

# NIH Public Access

**Author Manuscript**

*Exp Biol Med (Maywood)*. Author manuscript; available in PMC 2014 June 09.

#### Published in final edited form as:

*Exp Biol Med (Maywood)*. 2013 February ; 238(2): 127–132. doi:10.1177/1535370213477602.

# **Weight control and cancer preventive mechanisms: role of IGF-1-mediated signaling pathways**

# **Linglin Xie**1,2 and **Weiqun Wang**<sup>2</sup>

<sup>1</sup>Department of Biochemistry and Molecular Biology, University of North Dakota, Grand Forks, ND 58202, USA

<sup>2</sup>Department of Human Nutrition, Kansas State University, Manhattan, KS 66506, USA

# **Abstract**

Overweight and obese not only increase the risk of cardiovascular disease and type-2 diabetes mellitus, but are also now known risk factors for a variety of cancers. Weight control, via dietary calorie restriction (DCR) and/or exercise, has been demonstrated to be beneficial for cancer prevention in various experimental models, but the underlying mechanisms are still not well defined. Recent studies conducted in a mouse skin carcinogenesis model show that weight loss induced a significant reduction of the circulating levels of IGF-1 and other hormones, including insulin and leptin, resulting in reduced IGF-1-dependent signaling pathways, i.e., Ras-MAPproliferation and Akt-PI3K-antiapoptosis. Selective targeting IGF-1 to Akt/mTOR and AMPK pathways, via negative energy balance, might inactivate cell cycle progression and ultimately suppress tumor development. This review highlights the current studies focused on the major role of reducing IGF-1-activated signaling via weight control as a potential cancer preventive mechanism.

#### **Keywords**

weight control; dietary calorie restriction; exercise; cancer prevention; IGF-1 signaling

# **Introduction**

Maintaining a healthy body weight emerges as a strategy to generally reduce cancers and has also been suggested to reduce cancer risk. With the increasing prevalence of obesity in both adults and children, it is important to identify the mechanism underlying cancer prevention via weight control. Current studies have focused on how understanding the molecular mechanisms of weight control, via dietary calorie restriction (DCR) and/or physical exercise, in both mouse model and human studies, prevent cancer. These studies will lead to future chemoprevention strategies for cancer.

Corresponding authors: Linglin Xie or Weiqun Wang, Department of Biochemistry and Molecular Biology, University of North Dakota, Grand Forks, ND 58202, USA; or Department of Human Nutrition, Kansas State University, Manhattan, KS 66506, USA. linglin.xie@med.und.edu or wwang@ksu.edu.

**Author contributions:** LX and WW were responsible for writing and revising the manuscript.

#### **Overweight/obesity as a risk factor of cancer**

It is well known that obesity has a substantial influence on the development of many chronic diseases, including cancer. Numerous prospective and case-control studies that address the effect of body weight on cancer risk, estimate that excess body weight and sedentary life style account for about 39% of endometrial, 25% of kidney, 11% of colon, 9% of postmenopausal breast cancer, and 5% of total cancer incidence.<sup>1</sup> The prevalence of obesity in the U.S. rose to approximately 25% and is projected to increase to more than 40% by 2030. In children and adolescents between the ages of 6 and 17 years, the prevalence of obesity appears to be increasing even more rapidly than in adults in industrialized and developing countries.<sup>2</sup> It has been suggested that those who are 25% over normal weight have a 33% greater cancer risk than those who maintain an ideal body weight.<sup>3</sup> Therefore, weight control to prevent obesity has been recommended as a strategy for reducing cancer risk by the American Cancer Society (ACS) as early as 1996.

In obese patients and animals, insulin/leptin resistance, inflammation, and changes in hormone and growth factor concentrations are found to be key pathogenic factors that lead to many types of cancer, including breast, colon, pancreas, and endometrium. These hormones, or growth factors, include insulin, leptin, and Insulin growth factor-1(IGF-1). They directly or indirectly provide a mitogenic effect in many cell types, especially in preneoplastic cells, by inducing proliferative and anti-apoptotic mechanisms.4–6 *In vitro* studies demonstrate that insulin, through binding to the insulin receptor, increases neoplastic proliferation at both physiological and pharmacological doses.<sup>7–9</sup> Recently, hyperinsulinemia associated with the IGF-1 signaling pathway has been widely studied, and the condition strongly stimulates the development and growth of several tumors, especially in breast cancer.<sup>10–12</sup> Hyperinsulinemia is associated with increased circulating free sex hormones, such as estrogen and androgen, via inhibiting the hepatic production of sex hormone-binding globulin.<sup>13</sup> Altered adipokine production is also associated with insulin resistance. Adiponectin, one of the most abundant adipokines, is shown to be both antiangiogenic and anti-inflammatory, and is lower in circulation in both obese and cancer patients.14–18

A high level of serum leptin in obese patients has been associated with increased cellular proliferation and angiogenesis across a wide variety of cancer subtypes, including colon, prostate and breast cancer.<sup>19–25</sup> The role of leptin in vascular remodeling may be independent of or coupled with vascular endothelial growth factor (VEGF) and fibroblast growth factor (FGF).26 Additionally, leptin stimulates several types of pre-neoplastic and neoplastic cells by mediating IGF-1R, resulting proliferation<sup>27–29</sup> and/or anti-apoptosis.<sup>30–32</sup>

Levels of proinflammatory cytokines, including IL-6, TNF-α and IL-1β, are found to be higher in obese mice and humans. Adipose tissue is a major source for these cytokines, contributing up to 35% of circulating IL-6.<sup>33</sup> Levels of these cytokines are also elevated in cancer patients.34,35 Although it is still unclear how these cytokines play a combined role in tumorigenesis, activation of NF- $\kappa$ B and STAT3 seem to be likely associated.<sup>36–41</sup>

#### **Weight control via DCR inhibits tumor development in rodents**

DCR is the most efficient method of weight control. The first work to show that incidence of tumors in mice positively correlated with food intake was published in 1944 by Tui and colleagues.42 To date, DCR has been the most widely studied and most potent, broadly acting dietary intervention for cancer prevention in various experimental models.<sup>43,44</sup> DCR is a dietary regimen that restricts calorie intake without malnutrition (usually by 20%–40% relative to *ad libitum*-fed controls from lipids and carbohydrates, but same amount of proteins, micronutrients and minerals, etc.). DCR-fed *wildtype* animals are typically healthier, live longer, and more active than their *ad libitum*-fed counterparts<sup>45</sup>.

