

HHS Public Access

Neurobiol Aging. Author manuscript; available in PMC 2015 September 01.

Published in final edited form as:

Author manuscript

Neurobiol Aging. 2014 September ; 35(9): 2096–2106. doi:10.1016/j.neurobiolaging.2014.03.006.

¹⁸F-fluorodeoxyglucose positron emission tomography, aging, and apolipoprotein E genotype in cognitively normal persons

David S. Knopmana,b,* , **Clifford R. Jack Jr.**b,c , **Heather J. Wiste**d, **Emily S. Lundt**d, **Stephen** D. Weigand^d, Prashanthi Vemuri^a, Val J. Lowe^c, Kejal Kantarci^c, Jeffrey L. Gunter^c, Matthew L. Senjem^c, Michelle M. Mielke^e, Rosebud O. Roberts^{a,e}, Bradley F. Boeve^{a,b}, and **Ronald C. Petersen**a,b,e

aDepartment of Neurology, Mayo Clinic and Foundation, Rochester, MN, USA

^bMayo Clinic Alzheimer's Disease Research Center, Mayo Clinic and Foundation, Rochester, MN, USA

^cDepartment of Radiology, Mayo Clinic and Foundation, Rochester, MN, USA

^dDivision of Biomedical Statistics and Informatics, Department of Health Sciences Research, Mayo Clinic and Foundation, Rochester, MN, USA

^eDivision of Epidemiology, Department of Health Sciences Research, Mayo Clinic and Foundation, Rochester, MN, USA

Abstract

Our objective was to examine associations between glucose metabolism, as measured by ^{18}F fluorodeoxyglucose positron emission tomography (FDG PET), and age and to evaluate the impact of carriage of an apolipoprotein E (*APOE*) ε4 allele on glucose metabolism and on the associations between glucose metabolism and age. We studied 806 cognitively normal (CN) and 70 amyloid-imaging-positive cognitively impaired participants (35 with mild cognitive impairment and 35 with Alzheimer's disease [AD] dementia) from the Mayo Clinic Study of Aging, Mayo Alzheimer's Disease Research Center and an ancillary study who had undergone

^{© 2014} Elsevier Inc. All rights reserved.

^{*}Corresponding author at: Department of Neurology, College of Medicine, Mayo Clinic, 200 First Street SW, Rochester, MN 55905, USA. Tel.: +1 507 538 1038; fax: +1 507 538 6012. knopman@mayo.edu (D.S. Knopman).

Disclosure statement

The authors except the following authors have no conflicts of interest to disclose. Dr Knopman serves as Deputy Editor for Neurology; served on a Data Safety Monitoring Board for Lilly Pharmaceuticals; will serve on a Data Safety Monitoring Board for Lundbeck Pharmaceuticals and for the DIAN study; served as a consultant to TauRx Pharmaceuticals, was an investigator in clinical trials sponsored by Baxter and TauRX Pharmaceuticals in the past 2 years; and receives research support from the NIH. Dr Jack serves on scientific advisory boards for Pfizer, Elan and/or Janssen AI, Eli Lilly & Company, GE Healthcare; receives research support from Baxter International Inc, Allon Therapeutics, Inc, the NIH and/or NIA, and the Alexander Family Alzheimer's Disease Research Professorship of the Mayo Foundation; and holds stock in Johnson & Johnson. Dr Mielke receives research grants from the NIH/NIA, Alzheimer Drug Discovery Foundation, Lewy Body Association, and the Driskill Foundation. Dr Lowe serves on scientific advisory boards for Bayer Schering Pharma and receives research support from GE Healthcare, Siemens Molecular Imaging, AVID Radiopharmaceuticals, and the NIH (NIA, NCI). Dr Kantarci receives research grants from the NIH/NIA. Dr Roberts receives research support from Abbvie Health and Economics Outcomes Research and from the NIH/NIA. Dr Boeve receives royalties from the publication of Behavioral Neurology of Dementia and receives research support from Cephalon, Inc, Allon Therapeutics, GE Healthcare, the NIH/NIA, and the Mangurian Foundation. Dr Petersen serves on scientific advisory boards for Pfizer, Inc, Janssen Alzheimer Immunotherapy, Elan Pharmaceuticals, and GE Healthcare; receives royalties from the publication of Mild Cognitive Impairment (Oxford University Press, 2003); and receives research support from the NIH/NIA.

structural MRI, FDG PET, and ${}^{11}C$ -Pittsburgh compound B (PiB) PET. Using partial volume corrected and uncorrected FDG PET glucose uptake ratios, we evaluated associations of regional FDG ratios with age and carriage of an *APOE* ε4 allele in CN participants between the ages of 30 and 95 years, and compared those findings with the cognitively impaired participants. In regionof-interest (ROI) analyses, we found modest but statistically significant declines in FDG ratio in most cortical and subcortical regions as a function of age. We also found a main effect of *APOE* ε4 genotype on FDG ratio, with greater uptake in ε4 noncarriers compared with carriers but only in the posterior cingulate and/or precuneus, lateral parietal, and AD-signature meta-ROI. The latter consisted of voxels from posterior cingulate and/or precuneus, lateral parietal, and inferior temporal. In age- and sex-matched CN participants the magnitude of the difference in partial volume corrected FDG ratio in the AD-signature meta-ROI for *APOE* ε4 carriers compared with noncarriers was about 4 times smaller than the magnitude of the difference between age- and sexmatched elderly *APOE* ε4 carrier CN compared with AD dementia participants. In an analysis in participants older than 70 years (31.3% of whom had elevated PiB), there was no interaction between PiB status and *APOE* ε4 genotype with respect to glucose metabolism. Glucose metabolism declines with age in many brain regions. Carriage of an *APOE* ε4 allele was associated with reductions in FDG ratio in the posterior cingulate and/or precuneus, lateral parietal, and AD-signature ROIs, and there was no interaction between age and *APOE* ε4 status. The posterior cingulate and/or precuneus and lateral parietal regions have a unique vulnerability to reductions in glucose metabolic rate as a function both of age and carriage of an *APOE* ε4 allele.

Keywords

Aging; Alzheimer's disease; FDG positron emission tomography; Apolipoprotein E

1. Introduction

Understanding the influence of aging on Alzheimer's disease (AD) biomarkers is essential to our ongoing effort to model the pathophysiology of AD (Jack et al., 2013). For ^{18}F fluorodeoxyglucose positron emission tomography (FDG PET), there has been controversy about whether brain glucose metabolism declines with aging in the absence of cognitive impairment. Early reports claimed that there were declines in glucose metabolism (Herholz et al., 2002; Loessner et al., 1995). Most subsequent studies but not all (Kalpouzos et al., 2009) claimed that declines in glucose metabolism over the adult age spectrum were abolished by partial volume correction (Ibanez et al., 2004; Kantarci et al., 2010; Yanase et al., 2005). Any analysis of the role of age on glucose metabolism must also take apolipoprotein E (*APOE*) genotype into account. Reductions in glucose metabolism in regions known to be affected in AD dementia (Chetelat et al., 2008; Mosconi, 2005; Nestor et al., 2003; Rabinovici et al., 2010) have been shown in cognitively normal middle-aged carriers of the *APOE* ε4 allele (Reiman et al., 1996, 2001).

