

Published in final edited form as:

J Biomech. 2014 June 27; 47(9): 1933–1940. doi:10.1016/j.jbiomech.2014.04.019.

Biomechanics and mechanobiology in functional tissue engineering

Farshid Guilak¹, David L. Butler², Steven A. Goldstein³, and Frank P.T. Baaijens⁴

¹Departments of Orthopaedic Surgery and Biomedical Engineering, Duke University Medical Center, Durham, North Carolina USA ²Department of Biomedical, Chemical and Environmental Engineering, University of Cincinnati, Cincinnati, Ohio USA ³Department of Orthopaedic Surgery, University of Michigan, Ann Arbor, Michigan USA ⁴Institute for Complex Molecular Systems, Eindhoven University of Technology, Eindhoven, The Netherlands

Abstract

The field of tissue engineering continues to expand and mature, and several products are now in clinical use, with numerous other preclinical and clinical studies underway. However, specific challenges still remain in the repair or regeneration of tissues that serve a predominantly biomechanical function. Furthermore, it is now clear that mechanobiological interactions between cells and scaffolds can critically influence cell behavior, even in tissues and organs that do not serve an overt biomechanical role. Over the past decade, the field of “functional tissue engineering” has grown as a subfield of tissue engineering to address the challenges and questions on the role of biomechanics and mechanobiology in tissue engineering. Originally posed as a set of principles and guidelines for engineering of load-bearing tissues, functional tissue engineering has grown to encompass several related areas that have proven to have important implications for tissue repair and regeneration. These topics include measurement and modeling of the *in vivo* biomechanical environment; quantitative analysis of the mechanical properties of native tissues, scaffolds, and repair tissues; development of rationale criteria for the design and assessment of engineered tissues; investigation of the effects biomechanical factors on native and repair tissues, *in vivo* and *in vitro*; and development and application of computational models of tissue growth and remodeling. Here we further expand this paradigm and provide examples of the numerous advances in the field over the past decade. Consideration of these principles in the design process will hopefully improve the safety, efficacy, and overall success of engineered tissue replacements.

© 2014 Elsevier Ltd. All rights reserved.

Correspondence address: Farshid Guilak, Ph.D., Duke University Medical Center, 375 MSRB, Box 3093, Durham, NC 27710 USA, Phone (919) 684-2521, Fax (919) 681-8490, guilak@duke.edu.

Publisher's Disclaimer: This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final citable form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

Conflict of Interest

Dr. Guilak is a founder of Cytek Therapeutics, Inc.

Keywords

regenerative medicine; gene therapy; biomaterials; cellular engineering

Introduction

Tissue engineering seeks to enable the repair or regeneration of tissues and organs through combinations of cells, biomaterial scaffolds, regulatory factors and environmental cues. This rapidly growing field primarily draws on various aspects of engineering, biomaterials, biology, and medicine, but has expanded to encompass a wide array of scientific disciplines needed to address the many challenges of the field. Despite many early advances (Langer and Vacanti, 1993), the field of tissue engineering encountered setbacks and delays in the translation of products to the clinic, with few commercial successes in early years of the field (Lysaght et al., 2008). However, the past decade has seen a significant expansion and maturation of the field, and there are now several tissue-engineered and cell-based products in clinical use as well as numerous preclinical studies and clinical trials underway.

In this regard, it is now clear that specific challenges still remain in the repair or regeneration of tissues that predominantly serve a biomechanical function. Furthermore, it is also becoming apparent that mechanobiological interactions between cells and scaffolds can have a critical influence on cell behavior, even in tissues and organs that do not serve an overt biomechanical role in the body. To highlight and address these issues and challenges, the United States National Committee on Biomechanics (USNCB) spearheaded an initiative on “functional tissue engineering”, which was originally posed as a set of principles and guidelines for tissue-engineering of load-bearing structures (Butler et al., 2000). The goals of the USNCB were primarily focused on: 1) increasing awareness among tissue engineers about the importance of biomechanical function when engineering tissues that serve biomechanical roles in the body; 2) identifying the structural and mechanical requirements needed for engineered tissues; and 3) encouraging tissue engineers to incorporate these functional criteria in the development and translation of tissue engineered products. This initiative was a catalyst for a session on “Functional Assessment of Engineered Tissues and Elements of Tissue Design” at the 2001 NIH BECON meeting on Reparative Medicine (Guilak, 2002; Guilak et al., 2002) and subsequently led to a workshop on this topic, funded in part by the National Science Foundation. This presentation and discussion from this workshop were summarized in a multi-chapter text that highlighted many different aspects of the field with a particular emphasis on the application of engineering principles to tissue engineering (Guilak et al., 2003).

Over the past decade, the field of functional tissue engineering has grown to encompass several related topics that have proven to have a critical influence on tissue repair and regeneration. The principles outlined previously for functional tissue engineering have also been advanced as well as recapitulated in more specific forms as they apply to particular tissues of the body (Baaijens et al., 2005; Butler et al., 2009; Butler et al., 2008; Guilak et al., 2001). Here we review and further expand these principles, and provide examples of some of the numerous advances that have occurred in the field over the past decade. As the

number of publications in this area has grown tremendously in the last decade, it is beyond the scope of this paper to review all of the outstanding studies in this area; rather, we have focused on several specific studies and review papers (with apologies to many authors whose studies were not included here). Consideration of these principles in the design process will hopefully improve the safety, efficacy, and overall success of engineered tissue replacements.

Principles of Functional Tissue Engineering

Many of the tissues of body – particularly in the musculoskeletal, cardiovascular, or pulmonary systems – have a primarily “biomechanical” function, such as load bearing, force generation or transmission, or fluid transport. In fact, the biomechanical properties of these tissues are critical to their normal *in vivo* behavior, and changes in tissue mechanical properties due to aging, injury or disease can result in significant tissue dysfunction.

In addition to the important role of biomechanics for these load-bearing (sometimes termed “structural”) tissues, it is now apparent that biomechanical and mechanobiological factors are critical for regulating cell behavior in virtually all other tissues of the body (Ingber, 2003). In this regard, understanding the role of physical factors in regulating cell growth, differentiation, and metabolism in “non-structural” (e.g., metabolic) organs and tissues has also become an important focus of functional tissue engineering. Thus, a number of significant questions remain on the role of biomechanics in tissue engineering, and these questions are relevant to most tissues and organ systems in the body (Table 1). Here we describe principles of functional tissue engineering that can serve as guidelines for developing and improving engineered systems for restoring tissue function. While this list continues to expand, the proposed principles are not meant to be exhaustive and continue to serve as a roadmap for further development of this paradigm.

1. Measurement and modeling of *in vivo* biomechanical histories in native and repair tissues

In attempting to define design parameters for the biomechanical function of repair tissues, knowledge of the mechanical context in which normal and repair tissues will serve for different *in vivo* activities will be required to establish patterns of activity and the limits of expected usage. In this regard, a further understanding of the mechanical “thresholds” that normal tissues encounter for different *in vivo* activities are critical to developing appropriate design criteria for tissue repairs/replacements that can meet functional demands. For many tissues, these measurements are difficult to make, but they establish the history and boundaries of expected usage and will help develop “safety factors” for tissue-engineered implants (Juncosa et al., 2003).

Over the past decade, significant advances have been made in such *in vivo* measurements for a number of tissues and organs, particularly in studies that have combined novel imaging methods with theoretical modeling. For example, in the musculoskeletal system, we now have a much better understanding of the range and history of stresses and strains placed on tissues such as tendons/ligaments (Juncosa et al., 2003; Taylor et al., 2013), articular cartilage (Adouni et al., 2012; Coleman et al., 2013), and bone (Al Nazer et al., 2012; Fritton

et al., 2000). Similarly, in the cardiovascular system, a number of studies have combined novel imaging methods with computational models to extend our knowledge of the flow- and pressure-induced stresses in blood vessels and heart valves (De Hart et al., 2003; Loerakker et al., 2013; Marom et al., 2013).

