



Research

Cite this article: Gerrish PJ, Colato A, Sniegowski PD. 2013 Genomic mutation rates that neutralize adaptive evolution and natural selection. *J R Soc Interface* 10: 20130329. <http://dx.doi.org/10.1098/rsif.2013.0329>

Received: 11 April 2013

Accepted: 8 May 2013

Subject Areas:

biomathematics, biophysics, medical physics

Keywords:

population genetics, mutagenesis, error threshold, Fisher's fundamental theorem, beneficial mutations

Author for correspondence:

Philip J. Gerrish

e-mail: pgerrish@unm.edu

Electronic supplementary material is available at <http://dx.doi.org/10.1098/rsif.2013.0329> or via <http://rsif.royalsocietypublishing.org>.

Genomic mutation rates that neutralize adaptive evolution and natural selection

Philip J. Gerrish^{1,2}, Alexandre Colato³ and Paul D. Sniegowski⁴

¹Department of Biology, Center for Evolutionary and Theoretical Immunology, University of New Mexico, 230 Castetter Hall, MSC03-2020, Albuquerque, NM 87131, USA

²Theoretical Biology and Biophysics, Los Alamos National Laboratory, MS K710, Los Alamos, NM 87545, USA

³Departamento de Ciências da Natureza, Matemática e Educação, Federal University of São Carlos (UFSCar), Araras, Brazil

⁴Department of Biology, 213 Leidy Laboratories, University of Pennsylvania, Philadelphia, PA 19104, USA

When mutation rates are low, natural selection remains effective, and increasing the mutation rate can give rise to an increase in adaptation rate. When mutation rates are high to begin with, however, increasing the mutation rate may have a detrimental effect because of the overwhelming presence of deleterious mutations. Indeed, if mutation rates are high enough: (i) adaptive evolution may be neutralized, resulting in a zero (or negative) adaptation rate despite the continued availability of adaptive and/or compensatory mutations, or (ii) natural selection may be neutralized, because the fitness of lineages bearing adaptive and/or compensatory mutations—whether established or newly arising—is eroded by excessive mutation, causing such lineages to decline in frequency. We apply these two criteria to a standard model of asexual adaptive evolution and derive mathematical expressions—some new, some old in new guise—delineating the mutation rates under which either adaptive evolution or natural selection is neutralized. The expressions are simple and require no *a priori* knowledge of organism- and/or environment-specific parameters. Our discussion connects these results to each other and to previous theory, showing convergence or equivalence of the different results in most cases.

1. Introduction

Even the simplest of living organisms are highly complex. Mutations—indiscriminate alterations of such complexity—are much more likely to be harmful than beneficial [1–3]. For an individual organism, therefore, an increase in the overall rate of mutation should be detrimental. In a population of organisms, however, natural selection disproportionately favours beneficial mutations, and the net effect of increasing the overall mutation rate is thus less clear.

1.1. Previous studies

Generally speaking, the population-level effects of increasing the mutation rate have been studied separately under two artificial assumptions: the absence of beneficial mutations, and infinite population size. Only a handful of studies have relaxed both assumptions.

1.1.1. Absence of beneficial mutations

When beneficial mutations are assumed to be absent, and population size is finite, fitness will undergo a slow but steady decline because of the sluggish but largely irreversible accumulation of deleterious mutations. This process is especially pronounced in asexual populations, and it was in this context that the process was first described by Muller [4] and later dubbed ‘Muller’s ratchet’ [5] and formalized by Haigh [6]. Under the relentless accumulation of deleterious mutations, fitness will decline monotonically. Most of the subsequent work on Muller’s ratchet has focused on the rate of the ratchet, different factors affecting this rate, and in particular factors or conditions that can cause this rate to become negligible (i.e. that

halt the ratchet) [7–15]. Increasing the genomic mutation rate can only accelerate Muller's ratchet.

1.1.2. Infinite population size

When population size is assumed to be infinite, populations whose adaptation is constrained, i.e. populations in which beneficial mutations can occur but that have a maximum attainable fitness, will eventually achieve an equilibrium fitness distribution shaped by the largely opposing forces of mutation and natural selection. Above a critical mutation rate dubbed the 'error threshold' [16,17], this distribution becomes remarkably flat, indicating that a genotype's equilibrium frequency is essentially independent of its fitness. This conversion to a state of random fitness dispersion is reminiscent of a phase transition [17–21] and, in its simplest formulation, the two are mathematically equivalent [22,23]. The simplest formulation of the error threshold has been called into question because of some unrealistic assumptions that are often perceived as strong assumptions, the most notable of which is the 'single-peak' fitness landscape assumption [24]. The error threshold has since been studied extensively and shown to exist under many different conditions that eliminate different assumptions, for example, allowing for recombination and departures from random mating [25–27], viral complementation [28], spatial structure and different modes of replication [29–34] and more realistic static and dynamic fitness landscapes [26,27,35–40] (but see Wiehe [41]).

1.1.3. Extinction

The two classes of models described earlier—Muller's ratchet and the error threshold—encompass most previous characterizations of mutational degradation processes. In their original formulations, and in most subsequent work, neither of these two classes of models explicitly accounts for demographic decline as a result of excess mutation. There has been some work, however, that has superimposed demography onto both Muller's ratchet [42–44] and error threshold [45,46] models, finding a positive feedback between these processes and demographic decline towards extinction. These models, however, are typically sensitive to organism-, environment- and time-dependent parameters. In particular, they require an assumption about the mapping between relative and absolute fitness—an assumption that is loaded with requisite assumptions about the organism and environment, both of which can change with time.

1.1.4. Finite populations with beneficial mutations

A few studies have addressed the effect of increasing the mutation rate when the two foregoing assumptions are relaxed, i.e. when beneficial mutations are accounted for and populations are finite. Under these more realistic conditions, the fitness decline due to Muller's ratchet can be cancelled out or even reversed by beneficial mutations, resulting in unchanging or increasing fitness. The effect of beneficial mutations on Muller's ratchet has been explored previously [47–49]; these studies focused on how the effects and relative fractions of beneficial versus deleterious mutations would affect the adaptation rate and whether that rate was positive or negative. In this study, we focus on how the genomic mutation rate affects the progress of adaptive evolution and the effectiveness of natural selection.

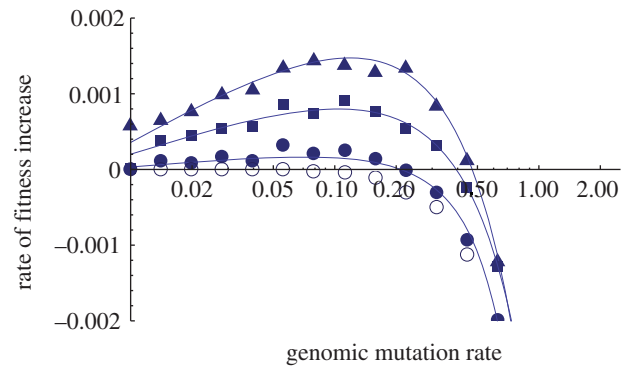


Figure 1. Time-averaged fitness gradients from simulations of adapting populations as a function of genomic mutation rate. Simulations are fully stochastic and individual-based; populations are asexual. (See the electronic supplementary material for details.) Each point represents the average of eight independent simulation runs. The fraction of mutations that are deleterious is constant at 0.1, and the effects of all mutations are drawn at random from an exponential distribution with mean 0.03. At high enough mutation rates, the rate of fitness increase becomes negative (indicating persistent fitness decline), from which inference of eventual extinction seems reasonable. Filled circles plot simulation results in which population size is 10 000 and the fraction of mutations that are beneficial is 10^{-5} ; filled squares plot simulation results in which population size is 50 000 and the fraction of mutations that are beneficial is 10^{-5} ; filled triangles plot simulation results in which population size is 10 000 and the fraction of mutations that are beneficial is 10^{-4} ; open circles plot simulation results in which population size is 10 000 and the fraction of mutations that are beneficial is zero (classical Muller's ratchet). (Online version in colour.)

1.2. Present study

1.2.1. Neutralizing adaptive evolution

When genomic mutation rate is low to begin with, an increase in this rate may be advantageous: the increased production of deleterious mutations can be of disproportionately small consequence, because natural selection tends to eliminate deleterious mutations from the population, whereas the increased production of rare beneficial mutations can be of disproportionately large consequence, because natural selection can cause the fixation of beneficial mutations from which the entire population benefits. Thus, if a population's overall mutation rate is low to begin with, then an increase in the mutation rate can increase the rate at which beneficial mutations are fixed, thereby increasing the adaptation rate, where adaptation is defined as increase in mean fitness. In other words, a positive correlation can exist between genomic mutation rate and adaptation rate.

When genomic mutation rate is high to begin with, however, an increase in this rate may be disadvantageous because of excess deleterious mutations. While the consequence of deleterious mutations is still disproportionately small, it is less so at high mutation rates, because deleterious mutations can be produced faster than natural selection can remove them. At high mutation rates, therefore, a negative correlation can exist between genomic mutation rate and adaptation rate.

The foregoing considerations indicate a non-monotonic relationship between mutation rate and adaptation rate, a relationship confirmed by simulation (figures 1 and 2). In this paper, we are interested in finding critical genomic mutation rates above which adaptation rate becomes negative. It seems reasonable to speculate that a negative adaptation rate, if sustained, would ultimately result in extinction.