We examined associations between glucose metabolism, as measured by FDG PET and age in a large number of cognitively normal (CN) individuals. We also evaluated the impact of carriage of an *APOE* ε4 allele on glucose metabolic rate and whether the associations

between FDG and age differed among *APOE* ε4 carriers and noncarriers. A group of persons with cognitive impairment due to of AD were also studied for comparison purposes.

2. Methods

2.1. Participants

We studied 806 CN participants who had undergone structural MRI, FDG PET, and PiB PET and had available APOE genotype data. They were drawn from 2 sources. The first was the Mayo Clinic Study of Aging (MCSA). The MCSA is a population-based study in Olmsted County MN that originally enrolled persons between the ages of 70 and 89 years. The initial recruitment and description of baseline imaging has been described (Jack et al., 2012; Roberts et al., 2008). After the initial wave of participants were recruited beginning in October 1, 2004, the group has subsequently been replenished with new participants in 2008, 2009, 2010, and 2011. In 2012, the MCSA was expanded to include persons between the ages of 50 and 69 years. These participants were also drawn randomly from Olmsted County, MN, USA. In both the older than 70 and the 50 to 69-year-olds, every participant without contraindications to magnetic resonance (MR) scanning was offered the opportunity to undergo MR and PET imaging. The participants in both these groups were diagnosed as being CN through a consensus process that used information from 3 sources: a mental status examination performed by a study physician, a Clinical Dementia Rating completed by a trained study coordinator that included an interview of an informant as well as the participant and a psychometric battery previously described for the CN group (Petersen et al., 2010; Roberts et al., 2008, 2012) and for the Mayo Alzheimer's Disease Research Center (Kantarci et al., 2008). The second source of participants was a convenience sample of Rochester, MN residents between the ages of 30 and 49 years who were specifically recruited via flyers and word of mouth to undergo brain imaging. This group of younger participants also received the same psychometric battery as the older cognitively normal persons; all scored in the normal range.

For purposes of comparison of the magnitude of the impact of APOE genotype versus being cognitively impaired on FDG ratios, we also included 35 individuals with AD dementia from the MCSA or the Mayo Clinic ADRC who were aged 70 years or older, *APOE* ε4 carriers, and amyloid imaging positive. These AD dementia participants were matched on sex and age ±5 years to 35 mild cognitive impairment (MCI) participants from the MCSA or Mayo ADRC who were *APOE* ε4 carriers and positive on amyloid imaging from the MCSA. Both MCI and AD dementia participants were also age- and sex-matched with *APOE* ε4 carrier CN participants. The MCI and AD dementia individuals were diagnosed via the same consensus process described previously. The diagnosis of MCI (Albert et al., 2011) and dementia because of AD (McKhann et al., 2011) followed standard criteria.

DNA extraction and *APOE* genotyping was performed for each subject using standard methods (Hixson and Vernier, 1990).

2.2. Human subjects protections

All study protocols were approved by the Mayo Clinic and Olmsted Medical Center Institutional Review Boards, and all cognitively normal participants provided signed

informed consent to participate in the study and in the imaging protocols. In participants with cognitive impairment, they and their accompanying family member jointly provided consent.

2.3. Imaging methods

Imaging methods for structural MR, FDG PET, and PiB PET were identical to those that have been described previously (Jack et al., 2008, 2012; Knopman et al., 2012). For FDG-PET scanning, participants were injected with 366–399 MBq of FDG and imaged after 30– 38 minutes, for an 8-minute image acquisition consisting of four 2-minute dynamic frames. During the 30-minute uptake period, participants were left undisturbed in a darkened room and instructed to rest quietly without activity with their eyes open. The scans were performed with eyes open. A CT image was obtained for attenuation correction. Regional cerebral glucose uptake at the static time point 30 minutes after FDG injection was expressed as a ratio of each region's FDG uptake to that of the pons, which we will refer to as the FDG ratio.

Quantitative image analysis for FDG PET was performed using our in-house fully automated image processing pipeline (Jack et al., 2008; Lopresti et al., 2005). Statistics on image voxel values were extracted from automatically labeled cortical regions of interest (ROIs) using an atlas (Tzourio-Mazoyer et al., 2002) modified in-house. There were 19 regions of interest after combining the left and right regions from the atlas. In addition to atlas-defined regions, we also identified a meta-region of interest (ROI) that we have labeled as an "AD signature meta-ROI" that included pre-defined voxels in the angular gyrus, posterior cingulate, and inferior temporal cortical regions from both hemispheres combined (Landau et al., 2011). The pons was used as the reference region for the atlas-defined ROIs as well as for the AD signature region.

Amyloid PET images were acquired using a GE Discovery RX PET/CT scanner. Subjects are injected with 292–729 MBq 11C PiB. The PiB PETscan consists of four 5-minute dynamic frames and was acquired from 40 minutes after injection (McNamee et al., 2009; Price et al., 2005). PiB PET scans were obtained on the same day 1 hour before the FDG PET scan. In the current analyses, amyloid PET results were used only to define the MCI and dementia because of AD comparison groups. A cortical PiB PETstandardized uptake value ratio (SUVr) was formed by combining the prefrontal, orbitofrontal, parietal, temporal, anterior cingulate, and posterior cingulate and/or precuneus ROI values normalized by the cerebellar gray matter ROI of the atlas. We defined an abnormal PiB scan as an SUVr > 1.5 (Jack et al., 2012).

All participants underwent MR scanning at 3T with a standardized protocol that included a 3D-MPRAGE sequence (Jack et al., 2008). MPRAGE images were corrected for image distortion and bias field (Gunter et al., 2009) as previously described (Jack et al., 2008). The MR scans were used for atrophy correction of the FDG images.

On MR scans we quantitated white matter hyperintensity volume and numbers of cortical and subcortical infarctions. The MR protocol included a fluid attenuated inversion recovery scan from which we assessed features of cerebrovascular disease. White matter hyper-

intensity burden was measured quantitatively using an algorithm developed in-house (Jack et al., 2001). Supratentorial subcortical and cortical infarctions greater than 1 cm were ascertained visually by highly experienced image analysts and confirmed by a radiologist (Kejal Kantarci).

