# Comparison of two serpins of *Clonorchis sinensis* by bioinformatics, expression, and localization in metacercaria

## Yabo Yang<sup>1,2</sup>, Dong Hu<sup>3</sup>, Lexun Wang<sup>4</sup>, Chi Liang<sup>2</sup>, Xuchu Hu<sup>2</sup>, Jin Xu<sup>2</sup>, Yan Huang<sup>2</sup>, Xinbing Yu<sup>2</sup>

<sup>1</sup>Department of Obstetrics and Gynecology, Sun Yat-Sen Memorial Hospital, Sun Yat-Sen University, Guangzhou, China, <sup>2</sup>Department of Parasitology, Key Laboratory for Tropical Diseases Control, Sun Yat-Sen University, Guangzhou, China, <sup>3</sup>Department of Medical Immunology, School of Medicine, Anhui University of Science and Technology, Huainan, China, <sup>4</sup>Department of Hematology, the Third Affiliated Hospital, Sun Yat-Sen University, Guangzhou, China

Clonorchiasis, which has been an important public health problem in China, is caused by ingestion of raw or undercooked fish contaminated by live metacercaria. Therefore, preventing fish from infecting is of great significance for controlling the disease. SERPINs (serine protease inhibitors) are well known as negative regulators of hemostasis, thrombolysis, and innate immune responses. In the present study, two full-length sequences encoding SERPIN were identified from metacercaria cDNA library of Clonorchis sinensis (C. sinensis) and were denominated as CsSERPIN and CsSERPIN3, respectively. Bioinformatics analysis showed that the two sequences shares 35.9% identity to each other. Both of the sequences have SERPIN domain and the greatest difference between the two domains is the reactive centre loop. Transmembrane region was found in CsSERPIN3 while not in CsSERPIN. The expression of the two CsSERPINs was significantly higher at the life stage of metacercaria than that of adult. The transcription levels of CsSERPIN and CsSERPIN3 at metacercaria stage were 3.249- and 11.314-fold of that at adult stage, respectively. Furthermore, the expression of CsSERPIN was 4.32-fold of that of CsSERPIN3 at metacercaria stage. Immunobiochemistry revealed that CsERPIN was dispersed at subtegument and oral sucker of metacercaria, while CsSERPIN3 localized intensely in the tegument of metacercaria of C. sinensis inside of the cyst wall. All these indicated that the CsSERPINs play important roles at metacercaria stage of the parasite. CsSERPIN may take part in regulation of endogenous serine proteinase and CsSERPIN3 may be involved in immune evasion and be a potential candidate for vaccine and drug target for clonorchiasis.

Keywords: Clonorchis sinensis, Serpin, Serine proteinase inhibitor, Metacercaria, Gene expression, Immunolocalization

#### Introduction

Clonorchiasis, caused by the liver fluke *Clonorchis* sinensis (*C. sinensis*), is one of the most prevalent parasitic diseases distributed in China, Korea, and Japan. As an important food-borne parasite, *C. sinensis* has afflicted more than 35 million people in Asia and approximately 15 million in China, creating a socioeconomic burden in epidemic regions.<sup>1,2</sup> Current evidences from experimental and epidemiological investigations have confirmed the association between *C. sinensis* and cholangiocarcinoma.<sup>2–4</sup> Moreover, long-term infections by liver flukes lead to other hepatobiliary diseases such as pyogenic cholangitis, cholelithiasis, cholecystitis, and hepatic fibrosis.<sup>5,6</sup> The consumption of raw or uncooked fish containing infectious metacercariae of *C. sinensis* is considered to be the major source of infection. Therefore, preventing fish from infection may be the best way of controlling *C. sinensis* infection.