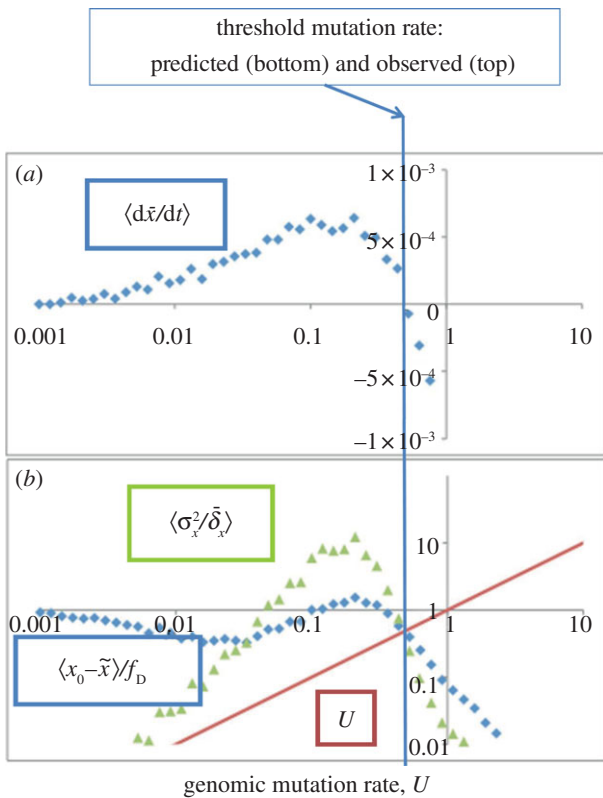


Figure 2. (a) Time-averaged adaptation rate as a function of genomic mutation rate. The point at which adaptation rate becomes negative marks the threshold mutation rate, indicated by the blue vertical line. (b) Predictions for the threshold mutation rate. Green triangles plot the variance threshold given by equation (2.1); blue diamonds plot the error threshold given by equation (2.4); red line plots genomic mutation rate U . Where threshold predictions intersect with the red line marks the predicted threshold mutation rate for these simulations, and coincides exactly with the observed threshold mutation rate in (a). Each point represents an average taken over the full time course of eight fully stochastic, individual-based simulations of evolving asexual populations. Population size was 10 000, fractions of mutations that were beneficial and deleterious were 0.001 and 0.5, respectively. Ten per cent of deleterious mutations were lethal; otherwise, beneficial and deleterious mutations were drawn from an exponential distribution with mean 0.03. Epistasis among deleterious mutations is synergistic with epistasis parameter 0.1 and epistasis exponent 5 (see the electronic supplementary material for epistasis function). Several similar plots with different sets of biological complexities are posted in the electronic supplementary material.

1.2.2. Neutralizing natural selection

Evolution by natural selection proceeds through the appearance and subsequent fixation of adaptive and/or compensatory mutations. When mutation rate is low, virtually all adaptive and/or compensatory mutations produced have *fixation potential*: all of them have the possibility, at least, of enduring the first few generations of random sampling (surviving genetic drift [50]), outcompeting other adaptive and/or compensatory mutations (surviving the Hill–Robertson effect [51] or clonal interference [52]) and spreading to fixation. This is because, with low mutation rates, progress to fixation is relatively unhindered by deleterious mutations.

As mutation rate increases, however, the *fixation potential* of adaptive and/or compensatory mutations is reduced: each such mutation finds a lineage whose growth is increasingly eroded by the accumulation of deleterious mutations. As mutation rate continues to increase, a point may be reached

at which adaptive and/or compensatory mutations lose their fixation potential altogether, thereby neutralizing natural selection. We explore three particularly telling indicators that this point has been reached: (i) the fittest genotype in the population (e.g. an adaptive mutant) decreases in frequency, (ii) the fittest genotype in the population has an equilibrium frequency (i.e. a mutation–selection balance frequency) very close to zero, and (iii) a newly arising fittest genotype is ultimately doomed to extinction with probability one.

1.2.3. A key innovation: dynamical insufficiency

In many of the previous investigations of mutational degradation processes, analogies are drawn to physical processes not least of which is the phase transition analogy. However, the analogous physical processes typically occur on short time scales during which the relevant parameters remain constant and convergence to equilibria occurs rapidly. This context affords the luxury of dynamically sufficient models and applicability of their steady-state analyses. In evolutionary biology, however, time scales are longer, relevant parameters cannot reliably be assumed to remain constant, and equilibria may rarely, if ever, be achieved. In the face of such long-term uncertainty, predictive accuracy seems unlikely; nevertheless, dynamically insufficient models may provide short-term predictive accuracy. Fisher’s ‘fundamental theorem of natural selection’ accurately predicts the evolution of fitness over the course of a single generation; by sacrificing dynamical sufficiency, this theorem achieves short-term predictive accuracy. Some of the conditions that we derive here (the more useful conditions) use variations of this approach; they depend on statistical properties of the population that, by virtue of their *intermediate* dynamical sufficiency, absorb contingencies and other surprises that are so characteristic of the biological world (see §3) and thereby may subsume many previous results that individually treat an array of different complexities and were derived under the purview of dynamical sufficiency.

1.2.4. Extinction

We stress that the work we present here only delineates conditions under which adaptive evolution or natural selection is neutralized. Demographic decline is *not* explicitly accounted for in our modelling here, and the few references we make to extinction (one of which is in §1.2.5) are therefore based on reasonable but nonetheless speculative inference. (We note that reference is made in §2.3.2 (criterion 2) to the extinction of individual beneficial lineages owing to *differential* or ‘relative’ fitness. This is very different from, and should not be confused with, whole-population extinction.)

1.2.5. Our default application

The discovery and development of the error threshold sparked the imagination of virologists, whose efforts to clear viral infections using antiviral drugs are bedevilled by the high mutation rates of many viruses. If mutation rate could be elevated even further through mutagenesis, then error threshold theory suggested that viral populations should undergo ‘informational collapse’, which has a dire ring to it, and suggested that populations might be driven extinct, thereby in a sense beating the virus at its own game [53–57]. As in our models, extinction in these earlier models is not explicit but may be considered a reasonable, if speculative, prediction (see §1.2.4).

Partly because of this historical context, we have adopted this particular application as our ‘default’ application: unless otherwise stated, we have in mind the general aim of neutralizing adaptive evolution and/or natural selection in an unwanted population (and by extrapolation, eradicating the unwanted population) through mutagenesis, and the inequalities we derive reflect this aim.

1.2.6. Outline of this study

In this study, we independently apply the two earlier-described criteria to a standard, general model of fitness evolution in order to derive the conditions under which adaptive evolution and natural selection are neutralized. (To guide the reader, we would point out that our main results are thus indicated by the word ‘condition’.) The conditions that we derive from criterion 1 range from sufficient to sufficient and necessary; however, it is the intermediate condition—called ‘sufficient and *somewhat* necessary’—that we believe is the most novel and perhaps the most practical. We apply criterion 2 both to a population in which the fittest genotype is resident (recovering the classical error threshold results in a new guise that lends itself to an alternative and perhaps more useful interpretation) and to one in which the fittest genotype is a newly arising beneficial mutant.

2. Results

2.1. The model

We use a standard model of adaptive evolution of an asexual population in which a genotype or class increases in log-frequency as the fitness of that genotype or class minus the mean fitness of the population. Mutation occurs among genotypes or classes as a diffusion process that is strongly biased in favour of deleterious mutations. Mathematical formulations of this model are given in appendix A.

In what follows, we use the bracket notation in addition to the overbar notation to denote averaging. Overbar notation is used to denote averaging over individuals in a population. (Because our analyses implicitly assume infinite population size, such averaging is equivalent to expectation.) Some examples are: x denotes fitness, and \bar{x} denotes the mean fitness of a population; δ_x denotes the effect of mutation on fitness, and $\bar{\delta}_x$ denotes its average taken over all possible mutated offspring (here, the ‘population’ is really a *potential* population). Bracket notation is used to denote averaging over time; for example, $\langle \bar{x} \rangle$ denotes the mean fitness of a population averaged over time. The models we use are continuous in time and thus our measure of fitness x corresponds to the log of fitness w used in classical population genetics (discrete-time) models.

2.2. Criterion 1: adaptive evolution is neutralized

Here, we use a formulation of our model that is continuous in both time and fitness. We ask under what conditions adaptation will move backwards, i.e. under what conditions population mean fitness will decrease in spite of an inexhaustible supply of beneficial mutations.

Assumptions of our model under this criterion are minimal: (i) no assumptions are made about the fitness landscape (except for the very weak assumption of ‘compact support’ of the mutation kernel; see appendix A), (ii) no equilibrium

assumptions are made, and (iii) our results here share the tautological flavour of Fisher’s theorem [58–60] (and the Price equation [61,62]) and in this sense are more akin to the theory of natural selection than to any particular model of evolution.