2.4. Atrophy correction

Given concerns that brain volume loss was likely to accentuate regional FDG ratio estimates (Ibanez et al., 2004; Kantarci et al., 2010; Yanase et al., 2005), we chose partial volume corrected analyses as our primary outcome measures. Nonpartial volume corrected data are presented in Appendix Table 2. In one variation of the automated ROI parcellation algorithm, partial volume correction was performed on the FDG images, using a 2 compartment model. Each subject's FDG images were co-registered with a 6 degrees of freedom affine registration to his or her MRI scan, which was then simultaneously spatially normalized and segmented into gray matter, white matter, and cerebrospinal fluid compartments, using the unified segmentation method of SPM5. The gray matter and white matter segmentations were saved in the subject's native space and combined to form a brain tissue probability mask, which was then re-sampled to the resolution of the PET image, and smoothed with a 6 mm FWHM Gaussian filter, to approximate the point spread function of the PET camera. Finally, each FDG PET voxel was divided by the corresponding value in this smoothed brain mask, closely following a published method (Meltzer et al., 2000).

2.5. Statistical methods

Linear regression was used to assess the relationship between FDG ratio and age and APOE ε4 genotype. For each region of interest, we fit a model with age, *APOE* ε4 carrier versus noncarrier and the interaction between age and APOE ε4 genotype as predictors of FDG ratio. Restricted cubic splines with 3 knots at ages 45, 65, and 80 years were used to allow for nonlinearity in the relationship between FDG ratio and age. We report *p*-values based on Wald tests. A *p*-value for the association between FDG ratio and age was based on a joint test of all the coefficients involving age (i.e., main effects and interactions) while a *p*-value for the association between FDG and APOE ε4 genotype was based on a joint test of all the coefficients involving *APOE* ε4 genotype.

To estimate the *APOE* ε4 carrier effect on FDG ratios in CN subjects, we matched 209 *APOE* ε4 carriers to noncarriers on sex and within the age of 5 years. *t* Tests were used to test for differences in FDG ratios between the carriers and noncarriers. To estimate the impact of cognitive impairment on FDG ratios, we compared FDG ratios across 3 diagnosis groups that had been matched for sex and age (as previously mentioned): CN ($n = 70$), MCI $(n = 35)$, and AD dementia $(n = 35)$. We used pairwise *t* tests to assess significant differences among the groups. FDG ratios were compared across the 3 diagnosis groups and pairwise *t* tests were used to assess significant differences.

3. Results

3.1. Age and FDG ratio

Table 1 summarizes the demographic features of the participants. In ROI analyses, FDG ratio with partial volume correction declined with advancing age in many regions of the brain including the AD signature meta-ROI, putamen, insula, thalamus, anterior cingulate, supplementary motor, lateral temporal, caudate, orbital frontal, posterior cingulate and/or precuneus, prefrontal, rolandic operculum, lateral parietal, primary visual, precentral gyrus, occipital, and medial temporal regions (Figs. 1 and 2, Appendix Table 1). While FDG was associated with age in many regions, the associations were modest. For an example, the estimated mean FDG ratio in the lateral temporal region for a 40-year-old *APOE* ε4 carrier was 1.62 compared with 1.50 for an 85-year-old *APOE* ε4 carrier, a decrease of only 0.12 SUVR over 45 years.

3.2. Cerebrovascular pathology

Because of the frontal predominance of the aging changes in FDG ratio, we speculated that some of the age differences might be related to the burden of cerebrovascular pathology. In a subset of 471 CN subjects who were over the age of 70 years and had available vascularlesion imaging data, we compared the FDG ratios in the orbito- frontal, prefrontal, and supplementary motor area among subjects by white matter hyperintensity burden quartiles, presence or absence of large cortical infarcts, and presence or absence of subcortical infarcts. None of the vascular features were significantly associated with differences in FDG ratio in these frontal cortical regions (data not shown).

3.3. APOE genotype and FDG ratio

APOE ε4 genotype was associated with lower FDG ratios in the cingulate and/or precuneus and lateral parietal regions and the AD-signature meta-ROI with partial volume correction. In partial volume uncorrected data, only the AD signature region difference remained marginally significant. (Figs. 1 and 2 and Appendix Tables 1 and 2). None of the age \times *APOE* ε4 interactions was significant indicating that the effect of *APOE* ε4 genotype was similar across all age ranges. However, in examining the subgroup of participants who were between the ages of 30 and 60 years ($n = 62$), there were no significant regional differences between *APOE* ε4 carriers and noncarriers. We had too few *APOE* ε4 homozygotes (n = 14) among the CN group to analyze separately.

3.4. Amyloid status, APOE genotype, and FDG ratio

Only 6 of our 167 (3.6%) participants younger than 70 years had PiB SUVr > 1.5 , while 200 of the 639 (31.3%) older participants had PiB SUVr > 1.5 . We examined the impact of high amyloid burden in our cognitive normal individuals older than 70 years. In Fig. 3, we show FDG ratio values from the AD signature meta-ROI for CN participants who had low (SUVr < 1.5) or high (SUVr ≥ 1.5) PiB retention, with or without *APOE* ε4 genotype. While the FDG ratio was lower in those with higher PiB SUVr (*p* = 0.03), the *APOE* ε4 carrier groups showed lower FDG ratios ($p < 0.001$) with no interaction between the 2 ($p = 0.67$).

3.5. Comparison of the impact of APOE ε**4 genotype and cognitive impairment on FDG ratios**

We wished to characterize and contrast the impact of both the carriage of an *APOE* ε4 allele on reductions FDG ratio in CN participants and the magnitude of reductions in FDG ratio observed in persons with cognitive impairment. The mean reduction in FDG ratio in the ADsignature meta-ROI between persons with AD dementia versus matched CN participants was nearly 4 times larger than the *APOE* ε4 allele effect in CN carriers versus noncarriers (Fig. 4 and Table 2). Reductions in FDG ratio in patients with MCI due to AD were similar to the magnitude of the *APOE* ε4 difference in CN carriers versus noncarriers (Kantarci et al., 2010).

4. Discussion

Our findings can be summarized in 4 points. First, we demonstrated statistically significant, though modest, age-related decrements in FDG ratio in a number of frontal and parietal regions even with partial volume correction. Second, carriage of at least one *APOE* ε4 allele was associated with reductions in FDG ratio in the posterior cingulate and/or precuneus and lateral parietal regions, including the AD-signature meta-ROI. The impact of carriage of one *APOE* ε4 allele versus none on FDG ratio in the AD signature region in cognitively normal individuals is about a quarter of the magnitude of that seen in AD dementia compared with age-matched normal. Third, FDG ratios in several regions including the AD signature meta-ROI were lower in *APOE* ε4 carriers across the age spectrum and showed no interaction with age. Fourth, the regional pattern of age-related FDG changes, *APOE* ε4-related FDG changes, and the changes occurring in AD dementia patients with elevated PiB retention have 2 loci in common: the posterior cingulate and/or precuneus and lateral parietal, even while the age-related changes were seen much more broadly throughout the brain. Thus, the pivotal role of these regions in the pathogenesis of AD may relate to their unique susceptibility to both aging and *APOE* ε4-related effects.

