endothelial cells involves RAGE and PECAM-1. *Am J Physiol Cell Physiol.* 2000; 279:C1772–1781. [PubMed: 11078691]
147. Yan SD, et al. RAGE and amyloid-beta peptide neurotoxicity in Alzheimer's disease. *Nature.* 1996; 382:685–691. [PubMed: 8751438]
148. Yan SF, Ramasamy R, Schmidt AM. The RAGE axis: a fundamental mechanism signaling danger to the vulnerable vasculature. *Circ Res.* 2010; 106:842–853. [PubMed: 20299674]
149. Mawuenyega KG, et al. Decreased clearance of CNS beta-amyloid in Alzheimer's disease. *Science.* 2010; 330:1774. Important study demonstrating faulty amyloid- β clearance from the brain in patients affected by Alzheimer's disease. [PubMed: 21148344]
150. Zlokovic BV, Deane R, Sagare AP, Bell RD, Winkler EA. Low-density lipoprotein receptor-related protein-1: a serial clearance homeostatic mechanism controlling Alzheimer's amyloid beta-peptide elimination from the brain. *J Neurochem.* 2010; 115:1077–1089. [PubMed: 20854368]
151. Deane R, et al. LRP/amyloid beta-peptide interaction mediates differential brain efflux of A β isoforms. *Neuron.* 2004; 43:333–344. [PubMed: 15294142]
152. Shibata M, et al. Clearance of Alzheimer's amyloid-ss(1–40) peptide from brain by LDL receptor-related protein-1 at the blood-brain barrier. *J Clin Invest.* 2000; 106:1489–1499. Pioneering study demonstrating that LRP-1 mediates amyloid- β clearance from the brain to blood across the blood-brain barrier. [PubMed: 11120756]
153. Bell RD, et al. Transport pathways for clearance of human Alzheimer's amyloid beta-peptide and apolipoproteins E and J in the mouse central nervous system. *J Cereb Blood Flow Metab.* 2007; 27:909–918. [PubMed: 17077814]

154. Jaeger LB, et al. Testing the neurovascular hypothesis of Alzheimer's disease: LRP-1 antisense reduces blood-brain barrier clearance, increases brain levels of amyloid-beta protein, and impairs cognition. *J Alzheimers Dis.* 2009; 17:553–570. [PubMed: 19433890]
155. Shinohara M, et al. Reduction of brain beta-amyloid (Abeta) by fluvastatin, a hydroxymethylglutaryl-CoA reductase inhibitor, through increase in degradation of amyloid precursor protein C-terminal fragments (APP-CTFs) and Abeta clearance. *J Biol Chem.* 2010; 285:22091–22102. [PubMed: 20472556]
156. Jaeger LB, et al. Lipopolysaccharide alters the blood-brain barrier transport of amyloid beta protein: a mechanism for inflammation in the progression of Alzheimer's disease. *Brain Behav Immun.* 2009; 23:507–517. [PubMed: 19486646]
157. Yamada K, et al. The low density lipoprotein receptor-related protein 1 mediates uptake of amyloid beta peptides in an in vitro model of the blood-brain barrier cells. *J Biol Chem.* 2008; 283:34554–34562. [PubMed: 18940800]
158. Nazer B, Hong S, Selkoe DJ. LRP promotes endocytosis and degradation, but not transcytosis, of the amyloid-beta peptide in a blood-brain barrier in vitro model. *Neurobiol Dis.* 2008; 30:94–102. [PubMed: 18289866]
159. Monro OR, et al. Substitution at codon 22 reduces clearance of Alzheimer's amyloid-beta peptide from the cerebrospinal fluid and prevents its transport from the central nervous system into blood. *Neurobiol Aging.* 2002; 23:405–412. [PubMed: 11959403]
160. Davis J, et al. Early-onset and robust cerebral microvascular accumulation of amyloid beta-protein in transgenic mice expressing low levels of a vasculotropic Dutch/Iowa mutant form of amyloid beta-protein precursor. *J Biol Chem.* 2004; 279:20296–20306. [PubMed: 14985348]
161. Deane R, et al. apoE isoform-specific disruption of amyloid beta peptide clearance from mouse brain. *J Clin Invest.* 2008; 118:4002–4013. [PubMed: 19033669]
162. DeMattos RB, et al. ApoE and clusterin cooperatively suppress Abeta levels and deposition: evidence that ApoE regulates extracellular Abeta metabolism in vivo. *Neuron.* 2004; 41:193–202. [PubMed: 14741101]
163. DeMattos RB, et al. Clusterin promotes amyloid plaque formation and is critical for neuritic toxicity in a mouse model of Alzheimer's disease. *Proc Natl Acad Sci U S A.* 2002; 99:10843–10848. [PubMed: 12145324]
164. Bading JR, et al. Brain clearance of Alzheimer's amyloid-beta40 in the squirrel monkey: a SPECT study in a primate model of cerebral amyloid angiopathy. *J Drug Target.* 2002; 10:359–368. [PubMed: 12164385]
165. Donahue JE, et al. RAGE, LRP-1, and amyloid-beta protein in Alzheimer's disease. *Acta Neuropathol.* 2006; 112:405–415. [PubMed: 16865397]
166. Cirrito JR, et al. P-glycoprotein deficiency at the blood-brain barrier increases amyloid-beta deposition in an Alzheimer disease mouse model. *J Clin Invest.* 2005; 115:3285–3290. [PubMed: 16239972]
167. Owen JB, et al. Oxidative modification to LDL receptor-related protein 1 in hippocampus from subjects with Alzheimer disease: implications for Abeta accumulation in AD brain. *Free Radic Biol Med.* 2010; 49:1798–1803. [PubMed: 20869432]
168. Behl M, et al. Lead-induced accumulation of beta-amyloid in the choroid plexus: role of low density lipoprotein receptor protein-1 and protein kinase C. *Neurotoxicology.* 2010; 31:524–532. [PubMed: 20488202]
169. Sagare AP, et al. Impaired Lipoprotein Receptor-Mediated Peripheral Binding of Plasma Amyloid-beta is an Early Biomarker for Mild Cognitive Impairment Preceding Alzheimer's Disease. *J Alzheimers Dis.* 2011; 24:25–34. [PubMed: 21157031]
170. Tamaki C, et al. Major involvement of low-density lipoprotein receptor-related protein 1 in the clearance of plasma free amyloid beta-peptide by the liver. *Pharm Res.* 2006; 23:1407–1416. [PubMed: 16779710]
171. Iwata N, et al. Metabolic regulation of brain Abeta by neprilysin. *Science.* 2001; 292:1550–1552. [PubMed: 11375493]

