

# $^{68}\text{Ga}$ -Labeled Radiopharmaceuticals for Positron Emission Tomography

Dinesh Shetty · Yun-Sang Lee · Jae Min Jeong

Received: 21 September 2010 / Accepted: 24 September 2010 / Published online: 12 October 2010  
© Korean Society of Nuclear Medicine 2010

**Abstract**  $^{68}\text{Ga}$  is a promising emerging radionuclide for positron emission tomography (PET). It is produced using a  $^{68}\text{Ge}/^{68}\text{Ga}$ -generator, and thus, would enable the cyclotron-independent distribution of PET. However, new  $^{68}\text{Ga}$ -labeled radiopharmaceuticals that can replace  $^{18}\text{F}$ -labeled agents like [ $^{18}\text{F}$ ]fluorodeoxyglucose (FDG) are needed. Most of the  $^{68}\text{Ga}$ -labeled derivatives currently used are peptide agents, but the developments of other agents, such as amino acid derivatives, nitroimidazole derivatives, and glycosylated human serum albumin, are being actively pursued in many laboratories. Thus, appearance of new  $^{68}\text{Ga}$ -labeled radiopharmaceuticals with high impact are expected in the near future. Here, we present an overview of  $^{68}\text{Ga}$ -labeled agents in terms of their clinical significances and relevances to the management of certain tumors, and pertinent pre-clinical developments.

**Keywords** Gallium-68 · PET · Peptide · DOTA · NOTA · BAPEN · MSA · Ga-68

## Introduction

The introduction of  $^{68}\text{Ga}$  positron emission tomography (PET) to clinical practice represents a developmental milestone in the functional and metabolic imaging fields,

and was facilitated by the cyclotron-independent availability of  $^{68}\text{Ga}$ , enabled by the use of the  $^{68}\text{Ge}/^{68}\text{Ga}$  radionuclide generator system [1, 2].  $^{68}\text{Ga}$  is an excellent positron emitter. It has the characteristics of low photon emission (1,077 keV, 3.22%) and 89% positron branching [3]. Recent studies have shown that some  $^{68}\text{Ga}$ -labeled peptides exhibited distinctly better images than their  $^{111}\text{In}$ -labeled analogues [4–6] and than  $^{18}\text{F}$ -based radiotracers [7]. Unlike other PET radioisotopes, like  $^{18}\text{F}$  or  $^{11}\text{C}$ , ionic  $\text{Ga}^{3+}$  cannot be bound covalently to targeting vectors but must be conjugated to a target vector using a bifunctional chelating agent (BCA). Nevertheless, the labeling can be done just prior to diagnostic examinations, rapidly with minimum loss of radioactivity. The only stable chemical form of Ga in solution at physiological conditions is  $\text{Ga}^{3+}$ , and this ion can form stable complexes with chelators that are either free or conjugated with macromolecules or small organic molecules [8].

There are two requirements for using gallium complexes as radiopharmaceuticals: (1) they should be resistant to hydrolysis (the formation of complexes with  $\text{OH}^-$ ) and (2) they should be more stable than the Ga(III)–transferrin complex, and thus, the labeled gallium complex must be stable in the presence of transferrin—a plasma protein. The large formation constant of Ga(III)–transferrin ( $\log K=20.3$ ) [9] and the high plasma concentration of this protein (0.25 g/100 ml) favor the thermodynamic exchange of Ga (III) complexes with transferrin in vivo, and thus, the majority of radioactive gallium complexes used as radiopharmaceuticals have high thermodynamic and kinetic stabilities.

Various  $^{68}\text{Ga}$ -labeled radiopharmaceuticals have been developed by conjugating BCAs to peptides, proteins, or

D. Shetty · Y.-S. Lee · J. M. Jeong (✉)  
Department of Nuclear Medicine, Institute of Radiation Medicine,  
Seoul National University College of Medicine,  
101 Daehangno Jongno-gu,  
Seoul 110-744, South Korea  
e-mail: jmjng@snu.ac.kr

small biological molecules *via* active esters, isothiocyanates, maleimides, hydrazides, or haloamides.  $^{68}\text{Ga}$ -1,4,7,10-tetra-azacyclododecane-1,4,7,10-tetraacetic acid–Tyr3-octreotide (DOTA-TOC),  $^{68}\text{Ga}$ -DOTA-1-Nal-octreotide (DOTA-NOC),  $^{68}\text{Ga}$ -DOTA-bombesin,  $^{68}\text{Ga}$ -1,4,7-triazacyclononanetriacetic acid (NOTA)-RGD,  $^{68}\text{Ga}$ -DOTA-albumin, and  $^{68}\text{Ga}$ -DOTA-human epidermal growth factor (hEGF) are examples of such agents (Fig. 1) [8, 10–15]. Similarly, some agents, such as  $^{68}\text{Ga}$ -[(4,6-MeO<sub>2</sub>sal)<sub>2</sub>-BAPEN]<sup>+</sup> and  $^{68}\text{Ga}$ -N<sub>2</sub>S<sub>2</sub>, are chelates of radioactive gallium and used for myocardial imaging [16–18].

Acyclic BCAs, such as ethylenediaminetetraacetic acid (EDTA), diethylenetriaminepentaacetic acid (DTPA), desferrioxamine (DFO), N,N'-di(2-hydroxybenzyl)ethylenediamine-N,N'-diacetic acid (HBED), and their derivatives, have been used for labeling macromolecules with <sup>111</sup>In, <sup>67</sup>Zn, <sup>68</sup>Ga, or <sup>90</sup>Y for tumor imaging and therapy [19–22]. However, most of these complexes have low *in vivo* and *in vitro* stabilities due to their tendency to undergo acid- or cation-promoted dissociation [23, 24]. These limitations are overcome by using macrocyclic BCAs, such as NOTA, DOTA, or 1,4,8,11-tetraazacyclotetradecanetetraacetic acid (TETA), which can form highly stable complexes with these radiometals (Fig. 2) [23, 25].