Decrements in glucose uptake per unit brain volume occurred as a function of age. Our sample size of CN individuals was much larger than prior studies (Ibanez et al., 2004; Kalpouzos et al., 2009; Kantarci et al., 2010; Yanase et al., 2005), which enabled us to detect declines with aging in the presence of considerable between-subject variability (Fig. 1). Declines in FDG ratio with age do not appear to be mediated by elevated brain amyloid levels, as the FDG declines appear to begin before the age of 60 years, well before there is appreciable accumulation of brain β-amyloid (Rodrigue et al., 2012; Rowe et al., 2010). The anatomic distribution of the age-related FDG declines was very broad but did not involve every region. The age-related pattern of regional declines in FDG ratio were widespread and therefore not simply a replica of that seen in AD. Further, the medial temporal lobe showed small age-related declines in FDG ratio. However, in contrast to a prior report (Kalpouzos et al., 2009), we found declines in the posterior cingulate and/or precuneus FDG ratios that were comparable to declines seen in the frontal regions. Cognitively normal individuals who have reduced FDG ratios in the AD signature regions, along with elevations of brain amyloid are at increased risk for cognitive decline (Knopman et al., 2012).

There were notably large declines in FDG ratios in the putamen and thalamus with age, the relevance of which to cognitive aging or disease is not clear. In familial AD, volume loss occurs in the thalamus, suggesting that the region is involved in AD-related neurodegeneration (Cash et al., 2013). To the best of our knowledge, changes in glucose metabolism in subcortical structures have not been described before.

We found reductions in FDG ratio with the possession of at least one *APOE* ε4 allele in the posterior cingulate and/or precuneus, lateral parietal, and an AD-signature meta-ROI that were not age-dependent (Fig. 2: there was a main effect of $APOE \varepsilon 4$ but no age nor age \times *APOE* ε4 interaction). Although we could not demonstrate reduced FDG ratios in our younger participants separately, the absence of age $\times APOE$ at interaction suggests that the FDG changes, in fact, begin at a young age. One group, working with participants (aged 50– 63 years) with a family history of AD dementia found the *APOE* ε4 heterozygotes exhibited declines over a 2 year interval in a number of cortical regions including ones in our AD signature meta-ROI (Reiman et al., 2001).

There was also no interaction between *APOE* ε4 carriage and PiB SUVr in our older participants, suggesting that the impact of *APOE* ε4 on FDG ratio was not simply mediated by β-amyloid burden. Rather, it appeared that amyloidosis and *APOE* ε4 carriage were additive in their impact on FDG hypometabolism. The separation in the mean FDG ratio values beginning in our youngest participants between ε4 carriers and noncarriers (as demonstrated by the lack of an age $\times APOE \in \mathcal{A}$ interaction) also suggests that brain amyloidosis was not mediating the impact of *APOE* ε4 carriage on FDG ratios. Some reports have found relative FDG hypermetabolism in association with amyloidosis in elderly normal individuals (Oh et al., 2014) or persons with MCI (Cohen et al., 2009). We did not observe absolute FDG hyper-metabolism in our elderly CN participants in relationship to amyloid status (Fig. 3). Such a phenomenon highlights the independence of the impact of *APOE* ε4 from amyloid burden on FDG.

Our findings are consistent with the assertion (Jagust et al., 2012) that the impact of *APOE* ε4 on glucose hypometabolism in the AD meta-ROI is not entirely because of amyloid deposition. Hypometabolism in the posterior cingulate and/or precuneus or lateral parietal regions in some asymptomatic persons may be expression of the role of *APOE* ε4 in neuronal function that is independent of its well-established relationship to amyloid deposition (Kok et al., 2009; Morris et al., 2010). Eventually, β-amyloidosis promotes worsening metabolic function in critical cortical regions (Knopman et al., 2013; Landau et al., 2012), but in young people decades before brain amyloid levels become elevated (Rodrigue et al., 2012), *APOE* ε4 genotype has a unique impact on cortical metabolism.

The large FDG hypometabolic changes in temporal, parietal, and posterior cingulate cortices in persons with AD dementia are well established (Chetelat et al., 2008; Mosconi, 2005; Nestor et al., 2003; Rabinovici et al., 2010). The magnitude of the FDG ratio declines in the AD-signature meta-ROI attributable to *APOE* ε4 carriage was about a quarter of that attributable to having AD dementia. Thus, while the regional predilection for the lateral parietal, posterior cingulate and/or precuneus, and AD-signature meta-ROI conferred by *APOE* ε4 carriage is strikingly similar to that of AD dementia, the magnitude is much

smaller. The most parsimonious explanation is that excess decrement in metabolic efficacy in the AD signature regions caused by APOE ε4 carriage makes it that much easier for amyloid-related brain pathology to worsen cortical metabolism to a critical level and therefore confers risk for future cognitive decline.

Brain volume loss with advancing age in cognitively normal people is a consistently observed phenomenon (Bakkour et al., 2013; Fjell et al., 2013; Salat et al., 2004; Sowell et al., 2003). Structural imaging studies also show that some regions experience volume loss with aging, whereas the medial temporal lobe declines appear to be more specific for AD. Frontal and parietal cortex, in contrast, are susceptible to both aging and AD changes (Bakkour et al., 2013). Although brain atrophy and FDG ratio changes over the age spectrum are similar but not identical, it was beyond the scope of the current work to correlate structural imaging changes that occur with age to the metabolic changes. However, we were unable to ignore the confounding effect of brain volume loss in the current analyses, because the calculation of voxel-wise glucose metabolic rates are critically dependent upon excluding cerebro-spinal fluid in measured voxels.

The confluence of aging and genetic influences on the integrity of the precuneus and/or posterior cingulate cortex and its invariant involvement in AD (Minoshima et al., 1997; Nestor et al., 2003) is probably not a coincidence. The posterior cingulate has the highest rate of glucose metabolism at rest of any cortical region (Ivancevic et al., 2000), perhaps because of its role as one of the major hubs of the default mode network (Buckner et al., 2008). One of the principles of posterior cingulate physiology is that it tends to be tonically active (Raichle et al., 2001) but deactivates when mental effort is required. *APOE* ε4 carriers may be less efficient in deactivation during cognitive activity (Filippini et al., 2009). In CN elders, deactivation with mental effort also failed to occur (Lustig et al., 2003). A hypothesis is that a failure to deactivate in AD signature regions puts additional metabolic strain on the region. An alternative hypothesis is that the metabolic inefficiency because of carriage of APOE ε4 contributed to failure to deactivate. Moreover, the posterior cingulate and lateral temporal regions are also sites of high amyloid load (Jack et al., 2008; Rowe et al., 2007; Wolk et al., 2009). Finally, the posterior cingulate and/or precuneus (and the lateral parietal and inferior temporal regions) are well positioned to initiate or facilitate trans-synaptic propagation of amyloid- or tau-related neurodegeneration (de Calignon et al., 2012; Liu et al., 2012) by virtue of their powerful interconnections with many other cortical regions.