Furthermore, these studies have been extended to the study of repair tissues as well, which likely experience an altered mechanical environment due to differences in activity or physiology (i.e., changes in gait, blood pressure), or due to differences in the mechanical properties of the implant as compared to those of the native tissues [e.g., (Awad et al., 2003; Butler et al., 2008; Defrate et al., 2006; Juncosa-Melvin et al., 2007; Van Canneyt et al., 2013)]. These studies have greatly extended the previous knowledge of the necessary requirements for tissue replacements that would be expected to withstand physiologic, or in many cases, pathologic loading conditions, and will hopefully provide important insights into future design criteria for implants (Nerurkar et al., 2010). However, it is important to note that most current models are designed to simulate “normal” tissues, and the major structural or compositional changes that occur in advanced stages of disease or degeneration may require more complex geometric or constitutive models that may not be appropriately simulated by current continuum models.

2. Understanding the biomechanical properties of native and repair tissues across all geometric scales

Within the context of understanding the functional demands of different tissues, it will be important to have a thorough understanding of the sub-failure and failure properties of native tissues – through the processes of development, injury, disease, repair, and aging (Butler et al., 2008). This information will guide the design and engineering of repair tissues that provide the appropriate functional properties. While the mechanical properties of native tissues have been studied extensively, there has continued to be major advances in our understanding of cell and tissue biomechanics over the past decade. These measurements have included both “structural” and “material” (i.e., “intrinsic”) properties. Structural properties reflect the overall functional requirements of a tissue or organ and include the influence of morphological parameters, such as tissue geometry. “Material” properties, which should reflect tissue properties independent of geometry or loading conditions, are of particular value for predictive theoretical models to allow description of the mechanical response of a tissue/organ to different loading conditions, potentially *in vivo*.

Due to their hierarchical structure and composition, most biological tissues have highly complex material properties (Table 2). The fundamental basis for these behaviors is an important area of study, particularly with the implicit goal of recreating tissue function from independent components (i.e., cells and biomaterials). Importantly, it remains to be determined which attributes of these material properties are essential for the normal, healthy function of different tissues, as well as for successful tissue-engineered replacements. Nonetheless, great strides have been made in the past decade in our understanding of the more complex constitutive behaviors of tissue that involve anisotropic (Groves et al., 2013; Isenberg et al., 2012; Nagel and Kelly, 2012; Sommer et al., 2013; Teng et al., 2012),

multiphasic (Azeloglu et al., 2008), viscoelastic (Shirakawa et al., 2013), nonlinear (Buckley et al., 2013), and transport (Motaghinasab et al., 2012) properties.

Another important area has been the adoption of several novel techniques that have allowed measurement of functional mechanical properties at the nano- and micro-scales. Not only do such techniques provide insight into the fundamental structure-function relationships of developing (Buskohl et al., 2012; Marturano et al., 2013) and mature (Beenakker et al., 2012; McLeod et al., 2013) native tissues, they now allow testing of scaffolds and engineered tissues across a broad range of geometries and scales that were not possible using standard macroscopic methods (Diekman et al., 2012; Gilchrist et al., 2011; Hammond and Kamm, 2013).

The combinations of the various mechanical testing methods that are now available have provided an unprecedented level of quantitative assessment of native and engineered tissues. Importantly, these findings further reveal the highly complex and unique structure function relationships present in most tissues, and emphasize the challenges inherent to replicating these characteristic in engineered tissues. For example, hydrated collagenous tissues possess anisotropic and nonlinear mechanical and transport properties that vary significantly with site (Elliott and Setton, 2001; Huang et al., 2005; Martufi and Gasser, 2011). Not only are these properties critical to the overall function of the tissue, but the interactions among the different phases can result in complex stress-strain and pressure fields that can only be predicted using inhomogeneous or fiber-reinforced models (Shirazi-Adl, 1989), particularly at the cellular level (Baer et al., 2003; Korhonen et al., 2008; Likhitpanichkul et al., 2005). Furthermore, the development of engineered tissue replacements that replicate this mechanical environment will likely require structures that mimic certain aspects of the native tissue architecture (Moutos et al., 2007; Nerurkar et al., 2008).

3. Prioritization of specific mechanical properties as design parameters for biomaterial scaffolds and engineered tissues

Despite the growing database of mechanical properties, structure, and composition of native tissue, the relative importance of the different properties in influencing the success of an engineered repair tissue is not fully understood. Given the difficulties in matching all of the complex behaviors and architecture of native tissues, a key issue in the development of engineered repairs will be the prioritization of various biomechanical properties as design parameters, as it will be difficult if not impossible to completely match all of the material properties of native tissues (Table 2). Nonetheless, it may not be necessary to match all of the material properties of native tissue *a priori* in view of the remodeling potential of the implanted tissue *in vivo* (Schmidt et al., 2010). Priority has often been placed on a single parameter such as the compressive or tensile modulus of a tissue (Butler et al., 2008; Xiao et al., 2013), yet the relative importance of all of the different properties of a tissue or its constituents is unknown. To better understand and refine these design criteria, it will be necessary to understand the trade-offs and interactions of different functional characteristics and the overall success criteria of the implant. Of course this is not a simple task due to the large combination of biomechanical properties and functional demands (Table 2), coupled with equally complex or undefined success criteria.

4. Development of biomaterials and scaffolds with prescribed biomechanical properties

Novel biomaterials and biomaterial structures can provide control of functional biomechanical and other physical properties at the macro-, micro-, and nano- levels. These biomechanical and biological interactions between cells and scaffolds can have a significant influence on cell behavior (Guilak et al., 2009), and thus, the potential for long-term success of engineered tissues. Major advances in the past decade has been in the improved understanding of these interactions, coupled with breakthroughs in the development of scaffolds with highly controlled mechanical properties at multiple scales (Boffito et al., 2014; Courtney et al., 2006; Fan et al., 2013; Pennella et al., 2013; Soliman et al., 2010).

For example, the adoption of rapid-prototyping techniques such as microfabrication, 3D printing, or stereolithography has allowed the fabrication of a variety of scaffolds with controlled architecture and mechanical properties (Butscher et al., 2011; Engelmayer et al., 2008; Hollister, 2005; Hoque et al., 2005; Mironov et al., 2009; Paten et al., 2013). Additionally, a range of advanced textile processes including 2D or 3D weaving, knitting, and braiding have been applied to the development of scaffolds and constructs with highly biomimetic properties, as well as the ability to provide cell-instructive signals (Almeida et al., 2013; Brunger et al., 2014; Freeman et al., 2011; Liberski et al., 2011; Moutos et al., 2007; van Lieshout et al., 2006). Other non-woven techniques such as spun-bond or melt-extrusion have been shown to be highly versatile in the formation of different fiber architectures and to readily allow for scale-up and manufacturing (Tuin et al., 2013; Zeugolis et al., 2009). Probably the most rapid and broad adoption of materials processing methods for tissue engineering has been the use of hydrodynamic processing methods such as electrospinning, which provides a versatile method of controlling fiber architecture and properties at the nano- and micro-scales (Boland et al., 2004; Chew et al., 2006; Courtney et al., 2006; Mauck et al., 2009; Vaz et al., 2005). Importantly, recent studies suggest that the geometrical and biomechanical properties of electrospun scaffolds can also modulate the immune response to the scaffold (Ballotta et al., 2014; Garg et al., 2013).

New advances in the area of hydrogels have led to the development of biomaterials with tunable properties at the bulk or the cellular level (Stahl et al., 2010; Tseng et al., 2013; Wang et al., 2010). Biomaterials that undergo phase transition from liquid to solid through exposure to light, temperature changes, or chemical crosslinking/degradation can provide versatile environments for controlling cell behavior while defining the mechanical environment and functional properties of the scaffold (Betre et al., 2006; Chao et al., 2010; Cuchiara et al., 2012; Dankers et al., 2005; Sharma et al., 2013). Furthermore, chemical functionalization of such materials with proteins or gene delivery systems has been shown to provide an additional level of design capability for scaffold function (DeLong et al., 2005; Francisco et al., 2014; Glass et al., 2014; Hume et al., 2011; Kisiel et al., 2013; Salimath and Garcia, 2014).

Finally, the combinations of different biomaterials into “composite” scaffolds may in fact provide unique systems for controlling the spatio-temporal mechanical properties that are characteristic of native tissues (Table 2). For example, combinations of fiber-gel systems can be created to reproduce the nonlinear, anisotropic, and viscoelastic behavior of different tissues (Ekaputra et al., 2008; Freeman et al., 2011; Hong et al., 2011; Moutos et al., 2010),

whereas combinations of different hydrogels to form interpenetrating networks (IPNs) can provide toughness and fracture resistance that is not possible from single network gels (Daniele et al., 2014; Liao et al., 2013; Rennerfeldt et al., 2013; Suri and Schmidt, 2009). It may also be important to consider temporal factors in the design criteria of bioscaffolds. Whether a tissue engineered scaffold should recapitulate the properties of the native tissue to be repaired or the properties of the tissue during earlier stages of development or repair has yet to be determined and certainly may vary with tissue type.