DCR inhibits various spontaneous tumor developments in experimental animals. In rodents, a 20–40% below the usual ad libitum intake initiated early in life led to approximately 20– 60% reduction in tumor incidence, including tumors of mammary, liver, colon, skin, pancreas, bladder, and leukemia.<sup>46,47</sup> In p53-deficient mice, both juvenile- and adultinitiated calorie restriction to 60% of *ad libitum* intake, significantly delayed tumor development.48,49 In *ApcMin* mice, calorie intake restricted by 40% of the *ad libitum*-fed mice reduced intestinal polyps by 57%.<sup>49,50</sup> DCR is also well documented to suppress carcinogen-induced carcinogenesis, like benzo( $\alpha$ )pyrene<sup>51</sup> and 7,12dimethylbenz( $\alpha$ )anthracene (DMBA)<sup>52</sup>.

#### **Weight control via exercise**

Epidemiological studies report that physical activity can reduce the risk of many types of cancers, especially cancer of the prostate, breast, endometrial, and lung.<sup>53</sup> Evidence suggests that 4–7 hours per week of moderate to vigorous physical activity is required for adequate risk reduction.53 However, in the U.S., adults are not achieving the recommended amounts of physical activity, according to ACS guidelines.54–59

Despite the large numbers of studies conducted, cancer prevention by physical activity in animal models is not consistent. This is largely due to the lack of precise quantitative characteristics of duration and intensity of exercise as well as the control of the dietary calorie intake. Therefore, the impact of exercise on cancer development should be considered in combination with an isocaloric diet.

#### **Decreased IGF-1 signaling in cancer prevention**

There are several hypotheses describing mechanisms by which weight control via DCR and/or exercise may reduce tumor development. Some hypotheses include decreased oncogene expression, improved DNA repair, enhanced scavenging of reactive oxygen species, and altered levels of cancer-related hormones.<sup>60</sup>

Hormone alteration seems to be a critical factor for cancer prevention by weight control, due to the significant role of hormones in regulating cellular growth. Previous researchers have found that the levels of IGF-1<sup>62–64</sup>, insulin<sup>61</sup>, and leptin<sup>61,65–67</sup> decreased significantly in rodents in response to DCR.

Xie and Wang Page 4

IGF-1 is a major endocrine and paracrine regulator of cellular growth and metabolism. Binding of IGF-1 to IGF-1R activates many signaling pathways, including Jun N-terminal Kinases (JNK), p38 MAPK and PI3K via activation of receptor tyrosine kinases and/or the Ras proto-oncogene, to mediate suppression of apoptosis and contribute proliferation and cell growth.<sup>74–78</sup> IGF-1R is overexpressed in many tumors.<sup>70,71</sup> Abundant epidemiologic evidence supports the hypothesis that IGF-1 is involved in several types of human cancers.68,69 Adult HK1.IGF-1 mice that overexpressed IGF-1 spontaneously developed papillomas faster than non-transgenic littermates.72,73 Adult HK1.IGF-1 mice showed enhanced signaling through the Akt/mTOR pathways<sup>79</sup>, which suggested a critical role of IGF-1 in activating Akt/mTOR pathway that regulates cell proliferation, survival and energy metabolism. Activated mTOR signaling through the Akt/mTOR pathway in Akt overexpressing mouse caused alterations in epidermal proliferation and differentiation.<sup>79</sup>

The mice are more sensitive to topical 12-O-tetradecanoylphorbol-13-acetate (TPA) treatment. Activation of IGF-1R also indirectly acts with other cancer-related molecules, such as p53, known to arrest cell growth and induce apoptosis with increased levels of p21 and reduced Bcl-2.<sup>80</sup>

Reduction in glucose and insulin levels, as well as IGF-1, has been well documented in DCR-fed mice from different labs. Levels of circulating IGF-1 are influenced by dietary energy intake, which may be due to changing growth hormone-regulated hepatic synthesis of IGF-1.81 Restoration of IGF-1 in p53-deficient mice reversed DCR-induced cancer protection, suggesting a requirement of reduced levels of IGF-I.16 While 20% DCR-fed SENCAR mice showed a significant reduction of plasma IGF-1 levels, a remarkable decrease of IGF-1-dependent Ras/MAPK and PI3K/Akt signaling was demonstrated in TPA-stimulated skin tissues. $82,83$  Both down-expression of PCNA as a biomarker of proliferation and up-expression of Caspase-3 as a biomarker of apoptosis were also subsequently found in those mice. $82,83$ 

Diet-induced changes in Akt/mTOR signaling have been reported in various organ tissues (i.e., liver, skin epidermis, and mammary fat pad). DCR-induced increase of AMP/ATP ratio may inhibit AMPK signaling that regulates mTOR activity and thereby inhibits the targeted protein production. These changes appear to be related to the reduction of circulating IGF-1 levels. In such study, 30% DCR regimen for 15–17 weeks is sufficient to lower circulating IGF-1 concentrations and inactivate Akt and mTOR signaling in multiple epithelial tissues, regardless of genetic background of the experimental mice.<sup>84</sup> These results suggest that DCR induced a decrease of IGF-1 should play a central role in response with negative energy balance-altered depression of mTOR activity.<sup>85</sup>