172. Qiu WQ, Folstein MF. Insulin, insulin-degrading enzyme and amyloid-beta peptide in Alzheimer's disease: review and hypothesis. *Neurobiol Aging*. 2006; 27:190–198. [PubMed: 16399206]
173. Melchor JP, Pawlak R, Strickland S. The tissue plasminogen activator-plasminogen proteolytic cascade accelerates amyloid-beta (A β) degradation and inhibits A β -induced neurodegeneration. *J Neurosci*. 2003; 23:8867–8871. [PubMed: 14523088]
174. Yin KJ, et al. Matrix metalloproteinases expressed by astrocytes mediate extracellular amyloid-beta peptide catabolism. *J Neurosci*. 2006; 26:10939–10948. [PubMed: 17065436]
175. Koistinaho M, et al. Apolipoprotein E promotes astrocyte colocalization and degradation of deposited amyloid-beta peptides. *Nat Med*. 2004; 10:719–726. [PubMed: 15195085]
176. Bacskai BJ, et al. Non-Fc-mediated mechanisms are involved in clearance of amyloid-beta in vivo by immunotherapy. *J Neurosci*. 2002; 22:7873–7878. [PubMed: 12223540]
177. Hickman SE, Allison EK, El Khoury J. Microglial dysfunction and defective beta-amyloid clearance pathways in aging Alzheimer's disease mice. *J Neurosci*. 2008; 28:8354–8360. [PubMed: 18701698]
178. Weller RO, Subash M, Preston SD, Mazanti I, Carare RO. Perivascular drainage of amyloid-beta peptides from the brain and its failure in cerebral amyloid angiopathy and Alzheimer's disease. *Brain Pathol*. 2008; 18:253–266. [PubMed: 18363936]
179. Brody DL, et al. Amyloid-beta dynamics correlate with neurological status in the injured human brain. *Science*. 2008; 321:1221–1224. [PubMed: 18755980]
180. Querfurth HW, LaFerla FM. Alzheimer's disease. *N Engl J Med*. 2010; 362:329–344. [PubMed: 20107219]
181. Hardy J. The amyloid hypothesis for Alzheimer's disease: a critical reappraisal. *J Neurochem*. 2009; 110:1129–1134. [PubMed: 19457065]
182. Lagier-Tourenne C, Cleveland DW. Neurodegeneration: An expansion in ALS genetics. *Nature*. 2010; 466:1052–1053. [PubMed: 20740002]
183. Ilieva H, Polyimenidou M, Cleveland DW. Non-cell autonomous toxicity in neurodegenerative disorders: ALS and beyond. *J Cell Biol*. 2009; 187:761–772. [PubMed: 19951898]
184. Elden AC, et al. Ataxin-2 intermediate-length polyglutamine expansions are associated with increased risk for ALS. *Nature*. 2010; 466:1069–1075. [PubMed: 20740007]
185. Gruzman A, et al. Common molecular signature in SOD1 for both sporadic and familial amyotrophic lateral sclerosis. *Proc Natl Acad Sci U S A*. 2007; 104:12524–12529. [PubMed: 17636119]
186. Boillee S, et al. Onset and progression in inherited ALS determined by motor neurons and microglia. *Science*. 2006; 312:1389–1392. Important study demonstrating that toxicity of a ALS-linked SOD1 mutant to microglia determines the life span in mice with this disease. [PubMed: 16741123]
187. Yamanaka K, et al. Astrocytes as determinants of disease progression in inherited amyotrophic lateral sclerosis. *Nat Neurosci*. 2008; 11:251–253. [PubMed: 18246065]
188. Beers DR, et al. Wild-type microglia extend survival in PU.1 knockout mice with familial amyotrophic lateral sclerosis. *Proc Natl Acad Sci U S A*. 2006; 103:16021–16026. [PubMed: 17043238]
189. Di Giorgio FP, Carrasco MA, Siao MC, Maniatis T, Eggan K. Non-cell autonomous effect of glia on motor neurons in an embryonic stem cell-based ALS model. *Nat Neurosci*. 2007; 10:608–614. [PubMed: 17435754]
190. Nagai M, et al. Astrocytes expressing ALS-linked mutated SOD1 release factors selectively toxic to motor neurons. *Nat Neurosci*. 2007; 10:615–622. [PubMed: 17435755]
191. Lambrechts D, et al. VEGF is a modifier of amyotrophic lateral sclerosis in mice and humans and protects motoneurons against ischemic death. *Nat Genet*. 2003; 34:383–394. [PubMed: 12847526]
192. Greenway MJ, et al. ANG mutations segregate with familial and 'sporadic' amyotrophic lateral sclerosis. *Nat Genet*. 2006; 38:411–413. [PubMed: 16501576]