5. Development of success criteria based on appropriate outcome measures

A critical step in achieving success in tissue-engineered repair and regenerative medicine will be the development of appropriate standards of clinical success and the completion of prospective outcomes studies to compare the safety and efficacy of different procedures. These criteria will of course need to be specific for each tissue or organ. Inherent in this process will be the further development of minimally invasive methods (e.g., imaging, biomarkers) that can be used for the assessment of tissue function *in vitro* as well as *in vivo* (Acosta Santamaria et al., 2013; Boerckel et al., 2012; DeJaco et al., 2012; Delaine-Smith et al., 2014; Hagenmuller et al., 2010; Kotecha et al., 2013; Marcos-Campos et al., 2012; Mariappan et al., 2010; Marturano et al., 2014; Vogl et al., 2010).

6. Investigation of the effects of mechanical factors on tissue repair *in vivo*

Once implanted, tissue-engineered constructs will be subjected to significant loads and deformations *in vivo*. Depending on the type of tissue/organ, the stress/strain environment may vary widely within the body. In this regard, both the mechanical and biological consequences of *in vivo* loading must be understood to improve the success of engineered repairs and to develop appropriate rehabilitation protocols following surgery to correct acute or chronic injuries (Boerckel et al., 2012; Juncosa et al., 2003; Willie et al., 2010).

From a mechanical standpoint, implanted tissue will need to eventually perform their prescribed function, presumably in a manner that restores native tissue function (although an exact replication of native tissues is likely to be difficult to achieve and may not be necessary assuming consideration of appropriate safety factors above functional load limits). Thus, the development of minimally invasive measures of tissue/organ function, and their application to engineered implants, will be vital to the understanding of the *in vivo* function of engineered tissues (summarized in the previous section).

From a biological, or “mechanobiological” standpoint, cells within implanted constructs will be similarly subjected to a complex biomechanical environment, that may consist of time- and spatially-varying stresses, strains, fluid pressure, fluid flow, and other biophysical parameters (Mow et al., 1994). It is now clear that such physical signals may significantly influence cellular growth, differentiation, and metabolism, and thus could play an important role in the long-term outcome of engineered tissues. Not only is a better understanding of the *in vivo* mechanical environment required, but additional data are needed on the response of cells to these signals and how they influence cell growth and differentiation.

7. The use of physical factors to enhance tissue regeneration *in vitro*

Mechanical factors play an important role in regulating cell physiology, and a wealth of evidence now shows that physical factors may be used to improve or accelerate tissue regeneration *in vitro*. The development and study of “bioreactors” for physical stimulation of engineered constructs has expanded massively over the past decade, with a variety of techniques showing tremendous promise in controlling cell growth, differentiation, and biological activity (Butler et al., 2009; Cimetta et al., 2013; Darling and Athanasiou, 2003; Hansmann et al., 2013; Rouwkema et al., 2011; Yeatts et al., 2013).

Early studies have shown that various mechanical stimuli, whether it be stretch, compression, pressure, or enhanced perfusion/transport could greatly enhance matrix formation in the context of tissue engineering [reviewed previously in (Butler et al., 2000)]. From these seminal studies, investigators have examined other types of physical stimuli such as combinations of different mechanical or physicochemical loading regimens that could even further improve construct qualities (Huang et al., 2012; Huey and Athanasiou, 2011; Jungreuthmayer et al., 2009; Meinel et al., 2004; Sampat et al., 2013; Schatti et al., 2011; Zhang et al., 2009). With further understanding of the role of the biomechanical microenvironment on cell behavior (Guilak et al., 2009; Reilly and Engler, 2010), other studies have examined the role of factors such as extracellular matrix topography or modulus within scaffolds as means of influencing local cellular responses (Dellatore et al., 2008; Gauvin et al., 2012; Guex et al., 2013; Kolambkar et al., 2014; Lim et al., 2011; Nii et al., 2013).

As the field has progressed, new studies have focused on determining the mechanisms of physical signal transduction (Liu and Lee, 2014; Mammoto et al., 2012; Pioletti, 2013). One goal of this approach has been to take advantage of chemical means to circumvent the standard bioreactor loading systems to directly stimulate cellular responses (O’Conor et al., 2014). With increased knowledge of the mechanotransduction cascades in primary and stem/progenitor cells, these approaches may serve as a means of controlling cell fate through physical means, as well accelerating tissue growth with or without other exogenous growth factors or gene delivery (Neumann et al., 2013; O’Conor et al., 2013; Shah et al., 2014; Song et al., 2013; Wang and Chen, 2013). In particular, few studies have used model systems to investigate the mechanobiology of repair tissues within environments that simulate important physiologic characteristics that may inhibit repair, such as aging, overloading or systemic diseases (i.e., inflammation, infection, obesity/diabetes, etc.) (Choe et al., 2006; Lima et al., 2009; Liu and Agarwal, 2010; Ousema et al., 2012).

8. Development and validation of computational models of tissue growth and remodeling

One of the key discriminating features of a living tissue replacement compared to synthetic substitutes is the ability to grow, and mechanical stress is believed to be an important regulator of this process. Experiments provide evidence for the ability of engineered tissues to grow *in vitro*, but the underlying mechanisms are poorly understood. Similarly, *in vivo*, the mechanical properties and architecture of engineered tissues will evolve in response to the biomechanical and biochemical environment. A thorough understanding of the remodeling mechanisms will enable definition of the required initial conditions of the

substitute to further mature to a healthy, functional tissue and to avoid adverse remodeling or degeneration.

In the past decade, the field of tissue engineering has seen tremendous advances in this area, particularly in the development of theoretical models of growth and remodeling that allow simulation of a subset of the numerous parameters that are potentially involved in tissue repair or regeneration (Hwang et al., 2013). The ability to simulate tissue responses to factors such as scaffold/hydrogel properties, transport (e.g., nutrients, waste products, oxygen, biomarkers, drugs, etc.), matrix accumulation, or mechanical stimuli could greatly decrease the number of *in vitro* and *in vivo* experiments that need to be run to most rapidly improve the properties of engineered constructs and grafts (Ateshian et al., 2014; Bovendeerd, 2012; Buskohl et al., 2012; Haider et al., 2011; Klisch et al., 2008; Loerakker et al., 2013; Niklason et al., 2010) and could potentially identify *in vitro* predictors of *in vivo* outcome (Butler et al., 2008).

Future Directions

The field of functional tissue engineering has experienced phenomenal growth over the past decade, with revolutionary advances in many of the areas outlined above. The rapid evolution of new technologies in fields such as cell and molecular biology, biomaterials, gene therapy, and nanotechnology, coupled with tremendous advances in computational methods and hardware, will undoubtedly have a significant impact on functional tissue engineering over the next decade. Many challenges still remain, of course, in the testing, scale-up, and manufacturing processes that will be required before these technologies can be translated to the clinic. Furthermore, packaging, storage and handling properties will also be critical parameters to include the design of engineered constructs. Nonetheless, the field of functional tissue engineering has come into its own and on its current path is likely to have a very promising future.

Acknowledgments

We thank members of the United States National Committee on Biomechanics (USNCB) subcommittee who participated in the adoption of the concept of Functional Tissue Engineering (Drs. Van C. Mow, Columbia University; Geert Schmid-Schonbein, University of California, San Diego; Louis J. Soslowsky, University of Pennsylvania; Dr. Robert Spilker, Rensselaer Polytechnic Institute; and Dr. Savio L. Woo, University of Pittsburgh). Supported in part by NIH grants AR50245, AG15768, AR48852, AR48182, AG46927, AR56943, the Collaborative Research Center, AO Foundation, Davos, Switzerland, the Arthritis Foundation, and the Nancy Taylor Foundation.