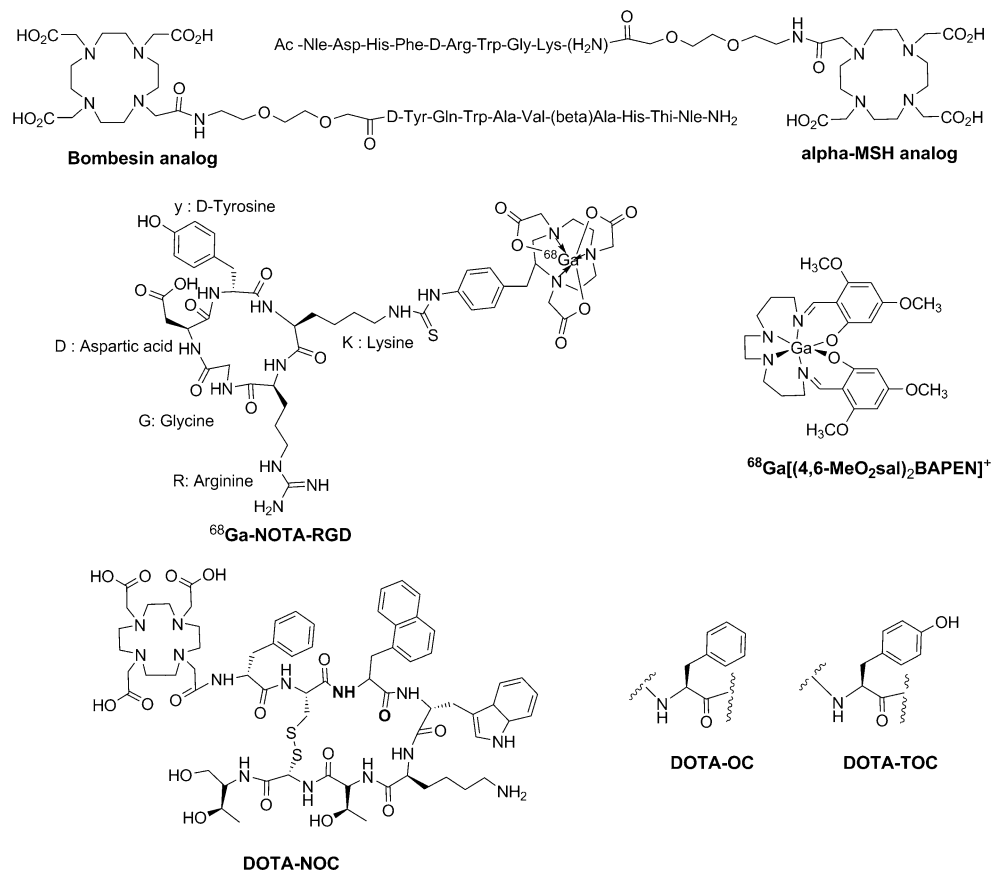
In this review, we present overviews of  $^{68}\text{Ga}$ -labeled agents in terms of their present clinical significances and their relevances to the management of certain tumors, and we include details of recent pre-clinical developments.

## $^{68}\text{Ga}$ -Peptides

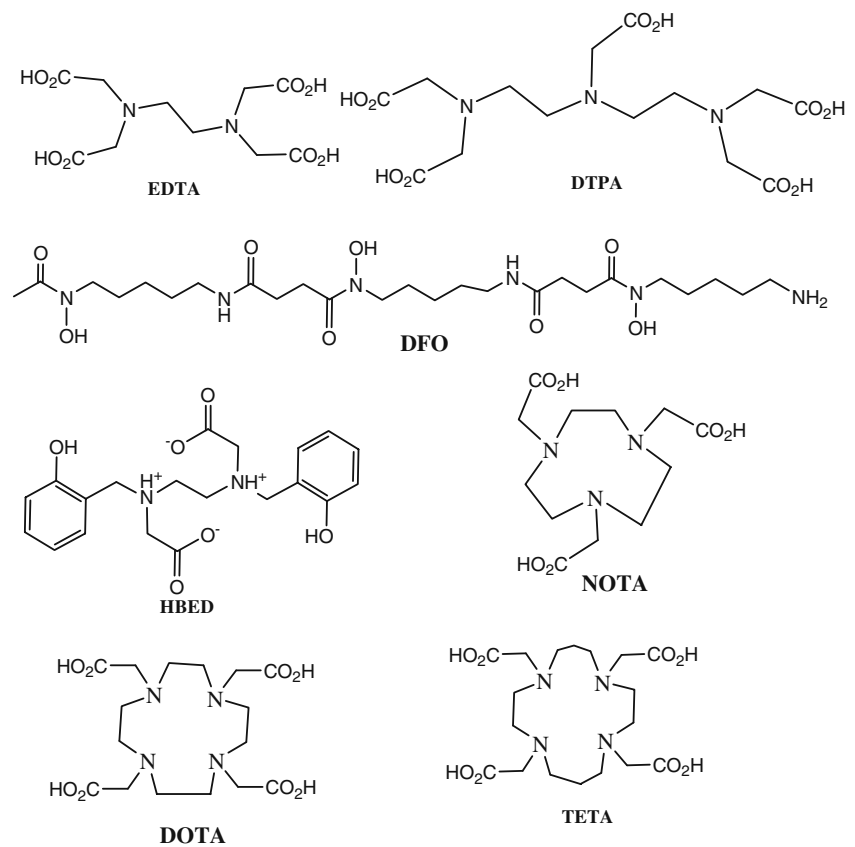
During the last decade, the developments of easy and economical production routes for radiolabeled peptides with rapid clearance and tissue penetration and low antigenicity, and the availabilities of simplified purification methods promoted their developments for diagnostic applications. Furthermore, the use of BCAs enabled peptides to be easily labeled with therapeutic radionuclides (<sup>90</sup>Y, <sup>177</sup>Lu).

Most of the early efforts made to label peptides targeted somatostatin and its derivatives. Somatostatin is a regulatory peptide and its action is mediated by membrane-bound receptors (SSTRs) that are present in normal human tissues, such as, in thyroid, brain, the gastrointestinal tract (GIT), pancreas, spleen, and kidneys [26], and they are also highly expressed in many different types of human tumors, notably neuroendocrine tumors (NET) [27], which in

**Fig. 1** Chemical structures of  $^{68}\text{Ga}$ -labeled radiopharmaceuticals



**Fig. 2** Some open chain and macrocyclic BCAs used for  $^{68}\text{Ga}$ -radiopharmaceutical synthesis



clinical practice are usually carcinoid tumors and pheochromocytomas. SSTRs are also expressed, to variable extents, in renal cell carcinoma, small cell lung cancer, breast cancer, prostate cancer, and in malignant lymphoma [4]. There are five SSTR subtypes, but subtype 2 (SSTR2), subtype 5 (SSTR5), and to a lesser extent, subtype 3 (SSTR3) have higher affinities than SSTR1 and 4, and thus, commercially available synthetic somatostatin analogues target these three high-affinity receptors [28]. These analogues are required because somatostatin is rapidly degraded by enzymes in vivo, as reflected by its short biological half-life, and thus, agents with high affinity for SSTR have been developed, which are resistant to enzyme degradation.