193. Oosthuysen B, et al. Deletion of the hypoxia-response element in the vascular endothelial growth factor promoter causes motor neuron degeneration. *Nat Genet.* 2001; 28:131–138. [PubMed: 11381259]
194. Mangialasche F, Solomon A, Winblad B, Mecocci P, Kivipelto M. Alzheimer's disease: clinical trials and drug development. *Lancet Neurol.* 2010; 9:702–716. [PubMed: 20610346]
195. Zlokovic BV, Griffin JH. Cytoprotective protein C pathways and implications for stroke and neurological disorders. *Trends Neurosci.* 2011; 34:198–209. [PubMed: 21353711]
196. Storkebaum E, et al. Treatment of motoneuron degeneration by intracerebroventricular delivery of VEGF in a rat model of ALS. *Nat Neurosci.* 2005; 8:85–92. [PubMed: 15568021]
197. Azzouz M, et al. VEGF delivery with retrogradely transported lentivector prolongs survival in a mouse ALS model. *Nature.* 2004; 429:413–417. [PubMed: 15164063]
198. A safety and tolerability study of intracerebroventricular administration of sNN0029 to patients with amyotrophic lateral sclerosis [online]. 2011. <http://clinicaltrials.gov/ct2/show/NCT00800501>
199. Kieran D, et al. Control of motoneuron survival by angiogenin. *J Neurosci.* 2008; 28:14056–14061. [PubMed: 19109488]
200. Lopez-Lopez C, Dietrich MO, Metzger F, Loetscher H, Torres-Aleman I. Disturbed cross talk between insulin-like growth factor I and AMP-activated protein kinase as a possible cause of vascular dysfunction in the amyloid precursor protein/presenilin 2 mouse model of Alzheimer's disease. *J Neurosci.* 2007; 27:824–831. [PubMed: 17251422]
201. Spuch C, et al. The effect of encapsulated VEGF-secreting cells on brain amyloid load and behavioral impairment in a mouse model of Alzheimer's disease. *Biomaterials.* 2010; 31:5608–5618. [PubMed: 20430437]
202. Jucker M. The benefits and limitations of animal models for translational research in neurodegenerative diseases. *Nat Med.* 2010; 16:1210–1214. [PubMed: 21052075]
203. Lo EH. Degeneration and repair in central nervous system disease. *Nat Med.* 2010; 16:1205–1209. [PubMed: 21052074]
204. Van Broeckhoven C. The future of genetic research on neurodegeneration. *Nat Med.* 2010; 16:1215–1217. [PubMed: 21052076]
205. de la Torre JC. Vascular risk factor detection and control may prevent Alzheimer's disease. *Ageing Res Rev.* 2010; 9:218–225. [PubMed: 20385255]
206. Luchsinger JA, et al. Relation of diabetes to mild cognitive impairment. *Arch Neurol.* 2007; 64:570–575. [PubMed: 17420320]
207. Iadecola C, Davisson RL. Hypertension and cerebrovascular dysfunction. *Cell Metab.* 2008; 7:476–484. [PubMed: 18522829]
208. Whitmer RA, et al. Central obesity and increased risk of dementia more than three decades later. *Neurology.* 2008; 71:1057–1064. [PubMed: 18367704]
209. Marchesi VT. Alzheimer's dementia begins as a disease of small blood vessels, damaged by oxidative-induced inflammation and dysregulated amyloid metabolism: implications for early detection and therapy. *FASEB J.* 2011; 25:5–13. [PubMed: 21205781]
210. Vermeer SE, et al. Silent brain infarcts and the risk of dementia and cognitive decline. *N Engl J Med.* 2003; 348:1215–1222. [PubMed: 12660385]
211. Snowdon DA, et al. Brain infarction and the clinical expression of Alzheimer disease. The Nun Study. *JAMA.* 1997; 277:813–817. [PubMed: 9052711]
212. Han MH, et al. Proteomic analysis of active multiple sclerosis lesions reveals therapeutic targets. *Nature.* 2008; 451:1076–1081. [PubMed: 18278032]