References

- Acosta Santamaria VA, Malve M, Duizabo A, Mena Tobar A, Gallego Ferrer G, Garcia Aznar JM, Doblare M, Ochoa I. Computational methodology to determine fluid related parameters of non regular three-dimensional scaffolds. *Ann Biomed Eng.* 2013; 41:2367–2380. [PubMed: 23807712]
- Adouni M, Shirazi-Adl A, Shirazi R. Computational biodynamics of human knee joint in gait: from muscle forces to cartilage stresses. *J Biomech.* 2012; 45:2149–2156. [PubMed: 22721726]
- Al Nazer R, Lanovaz J, Kawalilak C, Johnston JD, Kontulainen S. Direct *in vivo* strain measurements in human bone—a systematic literature review. *J Biomech.* 2012; 45:27–40. [PubMed: 21889149]
- Almeida LR, Martins AR, Fernandes EM, Oliveira MB, Mano JF, Correlo VM, Pashkuleva I, Marques AP, Ribeiro AS, Duraes NF, Silva CJ, Bonifacio G, Sousa RA, Oliveira AL, Reis RL. New

- biotextiles for tissue engineering: development, characterization and in vitro cellular viability. *Acta Biomater.* 2013; 9:8167–8181. [PubMed: 23727248]
- Ateshian GA, Nims RJ, Maas S, Weiss JA. Computational modeling of chemical reactions and interstitial growth and remodeling involving charged solutes and solid-bound molecules. *Biomech Model Mechanobiol.* 2014
- Awad HA, Boivin GP, Dressler MR, Smith FN, Young RG, Butler DL. Repair of patellar tendon injuries using a cell-collagen composite. *J Orthop Res.* 2003; 21:420–431. [PubMed: 12706014]
- Azeloglu EU, Albro MB, Thimmappa VA, Ateshian GA, Costa KD. Heterogeneous transmural proteoglycan distribution provides a mechanism for regulating residual stresses in the aorta. *Am J Physiol Heart Circ Physiol.* 2008; 294:H1197–1205. [PubMed: 18156194]
- Baaijens F, Bouten C, Hoerstrup S, Mol A, Driessen N, Boerboom R. Functional tissue engineering of the aortic heart valve. *Clin Hemorheol Microcirc.* 2005; 33:197–199. [PubMed: 16215285]
- Baer AE, Laursen TA, Guilak F, Setton LA. The micromechanical environment of intervertebral disc cells determined by a finite deformation, anisotropic, and biphasic finite element model. *J Biomech Eng.* 2003; 125:1–11. [PubMed: 12661192]
- Ballotta V, Driessen-Mol A, Bouten CV, Baaijens FP. Strain-dependent modulation of macrophage polarization within scaffolds. *Biomaterials.* 2014
- Beenakker JW, Ashcroft BA, Lindeman JH, Oosterkamp TH. Mechanical properties of the extracellular matrix of the aorta studied by enzymatic treatments. *Biophys J.* 2012; 102:1731–1737. [PubMed: 22768928]
- Betre H, Ong SR, Guilak F, Chilkoti A, Fermor B, Setton LA. Chondrocytic differentiation of human adipose-derived adult stem cells in elastin-like polypeptide. *Biomaterials.* 2006; 27:91–99. [PubMed: 16023192]
- Boerckel JD, Kolambkar YM, Stevens HY, Lin AS, Dupont KM, Guldborg RE. Effects of in vivo mechanical loading on large bone defect regeneration. *J Orthop Res.* 2012; 30:1067–1075. [PubMed: 22170172]
- Boffito M, Bernardi E, Sartori S, Ciardelli G, Sassi MP. A mechanical characterisation of polymer scaffolds and films at the macro- and nano-scale. *J Biomed Mater Res A.* 2014
- Boland ED, Matthews JA, Pawlowski KJ, Simpson DG, Wnek GE, Bowlin GL. Electrospinning collagen and elastin: preliminary vascular tissue engineering. *Front Biosci.* 2004; 9:1422–1432. [PubMed: 14977557]
- Bovendeerd PH. Modeling of cardiac growth and remodeling of myofiber orientation. *J Biomech.* 2012; 45:872–881. [PubMed: 22169149]
- Brunger JM, Huynh NP, Guenther CM, Perez-Pinera P, Moutos FT, Sanchez-Adams J, Gersbach CA, Guilak F. Scaffold-mediated lentiviral transduction for functional tissue engineering of cartilage. *Proc Natl Acad Sci U S A.* 2014; 111:E798–806. [PubMed: 24550481]
- Buckley MR, Sarver JJ, Freedman BR, Soslowsky LJ. The dynamics of collagen uncrimping and lateral contraction in tendon and the effect of ionic concentration. *J Biomech.* 2013; 46:2242–2249. [PubMed: 23876711]
- Buskohl PR, Gould RA, Butcher JT. Quantification of embryonic atrioventricular valve biomechanics during morphogenesis. *J Biomech.* 2012; 45:895–902. [PubMed: 22169154]
- Buskohl PR, Jenkins JT, Butcher JT. Computational simulation of hemodynamic-driven growth and remodeling of embryonic atrioventricular valves. *Biomech Model Mechanobiol.* 2012; 11:1205–1217. [PubMed: 22869343]
- Butler DL, Goldstein SA, Guilak F. Functional tissue engineering: the role of biomechanics. *J Biomech Eng.* 2000; 122:570–575. [PubMed: 11192376]
- Butler DL, Goldstein SA, Guldborg RE, Guo XE, Kamm R, Laurencin CT, McIntire LV, Mow VC, Nerem RM, Sah RL, Soslowsky LJ, Spilker RL, Tranquillo RT. The impact of biomechanics in tissue engineering and regenerative medicine. *Tissue Eng Part B Rev.* 2009; 15:477–484. [PubMed: 19583462]
- Butler DL, Juncosa-Melvin N, Boivin GP, Galloway MT, Shearn JT, Gooch C, Awad H. Functional tissue engineering for tendon repair: A multidisciplinary strategy using mesenchymal stem cells, bioscaffolds, and mechanical stimulation. *J Orthop Res.* 2008; 26:1–9. [PubMed: 17676628]

