

Complete Genome Sequence of a Virulent Strain, *Streptococcus iniae* ISET0901, Isolated from Diseased Tilapia

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***Streptococcus iniae* ISET0901 is a virulent strain isolated in 2007 from diseased tilapia. Its full genome is 2,070,856 bp. The availability of this genome will allow comparative genomics to identify virulence genes important for the pathogenesis of streptococcosis caused by *S. iniae*, as well as possible immunogens for vaccine development.**

Received 15 May 2014 Accepted 20 May 2014 Published 5 June 2014

Citation Pridgeon JW, Zhang D, Zhang L. 2014. Complete genome sequence of a virulent strain, *Streptococcus iniae* ISET0901, isolated from diseased tilapia. *Genome Announc.* 2(3):e00553-14. doi:10.1128/genomeA.00553-14.

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The Gram-positive bacterium *Streptococcus iniae* is a zoonotic pathogen that causes disease in both humans and fish (1–3). In aquaculture, *S. iniae* is a serious marine and freshwater fish pathogen causing significant economic losses (4, 5). Originally isolated from Amazon freshwater dolphin (*Inia geoffrensis*) in 1976 (6), *S. iniae* causes mortalities in >30 species of fish, including rainbow trout (*Oncorhynchus mykiss* [7]), barramundi (*Lates calcarifer* [8]), red drum (*Sciaenops ocellatus* [9]), flounder (*Paralichthys* spp. [10, 11]), and tilapia (*Oreochromis* spp. [12]). Strain *S. iniae* ISET0901 was cultured from diseased Nile tilapia (*Oreochromis niloticus*) during a disease outbreak in 2005 (13). Virulence studies (13) revealed that *S. iniae* ISET0901 is highly virulent to Nile tilapia. However, the virulence factors associated with the genome of *S. iniae* ISET0901 are unknown. Therefore, the complete genome sequence of *S. iniae* ISET0901 was determined in this study.

The genome of *S. iniae* ISET0901 was sequenced using the Illumina 1500 HiSeq platform. BioNumerics (Applied Maths) was used to assemble a total of 23,446,702 sequence reads, with an average length of 100.21 bp (estimated 1,134× coverage), using the complete genome of *S. iniae* SF1 (GenBank accession no. CP005941) as a reference. The assembled genome of *S. iniae* ISET0901 is 2,070,822 bp, with a G+C content of 36.8%. RNAmmer (14) predicted 12 copies of rRNA (4 copies of 5S RNA, 16S RNA, and 23S RNA), which is similar to that in the reference genome of *S. iniae* SF1 (15). The RAST server (16) predicted 1,982 coding sequences belonging to 303 subsystems, including 291 involved in carbohydrate catabolism, 149 in protein metabolism, 135 in the synthesis of amino acids and derivatives, 114 in cell wall and capsule synthesis, 95 in RNA metabolism, and 92 in DNA metabolism, including 77 in cofactors, vitamins, prosthetic groups, or pigments, 62 in nucleoside and nucleotide synthesis, 62 in fatty acid and lipid synthesis, 52 involved in virulence, disease, and defense, 47 in membrane transport, 37 in stress response, 31 in phosphorus metabolism, 26 in regulation and cell signaling, 7 in secondary metabolism, and 2 in motility and chemotaxis.

Nucleotide sequence accession number. The complete genome sequence of *S. iniae* ISET0901 was deposited at GenBank under the accession no. [CP007586](http://dx.doi.org/10.1128/genomeA.00553-14).

ACKNOWLEDGMENTS

This study was supported by the USDA/ARS CRIS project 6420-32000-024-00D.

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We thank James Becnel (USDA-ARS) and Laura Silo-Suh (Mercer University) for critical reviews of the manuscript. We thank Beth Peterman (USDA-ARS) for her technical support.