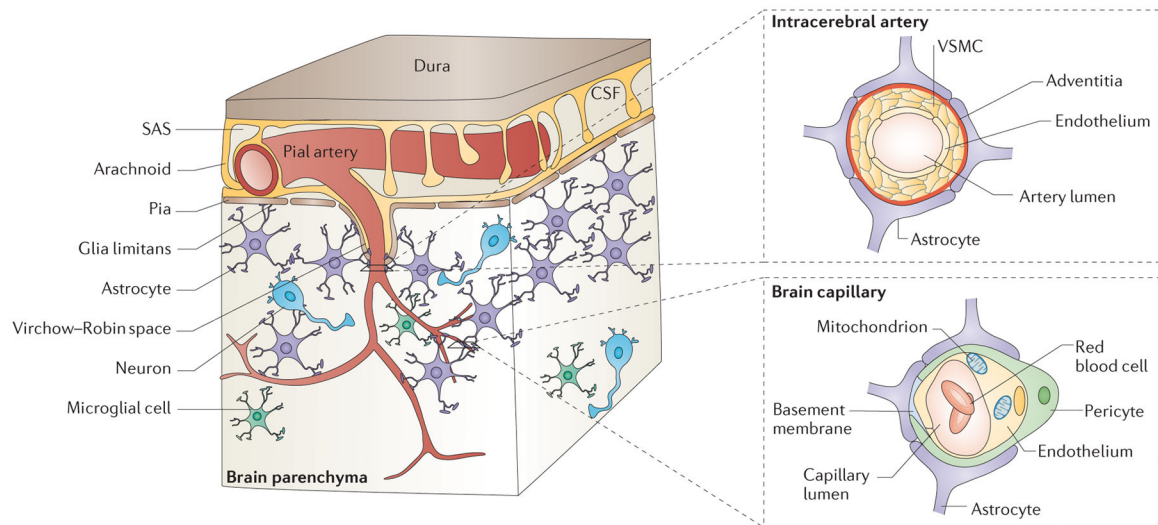


Figure 1. Cerebral microcirculation and the neurovascular unit

In the brain, pial arteries run through the subarachnoid space (SAS), which contains the cerebrospinal fluid (CSF). These vessels give rise to intracerebral arteries, which penetrate into brain parenchyma. Intracerebral arteries are separated from brain parenchyma by a single, interrupted layer of elongated fibroblast-like cells of the pia and the astrocyte-derived glia limitans membrane that forms the outer wall of the perivascular Virchow–Robin space. These arteries branch into smaller arteries and subsequently arterioles, which lose support from the glia limitans and give rise to pre-capillary arterioles and brain capillaries. In an intracerebral artery, the vascular smooth muscle cell (VSMC) layer occupies most of the vessel wall. At the brain capillary level, vascular endothelial cells and pericytes are attached to the basement membrane. Pericyte processes encase most of the capillary wall, and they communicate with endothelial cells directly through synapse-like contacts containing connexins and N-cadherin. Astrocyte end-foot processes encase the capillary wall, which is composed of endothelium and pericytes. Resting microglia have a ‘ramified’ shape and can sense neuronal injury.

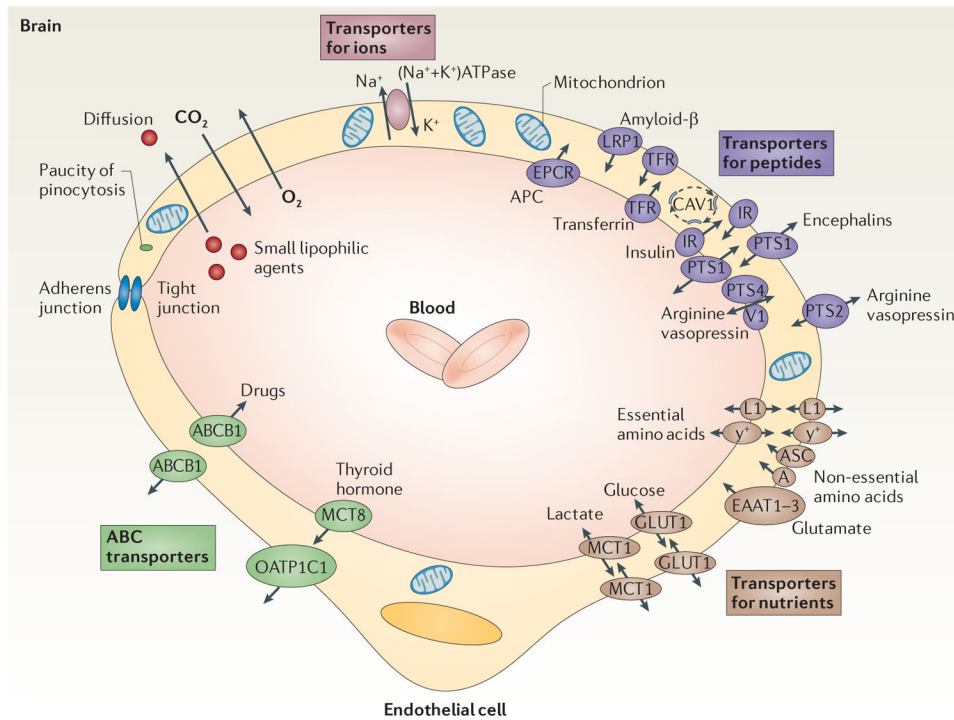


Figure 2. Blood–brain barrier transport mechanisms