Somatostatin analogues, such as, DOTA-TOC show better images than  $^{111}\text{In}$ -DTPA-octreotide, the most commonly used somatostatin analogue [29]. The phenylalanine residue at position 3 was replaced by tyrosine in DOTA-TOC, which makes the compound more hydrophilic, increases affinity for SSTR2, and increases uptake by SSTR2-positive tumors [30]. Other peptides have also been linked to DOTA, such as, DOTA-octreotate, which has high affinity for SSTR2 [28], and DOTA-lanreotide, which has high affinity for SSTR5. DOTA-NOC is the newest addition to these compounds, and has high affinity for SSTR2, SSTR3, and SSTR5. Furthermore, these DOTA-peptide products show high radiochemical purity, rapid renal clearance, and high

accumulation in tumors, and overall represent remarkable advances over standard peptides [31].

In parallel with the development of the clinical applications of  $^{68}\text{Ga}$ -labeled compounds, in vitro and animal testing of various chelators and compounds is on-going. Antunes et al. [4] demonstrated that gallium  $^{67}\text{Ga}$ - and  $^{68}\text{Ga}$ -DOTA-octapeptides have distinctly better pre-clinical pharmacological performances than  $^{111}\text{In}$ -labeled peptides, especially on SSTR2-expressing cells and in animal models. In particular,  $^{68}\text{Ga}$ -DFO-octreotide injected into rats bearing SSTR-positive pancreatic tumors demonstrated selective binding to tumor sites with a tumor to background ratio (T/B) of 5 [32]. Subsequently, several DOTA-SST analogues were evaluated in vivo, and  $^{68}\text{Ga}$ -DOTA-TOC and  $^{68}\text{Ga}$ -DOTA-NOC were found to be the most promising [33–36].

We previously found that the PET imaging agent,  $^{68}\text{Ga}$ -NOTA-RGD, can be used to visualize angiogenesis in ischemic tissue, and  $\alpha_v\beta_3$  integrin expression was found to play an important role [13]. Because angiogenesis is known to occur at ischemic lesions during cancer development, we used two animal biodistribution models, a hind limb ischemia and a SNU-C4 (a human colon cancer cell line) xenograft model. Significantly, higher tracer uptakes were observed in ischemic compared with nonischemic muscle tissues. Small-animal PET of mice bearing SNU-C4 xenografts injected with this tracer at 1 and 2 h post-

injection with or without cold c(RGDyK) showed specific uptake of tracer by tumors (Fig. 3). Furthermore, in a subsequent biodistribution study, tumor uptake of  $^{68}\text{Ga}$ -NOTA-RGD was  $5.2 \pm 1.0\%$  ID/g, and its tumor-to-blood ratio was  $10.4 \pm 4.8$ .

$^{68}\text{Ga}$  has also been successfully used to label melanocortin peptides, which have many physiologic functions; their receptors are expressed in several cell types, such as, cutaneous melanocytes, keratinocytes, fibroblasts, endothelial cells, antigen-presenting cells, and leukocytes. Melanoma is one of the tumors that can be successfully imaged with radiolabeled MSH, because melanoma overexpresses melanocortin receptor. In particular,  $^{68}\text{Ga}$ -DOTA-rhenium-cyclized alphanelanocyte-stimulating hormone (alpha-MSH) analogue [DOTA-ReCCMSH (Arg11)] is a promising agent for the early detection of melanoma in mice [37]. Likewise,  $^{68}\text{Ga}$ -DOTA-NAPamide, a short linear alpha-MSH analogue, has been reported to be superior to  $^{111}\text{In}$ -DOTA-MSH for the targeting of melanocortin type 1

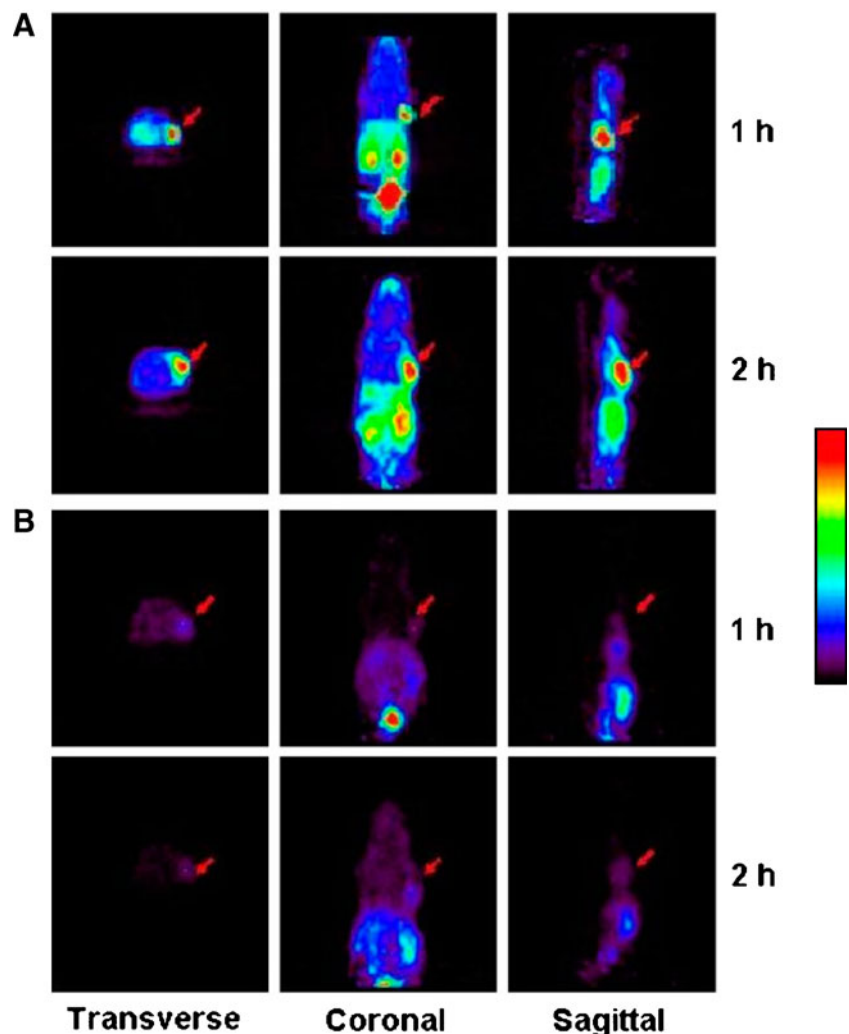
receptor in murine models of primary and metastatic melanoma [38]. However, receptor density in human melanoma is much lower than in murine tumor models, and thus, more work is needed to improve receptor affinity in man.

