# **Europe PMC Funders Group Author Manuscript Arterioscler Thromb Vasc Biol. Author manuscript; available in PMC 2014 June 17.**

Published in final edited form as: *Arterioscler Thromb Vasc Biol*. 2008 October ; 28(10): 1691–1693. doi:10.1161/ATVBAHA.108.173963.

# **Endothelial H2O2 A Bad Guy Turning Good?**

## **Wolfgang F. Graier** and

Institute of Molecular Biology and Biochemistry, Center of Molecular Medicine, Medical University Graz, Austria

#### **Markus Hecker**

Institute of Physiology and Pathophysiology, Division of Cardiovascular Physiology, University of Heidelberg, Germany

## **Abstract**

About a decade ago everyone who read, at least occasionally, the science part of the local newspapers or followed popular science broadcasts "knew" that reactive oxygen species (ROS) are dangerous. They were thought to be bad, really bad, and many looking for a healthier lifestyle took action against these nasty small molecules accused of harming lipids and proteins by taking antioxidants like vitamin E. Food industry teamed up with pharmaceutical companies and brought up functional food enriched or supplemented with trustful antioxidants in the fight against ROS, which seemed to be produced under pathological conditions with just one mission: to damage the body. Things have changed during recent years, and there is emerging evidence that ROS are not only continuously produced even in healthy individuals but also exert important physiological functions. While we are still in-between a period that provides exciting new findings on the physiology of ROS, one can already summarize our present knowledge rather accurately by citing Paracelsus' adage of "*Dosis (sola) facit venenum*" (*tertio defensio*, 1538).

> The present work of Edwards et al<sup>1</sup> is one of these exciting new reports that open our eyes to recognize the good side of the previously dammed ROS. This article deals with the effect of hydrogen peroxide  $(H_2O_2)$  on endothelium-derived hyperpolarization and is quite remarkable in several ways:

> First, the article is focused on EDHF (formerly termed endothelium-derived hyperpolarizing factor), a nitric oxide (NO) and prostacyclin-independent phenomenon of endotheliumdependent relaxation, of which the molecular mechanism and nature are still somewhat mysterious. Thus, this endothelium-dependent smooth muscle hyperpolarization has been attributed to either the formation and release of diffusible factors from the endothelium<sup>2</sup> such as potassium ions  $(K^+),^3$  cytochrome P450-derived arachidonic acid metabolites (epoxyeicosatrienoic acids),<sup>4,5</sup> anandamide,<sup>6</sup> or even  $H_2O_2^7$  but also to a direct electric

<sup>©2008</sup> American Heart Association, Inc.

Correspondence to Wolfgang F. Graier, PhD, Molecular and Cellular Physiology Research Unit, Institute of Molecular Biology and Biochemistry, Center of Molecular Medicine, Medical University Graz, Harrachgasse 21/III, A-8010 Graz, Austria. wolfgang.graier@meduni-graz.at.

**Disclosures** None.

coupling between endothelial and smooth muscle cells.<sup>8</sup> The latter work was largely guided and conducted by Dr Griffith and his team, who convincingly showed that gap junctions between endothelial and smooth muscle cells are a prerequisite to spread EDHF-type relaxations in many vascular beds.<sup>9</sup> In this context, it is interesting that despite the ongoing debate on the nature of EDHF—most probably there is more than one such "factor"—the authors used EDHF-type relaxation of isolated ring segments of the rat iliac artery as readout for endothelial cell EDHF generation in their study. For scientists who were involved in or witnessed the early days of NO research, such procedure provides a *déjà vu* of the bioassay experiments performed at the time when the nature of endothelium-derived relaxing factor (EDRF or NO) was still unknown.<sup>10</sup>

Second, the article addresses the rather unusual beneficial effect of  $H_2O_2$ —formerly denounced as one of the worst ROS—on the generation of EDHF in endothelial cells.  $H_2O_2$ represents the end product of the "detoxification" of superoxide anions  $(O_2^-)$  by 1 of 3 different superoxide dismutases (CuZn-SOD, SOD1; Mn-SOD, SOD2; EC-SOD, SOD3) in endothelial cells, so that its accumulation in the cell reflects an increase in  $O_2^-$  formation, which is most likely accompanied by a decreased bioavailability of NO because of its ultra fast reaction with  $O_2^{-11}$  This observation, along with previous findings demonstrating that EDHF-type relaxations frequently become detectable only or are accentuated in conditions of "endothelial dysfunction", ie, a reduced bioavailability of  $NO$ ,  $^{12}$  reinforces the view that at least 2 endothelium-derived vasodilator principles coexist whereby NO limits the generation of EDHF.13 EDHF-mediated relaxation may thus serve as a backup system and substitute for a diminished NO-mediated relaxation in conditions of "oxidative stress", ie, an increased vascular  $O_2^-$  formation, such as in, eg, diabetes mellitus.<sup>14</sup>