In addition, an important study has revealed that DCR contributes to mammalian cell survival by inducing Sirtuin 1 (SIRT1) deacetylase, which deacetylates the DNA repair factor Ku70. Ku70 can prevent the proapoptotic factor Bax from mitochondrial entry, therefore inhibiting stress induced apoptosis.86 This impact is probably mediated by insulin/ IGF-1 because treatment with insulin and/or IGF-1 can reduce SIRT1 deacetylase expression.<sup>86</sup>

In humans, 15 weeks of moderate exercise changes the levels of fasting insulin, glucose, IGF-I, IGF-II, IGFBP-1, IGFBP-3, and IGF-I:IGFBP-3 molar ratio.87,88 However, a 12 month exercise regimen did not change IGF-1 and IGFBG-3 levels in postmenopausal women.89 Regular exercise alters the serum IGF-1 axis *in vivo* and reduces prostate (LNCaP) tumor cell proliferation by enhancing the function of the p53 gene<sup>90,91</sup> In SENCAR mice, weight loss by 10-week physical activity with iso-caloric intake as sedentary controls was able to inhibit PI3K signaling and increase caspase-3 activity.<sup>82</sup> These effects were partially reversed by IGF-1 restoration.<sup>82</sup> Microarray analysis comparing TPA-induced gene expression profiles in DCR- or exercise-treated mouse skin tissues revealed 411 genes affected by DCR versus only 67 affected by exercise with iso-caloric intake, including PI3K and MAPK pathway genes. $82-83$  Similar results of mammary gland gene expression are found in C57BL/6 mice with a 30% DCR versus exercise.<sup>92</sup> The increased expression of fatty acid elongase-1 in treadmill exercised mice with iso-caloric intake suggests that exercise may affect the phospholipid profile.<sup>93</sup> In addition, a lipidomics study using electrospray ionization-tandem mass spectrometry demonstrated that 57 phospholipids were significantly changed among a total of 338 species detected, and 25 species were closely related to exercise by a stepwise discriminant analysis.<sup>93–94</sup> These combined results indicate that DCR and/or exercise may target IGF-1-dependent signaling directly for a potential cancer prevention.

# **IGF-1 signaling as a potential target for cancer prevention**

Effective prevention and treatment strategies are urgently needed for anti-tumorigenesis. Considering the central role that IGF-1 played on cancer development, decreasing IGF-1 signaling either by chemical intervention or genetic interference has been intensively studied. Liver IGF-1-deficient (LID) mice have been utilized to mimic the effect of DCR on reducing circulating IGF-1.95 These mice had about 75% reduction in the level of circulating IGF-1. Repressed IGF-1R, EGFR and Akt/mTOR were observed in the skin tissue of LID mice after TPA treatment. LID mice initiated with DMBA and promoted with TPA, developed fewer and slower growing of papillomas compared with untreated littermates.<sup>95</sup> In another study that tested if DCR effect can be replicated by chemotherapy, mice transplanted with Panc02 murine pancreatic cancer cell were treated with either DCR or rapamycin for 20 weeks.<sup>96</sup> Rapamycin treatment  $(2.5 \text{ mg/kg}$  intraperitoneal every other day) did not decrease body weight, IGF-1 or leptin level, unlike DCR, but inhibited glucose responsiveness. Mice that received rapamycin had depressed mTOR signaling and had significantly reduced tumor volume compared to untreated mice, although to a lesser extent than DCR-fed mice. These results suggest that the downstream modulators of IGF-1 pathway can be a potential target for cancer prevention supplements.

### **Conclusion**

Many epidemiological, clinical, and experimental studies have revealed a positive relationship between weight control via DCR or physical activity, and the frequency of various cancers. Reductions in the levels of circulating IGF-1 and other mitogenic hormones/growth factors, including insulin and leptin induced by DCR or physical activity, efficiently inactivate the downstream signaling pathways via IGF-1R as summarized in

details in Figure 1. Future studies aimed at further elucidating the mechanisms underlying the cancer prevention possibilities of weight control may finally lead to efficient pharmacologic approaches that can be used alone or combined with weight control.

#### **Acknowledgments**

WW is supported in part by grants from NIH R15CA167678, NIH-INBRE P20RR16475, NIH R01CA106397, and the Terry Johnson Center for Basic Cancer Research, Kansas State University. LX is supported by grants from NIH National Center for Research Resources 5P20RR016471-12/8 and P20GM103442-12. WW also provides a journal contribution #13-055-J of the Kansas Agricultural Experiment Station.