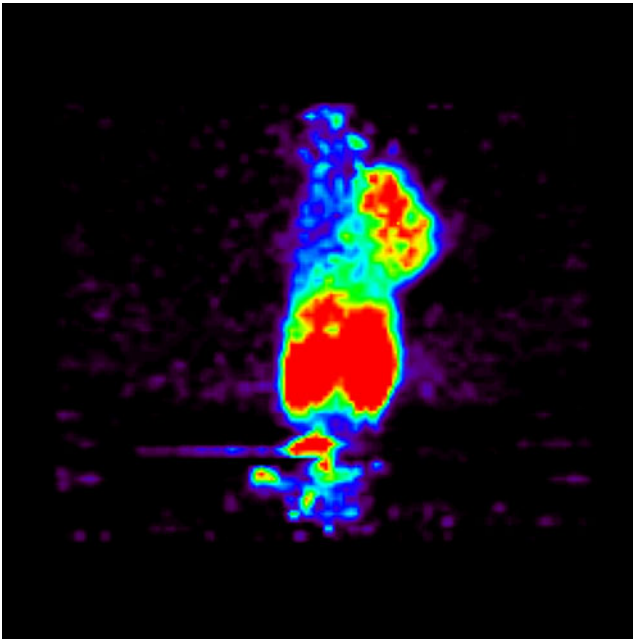
Promising pre-clinical studies using the DOTA-analogues of several other peptides, including substance P [39], neurotensin [40], and cholecystokinin (CCK) [41], have also been conducted.

DOTA-conjugated bombesin analogues labeled with  $^{68}\text{Ga}$  have been demonstrated to be promising imaging agents and to be useful for the targeted radionuclide treatment of bombesin receptor-positive tumors. These receptors have been reported to be overexpressed in invasive primary prostate carcinoma, associated lymph nodes, breast cancer and gastrointestinal stromal tumor [42, 43].

In terms of the assessment of infection and inflammation,  $^{68}\text{Ga}$ -DOTA-VAP-P1, a peptide inhibitor of vascular

**Fig. 3** Small-animal PET of  $^{68}\text{Ga}$ -NOTA-RGD in mice bearing SNU-C4 xenografts at 1 and 2 h after injection without or with cold c(RGDyK) (**a** and **b**, respectively). Arrows indicate tumor positions. Reprinted by permission of the Society of Nuclear Medicine from: Jeong JM, et al. (2008) Preparation of a promising angiogenesis PET imaging agent:  $^{68}\text{Ga}$ -labeled c(RGDyK)-isothiocyanatobenzyl-1,4,7-triazacyclononane-1,4,7-triacetic acid and feasibility studies in mice. *J Nucl Med* 49:830–836





**Fig. 4** Micro-PET image of  $^{68}\text{Ga}$ -NOTA-aminoalanine in mouse colon cancer (CT-26) showing clear uptake by the cancer lesion located at the right shoulder

adhesion protein 1/semicarbazine sensitive amine oxidase (VAP-1/SSAO), is a promising agent for the assessment of inflammatory reactions in healing bones [44]. In addition, studies have suggested that  $^{68}\text{Ga}$  PET imaging might be useful in experimental osteomyelitis caused by *Staphylococcus aureus* [45].

### Non-peptide Agents Labeled with $^{68}\text{Ga}$

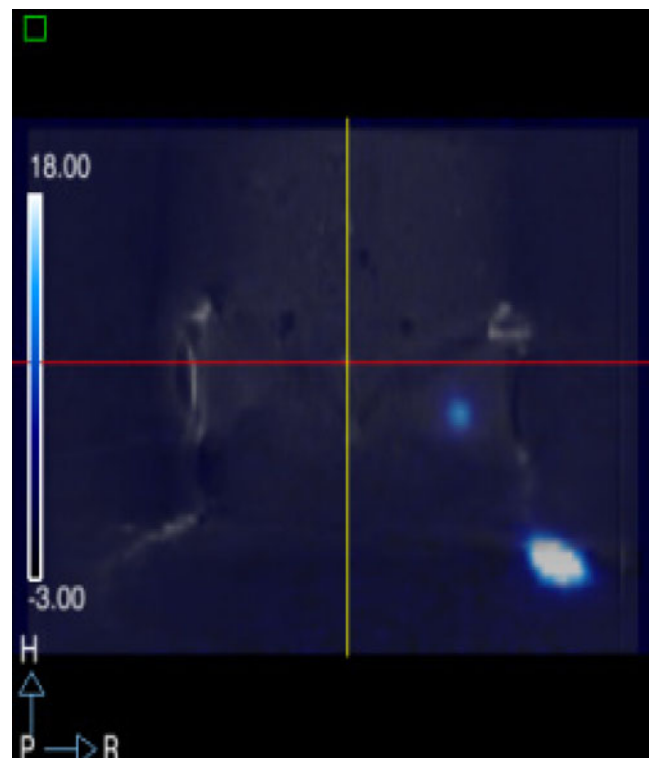
Most non-peptide agents labeled with  $^{68}\text{Ga}$  are complexes that are not conjugated to specific ligands. However, some non-peptide agents are conjugated with ligands via BCA. For example, three different forms of antisense oligonucleotides targeting activated human K-ras oncogene labeled with  $^{68}\text{Ga}$  have been shown to provide a convenient means for in vivo imaging and quantification of oligonucleotide biokinetics in living animals [46].