Surprisingly, just about two decades ago or so it was realized that molecular oxygen not only is essential for aerobic metabolism but also gives rise to the formation of ROS which can act both as powerful defensive arms, eg, against invading microorganisms, and as intercellular or intracellular signaling molecules. Most prominent among these ROS is  $O_2^-$ , generated by Nox-2– or Nox-4–containing NADPH oxidase in endothelial cells,<sup>15</sup> and its primary SOD-derived or nonenzymatic dismutation product  $H_2O_2$ .<sup>16</sup> Both molecules, either directly or indirectly, eg, by way of secondary peroxynitrite formation with NO  $(O_2^-)$  and through metal-catalyzed generation of hydroxyl radicals  $(H_2O_2)$ , are capable of altering proteins chemically thus influencing their function. To date many signal transduction pathways have been characterized that operate at least in part through enzymatic ROS formation and consecutive protein modification, thereby eliciting changes in gene expression, cell migration, and proliferation, but also in ion channel activity.<sup>17</sup> Chief modifications comprise a direct oxidation of the target protein, namely that of amino acids with a thiol group such as cysteine, oxidative glycation, and carbonylation.<sup>18-20</sup> In this context, it is noteworthy that not the oxidation of cysteine residues, as favored by the authors in the discussion, but oxidative protein carbonylation may represent the most frequent type of protein modification in conditions of oxidative stress.

Oxidative carbonylation preferentially occurs at the amino acids proline, threonine, lysine, and arginine, presumably through a metal-catalyzed activation of hydrogen peroxide to a reactive intermediate, and according to a recent study conducted with mammalian vascular

*Arterioscler Thromb Vasc Biol*. Author manuscript; available in PMC 2014 June 17.

smooth muscle cells the carbonylated proteins can—comparable with the reduction of oxidized thiols—be decarbonylated as well, thus offering a whole host of novel possibilities for intracellular signal transduction.<sup>18,20</sup> In this context, it would be quite interesting to find out whether the sensitivity of the inositol-1,4,5-trisphosphate (inositol triphosphate  $[IP_3]$ )– stimulated calcium ion  $(Ca^{2+})$  release channel in the endoplasmic reticulum (ER) of endothelial cells can in fact be altered by oxidation of critical sulfhydryl groups, as suggested by the authors, or by (reversible) oxidative carbonylation. In this way, the potentially harmful ROS may be transformed into physiologically important second messengers that, eg, help to maintain or even improve endothelial cell  $Ca^{2+}$  homeostasis.

Third, and perhaps most importantly, the article provides evidence that  $H_2O_2$  enhances EDHF formation through its stimulatory effect on endothelial  $Ca^{2+}$  homeostasis. While an effect of ROS on endothelial  $Ca^{2+}$  signaling per se is not unprecedented,<sup>17</sup> the study by Edwards et al is insofar a surprise that the SERCA inhibitor cyclopiazonic acid (CPA) was used.<sup>1</sup> In most studies dealing with an impact of ROS on endothelial  $Ca^{2+}$  homeostasis so far, an increase in intracellular  $Ca^{2+}$  was evoked by IP<sub>3</sub>-generating agonists such as bradykinin, histamine, or ATP, whereas SERCA inhibitors were frequently used as receptorindependent control stimulus that is insensitive to ROS, at least in terms of intracellular  $Ca^{2+}$  mobilization. The data obtained by the authors argue for a sensitization by H<sub>2</sub>O<sub>2</sub> of IP<sub>3</sub> receptors resulting in an accelerated  $Ca^{2+}$  leakage from the ER that becomes evident when SERCA activity is blocked (Figure). Although this hypothesis seems conclusive, the observation that  $H_2O_2$  augments CPA-triggered intracellular  $Ca^{2+}$  mobilization only at concentrations of the SERCA inhibitor below or equal to 10 *μ*mol/L may deserve further attention. In fact, passive leakage of  $Ca^{2+}$  should be even more pronounced when SERCA activity is blocked more effectively. On the other hand, as demonstrated by the authors, higher concentrations of CPA are also more effective in depleting the intracellular  $Ca^{2+}$ store so that no additional effect by  $H_2O_2$  might be detectable. ER IP<sub>3</sub> receptors, however, have been shown to be closely linked, on a functional level, with mitochondrial  $Ca^{2+}$ uniporters resulting in the establishment of a concerted interorganelle  $Ca^{2+}$  cycling<sup>21</sup> for which some SERCA activity would seem to be essential, eg, by affecting mitochondrial  $Ca^{2+}$  handling. Another important aspect to be solved is whether H<sub>2</sub>O<sub>2</sub>, besides its effect on basal IP<sub>3</sub> receptor activity (ie,  $Ca^{2+}$  leakage through the channel), also affects its IP<sub>3</sub>induced opening.