2.2.1. Sufficient and sufficient/necessary conditions

Adaptive evolution is neutralized when the long-term tendency of absolute fitness is to decrease, despite the availability of adaptive and/or compensatory mutations. A sufficient but not necessary version of this condition imposes $d\bar{x}/dt < 0$ at all times, where \bar{x} is population mean fitness. The necessary and sufficient version of this condition is $\langle d\bar{x}/dt \rangle < 0$. These conditions imply (appendix A) that adaptive evolution will be neutralized and fitness will in fact decline if the relation

$$-U\bar{\delta}_x > \sigma_x^2 \quad (2.1)$$

holds persistently (sufficient) or at least on average (sufficient and necessary), where σ_x^2 is variance in fitness, U is genomic mutation rate and $\bar{\delta}_x$ is the average effect of mutation on fitness. (While this expression is given in terms of fitness, an equivalent expression is derived in terms of a fitness-related phenotype in the electronic supplementary material.) If the effects of beneficial and deleterious mutations are considered separately, then $\bar{\delta}_x = f_B m_B - f_D m_D$, where f_D and f_B are the fractions of all mutations that are deleterious and beneficial, respectively; m_D and m_B are the mean effects of deleterious and beneficial mutations on fitness, respectively. Biological considerations overwhelmingly support $f_D m_D \gg f_B m_B$, so the left-hand side of (2.1) will most likely be positive. By some estimates [63–68], f_B can be surprisingly high; however (i) this does not necessarily imply high values of $f_B m_B$ [65] and (ii) it seems unlikely that $f_B m_B$ would ever exceed $f_D m_D$, simply because the ways to damage a highly complex entity (such as a living organism) far outnumber the ways to improve it. In the very unlikely case that $f_B m_B > f_D m_D$, condition (2.1) would present a contradiction, and fitness decline would be impossible regardless of U . The accuracy of (2.1) and its robustness to several factors such as epistasis are illustrated by figure 2 and in the electronic supplementary material.

Critical mutation rate can be a moving target. As evidenced by (2.1), the critical mutation rate required to neutralize adaptive evolution is a function of the fitness variance. Increasing the mutation rate, however, will often cause a subsequent increase in fitness variance, in turn increasing the mutation rate required to satisfy (2.1). In fact, classical population genetics (accounting for deleterious mutations only), and work by Rouzine *et al.* [69,70] and Goyal *et al.* [49] (accounting for beneficial and deleterious mutations) all indicate that, for low to moderate mutation rates, the fitness variance should tend towards $-U\bar{\delta}_x$ following a perturbation in fitness and/or mutation rate. This suggests that an adjustment in the mutation rate (perhaps through increasing the dose of a mutagen, for example) to satisfy the condition $-U\bar{\delta}_x > \sigma_x^2$ will be followed by an increase in fitness variance such that $\sigma_x^2 \rightarrow -U\bar{\delta}_x$, thus necessitating a further increase in U in order to maintain the relation $-U\bar{\delta}_x > \sigma_x^2$. Frank & Slatkin [71] have pointed out that the tendency $\sigma_x^2 \rightarrow -U\bar{\delta}_x$ represents mutation–selection balance (in fact, they mention this in the context of phenotypic evolution but the same notion applies). Figuratively, the condition $-U\bar{\delta}_x > \sigma_x^2$ may

be thought of as a mutation rate that persistently tips the balance in favour of mutation; alternatively, it may be thought of as a mutation rate persistently high enough to prevent convergence to mutation–selection balance. As U is increased to maintain $-U\bar{\delta}_x > \sigma_x^2$ in a continually adapting population, σ_x^2 will eventually reach a maximal value (owing to finite population size) and, at this point, the value of U need not increase further to satisfy $-U\bar{\delta}_x > \sigma_{\max}^2$. In figure 3*b*, the genetic variance in fitness is measured in simulated populations every 100 generations, and $-U\bar{\delta}_x$ is set at 10 per cent above σ_x^2 , thereby maintaining $-U\bar{\delta}_x > \sigma_x^2$. For a long time, the positive feedback between mutation rate and fitness variance results in escalating adjustments to the mutation rate; after some time, however, the variance appears to achieve a maximum, so that the mutation rate required for continued fitness decline levels off.

2.2.2. Sufficient and somewhat necessary condition

So far, we have derived conditions that lie at opposite ends of the spectrum from sufficiency to sufficiency-and-necessity. From a practical standpoint, however, both are of limited utility. Condition (2.1) ensures declining fitness only for the current generation. The sufficient condition is that this relation hold persistently, but this condition may be frustratingly elusive because it fails to anticipate the change in fitness variance that typically follows an adjustment to the mutation rate. For this condition to be enforced in practice, therefore, frequent measurements of σ_x^2 would be required, followed by adjustments in U (e.g. by increasing the dose of a mutagen), if needed, to maintain the relation (2.1) (as in figure 3*a,b*). In practice, therefore, the sufficient condition amounts to a rather inconvenient protocol. The sufficient-and-necessary condition, that (2.1) holds on average, requires long-term future knowledge of population fitness that is generally not attainable in practice. Here, we derive conditions that lie somewhere in the middle of the spectrum from sufficiency to sufficiency-and-necessity and that have increased practical applicability.

To this end, we temper our sufficient and necessary condition: instead of requiring that the *long-term* average gradient oppose selection, we now require only that the *medium-term* average gradient oppose selection. We will denote this intermediate condition as $\langle d\bar{x}/dt \rangle_r < 0$, where r denotes the number of future generations over which to take the average. In order to enforce this condition, however, one needs a way to predict the near-future course of evolution; an algorithm for doing this is outlined in Gerrish & Sniegowski [72]. There, it is shown that prediction of the near-future course of evolution can be achieved by a time-discretization of a hierarchy of cumulant equations.

Using the equations for fitness evolution derived in Gerrish & Sniegowski [72] and imposing $\langle d\bar{x}/dt \rangle_r < 0$, the condition under which adaptive evolution is neutralized may be written as

$$-U\bar{\delta}_x > \frac{1}{r} \sum_{\tau=0}^{r-1} \kappa_2(\tau), \quad (2.2)$$

where the future fitness variances (or second cumulants), $\kappa_2(\tau) = \sigma_x^2(\tau)$, are computed from the set of recursions $\kappa_i(\tau+1) = \kappa_i(\tau) + \kappa_{i+1}(\tau) + Um_i$ as outlined in Gerrish & Sniegowski [72] (also, see appendix A); $\kappa_i(\tau)$ denotes the i th cumulant at generation τ ; $\tau = 0$ denotes the present generation (called ‘now’), $\tau = 1$ denotes

one generation from now, $\tau = 2$ denotes two generations from now, etc.; and r is the ‘predictive reach’, i.e. r is how many generations into the future the algorithm in Gerrish & Sniegowski [72] can be trusted to predict. In ongoing work, we have shown that this algorithm can be trusted to predict $d\bar{x}/dt$ over at least $r = 20$ generations in laboratory *Escherichia coli* populations, and roughly $r = 45$ generations in simulations [72]. An alternative condition that errs conservatively is: $-U\bar{\delta}_x > \max(\kappa_2(0), \kappa_2(1), \dots, \kappa_2(r))$. (See the electronic supplementary material for equivalent phenotypic expressions.) The appearance of these equations is deceptively simple because as U is changed, the predictions for $\kappa_2(\tau)$ will change, i.e. the equations look explicit when in fact they are implicit for U . (They are implicit for U because a certain degree of circularity is required by their intermediate dynamical sufficiency, which anticipates future changes in σ_x^2 without requiring knowledge of organismal and environmental parameters; in practice, this fact only imposes the slight inconvenience of having to use an iterative procedure in the calculations.)

2.3. Criterion 2: natural selection is neutralized

The approach that derives from this criterion takes its lead from statistical physics, where an ‘order parameter’ quantifies the degree of order present in the system at hand. Order in an evolving population is brought about through the action of natural selection on genetic variation. In evolution, a natural choice for an order parameter is the frequency of the fittest genotype. If natural selection is operational, the fittest genotype should persist at reasonable frequency despite recurrent mutation away from this genotype, and this frequency is thus indicative of the amount of order present in the population. As mutation rate increases, the frequency of the fittest genotype will decrease, indicating a decrease in the overall order present. At a sufficiently high mutation rate, the amount of order will approach zero.

Assumptions implicit under this condition again are minimal: (i) no assumptions are made about the fitness landscape (e.g. the ‘single-peak’ landscape used by many error-threshold models is not required here; curiously, Eigen’s original paper on the error threshold had a formulation similar to ours—as shown in §3—and also did not require a ‘single-peak’ landscape) and (ii) equilibrium is not assumed, although some of the results are obtained by solving for the equilibrium state.

2.3.1. Sufficient condition

Here, we have in mind a population that is heterogeneous and that is predominated by a fittest genotype whose frequency is u_0 . Our sufficient condition is derived by finding the mutation rate that causes the frequency of the fittest genotype to decrease relative to its mutational neighbours: $du_0/dt < 0$, persistently. Solving for the mutation rate that ensures this inequality gives rise to the condition

$$Uf_D > (x_0 - \bar{x}) \left(\frac{1}{1 - u_0} - \frac{1}{Lu_0} \right)^{-1}, \quad (2.3)$$

where L is the size of the deleterious genome, x_0 is the fitness of the fittest genotype; $\bar{x} = \sum_{j=1}^L x_j \hat{u}_j$, and $\bar{x} = \sum_{j=0}^L x_j \hat{u}_j$, from which we have the useful relation $(1 - u_0)(x_0 - \bar{x}) = x_0 - \bar{x}$. In a finite population, x_0 is the maximum fitness found in the population, and \bar{x} is the average fitness of everybody else: $\bar{x} = (1/\#S) \sum_{i \in S} x_i$, where S is the subset of the