In our prior work (Jack et al., 2012; Knopman et al., 2012) as well as in that of others (Landau et al., 2011), the cut-point for abnormal FDG PET was defined in relation to a demented group. There are advantages and disadvantages of that approach, the main disadvantage being that such a cut-point might be insensitive to pre-symptomatic alterations in brain glucose metabolism. The fact that a substantial fraction of AD dementia patients do not have abnormal FDG PET ratio (Lowe et al., 2013) raises another concern about defining a cut-point relative to dementia status. The discrepancy between a dementia-derived cutpoint and a young normal cut-point has been shown for amyloid imaging (Mormino et al., 2012), for example. In young normal individuals, a cut-point for abnormal amyloid levels was far below the cut-point derived relative to AD dementia. Thus, one of the motivations for this analysis was to derive a cut-point for FDG ratio in some region or regions that was

based on deviation from healthy youthful brains. However, the great variability in FDG ratio combined with the modest age-related changes make it unlikely that an abnormal FDG ratio can be defined with reference to normal aging. Instead, abnormal FDG ratio in the AD signature meta-ROI might need to be defined by the levels seen in persons with amyloidpositive AD dementia.

Acknowledgments

This work was supported by NIH grants P50 AG16574, U01 AG06786 and R01 AG11378, the Elsie and Marvin Dekelboum Family Foundation and the Robert H. and Clarice Smith and Abigail Van Buren Alzheimer's Disease Research Program of the Mayo Foundation. Dr Knopman took part in data collection, supervised analyses, generated the first and final drafts and takes overall responsibility for the data and the manuscript. Dr Jack took part in data collection, supervised analyses, and critically reviewed the manuscript. Ms. Wiste performed analyses and critically reviewed the manuscript. Ms. Lundt performed analyses and critically reviewed the manuscript. Mr. Weigand performed analyses and critically reviewed the manuscript. Dr Vemuri performed analyses of imaging data and critically reviewed the manuscript. Dr Lowe supervised collection of PET imaging, and critically reviewed the manuscript. Dr Kantarci performed analyses of imaging data and critically reviewed the manuscript. Dr Gunter performed analyses of imaging data. Mr. Senjem performed analyses of imaging data. Dr Roberts critically reviewed the manuscript. Dr Mielke critically reviewed the manuscript. Dr Boeve took part in data collection and critically reviewed the manuscript. Dr Petersen obtained funding, took part in data collection and critically reviewed the manuscript.

References

- Albert M, DeKosky ST, Dickson D, Dubois B, Feldman H, Fox NC, Gamst A, Holtzman D, Jagust WJ, Petersen RC, Snyder PJ, Phelps CH. The Diagnosis of Mild cognitive impairment due to Alzheimer's disease: Report of the National Institute on Aging and the Alzheimer's Association Workgroup. Alzheimer's Demen J Alzheimer's Assoc. 2011; 7:270–279.
- Bakkour A, Morris JC, Wolk DA, Dickerson BC. The effects of aging and Alzheimer's disease on cerebral cortical anatomy: specificity and differential relationships with cognition. Neuroimage. 2013; 76:332–344. [PubMed: 23507382]
- Buckner RL, Andrews-Hanna JR, Schacter DL. The brain's default network: anatomy, function, and relevance to disease. Ann NY Acad Sci. 2008; 1124:1–38. [PubMed: 18400922]
- Cash DM, Ridgway GR, Liang Y, Ryan NS, Kinnunen KM, Yeatman T, Malone IB, Benzinger TL, Jack CR Jr, Thompson PM, Ghetti BF, Saykin AJ, Masters CL, Ringman JM, Salloway SP, Schofield PR, Sperling RA, Cairns NJ, Marcus DS, Xiong C, Bateman RJ, Morris JC, Rossor MN, Ourselin S, Fox NC. The pattern of atrophy in familial Alzheimer disease: volumetric MRI results from the DIAN study. Neurology. 2013; 81:1425–1433. [PubMed: 24049139]
- Chetelat G, Desgranges B, Landeau B, Mezenge F, Poline JB, de la Sayette V, Viader F, Eustache F, Baron JC. Direct voxel-based comparison between grey matter hypometabolism and atrophy in Alzheimer's disease. Brain. 2008; 131:60–71. [PubMed: 18063588]
- Cohen AD, Price JC, Weissfeld LA, James J, Rosario BL, Bi W, Nebes RD, Saxton JA, Snitz BE, Aizenstein HA, Wolk DA, Dekosky ST, Mathis CA, Klunk WE. Basal cerebral metabolism may modulate the cognitive effects of Abeta in mild cognitive impairment: an example of brain reserve. J Neurosci. 2009; 29:14770–14778. [PubMed: 19940172]
- de Calignon A, Polydoro M, Suarez-Calvet M, William CM, Adamowicz DH, Kopeikina KJ, Pitstick R, Sahara N, Ashe KH, Carlson GA, Spires-Jones TL, Hyman BT. Propagation of tau pathology in a model of early Alzheimer's disease. Neuron. 2012; 73:685–697. [PubMed: 22365544]
- Filippini N, MacIntosh BJ, Hough MG, Goodwin GM, Frisoni GB, Smith SM, Matthews PM, Beckmann CF, Mackay CE. Distinct patterns of brain activity in young carriers of the APOEepsilon4 allele. Proc Natl Acad Sci USA. 2009; 106:7209–7214. [PubMed: 19357304]
- Fjell AM, McEvoy L, Holland D, Dale AM, Walhovd KB. Brain changes in older adults at very low risk for Alzheimer's disease. J Neurosci. 2013; 33:8237–8242. [PubMed: 23658162]