Fourth, calcium ions probably represent the most versatile (second) messengers in living nature. The physiological consequences of the observed augmentation by  $H_2O_2$  of the mobilization of intracellular  $Ca^{2+}$  most likely exceed those for the formation of EDHF by far. A more pronounced rise in endothelial  $Ca^{2+}$ , for example, has been associated with an elevated synthesis/release of the (potent) vasoconstrictor endothelin-1,22 which might counteract the vasodilator effect of EDHF on the vasculature, more precisely the smooth muscle cells of the media. Moreover, a continuously elevated release by  $H_2O_2$  of  $Ca^{2+}$  from the ER that is accompanied by an accumulation of  $Ca^{2+}$  in the mitochondria may on the one hand trigger an increased biosynthesis of  $ATP<sup>23</sup>$  but then, if the worst comes to the worst, may also result in mitochondria-induced apoptosis.<sup>24</sup> Furthermore, the cytosolic Ca<sup>2+</sup> level governs the activity of numerous signaling pathways and transcription factors, and

*Arterioscler Thromb Vasc Biol*. Author manuscript; available in PMC 2014 June 17.

maintenance of the  $Ca^{2+}$  concentration in the ER is a prerequisite for correct protein folding therein. In this respect, cellular  $Ca^{2+}$  homeostasis needs to be tightly regulated to avoid any uncontrolled activation of  $Ca^{2+}$ -senstive processes. Despite its beneficial effect on EDHF formation, the physiological or pathophysiological consequences of the enhanced leakage of the intracellular  $Ca^{2+}$  store in the presence of  $H_2O_2$  can thus not be properly surveyed at present and require further investigation.

So there is yet another solid piece of evidence that our judgment on the contribution of ROS in physiology and pathophysiology needs to be reconsidered. Moreover, the current work by Edwards et al<sup>1</sup> again points at  $Ca^{2+}$  as one of the major second messengers, particularly in endothelial cells, and supports the concept of EDHF as an important principle of vasodilatation and not just an epiphenomenon of the NO-mediated control of blood flow. One of several questions that nonetheless needs to be answered is where or what the endogenous source of  $H_2O_2$  in the native endothelial cells is. It may the mitochondria reacting to an increase in intracellular  $Ca^{2+}$ , the agonist-dependent NADPH oxidase assembled at the plasma membrane (Nox-2), or the constitutively active NADPH oxidase situated in the ER (Nox-4). In each case, effective enzymatic dismutation of the primary reaction product  $O_2^-$  to  $H_2O_2$  seems to be a must.

# **Acknowledgments**

#### **Sources of Funding**

Research in the laboratories of Drs Graier and Hecker is supported by the Austrian Science Funds (FWF; P20181- B05, F 3010-B05), the German Research Foundation (DFG; SFB/TR 23-C5, SFB 405-B17, GRK 880, HE 1587/9-1), and the Federal Ministry of Education and Research (BMBF; 01GR0822), respectively.

#### **References**

- 1. Edwards D, Li Y, Griffith T. Hydrogene peroxide potentiates the EDHF phenomenon by promoting endothelial Ca<sup>2+</sup> mobilization. Arterioscler Thromb Vasc Biol. 2008; 28:1774–1781. [PubMed: 18669883]
- 2. Busse R, Edwards G, Feletou M, Fleming I, Vanhoutte PM, Weston AH. EDHF: bringing the concepts together. Trends Pharmacol Sci. 2002; 23:374–380. [PubMed: 12377579]
- 3. Edwards G, Dora KA, Gardener MJ, Garland CJ, Weston AH. K+ is an endothelium-derived hyperpolarizing factor in rat arteries. Nature. 1998; 396:269–272. [PubMed: 9834033]
- 4. Hayabuchi Y, Nakaya Y, Matsuoka S, Kuroda Y. Endothelium-derived hyperpolarizing factor activates  $Ca^{2+}$ -activated  $K^+$  channels in porcine coronary artery smooth muscle cells. J Cardiovasc Pharmacol. 1998; 32:642–649. [PubMed: 9781934]
- 5. Fisslthaler B, Popp R, Kiss L, Potente M, Harder DR, Fleming I, Busse R. Cytochrome P450 2C is an EDHF synthase in coronary arteries. Nature. 1999; 401:493–497. [PubMed: 10519554]
- 6. Randall MD, Kendall DA. Involvement of a cannabinoid in endothelium-derived hyperpolarizing factor-mediated coronary vasorelaxation. Eur J Pharmacol. 1997; 335:205–209. [PubMed: 9369375]
- 7. Shimokawa H, Morikawa K. Hydrogen peroxide is an endotheliumderived hyperpolarizing factor in animals and humans. J Mol Cell Cardiol. 2005; 39:725–732. [PubMed: 16122755]
- 8. Griffith TM. Endothelium-dependent smooth muscle hyperpolarization: do gap junctions provide a unifying hypothesis? Br J Pharmacol. 2004; 141:881–903. [PubMed: 15028638]
- 9. Harris D, Martin PE, Evans WH, Kendall DA, Griffith TM, Randall MD. Role of gap junctions in endothelium-derived hyperpolarizing factor responses and mechanisms of K+-relaxation. Eur J Pharmacol. 2000; 402:119–128. [PubMed: 10940365]