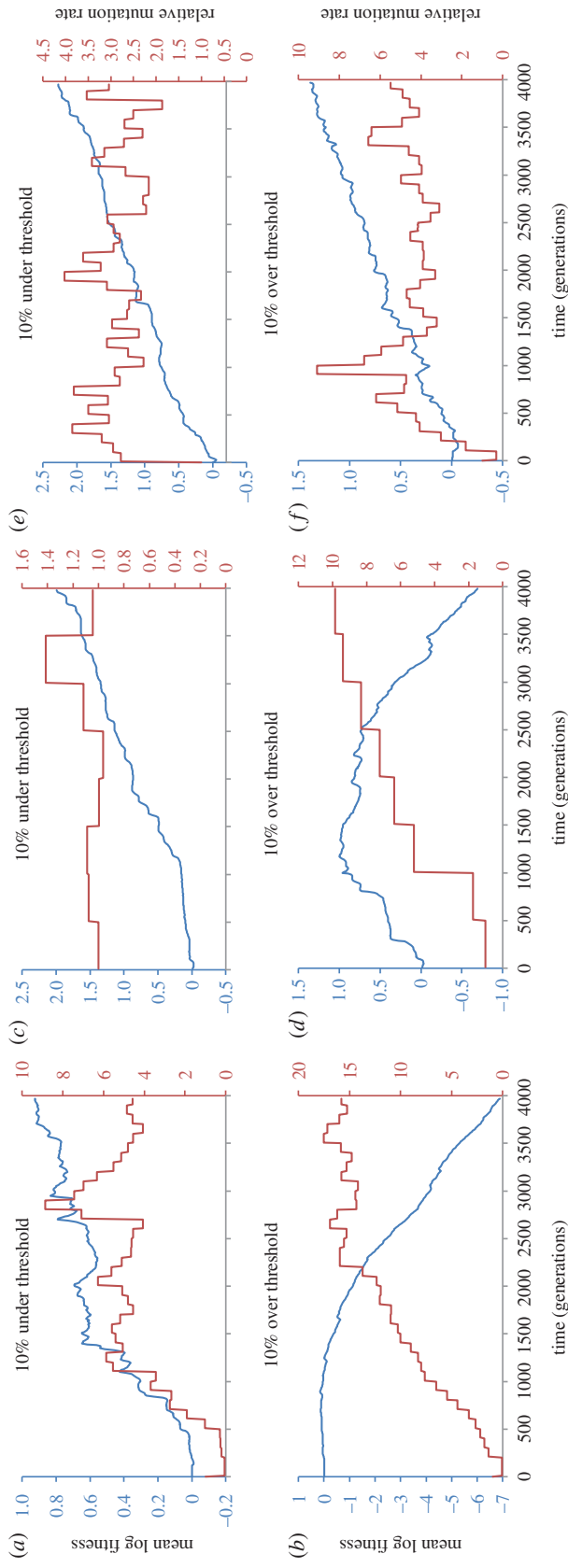


Figure 3. Real-time application of the different thresholds. Each panel plots a single representative simulation run. Simulations and parameters are the same as those in figure 1, with beneficial fraction set at 10^{-4} . (a,b) Variance threshold given by equation (2.1). Every 100 generations, fitness variance is measured and U is set equal to $-0.9\sigma_x^2/\bar{\delta}_x$ (a) and $-1.1\sigma_x^2/\bar{\delta}_x$ (b). (c,d) Variance-projection threshold given by equation (2.2). Every 500 generations, fitness measurements are used to compute cumulants $\kappa_1(\tau)$ from which the $\kappa_2(\tau)$ are calculated (appendix A), and U is set equal to $-0.9\frac{1}{\tau}\sum_{r=0}^{\tau-1}\kappa_2(\tau)/\bar{\delta}_x$ (c) and $-1.1\frac{1}{\tau}\sum_{r=0}^{\tau-1}\kappa_2(\tau)/\bar{\delta}_x$ (d). (e,f) Error threshold given by equation (2.4). Every 100 generations, U is set equal to $0.9(x_0 - \bar{x})/f_D$ (e) and $1.1(x_0 - \bar{x})/f_D$ (f). Our criterion 1 appears to perform better for such real-time application than criterion 2.

Table 1. Summary of neutralizing conditions.

conditions	adaptive evolution (criterion 1)	natural selection (criterion 2)
sufficient	$-U\bar{\delta}_x > \sigma_x^2$ persistently	$Uf_D \gtrsim x_0 - \bar{x}$ persistently ^a
sufficient and <i>somewhat</i> necessary	$-U\bar{\delta}_x > \frac{1}{r} \sum_{i=0}^{r-1} \kappa_2(i)$ intermittently	
sufficient and necessary	$-U\bar{\delta}_x > \sigma_x^2$ long-term average	$Uf_D \geq x_0 - \bar{x}$ long-term steady state

^aThis condition holds when $L \gg N$.

population that has fitness less than the maximum and #S is the number of individuals in that subset. If it is the case that the population is finite and $L \gg N$, then the term $1/Lu_0 \leq N/L \approx 0$, giving rise to the condition: $Uf_D \gtrsim (1 - u_0)(x_0 - \bar{x}) = x_0 - \bar{x}$ (reported in table 1). In our simulations, we assume an infinite genome ($L \rightarrow \infty$) and finite population size; under these conditions, $Uf_D > x_0 - \bar{x}$ is exact. In a continually adapting population, u_0 will be small most of the time, in which case this expression may be used interchangeably with: $Uf_D \gtrsim x_0 - \bar{x}$ (used in figure 3).

2.3.2. Sufficient and necessary conditions

(1) *Mutational degradation of an established fittest genotype.* Here, we have in mind a population that is heterogeneous but that has been predominated by a fittest lineage for some time. To determine the amount of order in this population, we compute its order parameter, \hat{u}_0 : the equilibrium frequency of this fittest lineage relative to its mutational neighbours (genotypes that differ from the fittest lineage by mutation). We are especially interested in what happens to the order parameter as mutation rate increases.

Analysis of the evolutionary model at equilibrium reveals that, indeed, the order parameter \hat{u}_0 decreases with increasing mutation rate (appendix A). The approach of \hat{u}_0 towards zero as U increases is characterized by an inflection point that becomes increasingly sharp as deleterious genome size L increases. The mutation rate at which the inflection point occurs is found by solving for the critical mutation rate U_c that satisfies $\partial^3 \hat{u}_0 / \partial U^3 = 0$. As L increases, $U_c f_D = \mu_c L \rightarrow x_0 - \bar{x}$, where f_D is again the fraction of mutations that are deleterious, and μ_c is the critical point mutation rate. From this result, natural selection may reasonably be expected to be neutralized when mutation exceeds the critical rate:

$$Uf_D \geq x_0 - \bar{x}. \quad (2.4)$$

This is the classical ‘error threshold’ result in new guise. It is an equilibrium result and its practical use would therefore require knowledge of long-term future states of the population. When long-term data are available this condition is very accurate and is robust to many factors (figure 2 and electronic supplementary material).

The equilibrium frequency of the fittest class or genotype at the ‘error threshold’, while greatly reduced, is still greater than the frequencies of neighbouring genotypes: at $\mu = \mu_c$, the fittest genotype has frequency $\hat{u}_0 \approx 1/L + \sqrt{1/L}$, whereas mutational neighbours have frequency $\hat{u}_i < 1/L$. This stands in contrast to common notions about the error threshold as creating a competitive reversal that leads to the subordination

and/or loss of the fittest genotype. In a finite population, the fittest genotype will be deterministically lost from the population at the error threshold only if $N \lesssim \sqrt{L}$. To put this condition in perspective, we consider a strain of *Escherichia coli* that has a genome of length $L \approx 4.6 \times 10^6$ base pairs; if we make the very conservative assumption that mutation at any position on the genome will affect fitness, then any population larger than $\sqrt{L} \approx 2145$ will deterministically retain the fittest genotype at the error threshold.

Despite the persistence and continued dominance of the fittest genotype, the error threshold nevertheless marks a point at which the frequencies of the different genotypes are so severely eroded by mutation that their frequencies are clearly not indicative of their fitness. This neutralizing of natural selection is apparent in the relation: $\text{cov}(u, x)|_{\mu=\mu_c} \approx \bar{s}/L$, where $\bar{s} \approx -\bar{\delta}_x$. For large genomes, therefore, the covariance between fitness and frequency—an indicator of the efficacy of natural selection—is very small at the error threshold (but still positive). Additionally, the extent to which natural selection has become ineffective is reflected by the amount of *disorder* present in the equilibrium population; a standard index of disorder is the Shannon entropy (measured, for example, for RNA viral quasi-species [34]) which, at the error threshold, is approximately equal to $\log_2 L$.

(2) *Mutational degradation of a newly arising fittest genotype.* Here, we have in mind an asexual population that is heterogeneous and in which a beneficial mutation emerges. This mutation creates a newly arising ‘fittest genotype’ whose subsequent growth depends on the persistence of that genotype within the growing lineage, despite recurrent mutation away from that genotype. The newly arising fittest genotype has fitness x_0 , and the rest of the population has average fitness \bar{x} , as before. In a single generation, the new lineage grows by a factor $R = e^{x_0 - \bar{x}} = e^{(x_0 - \bar{x})(1 - u_0)}$. Accumulation of deleterious mutations occurs most rapidly early in the growth of a lineage [9], when $u_0 \approx 0$, suggesting the approximation $R \approx e^{x_0 - \bar{x}}$. Previous studies show that genomic mutation rates that cause the degradation of the newly arising fittest genotype must satisfy $Uf_D \geq \log R$ [9,73,74]. The extinction of a newly arising fittest genotype is therefore predicted to occur when

$$Uf_D \gtrsim x_0 - \bar{x}. \quad (2.5)$$

Compare with (2.4). This result was originally derived for an independent asexual population growing without bound at discrete-time rate R [9] and was later re-derived in a way that more explicitly allowed for purifying selection and dubbed the ‘lethal mutagenesis’ threshold [73,74] for unboundedly growing viral and bacterial populations. This result should

also apply, however, to lineages growing within a population as a consequence of positive *relative fitness* ($x_0 - \tilde{x} > 0$). Finite population size restricts applicability to lineages that begin to decline in frequency before being affected by population size constraints, which seems likely to account for many such lineages when at or near the critical mutation rate (but see Gerrish *et al.* [75]). Those lineages that do achieve higher frequencies are likely to become fixed in the population, in which case the relevant condition was derived in the previous subsection: $Uf_D \geq x_0 - \tilde{x}$ (condition (2.4)). It thus seems reasonable to conjecture that whatever the maximum frequency achieved by the new lineage, the condition is well approximated by (2.5).