#### **Abbreviations used**



#### **References**

- 1. Bergstrom A, Pisani P, Tenet V, Wolk A, Adami HO. Overweight as an avoidable cause of cancer in Europe. Int J Cancer. 2001; 91:421–430. [PubMed: 11169969]
- 2. Vainio H, Kaaks R, Bianchini F. Weight control and physical activity in cancer prevention: international evaluation of the evidence. Eur J Cancer Prev. 2002; 11:S94–S100. [PubMed: 12570341]
- 3. Kritchevsky D. Diet and cancer: what's next? J Nutr. 2003; 133:3827S–3829S. [PubMed: 14608121]
- 4. Al-Aubaidy HA, Jelinek HF. Oxidative DNA damage and obesity in type 2 diabetes mellitus. Eur J Endocrinol. 2011; 164:899–904. [PubMed: 21436346]
- 5. Zhang H, Xie C, Spencer HJ, Zuo C, Higuchi M, Ranganathan G, Kern PA, Chou MW, Huang Q, Szczesny B, et al. Obesity and hepatosteatosis in mice with enhanced oxidative DNA damage processing in mitochondria. Am J Pathol. 2011; 178:1715–1727. [PubMed: 21435453]
- 6. Dai Q, Gao YT, Shu XO, Yang G, Milne G, Cai Q, Wen W, Rothman N, Cai H, Li H, et al. Oxidative stress, obesity, and breast cancer risk: results from the Shanghai Women's Health Study. J Clin Oncol. 2009; 27:2482–2488. [PubMed: 19380446]
- 7. Stuart CA, Prince MJ, Peters EJ, Smith FE, Townsend CM 3rd, Poffenbarger PL. Insulin receptor proliferation: a mechanism for tumor-associated hypoglycemia. J Clin Endocrinol Metab. 1986; 63:879–885. [PubMed: 3528210]
- 8. Pavelic K. Aplastic carcinoma in diabetic mice: hyperglycemia-suppressed proliferation rate and insulin synthesis by tumor cells. J Natl Cancer Inst. 1979; 62:139–141. [PubMed: 281569]
- 9. Osborne CK, Bolan G, Monaco ME, Lippman ME. Hormone responsive human breast cancer in long-term tissue culture: effect of insulin. Proc Natl Acad Sci U S A. 1976; 73:4536–4540. [PubMed: 1070004]