- Butscher A, Bohner M, Hofmann S, Gauckler L, Muller R. Structural and material approaches to bone tissue engineering in powder-based three-dimensional printing. *Acta Biomater.* 2011; 7:907–920. [PubMed: 20920616]
- Chao PH, Yodmuang S, Wang X, Sun L, Kaplan DL, Vunjak-Novakovic G. Silk hydrogel for cartilage tissue engineering. *J Biomed Mater Res B Appl Biomater.* 2010; 95:84–90. [PubMed: 20725950]
- Chew SY, Wen Y, Dzenis Y, Leong KW. The role of electrospinning in the emerging field of nanomedicine. *Curr Pharm Des.* 2006; 12:4751–4770. [PubMed: 17168776]
- Choe MM, Sporn PH, Swartz MA. Extracellular matrix remodeling by dynamic strain in a three-dimensional tissue-engineered human airway wall model. *Am J Respir Cell Mol Biol.* 2006; 35:306–313. [PubMed: 16601241]
- Cimetta E, Godier-Furnemont A, Vunjak-Novakovic G. Bioengineering heart tissue for in vitro testing. *Curr Opin Biotechnol.* 2013; 24:926–932. [PubMed: 23932513]
- Coleman JL, Widmyer MR, Leddy HA, Utturkar GM, Spritzer CE, Moorman CT 3rd, Guilak F, DeFrate LE. Diurnal variations in articular cartilage thickness and strain in the human knee. *J Biomech.* 2013; 46:541–547. [PubMed: 23102493]
- Courtney T, Sacks MS, Stankus J, Guan J, Wagner WR. Design and analysis of tissue engineering scaffolds that mimic soft tissue mechanical anisotropy. *Biomaterials.* 2006; 27:3631–3638. [PubMed: 16545867]
- Cuchiara MP, Gould DJ, McHale MK, Dickinson ME, West JL. Integration of Self-Assembled Microvascular Networks with Microfabricated PEG-Based Hydrogels. *Adv Funct Mater.* 2012; 22:4511–4518. [PubMed: 23536744]
- Daniele MA, Adams AA, Naciri J, North SH, Ligler FS. Interpenetrating networks based on gelatin methacrylamide and PEG formed using concurrent thiol click chemistries for hydrogel tissue engineering scaffolds. *Biomaterials.* 2014; 35:1845–1856. [PubMed: 24314597]
- Dankers PY, Harmsen MC, Brouwer LA, van Luyn MJ, Meijer EW. A modular and supramolecular approach to bioactive scaffolds for tissue engineering. *Nat Mater.* 2005; 4:568–574. [PubMed: 15965478]
- Darling EM, Athanasiou KA. Articular cartilage bioreactors and bioprocesses. *Tissue Eng.* 2003; 9:9–26. [PubMed: 12625950]
- De Hart J, Peters GW, Schreurs PJ, Baaijens FP. A three-dimensional computational analysis of fluid-structure interaction in the aortic valve. *J Biomech.* 2003; 36:103–112. [PubMed: 12485644]
- Defrate LE, van der Ven A, Boyer PJ, Gill TJ, Li G. The measurement of the variation in the surface strains of Achilles tendon grafts using imaging techniques. *J Biomech.* 2006; 39:399–405. [PubMed: 16389080]
- Dejaco A, Komlev VS, Jaroszewicz J, Swieszkowski W, Hellmich C. Micro CT-based multiscale elasticity of double-porous (pre-cracked) hydroxyapatite granules for regenerative medicine. *J Biomech.* 2012; 45:1068–1075. [PubMed: 22296936]
- Delaine-Smith RM, Green NH, Matcher SJ, Macneil S, Reilly GC. Monitoring fibrous scaffold guidance of three-dimensional collagen organisation using minimally-invasive second harmonic generation. *PLoS One.* 2014; 9:e89761. [PubMed: 24587017]
- Dellatore SM, Garcia AS, Miller WM. Mimicking stem cell niches to increase stem cell expansion. *Curr Opin Biotechnol.* 2008; 19:534–540. [PubMed: 18725291]
- DeLong SA, Moon JJ, West JL. Covalently immobilized gradients of bFGF on hydrogel scaffolds for directed cell migration. *Biomaterials.* 2005; 26:3227–3234. [PubMed: 15603817]
- Diekmann BO, Christoforou N, Willard VP, Sun H, Sanchez-Adams J, Leong KW, Guilak F. Cartilage tissue engineering using differentiated and purified induced pluripotent stem cells. *Proc Natl Acad Sci U S A.* 2012; 109:19172–19177. [PubMed: 23115336]
- Ekaputra AK, Prestwich GD, Cool SM, Huttmacher DW. Combining electrospun scaffolds with electrospayed hydrogels leads to three-dimensional cellularization of hybrid constructs. *Biomacromolecules.* 2008; 9:2097–2103. [PubMed: 18646822]
- Elliott DM, Setton LA. Anisotropic and inhomogeneous tensile behavior of the human annulus fibrosus: experimental measurement and material model predictions. *J Biomech Eng.* 2001; 123:256–263. [PubMed: 11476369]

- Engelmayr GC Jr, Cheng M, Bettinger CJ, Borenstein JT, Langer R, Freed LE. Accordion-like honeycombs for tissue engineering of cardiac anisotropy. *Nat Mater.* 2008; 7:1003–1010. [PubMed: 18978786]
- Fan R, Bayoumi AS, Chen P, Hobson CM, Wagner WR, Mayer JE Jr, Sacks MS. Optimal elastomeric scaffold leaflet shape for pulmonary heart valve leaflet replacement. *J Biomech.* 2013; 46:662–669. [PubMed: 23294966]
- Francisco AT, Hwang PY, Jeong CG, Jing L, Chen J, Setton LA. Photocrosslinkable laminin-functionalized polyethylene glycol hydrogel for intervertebral disc regeneration. *Acta Biomater.* 2014; 10:1102–1111. [PubMed: 24287160]
- Freeman JW, Woods MD, Cromer DA, Ekwueme EC, Andric T, Atiemo EA, Bijoux CH, Laurencin CT. Evaluation of a hydrogel-fiber composite for ACL tissue engineering. *J Biomech.* 2011; 44:694–699. [PubMed: 21111422]
- Fritton SP, McLeod KJ, Rubin CT. Quantifying the strain history of bone: spatial uniformity and self-similarity of low-magnitude strains. *J Biomech.* 2000; 33:317–325. [PubMed: 10673115]
- Garg K, Pullen NA, Oskeritizian CA, Ryan JJ, Bowlin GL. Macrophage functional polarization (M1/M2) in response to varying fiber and pore dimensions of electrospun scaffolds. *Biomaterials.* 2013; 34:4439–4451. [PubMed: 23515178]
- Gauvin R, Chen YC, Lee JW, Soman P, Zorlutuna P, Nichol JW, Bae H, Chen S, Khademhosseini A. Microfabrication of complex porous tissue engineering scaffolds using 3D projection stereolithography. *Biomaterials.* 2012; 33:3824–3834. [PubMed: 22365811]
- Gilchrist CL, Darling EM, Chen J, Setton LA. Extracellular matrix ligand and stiffness modulate immature nucleus pulposus cell-cell interactions. *PLoS One.* 2011; 6:e27170. [PubMed: 22087260]
- Glass KA, Link JM, Brunger JM, Moutos FT, Gersbach CA, Guilak F. Tissue-engineered cartilage with inducible and tunable immunomodulatory properties. *Biomaterials.* 2014 in press.
- Groves RB, Coulman SA, Birchall JC, Evans SL. An anisotropic, hyperelastic model for skin: experimental measurements, finite element modelling and identification of parameters for human and murine skin. *J Mech Behav Biomed Mater.* 2013; 18:167–180. [PubMed: 23274398]
- Guex AG, Birrer DL, Fortunato G, Tevaearai HT, Giraud MN. Anisotropically oriented electrospun matrices with an imprinted periodic micropattern: a new scaffold for engineered muscle constructs. *Biomed Mater.* 2013; 8:021001. [PubMed: 23343525]
- Guilak F. Functional tissue engineering: the role of biomechanics in reparative medicine. *Ann N Y Acad Sci.* 2002; 961:193–195. [PubMed: 12081896]
- Guilak F, Butler DL, Goldstein SA. Functional tissue engineering: the role of biomechanics in articular cartilage repair. *Clin Orthop Relat Res.* 2001:S295–305. [PubMed: 11603713]
- Guilak, F.; Butler, DL.; Mooney, D.; Goldstein, S. *Functional Tissue Engineering.* Springer-Verlag; New York: 2003.
- Guilak F, Cohen DM, Estes BT, Gimble JM, Liedtke W, Chen CS. Control of stem cell fate by physical interactions with the extracellular matrix. *Cell Stem Cell.* 2009; 5:17–26. [PubMed: 19570510]
- Guilak F, Kapur R, Sefton MV, Vandenberg HH, Koretsky AP, Kriete A, O’Keefe RJ. Functional assessment of engineered tissues and elements of tissue design: breakout session summary. *Ann N Y Acad Sci.* 2002; 961:207–209. [PubMed: 12081902]
- Hagenmuller H, Hitz M, Merkle HP, Meinel L, Muller R. Design and validation of a novel bioreactor principle to combine online micro-computed tomography monitoring and mechanical loading in bone tissue engineering. *Rev Sci Instrum.* 2010; 81:014303. [PubMed: 20113118]
- Haider MA, Olander JE, Arnold RF, Marous DR, McLamb AJ, Thompson KC, Woodruff WR, Haugh JM. A phenomenological mixture model for biosynthesis and linking of cartilage extracellular matrix in scaffolds seeded with chondrocytes. *Biomech Model Mechanobiol.* 2011; 10:915–924. [PubMed: 21213013]
- Hammond NA, Kamm RD. Mechanical characterization of self-assembling peptide hydrogels by microindentation. *J Biomed Mater Res B Appl Biomater.* 2013; 101:981–990. [PubMed: 23529940]