3. Discussion

3.1. Practical use of the equations

3.1.1. Why are accurate predictions desirable?

On the surface, it seems that if one has the ability to increase mutation rate, perhaps through the use of a chemical mutagen, then to drive a population extinct, one needs only to increase the mutation rate by a large amount, perhaps by administering a high dose of mutagen. The problem with this approach is that, in real populations, variation in mutation rate is inevitable, and resistance to a mutagen can appear. A large increase in the mutation rate can create strong selection pressure for a lowered mutation rate, and a reduction in the mutation rate may thus evolve in short order. Our own work with a mutator strain of *Escherichia coli* and a nucleoside analogue mutagen, together with several previous mutagenesis studies using different viral systems, shows that resistance to mutagens at high doses can evolve rapidly and through a number of different mechanisms [76–82]. If one could increase the mutation rate to a level that is high enough to cause extinction, but not too high, selection for resistance could, in principle, be reduced considerably and the evolution of resistance might be prevented. Accurate predictions for the critical mutation rate required for extinction may therefore aid in the practical implementation of chemical mutagenesis, and the evolution of resistance might be prevented. Indeed, our equations and simulations would suggest an improved protocol in which a mutagen is administered in incrementally increasing dose (reflected in figure 3).

3.1.2. Timeframe of applicability

The equations derived here are similar in their generality and robustness; however, they differ among themselves in one aspect of practical relevance, namely, their timeframe of applicability. Under criterion 1, this timeframe ranges from short-term (sufficient) to medium-term (sufficient and somewhat necessary) to long-term (sufficient and necessary). Under criterion 2, the timeframe is short-term (sufficient) or long-term (sufficient and necessary). The long-term results might potentially be applied approximately using a running-average approach that is necessarily somewhat arbitrary, but technically correct application of these results requires information about long-term future states of the population that would not be obtainable in practice. When the mutation rate is adjusted according to fitness measurements from a population taken in real time, the correct equations to use are the

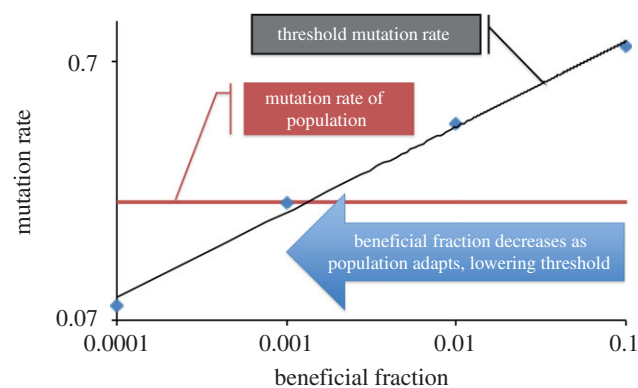


Figure 4. Schematic of how adaptive evolution and/or natural selection may be neutralized not as a result of increasing the mutation rate but as a result of a decreasing threshold mutation rate. The red line indicates the mutation rate of the population; the black line plots the threshold mutation rate as a function of the fraction of mutations that are beneficial (horizontal axis). The big blue arrow indicates that as a population adapts in a static environment, its supply of beneficial mutations is used up, resulting in a decreasing fraction of mutations that are beneficial. As this fraction decreases, the threshold mutation rate decreases, until eventually the threshold mutation rate is below the mutation rate of the population.

short-term and medium-term conditions. These conditions are applied in simulation studies of which representative runs are presented in figure 3; there, adjustments to the mutation rate are made in real time, and the short-term and medium-term conditions derived under criterion 1 (labelled ‘variance’ and ‘variance-projection’ thresholds, respectively) perform well, whereas the conditions derived under criterion 2 (error threshold) appear to be less well suited to such real-time application.

3.1.3. Adaptation in a static environment

A population adapting in a static environment typically has a limited, non-renewable supply of available beneficial mutations (barring intransitive interactions). As the population adapts, therefore, the supply of available beneficial mutations is slowly depleted; as a consequence, mean fitness may increase and subsequently decrease, and fitness variance may also change over time, thereby changing the minimal mutation rate prescribed by criterion 1. In particular, as a population adapts to a static environment, fitness variance should *decrease*, thereby decreasing the predicted threshold mutation rate. Eventually, the threshold mutation rate may decrease to a value that is below the population mutation rate such that further adaptive evolution is neutralized. This is shown schematically in figure 4: in static environments and, generally speaking, in environments where the supply of beneficial mutations can change over time, adaptive evolution or natural selection may be neutralized not as a result of changes in the mutation rate (i.e. changes in U) but as a result of changes in the requirements on the mutation rate (i.e. changes in $\delta_{\tilde{x}}$ and in $\sigma_{\tilde{x}}^2$).

3.2. Connections to previous theory

3.2.1. Fisher and Kimura

Fisher’s ‘fundamental theorem of natural selection’ states that, when x is defined as additive genetic fitness, $d\bar{x}/dt = \sigma_x^2$ quite

generally [59,83]. Fisher's theorem shows that this particular component of fitness can only increase (variance is a non-negative quantity); consequently, this component of fitness has accurately been called the 'adaptive engine' of natural selection [60]. This component of fitness, however, must be conserved over time, for example, in the transmission from parent to offspring for Fisher's theorem to apply. If there is a component of fitness that is not conserved, then to find the change in *total* mean fitness of the population (conserved and non-conserved), one must use the product and chain rules: $d\bar{x}/dt = \int \left(\frac{dx}{dt} u(x, t) + x \frac{d}{dt} u(x, t) \right) dx$ which, together with $\frac{d}{dt} u(x, t) = (x - \bar{x})u(x, t) + \frac{dx}{dt} \frac{\partial}{\partial x} u(x, t)$, yields $d\bar{x}/dt = \sigma_x^2 + \overline{dx/dt}$ —a fact pointed out by Kimura [84]. (We note that $\overline{dx/dt} = \int (d/dt) x u(x, t) dx$ denotes the mean change in 'individual' fitness, where the mean is taken over individuals in the population.) As a general rule, $\overline{dx/dt}$ will be negative, because random alterations in the organism or its environment are more likely to decrease the organism's fitness than to increase it. This simple calculation illustrates the fact that Fisher's fundamental theorem applies only to that subset of the population whose additive genic fitness is conserved over the period of time in question: only for this particular subset of the population are we guaranteed that mean fitness will not decrease.

Applying our criterion 1 to Kimura's equation yields a more general condition for the neutralizing of adaptive evolution:

$$-\overline{dx/dt} > \sigma_x^2 \quad (3.1)$$

must hold persistently or at least on average. Here, the mechanism of change in individual fitness over time is not specified. If we specify that the mechanism of change is mutation, then $\overline{dx/dt} = U \int_{\delta_x} \delta_x g(\delta_x, t) = U \overline{\delta_x}$, where $g(\delta_x, t)$ is the distribution of mutational effects on fitness, and we recover equation (2.1).

3.2.2. The error threshold

As previously stated, equations (2.4) and (2.5) are the error threshold in new guise. The original work on the error threshold due to Eigen [16] derives a minimum value for the 'quality factor'—the probability of complete fidelity of replication—that is needed to maintain the efficacy of natural selection and thus to support life. This minimum value is given by $Q_{\min} = (\bar{A}_{k \neq m} + \mathcal{D}_m - \bar{D}_{k \neq m}) / \mathcal{A}_m$ (equation II-45 in Eigen [16]), where \mathcal{A}_m and \mathcal{D}_m are the birth and death rates of the fittest genotype (the 'master sequence'), respectively, $\bar{A}_{k \neq m}$ and $\bar{D}_{k \neq m}$ are the mean birth and death rates of the rest of the population (individuals that do not carry the 'master sequence'). The quantity $\mathcal{A}_m(1 - Q_{\min})$ is the expected number of deleterious mutants produced by a single replication event of the fittest genotype. This quantity, in our notation, is Uf_D ; furthermore, $\mathcal{A}_m - \mathcal{D}_m$ is equivalent to our x_0 and $\bar{A}_{k \neq m} - \bar{D}_{k \neq m}$ is equivalent to our \tilde{x} . Eigen's result may thus be rewritten in our notation as requiring $Uf_D < x_0 - \tilde{x}$ for the effectiveness of natural selection to be maintained, or conversely, $Uf_D \geq x_0 - \tilde{x}$ for natural selection to be neutralized.