Small lipophilic drugs, oxygen and carbon dioxide diffuse across the blood–brain barrier (BBB), whereas ions require ATP-dependent transporters such as the $(\text{Na}^+\text{K}^+)\text{ATPase}$. Transporters for nutrients include the glucose transporter 1 (GLUT1; also known as solute carrier family 2, facilitated glucose transporter member 1 (SLC2A1)), the lactate transporter monocarboxylate transporter 1 (MCT1) and the L1 and γ^+ transporters for large neutral and cationic essential amino acids, respectively. These four transporters are expressed at both the luminal and abluminal membranes. Non-essential amino acid transporters (the alanine, serine and cysteine preferring system (ASC), and the alanine preferring system (A)) and excitatory amino acid transporter 1 (EAAT1), EAAT2 and EAAT3 are located at the abluminal side. The ATP-binding cassette (ABC) efflux transporters that are found in the endothelial cells include multidrug resistance protein 1 (ABCB1; also known as ATP-binding cassette subfamily B member 1) and solute carrier organic anion transporter family member 1C1 (OATP1C1). Finally, transporters for peptides or proteins include the endothelial protein C receptor (EPCR) for activated protein C (APC); the insulin receptors (IRs) and the transferrin receptors (TFRs), which are associated with caveolin 1 (CAV1); low-density lipoprotein receptor-related protein 1 (LRP1) for amyloid- β , peptide transport system 1 (PTS1) for enkephalins; and the PTS2 and PTS4–vasopressin V1a receptor (V1AR) for arginine vasopressin.