Various  $^{68}\text{Ga}$ -labeled agents have been developed using NOTA as the basic chelating agent, because of the high stability of the chelate formed; for example, the log  $K$  value of gallium-NOTA [47, 48] has been reported to be 30.98, which is much higher than that of gallium-DOTA (21.33) [49]. DOTA has eight binding sites available for complexing with metals, but NOTA has only six, and thus, no free binding site remains after complexing NOTA with gallium, which requires all six binding sites. However, it has been shown that amide oxygen or nitrogen can bind with gallium [50], and as a result, various amino acid derivatives conjugated with NOTA have been developed for imaging

cancers (Fig. 4) [51]. In addition, another amino acid derivatives conjugated with DO2A and DO3A have also been reported [52].

An in vitro cellular uptake and biodistribution study in breast cancer-bearing rats, using  $^{68}\text{Ga}$  metronidazole and ethylenedicycysteine (EC) as a chelator, demonstrated the feasibility of this tracer for the assessment of tumor hypoxia [53]. Conjugates of nitroimidazole and NOTA derivatives have also been developed [54]. The molecular imaging of the functional transport activity of multidrug resistance (MDR1) P-glycoprotein (Pgp) using  $^{67}\text{Ga}/^{68}\text{Ga}$ -(3-ethoxy-ENBDMPI) may also enable non-invasive monitoring of the blood-brain barrier, chemotherapeutic regimens, and MDR1 gene treatment protocols in vivo [55]. In addition,  $^{68}\text{Ga}$ -mannosylated human serum albumin (MSA) has been reported to be a promising agent for sentinel node detection (Fig. 5) [56]. MSA targets the mannose receptor of macrophages present in lymph nodes after subcutaneous injection. Furthermore, this heat labile agent can be labeled straightforwardly at room temperature, because NOTA is used as the BCA.

We have formulated  $^{68}\text{Ga}$ -BAPEN [ $^{68}\text{Ga}$ -Tris(4,6-dimethoxy-salicylaldehyde)-N,N'-bis(3-aminopropyl)-N,N'-ethylenediamine] as a kit for myocardial PET imaging and biodistribution studies [57]. This kit allows  $^{68}\text{Ga}$ -labeled agents to be easily prepared.



**Fig. 5** A micro-PET-CT image of  $^{68}\text{Ga}$ -MSA after injection into a mouse footpad. Radioactivity was present at the inguinal lymph node



## Clinical Applications of $^{68}\text{Ga}$ Peptides

Hofmann et al. [58] presented the first impressive  $^{68}\text{Ga}$ -DOTA-TOC images of neuroendocrine tumors, compared with  $^{111}\text{In}$ -octreotide scintigraphy, in eight patients with carcinoid tumors.  $^{68}\text{Ga}$ -DOTA-TOC identified all lesions, whereas  $^{111}\text{In}$ -octreotide identified only 85%. Furthermore, quantitative analysis of these lesions showed higher tumor to non-tumor contrast ratios and low kidney accumulation for  $^{68}\text{Ga}$ -DOTA-TOC PET imaging [4]. The pharmacokinetics of  $^{68}\text{Ga}$ -DOTA-TOC and  $^{18}\text{F}$ -fluorodeoxyglucose (FDG) in metastatic NET patients demonstrated uptakes by 57 of 63 lesions and by 43 of 63 lesions, respectively [59]. Another comparison between these two radiopharmaceuticals in a small group of patients ( $n=4$ ) with metastatic NET showed that  $^{68}\text{Ga}$ -DOTA-TOC was better at depicting smaller lesions with low tracer uptake, especially when tumors bore somatostatin receptors at low densities [60].

A comparative study between  $^{68}\text{Ga}$ -DOTA-TOC PET and  $^{99\text{m}}\text{Tc}$ -HYNIC-octreotide was performed by Gabriel et al. [6] in 88 patients with a neuroendocrine tumor.  $^{68}\text{Ga}$ -DOTA-TOC PET showed a sensitivity of 97%, a specificity of 92%, and an overall accuracy of 96%, which were significantly higher than those of single photon emission computed tomography (SPECT) using  $^{99\text{m}}\text{Tc}$ -labeled tracer (Fig. 3).

A viability study of  $^{68}\text{Ga}$ -DOTA-TATE for pheochromocytoma PET imaging was conducted in patients who had previously undergone surgical resection of malignant pheochromocytomas.  $^{68}\text{Ga}$ -DOTA-TATE was positive in all five patients studied, whereas only three patients were positive by  $^{123}\text{I}$ -metaiodobenzylguanidine (MIBG) scan [61, 62]. Due to the difficulties of diagnosing meningioma by computed tomography (CT) and magnetic resonance imaging (MRI), other methods of characterizing these intracranial lesions are being sought. Imaging meningiomas with  $^{68}\text{Ga}$ -DOTA-TOC can provide useful clinical indications, because SSTR2 is highly expressed in most meningiomas. The first use of  $^{68}\text{Ga}$ -DOTA-TOC in meningiomas in three patients was reported by Milker-Zabel and co-workers; the same group evaluated its kinetic parameters in meningioma [63].

## Conclusion

The use of a  $^{68}\text{Ge}/^{68}\text{Ga}$ -generator that can consistently supply  $^{68}\text{Ga}$ , which has a half-life of 271 days, provides a convenient way of producing  $^{68}\text{Ga}$  for more than a year. Furthermore, the cost of the generator is comparable with those of other radionuclides used for PET. In addition, diagnostic approaches based on  $^{68}\text{Ga}$ -labeled agents have the additional advantage of facilitating treatment. For

example, when a diagnostic scan is positive, these agents can be labeled with therapeutic radionuclides, such as, yttrium-90, lutetium-177, or rhenium-188. Ongoing experimental work suggests the feasibility of  $^{68}\text{Ga}$  labeling with different biomolecules for the imaging of different tumors, myocardium, and infection. The commercial availability of  $^{68}\text{Ge}/^{68}\text{Ga}$ -generator will undoubtedly encourage further preclinical research and clinical studies, and may open the door to new possibilities for PET.