- 10. Dunn SE, Hardman RA, Kari FW, Barrett JC. Insulin-like growth factor 1 (IGF-1) alters drug sensitivity of HBL100 human breast cancer cells by inhibition of apoptosis induced by diverse anticancer drugs. Cancer Res. 1997; 57:2687–2693. [PubMed: 9205078]
- 11. Dunn SE, Kari FW, French J, Leininger JR, Travlos G, Wilson R, Barrett JC. Dietary restriction reduces insulin-like growth factor I levels, which modulates apoptosis, cell proliferation, and tumor progression in p53-deficient mice. Cancer Res. 1997; 57:4667–4672. [PubMed: 9354418]
- 12. Jernstrom H, Sandberg T, Bageman E, Borg A, Olsson H. Insulin-like growth factor-1 (IGF1) genotype predicts breast volume after pregnancy and hormonal contraception and is associated with circulating IGF-1 levels: implications for risk of early-onset breast cancer in young women from hereditary breast cancer families. Br J Cancer. 2005; 92:857–866. [PubMed: 15756256]
- 13. Poretsky L, Kalin MF. The gonadotropic function of insulin. Endocr Rev. 1987; 8:132–141. [PubMed: 3301317]
- 14. Goodwin PJ, Ennis M, Pritchard KI, Trudeau ME, Koo J, Taylor SK, Hood N. Insulin- and obesity-related variables in early-stage breast cancer: correlations and time course of prognostic associations. J Clin Oncol. 2012; 30:164–171. [PubMed: 22162568]
- 15. Goodwin PJ, Stambolic V. Obesity and insulin resistance in breast cancer--chemoprevention strategies with a focus on metformin. Breast. 2011; 20:S31–S35. [PubMed: 22015290]
- 16. Gallagher EJ, LeRoith D. Insulin, insulin resistance, obesity, and cancer. Curr Diab Rep. 2010; 10:93–100. [PubMed: 20425567]
- 17. Smith MR, Lee H, Fallon MA, Nathan DM. Adipocytokines, obesity, and insulin resistance during combined androgen blockade for prostate cancer. Urology. 2008; 71:318–322. [PubMed: 18308111]
- 18. Rose DP, Komninou D, Stephenson GD. Obesity, adipocytokines, and insulin resistance in breast cancer. Obes Rev. 2004; 5:153–165. [PubMed: 15245384]
- 19. Koda M, Sulkowska M, Kanczuga-Koda L, Cascio S, Colucci G, Russo A, Surmacz E, Sulkowski S. Expression of the obesity hormone leptin and its receptor correlates with hypoxia-inducible factor-1 alpha in human colorectal cancer. Ann Oncol. 2007; 18:vi116–vi119. [PubMed: 17591803]
- 20. Chung WK, Leibel RL. The links between obesity, leptin, and prostate cancer. Cancer J. 2006; 12:178–181. [PubMed: 16803674]
- 21. Stattin P, Lukanova A, Biessy C, Soderberg S, Palmqvist R, Kaaks R, Olsson T, Jellum E. Obesity and colon cancer: does leptin provide a link? Int J Cancer. 2004; 109:149–152. [PubMed: 14735482]
- 22. Kos K, Wong S, Tan B, Gummesson A, Jernas M, Franck N, Kerrigan D, Nystrom FH, Carlsson LM, Randeva HS, et al. Regulation of the fibrosis and angiogenesis promoter SPARC/osteonectin in human adipose tissue by weight change, leptin, insulin, and glucose. Diabetes. 2009; 58:1780– 1788. [PubMed: 19509023]
- 23. Anagnostoulis S, Karayiannakis AJ, Lambropoulou M, Efthimiadou A, Polychronidis A, Simopoulos C. Human leptin induces angiogenesis in vivo. Cytokine. 2008; 42:353–357. [PubMed: 18448353]
- 24. Park HY, Kwon HM, Lim HJ, Hong BK, Lee JY, Park BE, Jang Y, Cho SY, Kim HS. Potential role of leptin in angiogenesis: leptin induces endothelial cell proliferation and expression of matrix metalloproteinases in vivo and in vitro. Exp Mol Med. 2001; 33:95–102. [PubMed: 11460888]
- 25. Hoda MR, Keely SJ, Bertelsen LS, Junger WG, Dharmasena D, Barrett KE. Leptin acts as a mitogenic and antiapoptotic factor for colonic cancer cells. Br J Surg. 2007; 94:346–354. [PubMed: 17212381]
- 26. Cao R, Brakenhielm E, Wahlestedt C, Thyberg J, Cao Y. Leptin induces vascular permeability and synergistically stimulates angiogenesis with FGF-2 and VEGF. Proc Natl Acad Sci U S A. 2001; 98:6390–6395. [PubMed: 11344271]
- 27. Ogunwobi OO, Beales IL. Cyclo-oxygenase-independent inhibition of apoptosis and stimulation of proliferation by leptin in human colon cancer cells. Dig Dis Sci. 2007; 52:1934–1945. [PubMed: 17406816]
- 28. Pai R, Lin C, Tran T, Tarnawski A. Leptin activates STAT and ERK2 pathways and induces gastric cancer cell proliferation. Biochem Biophys Res Commun. 2005; 331:984–992. [PubMed: 15882975]
- 29. Miyazaki T, Bub JD, Iwamoto Y. c-Jun NH(2)-terminal kinase mediates leptin-stimulated androgen-independent prostate cancer cell proliferation via signal transducer and activator of transcription 3 and Akt. Biochim Biophys Acta. 2008; 1782:593–604. [PubMed: 18718531]
- 30. Naviglio S, Di Gesto D, Romano M, Sorrentino A, Illiano F, Sorvillo L, Abbruzzese A, Marra M, Caraglia M, Chiosi E, et al. Leptin enhances growth inhibition by cAMP elevating agents through apoptosis of MDA-MB-231 breast cancer cells. Cancer Biol Ther. 2009; 8:1183–1190. [PubMed: 19662684]
- 31. Shen Y, Wang Q, Zhao Q, Zhou J. Leptin promotes the immune escape of lung cancer by inducing proinflammatory cytokines and resistance to apoptosis. Mol Med Report. 2009; 2:295–299.
- 32. Rouet-Benzineb P, Aparicio T, Guilmeau S, Pouzet C, Descatoire V, Buyse M, Bado A. Leptin counteracts sodium butyrate-induced apoptosis in human colon cancer HT-29 cells via NF-kappaB signaling. J Biol Chem. 2004; 279:16495–16502. [PubMed: 14752104]
- 33. Fenton JI, Birmingham JM. Adipokine regulation of colon cancer: adiponectin attenuates interleukin-6-induced colon carcinoma cell proliferation via STAT-3. Mol Carcinog. 2010; 49:700–709. [PubMed: 20564347]
- 34. Kai H, Kitadai Y, Kodama M, Cho S, Kuroda T, Ito M, Tanaka S, Ohmoto Y, Chayama K. Involvement of proinflammatory cytokines IL-1beta and IL-6 in progression of human gastric carcinoma. Anticancer Res. 2005; 25:709–713. [PubMed: 15868900]
- 35. Punnonen J, Heinonen PK, Kuoppala T, Jansen CT, Punnonen R. Production of interleukin-1 beta and tumour necrosis factor-alpha in patients with benign or malignant ovarian tumours. J Cancer Res Clin Oncol. 1991; 117:587–592. [PubMed: 1744165]
- 36. Syed V, Ulinski G, Mok SC, Ho SM. Reproductive hormone-induced, STAT3-mediated interleukin 6 action in normal and malignant human ovarian surface epithelial cells. J Natl Cancer Inst. 2002; 94:617–629. [PubMed: 11959895]
- 37. Sriuranpong V, Park JI, Amornphimoltham P, Patel V, Nelkin BD, Gutkind JS. Epidermal growth factor receptor-independent constitutive activation of STAT3 in head and neck squamous cell carcinoma is mediated by the autocrine/paracrine stimulation of the interleukin 6/gp130 cytokine system. Cancer Res. 2003; 63:2948–2956. [PubMed: 12782602]
- 38. Li Q, Zhou XD, Kolosov VP, Perelman JM. Nicotine reduces TNF-alpha expression through a alpha7 nAChR/MyD88/NF-kB pathway in HBE16 airway epithelial cells. Cell Physiol Biochem. 2011; 27:605–612. [PubMed: 21691078]
- 39. Ryazantseva NV, Novitskii VV, Zhukova OB, Biktasova AK, Chechina OE, Sazonova EV, Radzivil TT, Wice AN, Chasovskikh NY. Role of NF-kB, p53, and p21 in the regulation of TNFalpha mediated apoptosis of lymphocytes. Bull Exp Biol Med. 2010; 149:50–53. [PubMed: 21113457]
- 40. Crinelli R, Antonelli A, Bianchi M, Gentilini L, Scaramucci S, Magnani M. Selective inhibition of NF-kB activation and TNF-alpha production in macrophages by red blood cell-mediated delivery of dexamethasone. Blood Cells Mol Dis. 2000; 26:211–222. [PubMed: 10950941]
- 41. Xu YX, Pindolia KR, Janakiraman N, Chapman RA, Gautam SC. Curcumin inhibits IL1 alpha and TNF-alpha induction of AP-1 and NF-kB DNA-binding activity in bone marrow stromal cells. Hematopathol Mol Hematol. 1997; 11:49–62. [PubMed: 9439980]
- 42. Tui C, Wright AM, Mulholland JH, Carabba V, Barcham I, Vinci VJ. Studies on Surgical Convalescence I-Sources of Nitrogen Loss Postgastrectomy and Effect of High Amino-Acid and High Caloric Intake on Convalescence. Ann Surg. 1944; 120:99–122. [PubMed: 17858477]
- 43. Hursting SD, Kari FW. The anti-carcinogenic effects of dietary restriction: mechanisms and future directions. Mutat Res. 1999; 443:235–249. [PubMed: 10415442]
- 44. Hursting SD, Lashinger LM, Colbert LH, Rogers CJ, Wheatley KW, Nunez NP, Mahabir S, Barrett JC, Forman MR, Perkins SN. Energy balance and carcinogenesis: underlying pathways and targets for intervention. Curr Cancer Drug Targets. 2007; 7:484–491. [PubMed: 17691908]