REFERENCES

1. Sun JR, Yan JC, Yeh CY, Lee SY, Lu JJ. 2007. Invasive infection with *Streptococcus iniae* in Taiwan. *J. Med. Microbiol.* 56:1246–1249. <http://dx.doi.org/10.1099/jmm.0.47180-0>.
2. Lau SK, Woo PC, Luk WK, Fung AM, Hui WT, Fong AH, Chow CW, Wong SS, Yuen KY. 2006. Clinical isolates of *Streptococcus iniae* from Asia are more mucoid and beta-hemolytic than those from North America. *Diagn. Microbiol. Infect. Dis.* 54:177–181. <http://dx.doi.org/10.1016/j.diagmicrobio.2005.09.012>.
3. Miller JD, Neely MN. 2005. Large-scale screen highlights the importance of capsule for virulence in the zoonotic pathogen *Streptococcus iniae*. *Infect. Immun.* 73:921–934. <http://dx.doi.org/10.1128/IAI.73.2.921-934.2005>.
4. Agnew W, Barnes AC. 2007. *Streptococcus iniae*: an aquatic pathogen of global veterinary significance and a challenging candidate for reliable vaccination. *Vet. Microbiol.* 122:1–15. <http://dx.doi.org/10.1016/j.vetmic.2007.03.002>.
5. El Aamri F, Caballero MJ, Real F, Acosta F, Déniz S, Román L, Padilla D. *Streptococcus iniae* in gilthead seabream (*Sparus aurata*, L.) and red porgy (*Pagrus pagrus*, L.): ultrastructural analysis. *Vet. Pathol.*, in press.
6. Pier GB, Madin SH. 1976. *Streptococcus iniae* sp. nov., a beta-hemolytic streptococcus isolated from an Amazon freshwater dolphin, *Inia geoffrensis*. *Int. J. Syst. Bacteriol.* 26:545–553. <http://dx.doi.org/10.1099/00207713-26-4-545>.
7. Eyangor M, Tekoah Y, Shapira R, Hurvitz A, Zlotkin A, Lublin A, Eldar A. 2008. Emergence of novel *Streptococcus iniae* exopolysaccharide-

- producing strains following vaccination with nonproducing strains. *Appl. Environ. Microbiol.* 74:6892–6868. <http://dx.doi.org/10.1128/AEM.00853-08>.
8. Bromage ES, Thomas A, Owens L. 1999. *Streptococcus iniae*, a bacterial infection in barramundi *Lates calcarifer*. *Dis. Aquat. Organ.* 36:177–181. <http://dx.doi.org/10.3354/dao036177>.
 9. Eldar A, Perl S, Frelier PF, Bercovier H. 1999. Red drum *Sciaenops ocellatus* mortalities associated with *Streptococcus iniae* infection. *Dis. Aquat. Organ.* 36:121–127. <http://dx.doi.org/10.3354/dao036121>.
 10. Nho SW, Shin GW, Park SB, Jang HB, Cha IS, Ha MA, Kim YR, Park YK, Dalvi RS, Kang BJ, Joh SJ, Jung TS. 2009. Phenotypic characteristics of *Streptococcus iniae* and *Streptococcus parauberis* isolated from olive flounder (*Paralichthys olivaceus*). *FEMS Microbiol. Lett.* 293:20–27. <http://dx.doi.org/10.1111/j.1574-6968.2009.01491.x>.
 11. Nguyen HT, Kanai K. 1999. Selective agar for the isolation of *Streptococcus iniae* from Japanese flounder, *Paralichthys olivaceus*, and its cultural environment. *J. Appl. Microbiol.* 86:769–776. <http://dx.doi.org/10.1046/j.1365-2672.1999.00724.x>.
 12. Zhou SM, Xie MQ, Zhu XQ, Ma Y, Tan ZL, Li AX. 2008. Identification and genetic characterization of *Streptococcus iniae* strains isolated from diseased fish in China. *J. Fish Dis.* 31:869–875. <http://dx.doi.org/10.1111/j.1365-2761.2008.00954.x>.
 13. Pridgeon JW, Klesius PH. 2011. Development and efficacy of a novobiocin-resistant *Streptococcus iniae* as a novel vaccine in Nile tilapia (*Oreochromis niloticus*). *Vaccine* 29:5986–5993. <http://dx.doi.org/10.1016/j.vaccine.2011.06.036>.
 14. Lagesen K, Hallin P, Rødland EA, Staerfeldt HH, Rognes T, Ussery DW. 2007. RNAmmer: consistent and rapid annotation of ribosomal RNA genes. *Nucleic Acids Res.* 35:3100–3108. <http://dx.doi.org/10.1093/nar/gkm160>.
 15. Zhang BC, Zhang J, Sun L. 2014. *Streptococcus iniae* SF1: complete genome sequence, proteomic profile, and immunoprotective antigens. *PLoS One* 9:e91324. <http://dx.doi.org/10.1371/journal.pone.0091324>.
 16. Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsmma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST Server: Rapid Annotations using Subsystems Technology. *BMC Genomics* 9:75. <http://dx.doi.org/10.1186/1471-2164-9-75>.