## References

- Breeman WA, Verbruggen AM. The  $^{68}\text{Ge}/^{68}\text{Ga}$  generator has high potential, but when can we use  $^{68}\text{Ga}$ -labelled tracers in clinical routine? *Eur J Nucl Med Mol Imaging*. 2007;34:978–81.
- Ehrhardt GJ, Welch MJ. A new germanium-63/gallium-68 generator. *J Nucl Med*. 1978;19:925–9.
- Zhemosekov KP, Filosofov DV, Baum RP, Aschoff P, Bihl H, Razbash AA, et al. Processing of generator-produced  $^{68}\text{Ga}$  for medical application. *J Nucl Med*. 2007;48:1741–8.
- Antunes P, Ginj M, Zhang H, Waser B, Baum RP, Reubi JC, et al. Are radiogallium-labelled DOTA-conjugated somatostatin analogues superior to those labelled with other radiometals? *Eur J Nucl Med Mol Imaging*. 2007;34:982–93.
- Buchmann I, Henze M, Engelbrecht S, Eisenhut M, Runz A, Schafer M, et al. Comparison of  $^{68}\text{Ga}$ -DOTATOC PET and  $^{111}\text{In}$ -DTPAOC (Octreoscan) SPECT in patients with neuroendocrine tumours. *Eur J Nucl Med Mol Imaging*. 2007;34:1617–26.
- Gabriel M, Decristoforo C, Kendler D, Dobrozemsky G, Heute D, Uprimny C, et al.  $^{68}\text{Ga}$ -DOTA-Tyr3-octreotide PET in neuroendocrine tumors: comparison with somatostatin receptor scintigraphy and CT. *J Nucl Med*. 2007;48:508–18.
- Ambrosini V, Tomassetti P, Castellucci P, Campana D, Montini G, Rubello D, et al. Comparison between  $^{68}\text{Ga}$ -DOTA-NOC and  $^{18}\text{F}$ -DOPA PET for the detection of gastro-entero-pancreatic and lung neuro-endocrine tumours. *Eur J Nucl Med Mol Imaging*. 2008;35:1431–8.
- Parker D. Tumour targeting with radiolabelled macrocycle-antibody conjugates. *Chem Soc Rev*. 1990;19:271–91.
- Harris WR, Pecoraro VL. Thermodynamic binding constants for gallium transferrin. *Biochemistry*. 1983;22:292–9.
- Chong HS, Ma X, Le T, Kwamena B, Milenic DE, Brady ED, et al. Rational design and generation of a bimodal bifunctional ligand for antibody-targeted radiation cancer therapy. *J Med Chem*. 2008;51:118–25.
- Chong HS, Song HA, Ma X, Milenic DE, Brady ED, Lim S, et al. Novel bimodal bifunctional ligands for radioimmunotherapy and targeted MRI. *Bioconjug Chem*. 2008;19:1439–47.
- Hoffend J, Mier W, Schuhmacher J, Schmidt K, Dimitrakopoulou-Strauss A, Strauss LG, et al. Gallium-68-DOTA-albumin as a PET blood-pool marker: experimental evaluation in vivo. *Nucl Med Biol*. 2005;32:287–92.
- Jeong JM, Hong MK, Chang YS, Lee YS, Kim YJ, Cheon GJ, et al. Preparation of a promising angiogenesis PET imaging agent:  $^{68}\text{Ga}$ -labeled c(RGDyK)-isothiocyanatobenzyl-1, 4, 7-triazacyclononane-1, 4, 7-triacetic acid and feasibility studies in mice. *J Nucl Med*. 2008;49:830–6.
- Sabatino G, Chinol M, Paganelli G, Papi S, Chelli M, Leone G, et al. A new biotin derivative-DOTA conjugate as a candidate for pretargeted diagnosis and therapy of tumors. *J Med Chem*. 2003;46:3170–3.