- Hansmann J, Groeber F, Kahlig A, Kleinhans C, Walles H. Bioreactors in tissue engineering – principles, applications and commercial constraints. *Biotechnol J*. 2013; 8:298–307. [PubMed: 23161827]
- Hollister SJ. Porous scaffold design for tissue engineering. *Nat Mater*. 2005; 4:518–524. [PubMed: 16003400]
- Hong Y, Huber A, Takanari K, Amoroso NJ, Hashizume R, Badylak SF, Wagner WR. Mechanical properties and in vivo behavior of a biodegradable synthetic polymer microfiber-extracellular matrix hydrogel biohybrid scaffold. *Biomaterials*. 2011; 32:3387–3394. [PubMed: 21303718]
- Hoque ME, Hutmacher DW, Feng W, Li S, Huang MH, Vert M, Wong YS. Fabrication using a rapid prototyping system and in vitro characterization of PEG-PCL-PLA scaffolds for tissue engineering. *J Biomater Sci Polym Ed*. 2005; 16:1595–1610. [PubMed: 16366339]
- Huang AH, Baker BM, Ateshian GA, Mauck RL. Sliding contact loading enhances the tensile properties of mesenchymal stem cell-seeded hydrogels. *Eur Cell Mater*. 2012; 24:29–45. [PubMed: 22791371]
- Huang CY, Stankiewicz A, Ateshian GA, Mow VC. Anisotropy, inhomogeneity, and tension-compression nonlinearity of human glenohumeral cartilage in finite deformation. *J Biomech*. 2005; 38:799–809. [PubMed: 15713301]
- Huey DJ, Athanasiou KA. Tension-compression loading with chemical stimulation results in additive increases to functional properties of anatomic meniscal constructs. *PLoS One*. 2011; 6:e27857. [PubMed: 22114714]
- Hume PS, Bowman CN, Anseth KS. Functionalized PEG hydrogels through reactive dip-coating for the formation of immunoactive barriers. *Biomaterials*. 2011; 32:6204–6212. [PubMed: 21658759]
- Hwang M, Garbey M, Berceli SA, Wu R, Jiang Z, Tran-Son-Tay R. Rule-based model of vein graft remodeling. *PLoS One*. 2013; 8:e57822. [PubMed: 23533576]
- Ingber DE. Mechanobiology and diseases of mechanotransduction. *Ann Med*. 2003; 35:564–577. [PubMed: 14708967]
- Isenberg BC, Backman DE, Kinahan ME, Jesudason R, Suki B, Stone PJ, Davis EC, Wong JY. Micropatterned cell sheets with defined cell and extracellular matrix orientation exhibit anisotropic mechanical properties. *J Biomech*. 2012; 45:756–761. [PubMed: 22177672]
- Juncosa N, West JR, Galloway MT, Boivin GP, Butler DL. In vivo forces used to develop design parameters for tissue engineered implants for rabbit patellar tendon repair. *J Biomech*. 2003; 36:483–488. [PubMed: 12600338]
- Juncosa-Melvin N, Matlin KS, Holdcraft RW, Nirmalanandhan VS, Butler DL. Mechanical stimulation increases collagen type I and collagen type III gene expression of stem cell-collagen sponge constructs for patellar tendon repair. *Tissue Eng*. 2007; 13:1219–1226. [PubMed: 17518715]
- Jungreuthmayer C, Jaasma MJ, Al-Munajjed AA, Zanghellini J, Kelly DJ, O'Brien FJ. Deformation simulation of cells seeded on a collagen-GAG scaffold in a flow perfusion bioreactor using a sequential 3D CFD-elastostatics model. *Med Eng Phys*. 2009; 31:420–427. [PubMed: 19109048]
- Kisiel M, Martino MM, Ventura M, Hubbell JA, Hilborn J, Ossipov DA. Improving the osteogenic potential of BMP-2 with hyaluronic acid hydrogel modified with integrin-specific fibronectin fragment. *Biomaterials*. 2013; 34:704–712. [PubMed: 23103154]
- Klisch SM, Asanbaeva A, Oungoulian SR, Masuda K, Thonar EJ, Davol A, Sah RL. A cartilage growth mixture model with collagen remodeling: validation protocols. *J Biomech Eng*. 2008; 130:031006. [PubMed: 18532855]
- Kolambkar YM, Bajin M, Wojtowicz A, Hutmacher DW, Garcia AJ, Guldborg RE. Nanofiber orientation and surface functionalization modulate human mesenchymal stem cell behavior in vitro. *Tissue Eng Part A*. 2014; 20:398–409. [PubMed: 24020454]
- Korhonen RK, Julkunen P, Wilson W, Herzog W. Importance of collagen orientation and depth-dependent fixed charge densities of cartilage on mechanical behavior of chondrocytes. *J Biomech Eng*. 2008; 130:021003. [PubMed: 18412490]
- Kotecha M, Klatt D, Magin RL. Monitoring cartilage tissue engineering using magnetic resonance spectroscopy, imaging, and elastography. *Tissue Eng Part B Rev*. 2013; 19:470–484. [PubMed: 23574498]

- Langer R, Vacanti JP. Tissue engineering. *Science*. 1993; 260:920–926. [PubMed: 8493529]
- Liao IC, Moutos FT, Estes BT, Zhao X, Guilak F. Composite three-dimensional woven scaffolds with interpenetrating network hydrogels to create functional synthetic articular cartilage. *Adv Funct Mater*. 2013; 23:5833–5839. [PubMed: 24578679]
- Liberski AR, Delaney JT Jr, Schafer H, Perelaer J, Schubert US. Organ weaving: woven threads and sheets as a step towards a new strategy for artificial organ development. *Macromol Biosci*. 2011; 11:1491–1498. [PubMed: 21916011]
- Likhitpanichkul M, Guo XE, Mow VC. The effect of matrix tension-compression nonlinearity and fixed negative charges on chondrocyte responses in cartilage. *Mol Cell Biomech*. 2005; 2:191–204. [PubMed: 16705865]
- Lim JY, Loisel AE, Lee JS, Zhang Y, Salvi JD, Donahue HJ. Optimizing the osteogenic potential of adult stem cells for skeletal regeneration. *J Orthop Res*. 2011; 29:1627–1633. [PubMed: 21509820]
- Lima EG, Tan AR, Tai T, Marra KG, DeFail A, Ateshian GA, Hung CT. Genipin enhances the mechanical properties of tissue-engineered cartilage and protects against inflammatory degradation when used as a medium supplement. *J Biomed Mater Res A*. 2009; 91:692–700. [PubMed: 19025982]
- Liu J, Agarwal S. Mechanical signals activate vascular endothelial growth factor receptor-2 to upregulate endothelial cell proliferation during inflammation. *J Immunol*. 2010; 185:1215–1221. [PubMed: 20548028]
- Liu YS, Lee OK. In search of the pivot point of mechanotransduction: mechanosensing of stem cells. *Cell Transplant*. 2014; 23:1–11. [PubMed: 24439034]
- Loerakker S, Argento G, Oomens CW, Baaijens FP. Effects of valve geometry and tissue anisotropy on the radial stretch and coaptation area of tissue-engineered heart valves. *J Biomech*. 2013; 46:1792–1800. [PubMed: 23786664]
- Loerakker S, Obbink-Huizer C, Baaijens FP. A physically motivated constitutive model for cell-mediated compaction and collagen remodeling in soft tissues. *Biomech Model Mechanobiol*. 2013
- Lysaght MJ, Jaklenec A, Deweerd E. Great expectations: private sector activity in tissue engineering, regenerative medicine, and stem cell therapeutics. *Tissue Eng Part A*. 2008; 14:305–315. [PubMed: 18333783]
- Mammoto A, Mammoto T, Ingber DE. Mechanosensitive mechanisms in transcriptional regulation. *J Cell Sci*. 2012; 125:3061–3073. [PubMed: 22797927]
- Marcos-Campos I, Marolt D, Petridis P, Bhumiratana S, Schmidt D, Vunjak-Novakovic G. Bone scaffold architecture modulates the development of mineralized bone matrix by human embryonic stem cells. *Biomaterials*. 2012; 33:8329–8342. [PubMed: 22901965]
- Mariappan YK, Glaser KJ, Ehman RL. Magnetic resonance elastography: a review. *Clin Anat*. 2010; 23:497–511. [PubMed: 20544947]
- Marom G, Peleg M, Halevi R, Rosenfeld M, Raanani E, Hamdan A, Haj-Ali R. Fluid-structure interaction model of aortic valve with porcine-specific collagen fiber alignment in the cusps. *J Biomech Eng*. 2013; 135:101001–101006. [PubMed: 23775457]
- Martufi G, Gasser TC. A constitutive model for vascular tissue that integrates fibril, fiber and continuum levels with application to the isotropic and passive properties of the infrarenal aorta. *J Biomech*. 2011; 44:2544–2550. [PubMed: 21862020]
- Marturano JE, Arena JD, Schiller ZA, Georgakoudi I, Kuo CK. Characterization of mechanical and biochemical properties of developing embryonic tendon. *Proc Natl Acad Sci U S A*. 2013; 110:6370–6375. [PubMed: 23576745]
- Marturano JE, Xylas JF, Sridharan GV, Georgakoudi I, Kuo CK. Lysyl oxidase-mediated collagen crosslinks may be assessed as markers of functional properties of tendon tissue formation. *Acta Biomater*. 2014; 10:1370–1379. [PubMed: 24316363]
- Mauck RL, Baker BM, Nerurkar NL, Burdick JA, Li WJ, Tuan RS, Elliott DM. Engineering on the straight and narrow: the mechanics of nanofibrous assemblies for fiber-reinforced tissue regeneration. *Tissue Eng Part B Rev*. 2009; 15:171–193. [PubMed: 19207040]