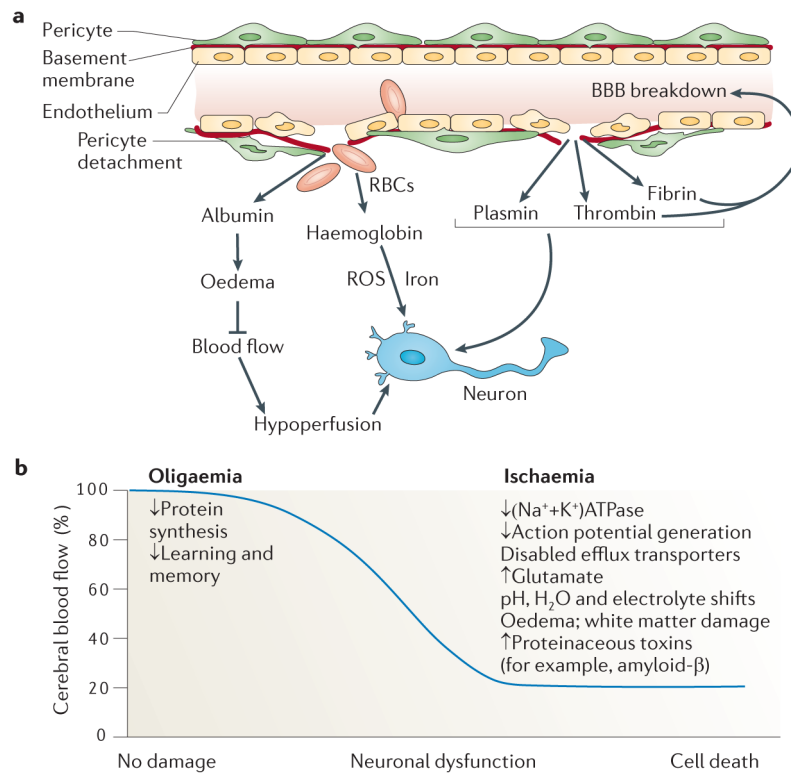


Figure 3. Vascular-mediated neuronal damage and neurodegeneration

a, Blood–brain barrier (BBB) breakdown that is caused by pericyte detachment leads to leakage of serum proteins and focal microhaemorrhages, with extravasation of red blood cells (RBCs). RBCs release haemoglobin, which is a source of iron. In turn, this metal catalyses the formation of toxic reactive oxygen species (ROS) that mediate neuronal injury. Albumin promotes the development of vasogenic oedema, contributing to hypoperfusion and hypoxia of the nervous tissue, which aggravates neuronal injury. A defective BBB allows several potentially vasculotoxic and neurotoxic proteins (for example, thrombin, fibrin and plasmin) to enter the brain. **b**, Progressive reductions in cerebral blood flow (CBF) lead to increasing neuronal dysfunction. Mild hypoperfusion, oligoemia, leads to a decrease in protein synthesis, whereas more-severe reductions in CBF, leading to hypoxia, cause an array of detrimental effects.

