

Complete Genome Sequence of the Attenuated Novobiocin-Resistant *Streptococcus iniae* Vaccine Strain ISNO

Julia W. Pridgeon,^a Dunhua Zhang,^a Lee Zhang^b

Aquatic Animal Health Research Unit, USDA, Auburn, Alabama, USA^a; Genomics and Sequencing Laboratory, Department of Entomology and Plant Pathology, Auburn University, Auburn, Alabama, USA^b

***Streptococcus iniae* ISNO is an attenuated novobiocin-resistant vaccine strain. Its full genome is 2,070,182 bp in length. The availability of this genome will allow comparative genomics to identify potential virulence genes important for pathogenesis of *S. iniae* and potential mechanisms associated with novobiocin resistance in this strain.**

Received 6 May 2014 Accepted 13 May 2014 Published 29 May 2014

Citation Pridgeon JW, Zhang D, Zhang L. 2014. Complete genome sequence of the attenuated novobiocin-resistant *Streptococcus iniae* vaccine strain ISNO. *Genome Announc.* 2(3):e00510-14. doi:10.1128/genomeA.00510-14.

Copyright © 2014 Pridgeon et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 3.0 Unported license](https://creativecommons.org/licenses/by/3.0/).

Address correspondence to Julia W. Pridgeon, Julia.Pridgeon@ars.usda.gov.

The Gram-positive bacterium *Streptococcus iniae* is a zoonotic pathogen that causes infections in both humans and fish (1–3). As a serious marine and freshwater fish pathogen, *S. iniae* causes significant economic losses to aquaculture (4, 5). This pathogen was originally isolated from Amazon freshwater dolphin (*Inia geoffrensis*) in 1976 (6). However, *S. iniae* causes diseases not only to dolphin, but also to many fish species, including rainbow trout (*Oncorhynchus mykiss* [7]), barramundi (*Lates calcarifer* [8]), red drum (*Sciaenops ocellatus* [9]), flounder (*Paralichthys* spp. [10, 11]), and tilapia (*Oreochromis* spp. [12]). A highly virulent strain, *S. iniae* ISET0901, was cultured from diseased Nile tilapia (*Oreochromis niloticus*) during a disease outbreak in 2005 (13). Through selection for resistance of *S. iniae* ISET0901 to novobiocin, an attenuated vaccine strain, *S. iniae* ISNO, was obtained (13). The vaccine strain *S. iniae* ISNO was found to be totally safe to Nile tilapia at various exposure doses and in back passage safety studies (13). In addition, the vaccine strain *S. iniae* ISNO offered Nile tilapia significant protection against *S. iniae* infections at a wide range of efficacious immunization doses (13). Furthermore, the vaccine strain *S. iniae* ISNO provided tilapia significant protection up to 6 months after a single vaccination (13). Compared to the virulent parent *S. iniae* ISET0901, the vaccine strain *S. iniae* ISNO was ~1,000-fold resistant to novobiocin. However, what changes occurred at the genomic DNA level were unknown. Therefore, the complete genome sequence of *S. iniae* ISNO was determined in this study.

The genome of *S. iniae* ISNO was sequenced using the Illumina 1500 HiSeq platform. BioNumerics (Applied Maths) was used to assemble a total of 15,690,266 sequence reads with an average length of 92.32 bp (estimated 700× coverage) using the complete genome of *S. iniae* SF1 (GenBank accession no. CP005941) as reference. The assembled genome of *S. iniae* ISNO is 2,070,182 bp, with G+C content of 36.8%. RNAmmer (14) predicted 12 copies of ribosomal RNA (4 copies of 5S RNA, 16S RNA, and 23S RNA, respectively), similar to reference genome *S. iniae* SF1 (15). The RAST server (16) predicted 1,980 coding sequences belonging to 303 subsystems, including 291 involved in carbohydrate catabo-

lism, 149 in protein metabolism, 135 in synthesis of amino acids and derivatives, 114 in cell wall and capsule synthesis, 95 in RNA metabolism, and 92 in DNA metabolism (including 77 in cofactors, vitamins, prosthetic groups, or pigments; 63 in nucleoside and nucleotide synthesis; 62 in fatty acid and lipid synthesis; 52 in virulence, disease, and defense; 47 in membrane transport; 37 in stress response; 31 in phosphorus metabolism; 26 in regulation and cell signaling; 7 in secondary metabolism; and 2 in motility and chemotaxis).

Nucleotide sequence accession number. The complete genome sequence of *S. iniae* ISNO was deposited at GenBank under the accession no. CP007587.

ACKNOWLEDGMENTS

This study was supported by the USDA/ARS CRIS project no. 6420-32000-024-00D.

The use of trade, firm, or corporate names in this publication is for the information and convenience of the reader. Such use does not constitute an official endorsement or approval by the U.S. Department of Agriculture or the Agricultural Research Service of any product or service to the exclusion of others that may be suitable.

We thank James Becnel (USDA-ARS) and Laura Silo-Suh (Mercer University) for critical reviews of the manuscript. We thank Beth Peterman (USDA-ARS) for her technical support.