- Gunter JL, Bernstein MA, Borowski BJ, Ward CP, Britson PJ, Felmlee JP, Schuff N, Weiner M, Jack CR. Measurement of MRI scanner performance with the ADNI phantom. Med Phys. 2009; 36:2193–2205. [PubMed: 19610308]
- Herholz K, Salmon E, Perani D, Baron JC, Holthoff V, Frolich L, Schonknecht P, Ito K, Mielke R, Kalbe E, Zundorf G, Delbeuck X, Pelati O, Anchisi D, Fazio F, Kerrouche N, Desgranges B, Eustache F, Beuthien-Baumann B, Menzel C, Schroder J, Kato T, Arahata Y, Henze M, Heiss WD. Discrimination between Alzheimer dementia and controls by automated analysis of multicenter FDG PET. Neuroimage. 2002; 17:302–316. [PubMed: 12482085]
- Hixson JE, Vernier DT. Restriction isotyping of human apolipoprotein E by gene amplification and cleavage with HhaI. J Lipid Res. 1990; 31:545–548. [PubMed: 2341813]
- Ibanez V, Pietrini P, Furey ML, Alexander GE, Millet P, Bokde AL, Teichberg D, Schapiro MB, Horwitz B, Rapoport SI. Resting state brain glucose metabolism is not reduced in normotensive healthy men during aging, after correction for brain atrophy. Brain Res Bull. 2004; 63:147–154. [PubMed: 15130704]
- Ivancevic V, Alavi A, Souder E, Mozley PD, Gur RE, Benard F, Munz DL. Regional cerebral glucose metabolism in healthy volunteers determined by fluordeoxyglucose positron emission tomography: appearance and variance in the transaxial, coronal, and sagittal planes. Clin Nucl Med. 2000; 25:596–602. [PubMed: 10944013]
- Jack CR Jr, Knopman DS, Jagust WJ, Petersen RC, Weiner MW, Aisen PS, Shaw LM, Vemuri P, Wiste HJ, Weigand SD, Lesnick TG, Pankratz VS, Donohue MC, Trojanowski JQ. Tracking pathophysiological processes in Alzheimer's disease: an updated hypothetical model of dynamic biomarkers. Lancet Neurol. 2013; 12:207–216. [PubMed: 23332364]
- Jack CR Jr, Knopman DS, Weigand SD, Wiste HJ, Vemuri P, Lowe V, Kantarci K, Gunter JL, Senjem ML, Ivnik RJ, Roberts RO, Rocca WA, Boeve BF, Petersen RC. An operational approach to National Institute on Aging-Alzheimer's Association criteria for preclinical Alzheimer disease. Ann Neurol. 2012; 71:765–775. [PubMed: 22488240]
- Jack CR Jr, Lowe VJ, Senjem ML, Weigand SD, Kemp BJ, Shiung MM, Knopman DS, Boeve BF, Klunk WE, Mathis CA, Petersen RC. 11C PiB and structural MRI provide complementary information in imaging of Alzheimer's disease and amnestic mild cognitive impairment. Brain. 2008; 131:665–680. [PubMed: 18263627]
- Jack CR Jr, O'Brien PC, Rettman DW, Shiung MM, Xu Y, Muthupillai R, Manduca A, Avula R, Erickson BJ. FLAIR histogram segmentation for measurement of leukoaraiosis volume. J Magn Reson Imaging. 2001; 14:668–676. [PubMed: 11747022]
- Jagust WJ, Landau SM. Apolipoprotein E, not Fibrillar beta-amyloid, Reduces cerebral glucose metabolism in Normal aging. J Neurosci. 2012; 32:18227–18233. [PubMed: 23238736]
- Kalpouzos G, Chetelat G, Baron JC, Landeau B, Mevel K, Godeau C, Barre L, Constans JM, Viader F, Eustache F, Desgranges B. Voxel-based mapping of brain gray matter volume and glucose metabolism profiles in normal aging. Neurobiol Aging. 2009; 30:112–124. [PubMed: 17630048]
- Kantarci K, Petersen RC, Przybelski SA, Weigand SD, Shiung MM, Whitwell JL, Negash S, Ivnik RJ, Boeve BF, Knopman DS, Smith GE, Jack CR Jr. Hippocampal volumes, proton magnetic resonance spectroscopy metabolites, and cerebrovascular disease in mild cognitive impairment subtypes. Arch Neurol. 2008; 65:1621–1628. [PubMed: 19064749]
- Kantarci K, Senjem ML, Lowe VJ, Wiste HJ, Weigand SD, Kemp BJ, Frank AR, Shiung MM, Boeve BF, Knopman DS, Petersen RC, Jack CR Jr. Effects of age on the glucose metabolic changes in mild cognitive impairment. AJNR Am J Neuroradiol. 2010; 31:1247–1253. [PubMed: 20299441]
- Knopman DS, Jack CR Jr, Wiste HJ, Weigand SD, Vemuri P, Lowe V, Kantarci K, Gunter JL, Senjem M, Ivnik RJ, Roberts RO, Boeve BF, Petersen RC. Short-term Clinical Outcomes for Stages of NIA-AA pre-clinical Alzheimer disease. Neurology. 2012; 78:1576–1582. [PubMed: 22551733]
- Knopman DS, Jack CR Jr, Wiste HJ, Weigand SD, Vemuri P, Lowe VJ, Kantarci K, Gunter JL, Senjem ML, Mielke MM, Roberts RO, Boeve BF, Petersen RC. Selective Worsening of brain Injury Biomarker Abnormalities in cognitively Normal elderly with β-amyloidosis. JAMA Neurol. 2013; 70:1030–1038. [PubMed: 23797806]
- Kok E, Haikonen S, Luoto T, Huhtala H, Goebeler S, Haapasalo H, Karhunen PJ. Apolipoprotein Edependent accumulation of Alzheimer disease-related lesions begins in middle age. Ann Neurol. 2009; 65:650–657. [PubMed: 19557866]