- 45. Hursting SD, Lavigne JA, Berrigan D, Perkins SN, Barrett JC. Calorie restriction, aging, and cancer prevention: mechanisms of action and applicability to humans. Annu Rev Med. 2003; 54:131–152. [PubMed: 12525670]
- 46. Albanes D. Total calories, body weight, and tumor incidence in mice. Cancer Res. 1987; 47:1987– 1992. [PubMed: 3828987]
- 47. Roebuck BD, Yager JD Jr, Longnecker DS, Wilpone SA. Promotion by unsaturated fat of azaserine-induced pancreatic carcinogenesis in the rat. Cancer Res. 1981; 41:3961–3966. [PubMed: 7285004]
- 48. Berrigan D, Perkins SN, Haines DC, Hursting SD. Adult-onset calorie restriction and fasting delay spontaneous tumorigenesis in p53-deficient mice. Carcinogenesis. 2002; 23:817–822. [PubMed: 12016155]
- 49. Hursting SD, Perkins SN, Brown CC, Haines DC, Phang JM. Calorie restriction induces a p53 independent delay of spontaneous carcinogenesis in p53-deficient and wild-type mice. Cancer Res. 1997; 57:2843–2846. [PubMed: 9230186]
- 50. Mai V, Colbert LH, Berrigan D, Perkins SN, Pfeiffer R, Lavigne JA, Lanza E, Haines DC, Schatzkin A, Hursting SD. Calorie restriction and diet composition modulate spontaneous intestinal tumorigenesis in Apc(Min) mice through different mechanisms. Cancer Res. 2003; 63:1752–1755. [PubMed: 12702556]
- 51. Chou MW, Kong J, Chung KT, Hart RW. Effect of caloric restriction on the metabolic activation of xenobiotics. Mutat Res. 1993; 295:223–235. [PubMed: 7507559]
- 52. Stewart JW, Koehler K, Jackson W, Hawley J, Wang W, Au A, Myers R, Birt DF. Prevention of mouse skin tumor promotion by dietary energy restriction requires an intact adrenal gland and glucocorticoid supplementation restores inhibition. Carcinogenesis. 2005; 26:1077–1084. [PubMed: 15746164]
- 53. Thune I, Furberg AS. Physical activity and cancer risk: dose-response and cancer, all sites and sitespecific. Med Sci Sports Exerc. 2001; 33:S530–S550. discussion S609-510. [PubMed: 11427781]
- 54. Kushi LH, Doyle C, McCullough M, Rock CL, Demark-Wahnefried W, Bandera EV, Gapstur S, Patel AV, Andrews K, Gansler T. American Cancer Society Guidelines on nutrition and physical activity for cancer prevention: reducing the risk of cancer with healthy food choices and physical activity. CA Cancer J Clin. 2012; 62:30–67. [PubMed: 22237782]
- 55. Kushi LH, Byers T, Doyle C, Bandera EV, McCullough M, McTiernan A, Gansler T, Andrews KS, Thun MJ. American Cancer Society Guidelines on Nutrition and Physical Activity for cancer prevention: reducing the risk of cancer with healthy food choices and physical activity. CA Cancer J Clin. 2006; 56:254–281. quiz 313-254. [PubMed: 17005596]
- 56. Byers T, Nestle M, McTiernan A, Doyle C, Currie-Williams A, Gansler T, Thun M. American Cancer Society guidelines on nutrition and physical activity for cancer prevention: Reducing the risk of cancer with healthy food choices and physical activity. CA Cancer J Clin. 2002; 52:92–119. [PubMed: 11929008]
- 57. Guidelines on diet, nutrition, and cancer prevention: reducing the risk of cancer with healthy food choices and physical activity. The American Cancer Society 1996 Advisory Committee on Diet, Nutrition, and Cancer Prevention. CA Cancer J Clin. 1996; 46:325–341. [PubMed: 8917019]
- 58. Doyle C, Kushi LH, Byers T, Courneya KS, Demark-Wahnefried W, Grant B, McTiernan A, Rock CL, Thompson C, Gansler T, et al. Nutrition and physical activity during and after cancer treatment: an American Cancer Society guide for informed choices. CA Cancer J Clin. 2006; 56:323–353. [PubMed: 17135691]
- 59. Brown JK, Byers T, Doyle C, Coumeya KS, Demark-Wahnefried W, Kushi LH, McTieman A, Rock CL, Aziz N, Bloch AS, et al. Nutrition and physical activity during and after cancer treatment: an American Cancer Society guide for informed choices. CA Cancer J Clin. 2003; 53:268–291. [PubMed: 14570227]
- 60. Jiang Y, Wang W. Potential mechnisms of cancer prevention by weight control. Biophy Rev Lett. 2008; 3:421–437.
- 61. Zhu M, Lee GD, Ding L, Hu J, Qiu G, de Cabo R, Bernier M, Ingram DK, Zou S. Adipogenic signaling in rat white adipose tissue: modulation by aging and calorie restriction. Exp Gerontol. 2007; 42:733–744. [PubMed: 17624709]