*Arterioscler Thromb Vasc Biol*. Author manuscript; available in PMC 2014 June 17.

- 10. Furchgott RF, Zawadzki JV. The obligatory role of endothelial cells in the relaxation of arterial smooth muscle by acetylcholine. Nature. 1980; 288:373–376. [PubMed: 6253831]
- 11. Graier WF, Posch K, Wascher TC, Kostner GM. Role of superoxide anions in changes of endothelial vasoactive response during acute hyperglycemia. Horm Metab Res. 1997; 29:622–626. [PubMed: 9497899]
- 12. Bauersachs J, Popp R, Hecker M, Sauer E, Fleming I, Busse R. Nitric oxide attenuates the release of endothelium-derived hyperpolarizing factor. Circulation. 1996; 94:3341–3347. [PubMed: 8989149]
- 13. Fleming I, Busse R. Endothelium-derived epoxyeicosatrienoic acids and vascular function. Hypertension. 2006; 47:629–633. [PubMed: 16490839]
- 14. Fleischhacker E, Esenabhalu VE, Spitaler M, Holzmann S, Skrabal F, Koidl B, Kostner GM, Graier WF. Human diabetes is associated with hyperreactivity of vascular smooth muscle cells due to altered subcellular  $Ca^{2+}$  distribution. Diabetes. 1999; 48:1323-1330. [PubMed: 10342823]
- 15. Brandes RP, Schröder K. Composition and functions of vascular nicotinamide adenine dinucleotide phosphate oxidases. Trends Cardiovasc Med. 2008; 18:15–19. [PubMed: 18206804]
- 16. Cai H. NAD(P)H oxidase-dependent self-propagation of hydrogen peroxide and vascular disease. Circ Res. 2005; 96:818–822. [PubMed: 15860762]
- 17. Spitaler MM, Graier WF. Vascular targets of redox signalling in diabetes mellitus. Diabetologia. 2002; 45:476–494. [PubMed: 12032623]
- 18. Dalle-Donne I, Aldini G, Carini M, Colombo R, Rossi R, Milzani A. Protein carbonylation, cellular dysfunction, and disease progression. J Cell Mol Med. 2006; 10:389–406. [PubMed: 16796807]
- 19. England K, Cotter TG. Direct oxidative modifications of signalling proteins in mammalian cells and their effects on apoptosis. Redox Rep. 2005; 10:237–245. [PubMed: 16354412]
- 20. Wong CM, Cheema AK, Zhang L, Suzuki YJ. Protein carbonylation as a novel mechanism in redox signaling. Circ Res. 2008; 102:310–318. [PubMed: 18079412]
- 21. Szabadkai G, Simoni AM, Rizzuto R. Mitochondrial  $Ca^{2+}$  uptake requires sustained  $Ca^{2+}$  release from the endoplasmic reticulum. J Biol Chem. 2003; 278:15153–15161. [PubMed: 12586823]
- 22. Brunner F, Stessel H, Simecek S, Graier W, Kukovetz WR. Effect of intracellular  $Ca^{2+}$ concentration on endothelin-1 secretion. FEBS Lett. 1994; 350:33–36. [PubMed: 8062919]
- 23. Jouaville LS, Pinton P, Bastianutto C, Rutter GA, Rizzuto R. Regulation of mitochondrial ATP synthesis by calcium: evidence for a long-term metabolic priming. Proc Natl Acad Sci U S A. 1999; 96:13807–13812. [PubMed: 10570154]
- 24. Demaurex N, Distelhorst C. Apoptosis the calcium connection. Science. 2003; 300:65–67. [PubMed: 12677047]



#### **Figure.**

Schematic illustration of the interplay between the intracellular generation of  $H_2O_2$  and intracellular  $Ca^{2+}$  mobilization from the endoplasmic reticulum (ER) as a consequence of the effect of  $H_2O_2$  on inositol-1,4,5 trisphosphate (IP<sub>3</sub>) receptors that was described by Edwards et al.