- McLeod MA, Wilusz RE, Guilak F. Depth-dependent anisotropy of the micromechanical properties of the extracellular and pericellular matrices of articular cartilage evaluated via atomic force microscopy. *J Biomech.* 2013; 46:586–592. [PubMed: 23062866]
- Meinel L, Karageorgiou V, Fajardo R, Snyder B, Shinde-Patil V, Zichner L, Kaplan D, Langer R, Vunjak-Novakovic G. Bone tissue engineering using human mesenchymal stem cells: effects of scaffold material and medium flow. *Ann Biomed Eng.* 2004; 32:112–122. [PubMed: 14964727]
- Mironov V, Visconti RP, Kasyanov V, Forgacs G, Drake CJ, Markwald RR. Organ printing: tissue spheroids as building blocks. *Biomaterials.* 2009; 30:2164–2174. [PubMed: 19176247]
- Motaghinasab S, Shirazi-Adl A, Urban JP, Parnianpour M. Computational pharmacokinetics of solute penetration into human intervertebral discs – effects of endplate permeability, solute molecular weight and disc size. *J Biomech.* 2012; 45:2195–2202. [PubMed: 22840491]
- Moutos FT, Estes BT, Guilak F. Multifunctional hybrid three-dimensionally woven scaffolds for cartilage tissue engineering. *Macromol Biosci.* 2010; 10:1355–1364. [PubMed: 20857388]
- Moutos FT, Freed LE, Guilak F. A biomimetic three-dimensional woven composite scaffold for functional tissue engineering of cartilage. *Nat Mater.* 2007; 6:162–167. [PubMed: 17237789]
- Mow, VC.; Bachrach, N.; Setton, LA.; Guilak, F. Stress, strain, pressure, and flow fields in articular cartilage. In: Mow, VC.; Guilak, F.; Tran-Son-Tay, R.; Hochmuth, R., editors. *Cell Mechanics and Cellular Engineering.* Springer Verlag; New York: 1994. p. 345-379.
- Nagel T, Kelly DJ. Remodelling of collagen fibre transition stretch and angular distribution in soft biological tissues and cell-seeded hydrogels. *Biomech Model Mechanobiol.* 2012; 11:325–339. [PubMed: 21611762]
- Nerurkar NL, Elliott DM, Mauck RL. Mechanical design criteria for intervertebral disc tissue engineering. *J Biomech.* 2010; 43:1017–1030. [PubMed: 20080239]
- Nerurkar NL, Mauck RL, Elliott DM. ISSLS prize winner: integrating theoretical and experimental methods for functional tissue engineering of the annulus fibrosus. *Spine (Phila Pa 1976).* 2008; 33:2691–2701. [PubMed: 19018251]
- Neumann AJ, Alini M, Archer CW, Stoddart MJ. Chondrogenesis of human bone marrow-derived mesenchymal stem cells is modulated by complex mechanical stimulation and adenoviral-mediated overexpression of bone morphogenetic protein 2. *Tissue Eng Part A.* 2013; 19:1285–1294. [PubMed: 23289669]
- Nii M, Lai JH, Keeney M, Han LH, Behn A, Imanbayev G, Yang F. The effects of interactive mechanical and biochemical niche signaling on osteogenic differentiation of adipose-derived stem cells using combinatorial hydrogels. *Acta Biomater.* 2013; 9:5475–5483. [PubMed: 23153761]
- Niklason LE, Yeh AT, Calle EA, Bai Y, Valentin A, Humphrey JD. Enabling tools for engineering collagenous tissues integrating bioreactors, intravital imaging, and biomechanical modeling. *Proc Natl Acad Sci U S A.* 2010; 107:3335–3339. [PubMed: 19955446]
- O’Conor CJ, Case N, Guilak F. Mechanical regulation of chondrogenesis. *Stem Cell Res Ther.* 2013; 4:61. [PubMed: 23809493]
- O’Conor CJ, Leddy HA, Benefield HC, Liedtke WB, Guilak F. TRPV4-mediated mechanotransduction regulates the metabolic response of chondrocytes to dynamic loading. *Proc Natl Acad Sci U S A.* 2014; 111:1316–1321. [PubMed: 24474754]
- Ousema PH, Moutos FT, Estes BT, Caplan AI, Lennon DP, Guilak F, Weinberg JB. The inhibition by interleukin 1 of MSC chondrogenesis and the development of biomechanical properties in biomimetic 3D woven PCL scaffolds. *Biomaterials.* 2012; 33:8967–8974. [PubMed: 22999467]
- Paten JA, Tilburey GE, Molloy EA, Zareian R, Trainor CV, Ruberti JW. Utility of an optically-based, micromechanical system for printing collagen fibers. *Biomaterials.* 2013; 34:2577–2587. [PubMed: 23352045]
- Pennella F, Cerino G, Massai D, Gallo D, Falvo D’Urso Labate G, Schiavi A, Deriu MA, Audenino A, Morbiducci U. A survey of methods for the evaluation of tissue engineering scaffold permeability. *Ann Biomed Eng.* 2013; 41:2027–2041. [PubMed: 23612914]
- Pioletti DP. Integration of mechanotransduction concepts in bone tissue engineering. *Comput Methods Biomech Biomed Engin.* 2013; 16:1050–1055. [PubMed: 23531231]