- 62. Fontana L, Weiss EP, Villareal DT, Klein S, Holloszy JO. Long-term effects of calorie or protein restriction on serum IGF-1 and IGFBP-3 concentration in humans. Aging Cell. 2008; 7:681–687. [PubMed: 18843793]
- 63. Lashinger LM, Malone LM, McArthur MJ, Goldberg JA, Daniels EA, Pavone A, Colby JK, Smith NC, Perkins SN, Fischer SM, et al. Genetic reduction of insulin-like growth factor-1 mimics the anticancer effects of calorie restriction on cyclooxygenase-2-driven pancreatic neoplasia. Cancer Prev Res (Phila). 2011; 4:1030–1040. [PubMed: 21593196]
- 64. Berrigan D, Lavigne JA, Perkins SN, Nagy TR, Barrett JC, Hursting SD. Phenotypic effects of calorie restriction and insulin-like growth factor-1 treatment on body composition and bone mineral density of C57BL/6 mice: implications for cancer prevention. In Vivo. 2005; 19:667–674. [PubMed: 15999532]
- 65. Rogozina OP, Bonorden MJ, Seppanen CN, Grande JP, Cleary MP. Effect of chronic and intermittent calorie restriction on serum adiponectin and leptin and mammary tumorigenesis. Cancer Prev Res (Phila). 2011; 4:568–581. [PubMed: 21257708]
- 66. Gauffin Cano MP, Van Nieuwenhove C, Chaila Z, Bazan C, Gonzalez S. Effects of short-term mild calorie restriction diet and renutrition with ruminant milks on leptin levels and other metabolic parameters in mice. Nutrition. 2009; 25:322–329. [PubMed: 19036563]
- 67. Chacon F, Esquifino AI, Perello M, Cardinali DP, Spinedi E, Alvarez MP. 24-hour changes in ACTH, corticosterone, growth hormone, and leptin levels in young male rats subjected to calorie restriction. Chronobiol Int. 2005; 22:253–265. [PubMed: 16021842]
- 68. Stattin P, Bylund A, Rinaldi S, Biessy C, Dechaud H, Stenman UH, Egevad L, Riboli E, Hallmans G, Kaaks R. Plasma insulin-like growth factor-I, insulin-like growth factor-binding proteins, and prostate cancer risk: a prospective study. J Natl Cancer Inst. 2000; 92:1910–1917. [PubMed: 11106682]
- 69. Chan JM, Stampfer MJ, Giovannucci E, Gann PH, Ma J, Wilkinson P, Hennekens CH, Pollak M. Plasma insulin-like growth factor-I and prostate cancer risk: a prospective study. Science. 1998; 279:563–566. [PubMed: 9438850]
- 70. Pandini G, Vigneri R, Costantino A, Frasca F, Ippolito A, Fujita-Yamaguchi Y, Siddle K, Goldfine ID, Belfiore A. Insulin and insulin-like growth factor-I (IGF-I) receptor overexpression in breast cancers leads to insulin/IGF-I hybrid receptor overexpression: evidence for a second mechanism of IGF-I signaling. Clin Cancer Res. 1999; 5:1935–1944. [PubMed: 10430101]
- 71. Belfiore A, Pandini G, Vella V, Squatrito S, Vigneri R. Insulin/IGF-I hybrid receptors play a major role in IGF-I signaling in thyroid cancer. Biochimie. 1999; 81:403–407. [PubMed: 10401676]
- 72. DiGiovanni J, Bol DK, Wilker E, Beltran L, Carbajal S, Moats S, Ramirez A, Jorcano J, Kiguchi K. Constitutive expression of insulin-like growth factor-1 in epidermal basal cells of transgenic mice leads to spontaneous tumor promotion. Cancer Res. 2000; 60:1561–1570. [PubMed: 10749124]
- 73. Bol DK, Kiguchi K, Gimenez-Conti I, Rupp T, DiGiovanni J. Overexpression of insulin-like growth factor-1 induces hyperplasia, dermal abnormalities, and spontaneous tumor formation in transgenic mice. Oncogene. 1997; 14:1725–1734. [PubMed: 9135074]
- 74. Kenchappa P, Yadav A, Singh G, Nandana S, Banerjee K. Rescue of TNFalpha-inhibited neuronal cells by IGF-1 involves Akt and c-Jun N-terminal kinases. J Neurosci Res. 2004; 76:466–474. [PubMed: 15114618]
- 75. Choi YS, Cho HY, Hoyt KR, Naegele JR, Obrietan K. IGF-1 receptor-mediated ERK/MAPK signaling couples status epilepticus to progenitor cell proliferation in the subgranular layer of the dentate gyrus. Glia. 2008; 56:791–800. [PubMed: 18338791]
- 76. Kundu AK, Nagaoka M, Chowdhury EH, Hirose S, Sasagawa T, Akaike T. IGF-1 induces growth, survival and morphological change of primary hepatocytes on a galactose-bared polymer through both MAPK and beta-catenin pathways. Cell Struct Funct. 2003; 28:255–263. [PubMed: 14586135]
- 77. Yuan TL, Cantley LC. PI3K pathway alterations in cancer: variations on a theme. Oncogene. 2008; 27:5497–5510. [PubMed: 18794884]
- 78. Engelman JA. Targeting PI3K signalling in cancer: opportunities, challenges and limitations. Nat Rev Cancer. 2009; 9:550–562. [PubMed: 19629070]