15. Tanaka K, Masuyama T, Hasegawa K, Tahara T, Mizuma H, Wada Y, et al. A submicrogram-scale protocol for biomolecule-based pet imaging by rapid 6p-azaelectrocyclization: visualization of sialic acid dependent circulatory residence of glycoproteins. *Angew Chem Int Ed*. 2008;47:102–5.
16. Tsang BW, Mathias CJ, Fanwick PE, Green MA. Structure-distribution relationships for metal-labeled myocardial imaging agents: comparison of a series of cationic gallium (III) complexes with hexadentate bis(salicylaldehyde) ligands. *J Med Chem*. 1994;37:4400–6.
17. Tsang BW, Mathias CJ, Green MA. A gallium-68 radiopharmaceutical that is retained in myocardium:  $^{68}\text{Ga}[(4, 6\text{-MeO}_2\text{sal})_2\text{BAPEN}]^+$ . *J Nucl Med*. 1993;34:1127–31.
18. Kung HF, Liu BL, Mankoff D, Kung MP, Billings JJ, Francesconi L, et al. A new myocardial imaging agent: synthesis, characterization, and biodistribution of gallium-68-BAT-TECH. *J Nucl Med*. 1990;31:1635–40.
19. Fichna J, Janecka A. Synthesis of target-specific radiolabeled peptides for diagnostic imaging. *Bioconjug Chem*. 2003;14:3–17.
20. Janoki G, Harwig J, Chanachai W, Wolf W.  $^{67}\text{Ga}$  desferrioxamine-HSA: synthesis of chelon protein conjugates using carbodiimide as a coupling agent. *Int J Appl Radiat Isot*. 1983;34:871–7.
21. Koizumi M, Endo K, Kunitatsu M, Sakahara H, Nakashima T, Kawamura Y, et al. Preparation of  $^{67}\text{Ga}$ -labeled antibodies using deferoxamine as a bifunctional chelate. An improved method. *J Immunol Methods*. 1987;104:93–102.
22. Mathias CJ, Sun YZ, Welch MJ, Connert JM, Philpott GW, Martell AE.  $\text{N,N}'$ -bis(2-hydroxybenzyl)-1-(4-bromoacetamidobenzyl)-1,2-ethylenediamine- $\text{N,N}'$ -diacetic acid: a new bifunctional chelate for radiolabeling antibodies. *Bioconjug Chem*. 1990;1:204–11.
23. Broan C, Cox J, Craig A, Katakly R, Parker D, Harrison A, et al. Structure and solution stability of indium and gallium complexes of 1,4,7-triazacyclononanetriacetate and of yttrium complexes of 1,4,7,10-tetraazacyclododecanetetracetate and related ligands: kinetically stable complexes for use in imaging and radioimmunotherapy. X-ray molecular structure of the indium and gallium complexes of 1,4,7-triazacyclononane-1,4,7-triacetic acid. *J Chem Soc Perkin Trans*. 1991;2:87–99.
24. Liu S, Edwards DS. Synthesis and characterization of two  $^{111}\text{In}$ -labeled DTPA-peptide conjugates. *Bioconjug Chem*. 2001;12:630–4.
25. Chong HS, Garmestani K, Ma D, Milenic DE, Overstreet T, Brechbiel MW. Synthesis and biological evaluation of novel macrocyclic ligands with pendent donor groups as potential yttrium chelators for radioimmunotherapy with improved complex formation kinetics. *J Med Chem*. 2002;45:3458–64.
26. Reubi JC, Schaer JC, Markwalder R, Waser B, Horisberger U, Laissue J. Distribution of somatostatin receptors in normal and neoplastic human tissues: recent advances and potential relevance. *Yale J Biol Med*. 1997;70:471–9.
27. Reubi JC. Regulatory peptide receptors as molecular targets for cancer diagnosis and therapy. *Q J Nucl Med*. 1997;41:63–70.
28. Schonbrunn A. Somatostatin receptors present knowledge and future directions. *Ann Oncol*. 1999;10:S17–21.
29. Reubi JC, Schar JC, Waser B, Wenger S, Heppeler A, Schmitt JS, et al. Affinity profiles for human somatostatin receptor subtypes SST1–SST5 of somatostatin radiotracers selected for scintigraphic and radiotherapeutic use. *Eur J Nucl Med*. 2000;27:273–82.
30. Kwekkeboom DJ, Kooij PP, Bakker WH, Macke HR, Krenning EP. Comparison of  $^{111}\text{In}$ -DOTA-Tyr3-octreotide and  $^{111}\text{In}$ -DTPA-octreotide in the same patients: biodistribution, kinetics, organ and tumor uptake. *J Nucl Med*. 1999;40:762–7.
31. Rufini V, Calcagni ML, Baum RP. Imaging of neuroendocrine tumors. *Semin Nucl Med*. 2006;36:228–47.
32. Smith-Jones PM, Stolz B, Bruns C, Albert R, Reist HW, Fridrich R, et al. Gallium-67/gallium-68-[DFO]-octreotide—a potential radiopharmaceutical for PET imaging of somatostatin receptor-positive tumors: synthesis and radiolabeling in vitro and preliminary in vivo studies. *J Nucl Med*. 1994;35:317–25.
33. Breeman WA, de Jong M, Kwekkeboom DJ, Valkema R, Bakker WH, Kooij PP, et al. Somatostatin receptor-mediated imaging and therapy: basic science, current knowledge, limitations and future perspectives. *Eur J Nucl Med*. 2001;28:1421–9.
34. Kwekkeboom DJ, Mueller-Brand J, Paganelli G, Anthony LB, Pauwels S, Kvols LK, et al. Overview of results of peptide receptor radionuclide therapy with 3 radiolabeled somatostatin analogs. *J Nucl Med*. 2005;46 Suppl 1:62S–6S.
35. Wild D, Macke HR, Waser B, Reubi JC, Ginj M, Rasch H, et al.  $^{68}\text{Ga}$ -DOTANOC: a first compound for PET imaging with high affinity for somatostatin receptor subtypes 2 and 5. *Eur J Nucl Med Mol Imaging*. 2005;32:724.
36. Wild D, Schmitt JS, Ginj M, Macke HR, Bernard BF, Krenning E, et al. DOTA-NOC, a high-affinity ligand of somatostatin receptor subtypes 2, 3 and 5 for labelling with various radiometals. *Eur J Nucl Med Mol Imaging*. 2003;30:1338–47.
37. Wei L, Miao Y, Gallazzi F, Quinn TP, Welch MJ, Vavere AL, et al. Gallium-68-labeled DOTA-rhenium-cyclized alpha-melanocyte-stimulating hormone analog for imaging of malignant melanoma. *Nucl Med Biol*. 2007;34:945–53.
38. Froidevaux S, Calame-Christe M, Schuhmacher J, Tanner H, Saffrich R, Henze M, et al. A gallium-labeled DOTA-alpha-melanocyte-stimulating hormone analog for PET imaging of melanoma metastases. *J Nucl Med*. 2004;45:116–23.
39. van Hagen PM, Breeman WA, Reubi JC, Postema PT, van den Anker-Lugtenburg PJ, Kwekkeboom DJ, et al. Visualization of the thymus by substance P receptor scintigraphy in man. *Eur J Nucl Med*. 1996;23:1508–13.
40. de Visser M, Janssen PJ, Srinivasan A, Reubi JC, Waser B, Erion JL, et al. Stabilised  $^{111}\text{In}$ -labelled DTPA- and DOTA-conjugated neurotensin analogues for imaging and therapy of exocrine pancreatic cancer. *Eur J Nucl Med Mol Imaging*. 2003;30:1134–9.
41. Behr TM, Behe MP. Cholecystokinin-B/Gastrin receptor-targeting peptides for staging and therapy of medullary thyroid cancer and other cholecystokinin-B receptor-expressing malignancies. *Semin Nucl Med*. 2002;32:97–109.
42. Reubi JC, Wenger S, Schmuckli-Maurer J, Schaer JC, Gugger M. Bombesin receptor subtypes in human cancers: detection with the universal radioligand  $^{125}\text{I}$ -[D-TYR(6), beta-ALA(11), PHE(13), NLE(14)] bombesin(6–14). *Clin Cancer Res*. 2002;8:1139–46.
43. Zhang H, Schuhmacher J, Waser B, Wild D, Eisenhut M, Reubi JC, et al. DOTA-PESIN, a DOTA-conjugated bombesin derivative designed for the imaging and targeted radionuclide treatment of bombesin receptor-positive tumours. *Eur J Nucl Med Mol Imaging*. 2007;34:1198–208.
44. Lankinen P, Mäkinen TJ, Pöyhönen TA, Virsu P, Salomaki S, Hakanen AJ, et al.  $^{68}\text{Ga}$ -DOTAVAP-P1 PET imaging capable of demonstrating the phase of inflammation in healing bones and the progress of infection in osteomyelitic bones. *Eur J Nucl Med Mol Imaging*. 2008;35:352–64.
45. Mäkinen TJ, Lankinen P, Pöyhönen T, Jalava J, Aro HT, Roivainen A. Comparison of  $^{18}\text{F}$ -FDG and  $^{68}\text{Ga}$  PET imaging in the assessment of experimental osteomyelitis due to *Staphylococcus aureus*. *Eur J Nucl Med Mol Imaging*. 2005;32:1259–68.
46. Roivainen A, Tolvanen T, Salomaki S, Lendvai G, Velikyán I, Numminen P, et al.  $^{68}\text{Ga}$ -labeled oligonucleotides for in vivo imaging with PET. *J Nucl Med*. 2004;45:347–55.
47. Clarke ET, Martell AE. Stabilities of the Fe(II), Ga(II) and In(III) chelates of  $\text{N,N,N}'$ -triazacyclononanetriacetic acid. *Inorg Chim Acta*. 1991;181:273–80.
48. Clarke ET, Martell AE. Potentiometric and spectrophotometric determination of the stabilities of In(III), Ga(II) and Fe(II) complexes of  $\text{N,N,N}'$ -tris(3,5-dimethyl-2-hydroxybenzyl)-1,4,7-triazacyclononane. *Inorg Chim Acta*. 1991;186:103–11.