- Landau SM, Harvey D, Madison CM, Koeppe RA, Reiman EM, Foster NL, Weiner MW, Jagust WJ. Associations between cognitive, functional, and FDG-PET measures of decline in AD and MCI. Neurobiol Aging. 2011; 32:1207–1218. [PubMed: 19660834]
- Landau SM, Mintun MA, Joshi AD, Koeppe RA, Petersen RC, Aisen PS, Weiner MW, Jagust WJ. Amyloid deposition, hypometabolism, and longitudinal cognitive decline. Ann Neurol. 2012; 72:578–586. [PubMed: 23109153]
- Liu L, Drouet V, Wu JW, Witter MP, Small SA, Clelland C, Duff K. Trans-synaptic spread of tau pathology in vivo. PLoS One. 2012; 7:e31302. Epub 2012 Feb 1. [PubMed: 22312444]
- Loessner A, Alavi A, Lewandrowski KU, Mozley D, Souder E, Gur RE. Regional cerebral function determined by FDG-PET in healthy volunteers: normal patterns and changes with age. J Nucl Med. 1995; 36:1141–1149. [PubMed: 7790936]
- Lopresti BJ, Klunk WE, Mathis CA, Hoge JA, Ziolko SK, Lu X, Meltzer CC, Schimmel K, Tsopelas ND, DeKosky ST, Price JC. Simplified quantification of Pittsburgh Compound B amyloid imaging PET studies: a comparative analysis. J Nucl Med. 2005; 46:1959–1972. [PubMed: 16330558]
- Lowe VJ, Peller PJ, Weigand SD, Montoya Quintero C, Tosakulwong N, Vemuri P, Senjem ML, Jordan L, Jack CR Jr, Knopman D, Petersen RC. Application of the National Institute on Aging-Alzheimer's Association AD criteria to ADNI. Neurology. 2013; 80:2130–2137. [PubMed: 23645596]
- Lustig C, Snyder AZ, Bhakta M, O'Brien KC, McAvoy M, Raichle ME, Morris JC, Buckner RL. Functional deactivations: change with age and dementia of the Alzheimer type. Proc Natl Acad Sci USA. 2003; 100:14504–14509. [PubMed: 14608034]
- McKhann GM, Knopman DS, Chertkow H, Hyman BT, Jack CRJ, Kawas CH, Klunk WE. The diagnosis of dementia due to Alzheimer's disease: Recommendations from the National Institute on Aging and the Alzheimer's Association workgroup. Alzheimer's Demen J Alzheimer's Assoc. 2011; 7:263–269.
- McNamee RL, Yee SH, Price JC, Klunk WE, Rosario B, Weissfeld L, Ziolko S, Berginc M, Lopresti B, Dekosky S, Mathis CA. Consideration of optimal time window for Pittsburgh compound B PET summed uptake measurements. J Nucl Med. 2009; 50:348–355. [PubMed: 19223409]
- Meltzer CC, Cantwell MN, Greer PJ, Ben-Eliezer D, Smith G, Frank G, Kaye WH, Houck PR, Price JC. Does cerebral blood flow decline in healthy aging? A PET study with partial-volume correction. J Nucl Med. 2000; 41:1842–1848. [PubMed: 11079492]
- Minoshima S, Giordani B, Berent S, Frey KA, Foster NL, Kuhl DE. Metabolic reduction in the posterior cingulate cortex in very early Alzheimer's disease. Ann Neurol. 1997; 42:85–94. [PubMed: 9225689]
- Mormino EC, Brandel MG, Madison CM, Rabinovici GD, Marks S, Baker SL, Jagust WJ. Not quite PIB-positive, not quite PIB-negative: Slight PIB elevations in elderly normal control subjects are biologically relevant. Neuroimage. 2012; 59:1152–1160. [PubMed: 21884802]
- Morris JC, Roe CM, Xiong C, Fagan AM, Goate AM, Holtzman DM, Mintun MA. APOE predicts amyloid-beta but not tau Alzheimer pathology in cognitively normal aging. Ann Neurol. 2010; 67:122–131. [PubMed: 20186853]
- Mosconi L. Brain glucose metabolism in the early and specific diagnosis of Alzheimer's disease. FDG-PET studies in MCI and AD. Eur J Nucl Med Mol Imaging. 2005; 32:486–510. [PubMed: 15747152]
- Nestor PJ, Fryer TD, Smielewski P, Hodges JR. Limbic hypometabolism in Alzheimer's disease and mild cognitive impairment. Ann Neurol. 2003; 54:343–351. [PubMed: 12953266]
- Oh H, Habeck C, Madison C, Jagust W. Covarying alterations in Abeta deposition, glucose metabolism, and gray matter volume in cognitively normal elderly. Hum Brain Mapp. 2014; 35:297–308. [PubMed: 22965806]
- Petersen RC, Roberts RO, Knopman DS, Geda YE, Cha RC, Pankratz VS, Boeve BF, Tangalos EG, Ivnik RJ, Rocca WA. Prevalence of mild cognitive impairment is higher in men than in women. The Mayo Clinic Study of Aging. Neurology. 2010; 75:889–897. [PubMed: 20820000]
- Price JC, Klunk WE, Lopresti BJ, Lu X, Hoge JA, Ziolko SK, Holt DP, Meltzer CC, DeKosky ST, Mathis CA. Kinetic modeling of amyloid binding in humans using PET imaging and Pittsburgh Compound-B. J Cereb Blood Flow Metab. 2005; 25:1528–1547. [PubMed: 15944649]

- Rabinovici GD, Furst AJ, Alkalay A, Racine CA, O'Neil JP, Janabi M, Baker SL, Agarwal N, Bonasera SJ, Mormino EC, Weiner MW, Gorno-Tempini ML, Rosen HJ, Miller BL, Jagust WJ. Increased metabolic vulnerability in early-onset Alzheimer's disease is not related to amyloid burden. Brain. 2010; 133:512–528. [PubMed: 20080878]
- Raichle ME, MacLeod AM, Snyder AZ, Powers WJ, Gusnard DA, Shulman GL. A default mode of brain function. Proc Natl Acad Sci USA. 2001; 98:676–682. [PubMed: 11209064]
- Reiman EM, Caselli RJ, Chen K, Alexander GE, Bandy D, Frost J. Declining brain activity in cognitively normal apolipoprotein E epsilon 4 heterozygotes: a foundation for using positron emission tomography to efficiently test treatments to prevent Alzheimer's disease. Proc Natl Acad Sci USA. 2001; 98:3334–3339. [PubMed: 11248079]
- Reiman EM, Caselli RJ, Yun LS, Chen K, Bandy D, Minoshima S, Thibodeau SN, Osborne D. Preclinical evidence of Alzheimer's disease in persons homozygous for the epsilon 4 allele for apolipoprotein E. N Engl J Med. 1996; 334:752–758. [PubMed: 8592548]
- Roberts RO, Geda YE, Knopman D, Cha R, Pankratz VS, Boeve B, Ivnik R, Tangalos E, Petersen RC, Rocca WA. The Mayo Clinic study of aging: Design and Sampling, Participation, Baseline measures and Sample Characteristics. Neuroepidemiology. 2008; 30:58–69. [PubMed: 18259084]
- Roberts RO, Geda YE, Knopman DS, Cha RH, Pankratz VS, Boeve BF, Tangalos EG, Ivnik RJ, Rocca WA, Petersen RC. The incidence of MCI differs by subtype and is higher in men: the Mayo Clinic Study of Aging. Neurology. 2012; 78:342–351. [PubMed: 22282647]
- Rodrigue KM, Kennedy KM, Devous MD Sr, Rieck JR, Hebrank AC, Diaz-Arrastia R, Mathews D, Park DC. beta-Amyloid burden in healthy aging: regional distribution and cognitive consequences. Neurology. 2012; 78:387–395. [PubMed: 22302550]
- Rowe CC, Ellis KA, Rimajova M, Bourgeat P, Pike KE, Jones G, Fripp J, Tochon-Danguy H, Morandeau L, O'Keefe G, Price R, Raniga P, Robins P, Acosta O, Lenzo N, Szoeke C, Salvado O, Head R, Martins R, Masters CL, Ames D, Villemagne VL. Amyloid imaging results from the Australian Imaging, Biomarkers and Lifestyle (AIBL) study of aging. Neurobiol Aging. 2010; 31:1275–1283. [PubMed: 20472326]
- Rowe CC, Ng S, Ackermann U, Gong SJ, Pike K, Savage G, Cowie TF, Dickinson KL, Maruff P, Darby D, Smith C, Woodward M, Merory J, Tochon-Danguy H, O'Keefe G, Klunk WE, Mathis CA, Price JC, Masters CL, Villemagne VL. Imaging beta-amyloid burden in aging and dementia. Neurology. 2007; 68:1718–1725. [PubMed: 17502554]
- Salat DH, Buckner RL, Snyder AZ, Greve DN, Desikan RS, Busa E, Morris JC, Dale AM, Fischl B. Thinning of the cerebral cortex in aging. Cereb Cortex. 2004; 14:721–730. [PubMed: 15054051]
- Sowell ER, Peterson BS, Thompson PM, Welcome SE, Henkenius AL, Toga AW. Mapping cortical change across the human life span. Nat Neurosci. 2003; 6:309–315. [PubMed: 12548289]
- Tzourio-Mazoyer N, Landeau B, Papathanassiou D, Crivello F, Etard O, Delcroix N, Mazoyer B, Joliot M. Automated anatomical labeling of activations in SPM using a macroscopic anatomical parcellation of the MNI MRI single-subject brain. Neuroimage. 2002; 15:273–289. [PubMed: 11771995]
- Wolk DA, Price JC, Saxton JA, Snitz BE, James JA, Lopez OL, Aizenstein HJ, Cohen AD, Weissfeld LA, Mathis CA, Klunk WE, Dekoskym ST. Amyloid imaging in mild cognitive impairment subtypes. Ann Neurol. 2009; 65:557–568. [PubMed: 19475670]
- Yanase D, Matsunari I, Yajima K, Chen W, Fujikawa A, Nishimura S, Matsuda H, Yamada M. Brain FDG PET study of normal aging in Japanese: effect of atrophy correction. Eur J Nucl Med Mol Imaging. 2005; 32:794–805. [PubMed: 15759148]

Fig. 1.