- 79. Segrelles C, Lu J, Hammann B, Santos M, Moral M, Cascallana JL, Lara MF, Rho O, Carbajal S, Traag J, et al. Deregulated activity of Akt in epithelial basal cells induces spontaneous tumors and heightened sensitivity to skin carcinogenesis. Cancer Res. 2007; 67:10879–10888. [PubMed: 18006833]
- 80. Vanamala J, Reddivari L, Radhakrishnan S, Tarver C. Resveratrol suppresses IGF-1 induced human colon cancer cell proliferation and elevates apoptosis via suppression of IGF-1R/Wnt and activation of p53 signaling pathways. BMC Cancer. 2010; 10:238. [PubMed: 20504360]
- 81. Hursting SD, Switzer BR, French JE, Kari FW. The growth hormone: insulin-like growth factor 1 axis is a mediator of diet restriction-induced inhibition of mononuclear cell leukemia in Fischer rats. Cancer Res. 1993; 53:2750–2757. [PubMed: 8389243]
- 82. Xie L, Jiang Y, Ouyang P, Chen J, Doan H, Herndon B, Sylvester JE, Zhang K, Molteni A, Reichle M, et al. Effects of dietary calorie restriction or exercise on the PI3K and Ras signaling pathways in the skin of mice. J Biol Chem. 2007; 282:28025–28035. [PubMed: 17646168]
- 83. Lu J, Xie L, Sylvester J, Wang J, Bai J, Baybutt R, Wang W. Different gene expression of skin tissues between mice with weight controlled by either calorie restriction or physical exercise. Exp Biol Med (Maywood). 2007; 232:473–480. [PubMed: 17392482]
- 84. Moore T, Beltran L, Carbajal S, Strom S, Traag J, Hursting SD, DiGiovanni J. Dietary energy balance modulates signaling through the Akt/mammalian target of rapamycin pathways in multiple epithelial tissues. Cancer Prev Res (Phila). 2008; 1:65–76. [PubMed: 19138937]
- 85. Pfaffenbach KT, Pong M, Morgan TE, Wang H, Ott K, Zhou B, Longo VD, Lee AS. GRP78/BiP is a novel downstream target of IGF-1 receptor mediated signaling. J Cell Physiol. 2012; 227:3803–3811. [PubMed: 22422508]
- 86. Cohen HY, Miller C, Bitterman KJ, Wall NR, Hekking B, Kessler B, Howitz KT, Gorospe M, de Cabo R, Sinclair DA. Calorie restriction promotes mammalian cell survival by inducing the SIRT1 deacetylase. Science. 2004; 305:390–392. [PubMed: 15205477]
- 87. Haydon AM, Macinnis RJ, English DR, Morris H, Giles GG. Physical activity, insulin-like growth factor 1, insulin-like growth factor binding protein 3, and survival from colorectal cancer. Gut. 2006; 55:689–694. [PubMed: 16299029]
- 88. Fairey AS, Courneya KS, Field CJ, Bell GJ, Jones LW, Mackey JR. Effects of exercise training on fasting insulin, insulin resistance, insulin-like growth factors, and insulin-like growth factor binding proteins in postmenopausal breast cancer survivors: a randomized controlled trial. Cancer Epidemiol Biomarkers Prev. 2003; 12:721–727. [PubMed: 12917202]
- 89. McTiernan A, Sorensen B, Yasui Y, Tworoger SS, Ulrich CM, Irwin ML, Rudolph RE, Stanczyk FZ, Schwartz RS, Potter JD. No effect of exercise on insulin-like growth factor 1 and insulin-like growth factor binding protein 3 in postmenopausal women: a 12-month randomized clinical trial. Cancer Epidemiol Biomarkers Prev. 2005; 14:1020–1021. [PubMed: 15824183]
- 90. Leung PS, Aronson WJ, Ngo TH, Golding LA, Barnard RJ. Exercise alters the IGF axis in vivo and increases p53 protein in prostate tumor cells in vitro. J Appl Physiol. 2004; 96(2):450–454. [PubMed: 14715676]
- 91. Barnard RJ, Leung PS, Aronson WJ, Cohen P, Golding LA. A mechanism to explain how regular exercise might reduce the risk for clinical prostate cancer. Eur J Cancer Prev. 2007; 16:415–421. [PubMed: 17923812]
- 92. Padovani M, Lavigne JA, Chandramouli GV, Perkins SN, Barrett JC, Hursting SD, Bennett LM, Berrigan D. Distinct effects of calorie restriction and exercise on mammary gland gene expression in C57BL/6 mice. Cancer Prev Res (Phila). 2009; 2:1076–1087. [PubMed: 19952363]
- 93. Ouyang P, Jiang Y, Doan HM, Xie L, Vasquez D, Welti R, Su X, Lu N, Herndon B, Yang SS, et al. Weight Loss via exercise with controlled dietary intake may affect phospholipid profile for cancer prevention in murine skin tissues. Cancer Prev Res (Phila). 2010; 3:466–477. [PubMed: 20233900]
- 94. Jiang Y, Ma H, Su X, Chen J, Xu J, Standard J, Lin D, Wang W. IGF-1 mediates exercise-induced phospholipid alteration in the murine skin tissues. Journal of Nutrition and Food Science. 2012 S2-003.

- 95. Moore T, Carbajal S, Beltran L, Perkins SN, Yakar S, Leroith D, Hursting SD, Digiovanni J. Reduced susceptibility to two-stage skin carcinogenesis in mice with low circulating insulin-like growth factor I levels. Cancer Res. 2008; 68:3680–3688. [PubMed: 18483250]
- 96. Lashinger LM, Malone LM, Brown GW, Daniels EA, Goldberg JA, Otto G, Fischer SM, Hursting SD. Rapamycin partially mimics the anticancer effects of calorie restriction in a murine model of pancreatic cancer. Cancer Prev Res (Phila). 2011; 4:1041–1051. [PubMed: 21593197]

Xie and Wang Page 13



**Figure 1.**

Schematic illustration of IGF-1- and/or leptin-induced signaling pathways that may be targeted by weight control via dietary calorie restriction and exercise.