In work subsequent to Eigen's original publication, the varied presentations of his error threshold result are usually rearrangements of this simple expression: $q_{\min}^L = \sigma^{-1}$, where q_{\min} is the minimum per-nucleotide

replication fidelity required for survival ($q_{\min} = 1 - \mu_c$), L is the length of the deleterious genome, and σ is the 'superiority parameter', defined as $\sigma = 1/Q_{\min}$. Rewriting reveals an interesting biological requirement: $\log q_{\min}^L \approx -\mu_c L$ gives rise to the relation

$$\mu_c \approx (\text{something})/L. \quad (3.2)$$

This inverse relation between μ_c and L intrigued its discoverers to the extent that the 'something' was all but ignored. It was since discovered, however, that observations of μL are surprisingly constant across microbial taxa [85,86] (indeed, it has been conjectured that this is the case precisely because of the inverse relation between μ_c and L). The relative constancy of μL across taxa suggests that the 'something' may in fact be quite relevant to the fate of a population; furthermore, L will probably not change on time scales pertinent to extinction-by-mutation. These considerations shift the focus to σ . Its name together with its traditional presentation obfuscates the fact that σ is a population-dependent *quantity* and not an organism-dependent *parameter*. Our new presentation of this old result shifts the emphasis from critical *point* mutation rate μ_c versus genome length L to critical *genomic* mutation rate $\mu_c L$ versus the myriad biological, ecological and environmental factors that are not explicitly part of the equation but that are absorbed by the quantity σ or, in our formulation, $x_0 - \tilde{x}$.

3.3. Connections among results presented here

Our first criterion is the sustained decline of absolute fitness, whereas our second criterion is the inefficacy of natural selection. We now show that, despite these perhaps disparate criteria, the resulting conditions for extinction connect through classical population genetics. Criterion 1 gives rise to the condition $-U\overline{\delta_x} > \sigma_x^2$, and criterion 2 gives rise to the condition $Uf_D \geq x_0 - \tilde{x}$. Our comparison of these two results proceeds by multiplying both sides of the second condition by m_D to obtain $Uf_D m_D \geq (x_0 - \tilde{x})m_D$. First, we note that the left-hand side is $Uf_D m_D \approx -U\overline{\delta_x}$ because most mutations are deleterious. Next, we focus on the right-hand side of the inequality. As a population approaches the error threshold (i.e. as this inequality approaches equality), the size of the fittest class approaches zero and it is the case that $(1/\#S) \sum_{i \in S} x_i \rightarrow (1/N) \sum_{i=1}^N x_i$, or $\tilde{x} \rightarrow \bar{x}$. The quantity $x_0 - \bar{x}$ is known in classical population genetics as the genetic load, and it is known to converge to the deleterious mutation rate Uf_D . Furthermore, it is known that if mutations are assumed to have a fixed deleterious effect, m_D , then the number of accumulated mutations becomes Poisson distributed with mean Uf_D/m_D [6]. The variance in number of accumulated mutations is the same as the mean, and the variance in *fitness* is therefore $\sigma_x^2 = (Uf_D/m_D)m_D^2 = Uf_D m_D$. As the error threshold is approached, therefore, the right-hand side becomes $(x_0 - \tilde{x})m_D \rightarrow (x_0 - \bar{x})m_D = Uf_D m_D = \sigma_x^2$.

3.4. Borrowed robustness

Fisher's fundamental theorem of natural selection is known to be extraordinarily accurate in spite of numerous complexities that are characteristic of real populations. Because (2.1) is implicit in the results of Fisher and Kimura, therefore, we expect these results to be quite robust to numerous biological complexities. Furthermore, the convergence we have demonstrated between (2.1), (2.3) and (2.5) leads us to believe that

the classical error threshold result is similarly robust, although it does not appear to perform as well in real time (figure 3). Figure 2 together with the plots we have posted in the electronic supplementary material—and many others not posted—demonstrate the robustness of (2.1) (and by inference (2.2)) to a wide range of complexities, including finite genome effects, the effects of finite population size (including Muller's ratchet), epistatic interactions among mutations, environmental noise (random changes in fitness caused by unspecified factors), an evolving mutational robustness modifier, compensatory mutations whose rate increases with decreasing fitness, an evolving mutation rate modifier and a fraction of mutations that are lethal.

Special thanks to Cristian Batista for insightful explanations of the error threshold as a phase transition, to Isabel Gordo for helping make connections among the different theories and to Claus Wilke for helpful comments and clarifications. We also thank Michael Lässig, Paul Joyce, Alan Perelson, Boris Shraiman, Sidhartha Goyal, Daniel Balick, Nico Stollenwerk, Gabriela Gomes, Ana Margarida Sousa, Jorge Carneiro and Josep Sardanyés for helpful discussions, and two anonymous reviewers for helpful comments. Much of this research was developed thanks to fertile environments provided by two institutes: the Kavli Institute for Theoretical Physics in Santa Barbara, CA (2011 Microbial and Viral Evolution workshop), and the Instituto Gulbenkian de Ciências in Oeiras, Portugal. This work was supported by the US National Institutes of Health grants: R01 GM079843-01 (P.J.G./P.D.S.), R01 GM079483-02S1 (P.J.G./P.D.S.), a seed grant through 1P20RR18754 (Center for Evolutionary and Theoretical Immunology) (P.J.G.), UM1-AI100645-01 (Center for HIV/AIDS Vaccine Immunology-Immunogen Design; P.J.G.); and European Commission grant no. FP7 231807 (P.J.G.).

Appendix A

The results we describe in the main text derive from two manifestations of a standard model of evolution described verbally in §2.1. Here, we give the mathematical details of those manifestations.

A.1. Model in continuous fitness for criterion 1

We let $u(x, t)$ denote the density of individuals in the population with log-fitness x at time t . Mutation can create 'jumps' in log-fitness whose size has probability density $g(\phi, t)$ at time t . Under selection and mutation, a population's evolution is described by:

$$\frac{\partial}{\partial t} u(x, t) = (x - \bar{x})u(x, t) + U \left(\int_{-\infty}^{+\infty} u(x - \phi, t) g(\phi, t) d\phi - u(x, t) \right), \quad (\text{A } 1)$$

where $\bar{x} = \int_{-\infty}^{+\infty} xu(x, t) dx$. If we apply the standard diffusion approximation to the mutation term, then this equation becomes

$$\frac{\partial}{\partial t} u(x, t) = (x - \bar{x})u(x, t) + U \mathbf{M} u(x, t). \quad (\text{A } 2)$$

Mutation operator, $\mathbf{M} = D_x \frac{\partial^2}{\partial x^2} - d_x \frac{\partial}{\partial x}$, where $D_x = \frac{1}{2} f_B (m_B^2 + \sigma_B^2) + \frac{1}{2} f_D (m_D^2 + \sigma_D^2)$ and $d_x = f_B m_B - f_D m_D$; f_B is the fraction of all mutations that are beneficial ('beneficial fraction'), f_D is the deleterious fraction; m_B and m_D are the mean effects of beneficial and deleterious mutations on fitness, respectively; σ_B^2 and σ_D^2 are the variances in those effects. We multiply both sides of (A 2) by x and integrate over all x to

obtain $\dot{\bar{x}} = \sigma_x^2 + U \int_{-\infty}^{+\infty} \mathbf{M} x u(x, t) dx$. Under the reasonable assumption that $u(x, t)$ has compact support in x , integration by parts gives $\dot{\bar{x}} = \sigma_x^2 + U \bar{\delta}_x$, where $\bar{\delta}_x = f_B m_B - f_D m_D$. The condition $\dot{\bar{x}} < 0$ reflects the neutralizing of adaptive evolution and is met when $-U \bar{\delta}_x > \sigma_x^2$.

A.2. Model in discrete fitness for criterion 2

As an indication of the amount of order in the system at hand, we would like to know the frequency of the fittest genotype relative to its mutational neighbours. The dynamics of this genotype and its mutational neighbours (genotypes that differ from the fittest genotype by mutation) are given by this set of equations:

$$\dot{u}_i = \left(x_i - \sum_{j=0}^L x_j u_j \right) u_i - L \mu u_i + \mu \sum_{j \neq i} u_j, \quad (\text{A } 3)$$

where u_0 is the frequency of the fittest genotype (the order parameter), u_i is the frequency of mutational neighbour, $i = 1, 2, 3, \dots, L$, x_i is fitness of genotype i , and μ is point mutation rate.

The equation for the fittest genotype u_0 may be written as

$$\dot{u}_0 = (x_0 - \bar{x})(1 - u_0)u_0 - \mu L u_0 + \mu(1 - u_0), \quad (\text{A } 4)$$

where x_0 is the fitness of the fittest genotype and \bar{x} is the average fitness of everybody else: $\bar{x} = \sum_{j=1}^L x_j u_j / (1 - u_0)$. We note that $x_0 - \bar{x}$ is not relative fitness; a possible interpretation of the value $x_0 - \bar{x}$ is that it is the reproductive 'pay-off' in a game played by the fittest genotype against everybody else.

A.3. Calculating the 'sufficient and somewhat necessary' conditions under criterion 1

To compute the 'sufficient and somewhat necessary' conditions requires projection of cumulants $\kappa_i(\tau)$ over a period of r generations into the future. Recurrence relations that do this are developed in Gerrish & Sniegowski [72].

The terms of the sum in (2.2) are computed from the recurrence relation: $\kappa_i(\tau) = \kappa_i(\tau - 1) + \kappa_{i+1}(\tau - 1) + U m_i$ for all $i \geq 1$, where $\kappa_i(\tau)$ is the i th cumulant in fitness at a time τ generations from now, U is genomic mutation rate and m_i is the i th raw moment of the distribution of mutational effects on fitness.

The practical implementation of condition (2.2) requires some care. The procedure outlined in Gerrish & Sniegowski [72] provides methods for estimating the m_j . These parameters cannot be estimated separately from U ; only their products $U m_j$ can be estimated, if the equations are left in non-parametric form. The obvious remedy is to make the equations parametric by writing the known expressions for the moments of an assumed distribution in place of m_j . Then, the parameters to be estimated are U and the limited number of parameters of the assumed distribution, and U can then be estimated separately. If one's objective is to monitor a population's risk of extinction, or to drive a population extinct through mutagenesis, however, a less obvious remedy may apply. In such cases, absolute mutation rates may be irrelevant, and the effects of an increased (or decreased) mutation rate can be predicted by simply multiplying the estimates of $U m_j$ by the factor by which mutation rate is increased (or decreased). In such cases, therefore, the equations may be left in non-parametric form.