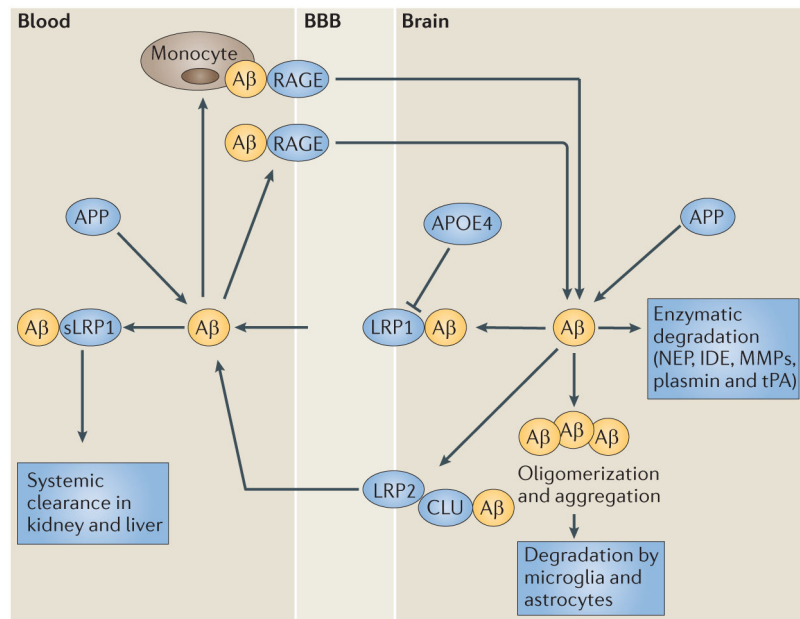


Figure 4. The role of blood–brain barrier transport in brain homeostasis of amyloid- β

Amyloid- β ($A\beta$) is produced from the amyloid- β precursor protein (APP), both in the brain and in peripheral tissues. Clearance of amyloid- β from the brain normally maintains its low levels in the brain. This peptide is cleared across the blood–brain barrier (BBB) by the low-density lipoprotein receptor-related protein 1 (LRP1). LRP1 mediates rapid efflux of a free, unbound form of amyloid- β and of amyloid- β bound to apolipoprotein E2 (APOE2), APOE3 or α 2-macroglobulin (not shown) from the brain's interstitial fluid into the blood, and APOE4 inhibits such transport. LRP2 eliminates amyloid- β that is bound to clusterin (CLU; also known as apolipoprotein J (APOJ)) by transport across the BBB, and shows a preference for the 42-aminoacid form of this peptide. ATP-binding cassette subfamily A member 1 (ABCA1; also known as cholesterol efflux regulatory protein) mediates amyloid- β efflux from the brain endothelium to blood across the luminal side of the BBB (not shown). Cerebral endothelial cells, pericytes, vascular smooth muscle cells, astrocytes, microglia and neurons express different amyloid- β -degrading enzymes, including neprilysin (NEP), insulin-degrading enzyme (IDE), tissue plasminogen activator (tPA) and matrix metalloproteinases (MMPs), which contribute to amyloid- β clearance. In the circulation, amyloid- β is bound mainly to soluble LRP1 (sLRP1), which normally prevents its entry into the brain. Systemic clearance of amyloid- β is mediated by its removal by the liver and kidneys. The receptor for advanced glycation end products (RAGE) provides the key mechanism for influx of peripheral amyloid- β into the brain across the BBB either as a free, unbound plasma-derived peptide and/or by amyloid- β -laden monocytes. Faulty vascular clearance of amyloid- β from the brain and/or an increased re-entry of peripheral amyloid- β across the blood vessels into the brain can elevate amyloid- β levels in the brain parenchyma and around cerebral blood vessels. At pathophysiological concentrations, amyloid- β forms neurotoxic oligomers and also self-aggregates, which leads to the development of cerebral β -amyloidosis and cerebral amyloid angiopathy.