49. Clarke ET, Martell AE. Stabilities of trivalent metal-ion complexes of the tetraacetate derivatives of 12-membered, 13-membered and 14-membered tetraazamacrocycles. *Inorg Chim Acta*. 1991;190:37–46.
50. Shetty D, Jeong JM, Hoigebazar L, Lee YS, Lee DS, Chung JK, et al. (2010) Formation and characterization of gallium(III) complexes with monoamide derivatives of 1,4,7-triazacyclononane-1,4,7-triacetic acid: a pH dependant structural study. *Eur J Inorg Chem* 2010:In press.
51. Shetty D, Ju CH, Kim YJ, Lee JY, Lee YS, Lee DS, et al. (2010) Synthesis and evaluation of macrocyclic amino acid derivatives for tumor imaging by gallium-68 positron emission tomography. *Bioorg Med Chem* 18. doi:10.1016/j.bmc.2010.09.022
52. Shetty D, Ju CH, Lee YS, Jeong SY, Choi JY, Yang BY, et al. (2010) Synthesis of novel  $^{68}\text{Ga}$  labeled amino acid derivatives for positron emission tomography of cancer cells. *Nucl Med Biol* 37. doi:10.1016/j.nucmedbio.2010.06.003
53. Ito M, Yang DJ, Mawlawi O, Mendez R, Oh CS, Azhdarinia A, et al. PET and planar imaging of tumor hypoxia with labeled metronidazole. *Acad Radiol*. 2006;13:598–609.
54. Hoigebazar L, Jeong JM, Choi SY, Choi JY, Shetty D, Lee YS, et al. Synthesis and characterization of nitroimidazole derivatives for  $^{68}\text{Ga}$ -labeling and testing in tumor xenografted mice. *J Med Chem*. 2010;53:6378–85.
55. Sharma V, Prior JL, Belinsky MG, Kruh GD, Piwnica-Worms D. Characterization of a  $^{67}\text{Ga}/^{68}\text{Ga}$  radiopharmaceutical for SPECT and PET of MDR1 P-glycoprotein transport activity in vivo: validation in multidrug-resistant tumors and at the blood-brain barrier. *J Nucl Med*. 2005;46:354–64.
56. Choi JY, Yoo BC, Kim K, Kim Y, Yang BY, Lee YS, et al. (2010) Development of  $^{68}\text{Ga}$ -labeled mannosylated human serum albumin (MSA) as a lymph node imaging agent for positron emission tomography. *Nucl Med Biol* 37. doi:10.1016/j.nucmedbio.2010.09.010
57. Yang BY, Jeong JM, Kim YJ, Choi JY, Lee YS, Lee DS, et al. Formulation of  $^{68}\text{Ga}$  BAPEN kit for myocardial positron emission tomography imaging and biodistribution study. *Nucl Med Biol*. 2009;37:149–55.
58. Hofmann M, Maecke H, Borner R, Weckesser E, Schoffski P, Oei L, et al. Biokinetics and imaging with the somatostatin receptor PET radioligand  $^{68}\text{Ga}$ -DOTATOC: preliminary data. *Eur J Nucl Med*. 2001;28:1751–7.
59. Koukouraki S, Strauss LG, Georgoulas V, Schuhmacher J, Haberkorn U, Karkavitsas N, et al. Evaluation of the pharmacokinetics of  $^{68}\text{Ga}$ -DOTATOC in patients with metastatic neuroendocrine tumours scheduled for  $^{90}\text{Y}$ -DOTATOC therapy. *Eur J Nucl Med Mol Imaging*. 2006;33:460–6.
60. Kowalski J, Henze M, Schuhmacher J, Macke HR, Hofmann M, Haberkorn U. Evaluation of positron emission tomography imaging using [ $^{68}\text{Ga}$ ]-DOTA-D Phe(1)-Tyr(3)-Octreotide in comparison to [ $^{111}\text{In}$ ]-DTPAOC SPECT. First results in patients with neuroendocrine tumors. *Mol Imaging Biol*. 2003;5:42–8.
61. Win Z, Al-Nahhas A, Towey D, Todd JF, Rubello D, Lewington V, et al.  $^{68}\text{Ga}$ -DOTATATE PET in neuroectodermal tumours: first experience. *Nucl Med Commun*. 2007;28:359–63.
62. Win Z, Rahman L, Murrell J, Todd J, Al-Nahhas A. The possible role of  $^{68}\text{Ga}$ -DOTATATE PET in malignant abdominal paraganglioma. *Eur J Nucl Med Mol Imaging*. 2006;33:506.
63. Henze M, Dimitrakopoulou-Strauss A, Milker-Zabel S, Schuhmacher J, Strauss LG, Doll J, et al. Characterization of  $^{68}\text{Ga}$ -DOTA-D-Phe1-Tyr3-octreotide kinetics in patients with meningiomas. *J Nucl Med*. 2005;46:763–9.