1. Carrasco P, de la Iglesia F, Elena SF. 2007 Distribution of fitness and virulence effects caused by single-nucleotide substitutions in tobacco etch virus. *J. Virol.* **81**, 12 979–12 984. (doi:10.1128/JVI.00524-07)
2. Sanjuán R. 2010 Mutational fitness effects in RNA and single-stranded DNA viruses: common patterns revealed by site-directed mutagenesis studies. *Phil. Trans. R. Soc. B* **365**, 1975–1982. (doi:10.1098/rstb.2010.0063)
3. Sanjuán R, Moya A, Elena SF. 2004 The distribution of fitness effects caused by single-nucleotide substitutions in an RNA virus. *Proc. Natl Acad. Sci. USA* **101**, 8396–8401. (doi:10.1073/pnas.0400146101)
4. Muller H. 1932 Some genetic aspects of sex. *Am. Nat.* **66**, 118–138. (doi:10.1086/280418)
5. Felsenstein J. 1974 The evolutionary advantage of recombination. *Genetics* **78**, 737–756.
6. Haigh J. 1978 The accumulation of deleterious genes in a population: Muller's ratchet. *Theor. Popul. Biol.* **14**, 251–267. (doi:10.1016/0040-5809(78)90027-8)
7. Andersson D, Hughes D. 1996 Muller's ratchet decreases fitness of a DNA-based microbe. *Proc. Natl Acad. Sci. USA* **93**, 906–907. (doi:10.1073/pnas.93.2.906)
8. Chao L, Tran T, Matthews C. 1992 Muller's ratchet and the advantage of sex in the RNA virus 6. *Evolution* **46**, 289–299. (doi:10.2307/2409851)
9. Fontanari JF, Colato A, Howard RS. 2003 Mutation accumulation in growing asexual lineages. *Phys. Rev. Lett.* **91**, 218101. (doi:10.1103/PhysRevLett.91.218101)
10. Gordo I, Charlesworth B. 2000 On the speed of Muller's ratchet. *Genetics* **156**, 2137–2140.
11. Green DM. 1990 Muller's Ratchet and the evolution of supernumerary chromosomes. *Genome* **33**, 818–824. (doi:10.1139/g90-123)
12. Heller R, Smith JM. 2009 Does Muller's ratchet work with selfing? *Genet. Res.* **32**, 289–293. (doi:10.1017/S0016672300018784)
13. Kondrashov AS. 1984 Deleterious mutations as an evolutionary factor: 1. The advantage of recombination. *Genet. Res.* **44**, 199–217. (doi:10.1017/S0016672300026392)
14. Loewe L, Lamatsch DK. 2008 Quantifying the threat of extinction from Muller's ratchet in the diploid Amazon molly (*Poecilia formosa*). *BMC Evol. Biol.* **8**, 88. (doi:10.1186/1471-2148-8-88)
15. Yuste E, Sanchez-Palmino S, Casado C, Domingo E, López-Galíndez C. 1999 Drastic fitness loss in human immunodeficiency virus type 1 upon serial bottleneck events. *J. Virol.* **73**, 2745–2751.
16. Eigen M. 1971 Self organization of matter and the evolution of biological macro-molecules. *Die Naturwissenschaften* **58**, 465–523. (doi:10.1007/BF00623322)
17. Eigen M. 2002 Error catastrophe and antiviral strategy. *Proc. Natl Acad. Sci. USA* **99**, 13 374–13 376. (doi:10.1073/pnas.212514799)
18. Biebricher CK, Eigen M. 2005 The error threshold. *Virus Res.* **107**, 117–127. (doi:10.1016/j.virusres.2004.11.002)
19. Biebricher CK, Eigen M. 2006 What is a quasispecies? *Curr. Top. Microbiol. Immunol.* **299**, 1–31. (doi:10.1007/3-540-26397-7_1)
20. Eigen M. 2000 Natural selection: a phase transition? *Biophys. Chem.* **85**, 101–123. (doi:10.1016/S0301-4622(00)00122-8)
21. Wolff A, Krug J. 2009 Robustness and epistasis in mutation–selection models. *Phys. Biol.* **6**, 036007. (doi:10.1088/1478-3975/6/3/036007)
22. Hermisson J, Redner O, Wagner H, Baake E. 2002 Mutation–selection balance: ancestry, load, and maximum principle. *Theor. Popul. Biol.* **62**, 9–46. (doi:10.1006/tpbi.2002.1582)
23. Tarazona P. 1992 Error thresholds for molecular quasispecies as phase transitions: from simple landscapes to spin-glass models. *Phys. Rev. A* **45**, 6038–6050. (doi:10.1103/PhysRevA.45.6038)
24. Summers J, Litwin S. 2006 Examining the theory of error catastrophe. *J. Virol.* **80**, 20–26. (doi:10.1128/JVI.80.1.20-26.2006)
25. Boerlijst MC, Bonhoeffer S, Nowak MA. 1996 Viral quasi-species and recombination. *Proc. R. Soc. Biol. Sci.* **263**, 1577–1584. (doi:10.1098/rspb.1996.0231)
26. Nilsson M, Snoad N. 2000 Error thresholds for quasispecies on dynamic fitness landscapes. *Phys. Rev. Lett.* **84**, 191–194. (doi:10.1103/PhysRevLett.84.191)
27. Ochoa G, Harvey I. 1999 Recombination and error thresholds in finite populations. In *Foundations of genetic algorithms 5* (eds W Banzhaf, C Reeves), pp. 245–264. San Francisco, CA: Morgan Kaufmann Publishers, Inc.
28. Sardanyés J, Elena SF. 2010 Error threshold in RNA quasispecies models with complementation. *J. Theor. Biol.* **265**, 278–286. (doi:10.1016/j.jtbi.2010.05.018)
29. Elena SF, Solé RV, Sardanyés J. 2010 Simple genomes, complex interactions: epistasis in RNA virus. *Chaos* **20**, 026106. (doi:10.1063/1.3449300)
30. Sardanyés J, Elena SF. 2011 Quasispecies spatial models for RNA viruses with different replication modes and infection strategies. *PLoS ONE* **6**, e24884. (doi:10.1371/journal.pone.0024884)
31. Sardanyés J, Martínez F, Darós J-A, Elena SF. 2012 Dynamics of alternative modes of RNA replication for positive-sense RNA viruses. *J. R. Soc. Interface* **9**, 768–776. (doi:10.1098/rsif.2011.0471)
32. Sardanyés J, Solé RV. 2006 Bifurcations and phase transitions in spatially extended two-member hypercycles. *J. Theor. Biol.* **243**, 468–482. (doi:10.1016/j.jtbi.2006.07.014)
33. Sardanyés J, Solé RV, Elena SF. 2009 Replication mode and landscape topology differentially affect RNA virus mutational load and robustness. *J. Virol.* **83**, 12 579–12 589. (doi:10.1128/JVI.00767-09)
34. Solé R, Sardanyés J, Diez J, Mas A. 2006 Information catastrophe in RNA viruses through replication thresholds. *J. Theor. Biol.* **240**, 353–359. (doi:10.1016/j.jtbi.2005.09.024)
35. Bonhoeffer S, Stadler P. 1993 Error thresholds on correlated fitness landscapes. *J. Theor. Biol.* **164**, 359–372. (doi:10.1006/jtbi.1993.1160)
36. Gerrish PJ. 2009 Some observations about the nearest-neighbor model of the error threshold. *AIP Conf. Proc.* **1168**, 1564–1568. (doi:10.1063/1.3241401)
37. Takeuchi N, Hogeweg P. 2007 Error-threshold exists in fitness landscapes with lethal mutants. *BMC Evol. Biol.* **7**, 15. (doi:10.1186/1471-2148-7-15)
38. Takeuchi N, Poorthuis PH, Hogeweg P. 2005 Phenotypic error threshold; additivity and epistasis in RNA evolution. *BMC Evol. Biol.* **5**, 9. (doi:10.1186/1471-2148-5-9)
39. Tejero H, Marín A, Montero F. 2010 Effect of lethality on the extinction and on the error threshold of quasispecies. *J. Theor. Biol.* **262**, 733–741. (doi:10.1016/j.jtbi.2009.10.011)
40. Wilke CO, Ronnewinkel C, Martinetz T. 2001 Dynamic fitness landscapes in molecular evolution. *Phys. Rep.* **349**, 395–446. (doi:10.1016/S0370-1573(00)00118-6)
41. Wiehe T. 1997 Model dependency of error thresholds: the role of fitness functions and contrasts between the finite and infinite sites models. *Genet. Res.* **69**, 127–136. (doi:10.1017/S0016672397002619)
42. Gabriel W, Lynch M, Burger R. 1993 Muller's ratchet and mutational melt-downs. *Evolution* **47**, 1744–1757. (doi:10.2307/2410218)
43. Lynch M, Burger R, Butcher D, Gabriel W. 1993 The mutational meltdown in asexual populations. *J. Hered.* **84**, 339–344.
44. Zeyl C, Mizesko M, de Visser JA. 2001 Mutational meltdown in laboratory yeast populations. *Evolution* **55**, 909–917. (doi:10.1554/0014-3820(2001)055[0909:MMILYP]2.0.CO;2)
45. Bagnoli F, Bezzi M. 1998 Eigen's error threshold and mutational meltdown in a quasispecies model. *Int. J. Mod. Phys.* **9**, 999–1005. (doi:10.1142/S0129183198000935)
46. Malarz K, Tiggemann D. 1998 Dynamics in Eigen quasispecies model. *Int. J. Mod. Phys. C* **9**, 481–490. (doi:10.1142/S0129183198000376)
47. Bachtrog D, Gordo I. 2004 Adaptive evolution of asexual populations under Muller's ratchet. *Evolution* **58**, 1403–1413. (doi:10.1554/03-595)
48. Balick D, Goyal S, Jerison E, Neher R, Shraiman B, Desai M. 2012 Rare beneficial mutations can halt Muller's ratchet. *Bull. Am. Phys. Soc.* **57**, abstract BAPS.2012.MAR.B51.5.
49. Goyal S, Balick DJ, Jerison ER, Neher RA, Shraiman BI, Desai MM. 2012 Dynamic mutation–selection balance as an evolutionary attractor. *Genetics* **191**, 1309–1319. (doi:10.1534/genetics.112.141291)
50. Haldane JBS. 1927 The mathematical theory of natural and artificial selection. *Proc. Camb. Phil. Soc.* **23**, 838–844. (doi:10.1017/S0305004100015644)