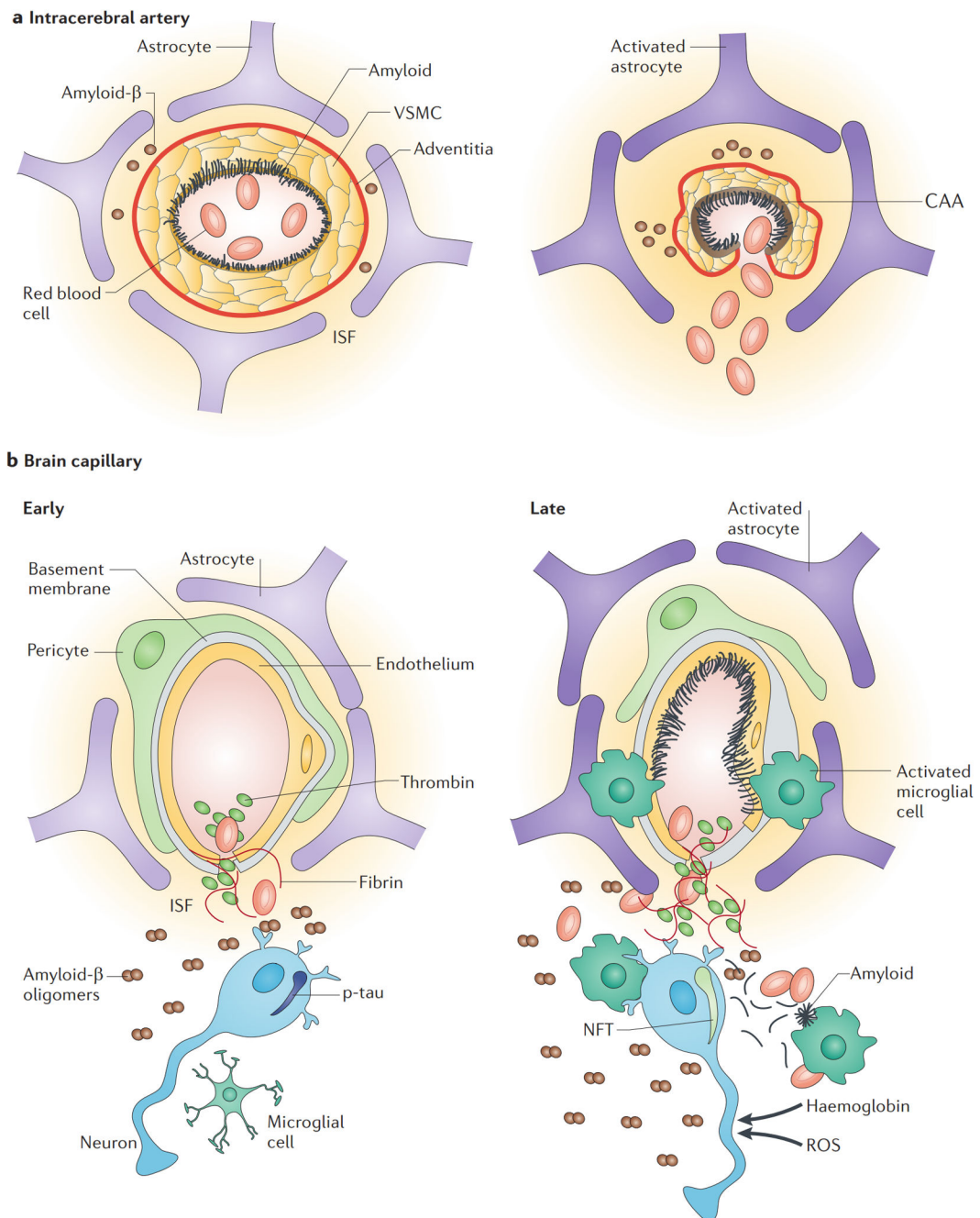


Figure 5. A model of vascular damage in Alzheimer's disease

a, In the early stages of Alzheimer's disease, small pial and intracerebral arteries develop a hypercontractile phenotype that underlies dysregulated cerebral blood flow (CBF). This phenotype is accompanied by diminished amyloid- β clearance by the vascular smooth muscle cells (VSMCs). In the later phases of Alzheimer's disease, amyloid deposition in the walls of intracerebral arteries leads to cerebral amyloid angiopathy (CAA), pronounced reductions in CBF, atrophy of the VSMC layer and rupture of the vessels causing microbleeds. **b,** At the level of capillaries in the early stages of Alzheimer's disease, blood-

brain barrier (BBB) dysfunction leads to a faulty amyloid- β clearance and accumulation of neurotoxic amyloid- β oligomers in the interstitial fluid (ISF), microhaemorrhages and accumulation of toxic blood-derived molecules (that is, thrombin and fibrin), which affect synaptic and neuronal function. Hyperphosphorylated tau (p-tau) accumulates in neurons in response to hypoperfusion and/or rising amyloid- β levels. At this point, microglia begin to sense neuronal injury. In the later stages of the disease in brain capillaries, microvascular degeneration leads to increased deposition of basement membrane proteins and perivascular amyloid. The deposited proteins and amyloid obstruct capillary blood flow, resulting in failure of the efflux pumps, accumulation of metabolic waste products, changes in pH and electrolyte composition and, subsequently, synaptic and neuronal dysfunction. Neurofibrillary tangles (NFTs) accumulate in response to ischaemic injury and rising amyloid- β levels. Activation of microglia and astrocytes is associated with a pronounced inflammatory response. ROS, reactive oxygen species.