- Reilly GC, Engler AJ. Intrinsic extracellular matrix properties regulate stem cell differentiation. *J Biomech.* 2010; 43:55–62. [PubMed: 19800626]
- Rennerfeldt DA, Renth AN, Talata Z, Gehrke SH, Detamore MS. Tuning mechanical performance of poly(ethylene glycol) and agarose interpenetrating network hydrogels for cartilage tissue engineering. *Biomaterials.* 2013; 34:8241–8257. [PubMed: 23932504]
- Rouwkema J, Gibbs S, Lutolf MP, Martin I, Vunjak-Novakovic G, Malda J. In vitro platforms for tissue engineering: implications for basic research and clinical translation. *J Tissue Eng Regen Med.* 2011; 5:e164–167. [PubMed: 21774080]
- Salimath AS, Garcia AJ. Biofunctional hydrogels for skeletal muscle constructs. *J Tissue Eng Regen Med.* 2014
- Sampat SR, Dermksian MV, Oungouljian SR, Winchester RJ, Bulinski JC, Ateshian GA, Hung CT. Applied osmotic loading for promoting development of engineered cartilage. *J Biomech.* 2013; 46:2674–2681. [PubMed: 24035014]
- Schatti O, Grad S, Goldhahn J, Salzmann G, Li Z, Alini M, Stoddart MJ. A combination of shear and dynamic compression leads to mechanically induced chondrogenesis of human mesenchymal stem cells. *Eur Cell Mater.* 2011; 22:214–225. [PubMed: 22048899]
- Schmidt D, Dijkman PE, Driessen-Mol A, Stenger R, Mariani C, Puolakka A, Rissanen M, Deichmann T, Odermatt B, Weber B, Emmert MY, Zund G, Baaijens FP, Hoerstrup SP. Minimally-invasive implantation of living tissue engineered heart valves: a comprehensive approach from autologous vascular cells to stem cells. *J Am Coll Cardiol.* 2010; 56:510–520. [PubMed: 20670763]
- Shah N, Morsi Y, Manasseh R. From mechanical stimulation to biological pathways in the regulation of stem cell fate. *Cell Biochem Funct.* 2014
- Sharma B, Fermanian S, Gibson M, Unterman S, Herzka DA, Cascio B, Coburn J, Hui AY, Marcus N, Gold GE, Elisseeff JH. Human cartilage repair with a photoreactive adhesive-hydrogel composite. *Sci Transl Med.* 2013; 5:167ra166.
- Shirakawa H, Furusawa K, Fukui A, Tadano S, Sasaki N. Changes in the viscoelastic properties of cortical bone by selective degradation of matrix protein. *J Biomech.* 2013; 46:696–701. [PubMed: 23261016]
- Shirazi-Adl A. On the fibre composite material models of disc annulus--comparison of predicted stresses. *J Biomech.* 1989; 22:357–365. [PubMed: 2745470]
- Soliman S, Pagliari S, Rinaldi A, Forte G, Fiaccavento R, Pagliari F, Franzese O, Minieri M, Di Nardo P, Licocchia S, Traversa E. Multiscale three-dimensional scaffolds for soft tissue engineering via multimodal electrospinning. *Acta Biomater.* 2010; 6:1227–1237. [PubMed: 19887125]
- Sommer G, Eder M, Kovacs L, Pathak H, Bonitz L, Mueller C, Regitnig P, Holzapfel GA. Multiaxial mechanical properties and constitutive modeling of human adipose tissue: a basis for preoperative simulations in plastic and reconstructive surgery. *Acta Biomater.* 2013; 9:9036–9048. [PubMed: 23811521]
- Song MJ, Dean D, Knothe Tate ML. Mechanical modulation of nascent stem cell lineage commitment in tissue engineering scaffolds. *Biomaterials.* 2013; 34:5766–5775. [PubMed: 23660249]
- Stahl PJ, Romano NH, Wirtz D, Yu SM. PEG-based hydrogels with collagen mimetic peptide-mediated and tunable physical cross-links. *Biomacromolecules.* 2010; 11:2336–2344. [PubMed: 20715762]
- Suri S, Schmidt CE. Photopatterned collagen-hyaluronic acid interpenetrating polymer network hydrogels. *Acta Biomater.* 2009; 5:2385–2397. [PubMed: 19446050]
- Taylor KA, Cutcliffe HC, Queen RM, Utturkar GM, Spritzer CE, Garrett WE, DeFrate LE. In vivo measurement of ACL length and relative strain during walking. *J Biomech.* 2013; 46:478–483. [PubMed: 23178040]
- Teng Z, Trabelsi O, Ochoa I, He J, Gillard JH, Doblare M. Anisotropic material behaviours of soft tissues in human trachea: an experimental study. *J Biomech.* 2012; 45:1717–1723. [PubMed: 22534565]
- Tseng H, Cuchiara ML, Durst CA, Cuchiara MP, Lin CJ, West JL, Grande-Allen KJ. Fabrication and mechanical evaluation of anatomically-inspired quasilaminate hydrogel structures with layer-specific formulations. *Ann Biomed Eng.* 2013; 41:398–407. [PubMed: 23053300]

- Tuin SA, Pourdeyhimi B, Lobo EG. Interconnected, Microporous Hollow Fibers for Tissue Engineering: Commercially Relevant, Industry Standard Scale-up Manufacturing. *J Biomed Mater Res A*. 2013
- Van Canneyt K, Morbiducci U, Elout S, De Santis G, Segers P, Verdonck P. A computational exploration of helical arterio-venous graft designs. *J Biomech*. 2013; 46:345–353. [PubMed: 23159095]
- van Lieshout MI, Vaz CM, Rutten MC, Peters GW, Baaijens FP. Electrospinning versus knitting: two scaffolds for tissue engineering of the aortic valve. *J Biomater Sci Polym Ed*. 2006; 17:77–89. [PubMed: 16411600]
- Vaz CM, van Tuijl S, Bouten CV, Baaijens FP. Design of scaffolds for blood vessel tissue engineering using a multi-layering electrospinning technique. *Acta Biomater*. 2005; 1:575–582. [PubMed: 16701837]
- Vogl TJ, Then C, Naguib NN, Nour-Eldin NE, Larson M, Zangos S, Silber G. Mechanical soft tissue property validation in tissue engineering using magnetic resonance imaging experimental research. *Acad Radiol*. 2010; 17:1486–1491. [PubMed: 20926314]
- Wang LS, Chung JE, Chan PP, Kurisawa M. Injectable biodegradable hydrogels with tunable mechanical properties for the stimulation of neurogenesis differentiation of human mesenchymal stem cells in 3D culture. *Biomaterials*. 2010; 31:1148–1157. [PubMed: 19892395]
- Wang YK, Chen CS. Cell adhesion and mechanical stimulation in the regulation of mesenchymal stem cell differentiation. *J Cell Mol Med*. 2013; 17:823–832. [PubMed: 23672518]
- Willie BM, Yang X, Kelly NH, Han J, Nair T, Wright TM, van der Meulen MC, Bostrom MP. Cancellous bone osseointegration is enhanced by in vivo loading. *Tissue Eng Part C Methods*. 2010; 16:1399–1406. [PubMed: 20367497]
- Xiao Y, Friis EA, Gehrke SH, Detamore MS. Mechanical testing of hydrogels in cartilage tissue engineering: beyond the compressive modulus. *Tissue Eng Part B Rev*. 2013; 19:403–412. [PubMed: 23448091]
- Yeatts AB, Choquette DT, Fisher JP. Bioreactors to influence stem cell fate: augmentation of mesenchymal stem cell signaling pathways via dynamic culture systems. *Biochim Biophys Acta*. 2013; 1830:2470–2480. [PubMed: 22705676]
- Zeugolis DI, Paul GR, Attenburrow G. Cross-linking of extruded collagen fibers--a biomimetic three-dimensional scaffold for tissue engineering applications. *J Biomed Mater Res A*. 2009; 89:895–908. [PubMed: 18465819]
- Zhang X, Wang X, Keshav V, Johanas JT, Leisk GG, Kaplan DL. Dynamic culture conditions to generate silk-based tissue-engineered vascular grafts. *Biomaterials*. 2009; 30:3213–3223. [PubMed: 19232717]

Table 1
Questions on the role of biomechanics in tissue engineering

-
- What are the ranges of stress and strain that native or repair tissues experience *in vivo* under physiologic or pathologic conditions?
 - What are the mechanical properties of native, diseased, or repair tissues at multiple scales?
 - Which of these properties should be used as design parameters in the development of engineered tissues?
 - What are the mechanical properties of biomaterials, scaffolds, and engineered tissues (at multiple scales) and how can we control these properties as design parameters?
 - When evaluating the biomechanical function of tissue-engineered repairs, how do we define success?
 - Can mechanical stimulation (i.e., in bioreactors) improve implant properties before implantation?
 - How do physical factors influence the behavior of cells and engineered constructs once they are implanted *in vivo*?
 - How do physical interactions with the microenvironment regulate the growth, differentiation, and metabolism of either primary or stem/progenitor cells?
 - How do biomechanical factors influence growth and remodeling of native or repair tissues?
 - What parameters and constitutive models can be used to predict the growth and remodeling of engineered tissues?
 - How do other genetic and environmental factors (e.g., co-morbidities, inflammation, etc.) interact with biomechanical factors to modulate tissue-engineered repair?
-

Table 2
Functional Properties of Natural And Engineered Tissues [adapted from (Butler et al., 2000), with permission]

Anisotropy (Properties vary with direction)
Tensile, compressive, shear moduli
Hydraulic permeability
Diffusion and other transport properties
Friction and wear
Failure stress and strain
Fatigue life
Inhomogeneity (Properties vary with site)
Potentially all properties
Morphology
Congruence
Microstructure
Nanotopography
Nonlinearity
Material nonlinearity
Tension-compression nonlinearity
Nonlinear permeability
Nonlinear diffusivity and deformation-diffusion coupling
Nonlinear viscoelasticity
Coupling of normal and shear stresses
Friction and wear
Physicochemical-mechanical coupling
Electrokinetic effects
Mechano-osmotic coupling
Mechano-electric coupling (e.g., piezoelectricity)
Residual stresses
Swelling
Tribological Properties
Frictional coefficient
Wear properties
Adhesive properties
Hardness
Viscoelasticity
Multiphase or poroelastic
Energy dissipation
Intrinsic material viscoelasticity
Fluid viscosity