REFERENCES

1. Sun JR, Yan JC, Yeh CY, Lee SY, Lu JJ. 2007. Invasive infection with *Streptococcus iniae* in Taiwan. *J. Med. Microbiol.* 56:1246–1249. <http://dx.doi.org/10.1099/jmm.0.47180-0>.
2. Lau SK, Woo PC, Luk WK, Fung AM, Hui WT, Fong AH, Chow CW, Wong SS, Yuen KY. 2006. Clinical isolates of *Streptococcus iniae* from Asia are more mucoid and beta-hemolytic than those from North America. *Diagn. Microbiol. Infect. Dis.* 54:177–181. <http://dx.doi.org/10.1016/j.diagmicrobio.2005.09.012>.
3. Miller JD, Neely MN. 2005. Large-scale screen highlights the importance of capsule for virulence in the zoonotic pathogen *Streptococcus iniae*. *Infect. Immun.* 73:921–934. <http://dx.doi.org/10.1128/IAI.73.2.921-934.2005>.
4. Agnew W, Barnes AC. 2007. *Streptococcus iniae*: an aquatic pathogen of global veterinary significance and a challenging candidate for reliable vac-

- cination. *Vet. Microbiol.* 122:1–15. <http://dx.doi.org/10.1016/j.vetmic.2007.03.002>.
5. El Aamri F, Caballero MJ, Real F, Acosta F, Déniz S, Román L, Padilla D. 4 February 2014. *Streptococcus iniae* in gilthead seabream (*Sparus aurata*, L.) and red porgy (*Pagrus pagrus*, L.): ultrastructural analysis. *Vet. Pathol.* <http://dx.doi.org/10.1177/0300985814520638>.
 6. Pier GB, Madin SH. 1976. *Streptococcus iniae* sp. nov., a beta-hemolytic *Streptococcus* isolated from an Amazon freshwater dolphin, *Inia geoffrensis*. *Int. J. Syst. Bacteriol.* 26:545–553. <http://dx.doi.org/10.1099/00207713-26-4-545>.
 7. Eynogor M, Tekoah Y, Shapira R, Hurvitz A, Zlotkin A, Lublin A, Eldar A. 2008. Emergence of novel *Streptococcus iniae* exopolysaccharide-producing strains following vaccination with nonproducing strains. *Appl. Environ. Microbiol.* 74:6892–6868. <http://dx.doi.org/10.1128/AEM.00853-08>.
 8. Bromage ES, Thomas A, Owens L. 1999. *Streptococcus iniae*, a bacterial infection in barramundi *Lates calcarifer*. *Dis. Aquat. Organ* 36:177–181. <http://dx.doi.org/10.3354/dao036177>.
 9. Eldar A, Perl S, Frelief PF, Bercovier H. 1999. Red drum *Sciaenops ocellatus* mortalities associated with *Streptococcus iniae* infection. *Dis. Aquat. Organ* 36:121–127. <http://dx.doi.org/10.3354/dao036121>.
 10. Nho SW, Shin GW, Park SB, Jang HB, Cha IS, Ha MA, Kim YR, Park YK, Dalvi RS, Kang BJ, Joh SJ, Jung TS. 2009. Phenotypic characteristics of *Streptococcus iniae* and *Streptococcus parauberis* isolated from olive flounder (*Paralichthys olivaceus*). *FEMS Microbiol. Lett.* 293:20–27. <http://dx.doi.org/10.1111/j.1574-6968.2009.01491.x>.
 11. Nguyen HT, Kanai K. 1999. Selective agars for the isolation of *Streptococcus iniae* from Japanese flounder, *Paralichthys olivaceus*, and its cultural environment. *J. Appl. Microbiol.* 86:769–776. <http://dx.doi.org/10.1046/j.1365-2672.1999.00724.x>.
 12. Zhou SM, Xie MQ, Zhu XQ, Ma Y, Tan ZL, Li AX. 2008. Identification and genetic characterization of *Streptococcus iniae* strains isolated from diseased fish in China. *J. Fish Dis.* 31:869–875. <http://dx.doi.org/10.1111/j.1365-2761.2008.00954.x>.
 13. Pridgeon JW, Klesius PH. 2011. Development and efficacy of a novobiocin-resistant *Streptococcus iniae* as a novel vaccine in Nile tilapia (*Oreochromis niloticus*). *Vaccine* 29:5986–5993. <http://dx.doi.org/10.1016/j.vaccine.2011.06.036>.
 14. Lagesen K, Hallin P, Rødland EA, Staerfeldt HH, Rognes T, Ussery DW. 2007. RNAmmer: consistent and rapid annotation of ribosomal RNA genes. *Nucleic Acids Res.* 35:3100–3108. <http://dx.doi.org/10.1093/nar/gkm160>.
 15. Zhang BC, Zhang J, Sun L. 2014. *Streptococcus iniae* SF1: complete genome sequence, proteomic profile, and immunoprotective antigens. *PLoS One* 9:e91324. <http://dx.doi.org/10.1371/journal.pone.0091324>.
 16. Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formisma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST Server: rapid annotations using subsystems technology. *BMC Genomics* 9:75. <http://dx.doi.org/10.1186/1471-2164-9-75>.