Scatterplots of FDG ratio with partial volume correction (PVCY) as a function of age in representative cortical regions. Estimated mean FDG ratio for each age is shown with a solid line for APOE ε4 carriers and a dashed line for APOE ε4 noncarriers. Estimates are from a linear regression model with age, APOE ε4 genotype, and their interaction. Age was fit with restricted cubic splines with 3 knots at ages 45, 65, and 80 years to allow for nonlinearity in the FDG-age relationship. The 7 regions depicted are medial temporal, lateral temporal, lateral parietal, prefrontal, primary visual cortex, posterior cingulate and/or precuneus, and the Alzheimer signature meta-ROI. Abbreviations: APOE, apolipoprotein E; FDG, 18Ffluorodeoxyglucose; ROI, region of interest.

Fig. 2.

Graphic illustration (left) and tabular listing of *p*-values (right) of regional FDG ratios for an average 40-year-old APOE ε4 carrier and noncarrier, and 85-year-old APOE ε4 carrier and noncarrier, where the FDG ratio values were derived from regression analyses. On the right are the *p*-values for the main effects of age and APOE ε4 carriage, and the *p*-value for the interaction term. The actual ratio values are given in the Appendix Table 1. "+" signs indicate APOE ε4 carriers and "o" signs indicate APOE ε4 noncarriers. Blue symbols indicate the 85-year-olds and gold symbols the 40-year=olds. Abbreviations: APOE, apolipoprotein E; FDG, ¹⁸F-fluorodeoxyglucose.

Fig. 3.

Boxplots of the FDG ratio (partial volume corrected [PVCY]) in the AD-signature meta-ROI among 639 elderly (age $\,$ 70) CN participants, stratified by low (SUVr < 1.5) or high (SUVr ≥ 1.5) PiB retention and *APOE* ε4 carriers and noncarriers. The only pairwise comparisons that reached a nominal level of significance (uncorrected for multiple comparisons) were low PiB/*APOE* ε4 noncarrier versus high PiB/*APOE* ε4 carrier (*p* < 0.001) and high PiB/*APOE* ε4 noncarrier versus high PiB/*APOE* ε4 carrier (*p* = 0.03). There were 365 low PiB/*APOE* ε4 noncarriers, 74 low PiB/*APOE* ε4 carriers, 115 high PiB/*APOE* ε4 noncarriers, and 85 high PiB/*APOE* ε4 carriers. Abbreviations: AD, Alzheimer's disease; APOE, apolipoprotein E; CN, cognitively normal; FDG, 18 F-fluorodeoxyglucose; PiB, 11 C-Pittsburgh compound B; ROI, region of interest; SUVr, standardized uptake value ratio.

Fig. 4.

Boxplots of the FDG ratio (partial volume corrected [PVCY]) in the AD-signature meta-ROI among elderly (age 70 years) CN, MCI, and AD subjects and among CN APOE ε4 carriers and noncarriers of all ages to demonstrate the relative AD dementia and APOE ε4 effects. AD dementia subjects who were APOE ε4 carriers and amyloid positive were matched to APOE ε4 carrier, amyloid positive MCI subjects, and APOE ε4 carrier CN subjects on age and sex. CN APOE ε4 carriers were matched on age and sex to APOE ε4 non-carriers. Abbreviations: AD, Alzheimer's disease; APOE, apolipoprotein E; CN, cognitively normal; FDG, 18F-fluorodeoxyglucose; MCI, mild cognitive impairment; ROI, region of interest.

Characteristics of participants

Table 1

Characteristic Young normal (N =35) MCSA 50 to 70-year-old normal (N = 143) MCSA >70-year-old normal (N = 628) All cognitively normal (N = 806) Age, y Median (IQR) 39 (34, 44) 64 (61, 68) 78 (75, 83) 76 (72, 82) Min, max 30, 50 51, 71 71, 95 30, 95 Male gender, no. (%) 18 (51) 78 (55) 342 (54) 342 (54) 438 (54) APOE ε4 positive, no. (%) 14 (40) 37 (26) 158 (25) 158 (25) 209 (26) Education, y Median (IQR) 16 (14, 17) 16 (14, 18) 14 (12, 16) 14 (12, 16) Min, max 12, 20 12, 20 8, 20 8, 20 8, 20

Key: APOE, apolipoprotein E; Max, maximum; MCSA, Mayo Clinic Study of Aging; Min, minimum.

Author Manuscript

Author Manuscript

 a AD dementia, MCI, and CN participants are APOE e4 carriers and age 70 or older. AD and MCI participants are also amyloid positive by PiB PET (SUVr > 1.5). *a*AD dementia, MCI, and CN participants are APOE ε4 carriers and age 70 or older. AD and MCI participants are also amyloid positive by PiB PET (SUVr > 1.5).

 $b_{\mbox{Mean}}$ differences in FDG ratios in AD signature meta-ROI with $p\mbox{-values}$ from t-tests. *b*Mean differences in FDG ratios in AD signature meta-ROI with *p*-values from t-tests.

Appendix Table 1 Appendix Table 1

Estimated mean FDG ratio and tests for significant differences in FDG by age and APOE ε4 genotype from linear regression models, partial volume Estimated mean FDG ratio and tests for significant differences in FDG by age and APOE $\varepsilon4$ genotype from linear regression models, partial volume corrected

Neurobiol Aging. Author manuscript; available in PMC 2015 September 01.

*a*Estimated FDG ratios for hypothetical participants based on regression model.

 4 Estimated FDG ratios for hypothetical participants based on regression model.

Author Manuscript

Author Manuscript

Appendix Table 2 **Appendix Table 2**

Estimated mean FDG ratio and tests for significant differences in FDG by age and APOE ε4 genotype from linear regression models, not partial volume Estimated mean FDG ratio and tests for significant differences in FDG by age and APOE $\varepsilon4$ genotype from linear regression models, not partial volume corrected

Neurobiol Aging. Author manuscript; available in PMC 2015 September 01.

*a*Estimated FDG ratios for hypothetical participants based on regression model.

 a Estimated FDG ratios for hypothetical participants based on regression model.