51. Hill WG, Robertson A. 1966 The effect of linkage on limits to artificial selection. *Genet. Res.* **8**, 269–294. (doi:10.1017/S0016672300010156)
52. Gerrish PJ, Lenski RE. 1998 The fate of competing beneficial mutations in an asexual population. *Genetica* **102–103**, 127–144. (doi:10.1023/A:1017067816551)
53. Anderson JP, Daifuku R, Loeb LA. 2004 Viral error catastrophe by mutagenic nucleosides. *Annu. Rev. Microbiol.* **58**, 183–205. (doi:10.1146/annurev.micro.58.030603.123649)
54. Bürger R. 1998 Lethal mutagenesis and evolutionary epidemiology. *Genetica* **102–103**, 279–298. (doi:10.1023/A:1017043111100)
55. Chen R, Quinones-Mateu ME, Mansky LM. 2004 Drug resistance, virus fitness and HIV-1 mutagenesis. *Curr. Pharm. Des.* **10**, 4065–4070. (doi:10.2174/1381612043382404)
56. Chen R, Yokoyama M, Sato H, Reilly C, Mansky LM. 2005 Human immunodeficiency virus mutagenesis during antiviral therapy: impact of drug-resistant reverse transcriptase and nucleoside and nonnucleoside reverse transcriptase inhibitors on human immunodeficiency virus type 1 mutation frequencies. *J. Virol.* **79**, 12 045–12 057. (doi:10.1128/JVI.79.18.12045-12057.2005)
57. Loeb LA, Essigmann JM, Kazazi F, Zhang J, Rose KD, Mullins JI. 1999 Lethal mutagenesis of HIV with mutagenic nucleoside analogs. *Proc. Natl Acad. Sci. USA* **96**, 1492–1497. (doi:10.1073/pnas.96.4.1492)
58. Ewens W. 1989 An interpretation and proof of the fundamental theorem of natural selection. *Theor. Popul. Biol.* **36**, 167–180. (doi:10.1016/0040-5809(89)90028-2)
59. Frank S, Slatkin M. 1992 Fisher's fundamental theorem of natural selection. *Trends Ecol. Evol.* **7**, 92–95. (doi:10.1016/0169-5347(92)90248-A)
60. Grafen A. 2003 Fisher the evolutionary biologist. *J. R. Stat. Soc. D* **52**, 319–329. (doi:10.1111/1467-9884.00362)
61. Frank SA. 1995 George Price's contributions to evolutionary genetics. *J. Theor. Biol.* **175**, 373–388. (doi:10.1006/jtbi.1995.0148)
62. Gardner A. 2008 The Price equation. *Curr. Biol.* **18**, R198–R202. (doi:10.1016/j.cub.2008.01.005)
63. Desai MM, Fisher DS. 2007 Beneficial mutation selection balance and the effect of linkage on positive selection. *Genetics* **176**, 1759–1798. (doi:10.1534/genetics.106.067678)
64. Joseph S, Hall D. 2004 Spontaneous mutations in diploid *Saccharomyces cerevisiae*: more beneficial than expected. *Genetics* **168**, 1817–1825. (doi:10.1534/genetics.104.033761)
65. Perfeito L, Fernandes L, Mota C, Gordo I. 2007 Adaptive mutations in bacteria: high rate and small effects. *Science* **317**, 813–815. (doi:10.1126/science.1142284)
66. Silander OK, Tenaillon O, Chao L. 2007 Understanding the evolutionary fate of finite populations: the dynamics of mutational effects. *PLoS Biol.* **5**, e94. (doi:10.1371/journal.pbio.0050094)
67. Sniegowski PD, Gerrish PJ. 2010 Beneficial mutations and the dynamics of adaptation in asexual populations. *Phil. Trans. R. Soc. B* **365**, 1255–1263. (doi:10.1098/rstb.2009.0290)
68. Wloch D, Szafraniec K, Borts R, Korona R. 2001 Direct estimate of the mutation rate and the distribution of fitness effects in the yeast *Saccharomyces cerevisiae*. *Genetics* **159**, 441–452.
69. Brunet E, Rouzine IM, Wilke CO. 2008 The stochastic edge in adaptive evolution. *Genetics* **179**, 603–620. (doi:10.1534/genetics.107.079319)
70. Rouzine IM, Wakeley J, Coffin JM. 2003 The solitary wave of asexual evolution. *Proc. Natl Acad. Sci. USA* **100**, 587–592. (doi:10.1073/pnas.242719299)
71. Frank SA, Slatkin M. 1990 The distribution of allelic effects under mutation and selection. *Genet. Res.* **55**, 111–117. (doi:10.1017/S0016672300025350)
72. Gerrish PJ, Sniegowski PD. 2012 Real time forecasting of near-future evolution. *J. R. Soc. Interface* **9**, 2268–2278. (doi:10.1098/rsif.2012.0119)
73. Bull JJ, Sanjuán R, Wilke CO. 2007 Theory of lethal mutagenesis for viruses. *J. Virol.* **81**, 2930–2939. (doi:10.1128/JVI.01624-06)
74. Bull JJ, Wilke CO. 2008 Lethal mutagenesis of bacteria. *Genetics* **180**, 1061–1070. (doi:10.1534/genetics.108.091413)
75. Gerrish PJ, Colato A, Perelson AS, Sniegowski PD. 2007 Complete genetic linkage can subvert natural selection. *Proc. Natl Acad. Sci. USA* **104**, 6266–6271. (doi:10.1073/pnas.0607280104)
76. Agudo R, Ferrer-Orta C, Arias A, de la Higuera I, Perales C, Pérez-Luque R, Verdagué N, Domingo E. 2010 A multi-step process of viral adaptation to a mutagenic nucleoside analogue by modulation of transition types leads to extinction-escape. *PLoS Pathog.* **6**, e1001072. (doi:10.1371/journal.ppat.1001072)
77. Arias A, Arnold JJ, Sierra M, Smidansky ED, Domingo E, Cameron CE. 2008 Determinants of RNA-dependent RNA polymerase (in)fidelity revealed by kinetic analysis of the polymerase encoded by a foot-and-mouth disease virus mutant with reduced sensitivity to ribavirin. *J. Virol.* **82**, 12 346–12 355. (doi:10.1128/JVI.01297-08)
78. Coffey LL, Beeharry Y, Borderia AV, Blanc H, Vignuzzi M. 2011 Arbovirus high fidelity variant loses fitness in mosquitoes and mice. *Proc. Natl Acad. Sci. USA* **108**, 16 038–16 043. (doi:10.1073/pnas.1111650108)
79. Domingo-Calap P, Pereira-Gómez M, Sanjuán R. 2012 Nucleoside analogue mutagenesis of a single-stranded DNA virus: evolution and resistance. *J. Virol.* **86**, 9640–9646. (doi:10.1128/JVI.00613-12)
80. Graci JD, Gnädig NF, Galarraza JE, Castro C, Vignuzzi M, Cameron CE. 2012 Mutational robustness of an RNA virus influences sensitivity to lethal mutagenesis. *J. Virol.* **86**, 2869–2873. (doi:10.1128/JVI.05712-11)
81. Pfeiffer JK, Kirkegaard K. 2003 A single mutation in poliovirus RNA-dependent RNA polymerase confers resistance to mutagenic nucleotide analogs via increased fidelity. *Proc. Natl Acad. Sci. USA* **100**, 7289–7294. (doi:10.1073/pnas.1232294100)
82. Sanjuán R, Cuevas JM, Furió V, Holmes EC, Moya A. 2007 Selection for robustness in mutagenized RNA viruses. *PLoS Genet.* **3**, e93. (doi:10.1371/journal.pgen.0030093)
83. Fisher RA. 1930 *The genetical theory of natural selection*. Oxford, UK: Clarendon Press.
84. Kimura M. 1958 On the change of population fitness by natural selection. *Heredity* **12**, 145–167. (doi:10.1038/hdy.1958.21)
85. Drake J. 1991 A constant rate of spontaneous mutation in DNA-based microbes. *Proc. Natl Acad. Sci. USA* **88**, 7160–7164. (doi:10.1073/pnas.88.16.7160)
86. Drake J. 1993 Rates of spontaneous mutation among RNA viruses. *Proc. Natl Acad. Sci. USA* **90**, 4171–4175. (doi:10.1073/pnas.90.9.4171)