Serpins are well known as negative regulators of haemostasis, thrombolysis, and innate immune responses.<sup>6</sup> In our previous study, one novel serpin named *Cs*SERPIN was identified and characterized.<sup>7,8</sup> It is significantly highly expressed in metacercaria stage. The biochemical activities showed that rCsproSERPIN could significantly inhibit trypsin, chymotrypsin, and thrombin, indicating that it may play an important role in the information of metacercariae. In the present study, another serpin gene was identified and expressed. The two sepins were compared

Correspondence to: X. B. Yu, Department of Parasitology, Key Laboratory for Tropical Diseases Control, Sun Yat-Sen University, Guangzhou 510080, China. Email: yuxbteam@163.com

Co-correspondence author: Y. Huang, Department of Parasitology, Key Laboratory for Tropical Disease Control, Sun Yat-Sen University, Guangzhou 510080, China. Email: huang66@mail.sysu.edu.cn

by bioinformatics, real-time PCR, localization in metacercariae, which might lay some cornerstone for evaluating *Cs*SERPINs as potential candidate for vaccine against clonorchiasis.

#### **Materials and Methods**

#### Animals

Six-week-old male Sprague–Dawley rats were purchased from the animal centre of Sun Yat-Sen University for experiments. The rats were housed in aseptic cages and fed with pelleted food and water. All experiments were performed on the basis of guidelines approved by the international animal ethical standard.

## Bioinformatics analysis of two cDNA sequence encoding C. sinensis serpin (Csserpin)

From the cDNA library of *C. sinensis* metacercaria, two different insert sequence, 133f12 and 84g02, were predicted of encoding homologue of serpins by BLASTx. The complete encoding sequence of *CsSERP* IN was analysed by Open Reading Frame Finder program at NCBI. The deduced protein sequences and the identities of them were analysed by Vector NTI suite software. Protein analysis was done with Proteomics and sequence analysis tools (http://ca.expasy.org/tools).

## Quantitative PCR analysis of two CsSERPINs at stage of adult and metacercaria

Adult worms were harvested from biliary tracts of infected cats and metacercariae were isolated from Pseudorasbora parva by pepsin digestion. Total RNA was extracted from adults and metacercariae, respectively, after being washed three times in phosphatebuffered saline (PBS) (pH 7.4). TRIZOL reagent (Invitrogen) was applied for extraction according to manufacturer's instructions and treated with DNase (Promega). cDNA was generated from total RNA with a PrimeScript<sup>TM</sup> RT-PCR Kit (Perfect Real Time) from TaKaRa using 1-2 µg of total RNA as template. Real-time PCR was performed using the SYBR<sup>®</sup> Premix ExTag<sup>TM</sup> kit (TaKaRa, Tokyo, Japan) according to the manufacturer's protocol. The amplification conditions were optimized for LightCycler 480 (Roche, Switzerland). Using Primer primier 5.0 software, the primers for CsSERPIN were designed as 5'-AGT AAC GAT GCG AAG CGA GAG C-3', 5'-ATA GAA CCC GAA GGA AGG AGG C-3'; for CsSERPIN3, they were 5'-CCA AAA TGT ATT CAT CTC CCC CC-3', 5'-GGC AAT GTG GCT TCT TCT AAC TC-3'. Beta-actin (GenBank accession no. EU109284, 323 bp) was amplified as the endogenous control<sup>9</sup> using the primers 5'-GGT GAC GCT GAA GTA TCC TAT TGA-3', 5'-CCA AAG CAT AGC CCT CGT AGA T-3', respectively. Data were analysed using LightCycler 480 Software and using the  $2^{-\Delta\Delta C_t}$  method.<sup>10</sup>

## Cloning, expression and purification of rCsproSERPIN in Escherichia coli

To get the recombinant plasmid of CsSERPIN3, the gene-encoding sequence was amplified by using specific primers as follows: sense, 5'-CGG CAT ATG ATG GAG AGT GAA ATG GAT T-3' and antisense, 5'-AAA CTC GAG CTA CAA GAC CTC AGG TTC-3' with Nde I/Xho I as restriction enzyme sites (underlined), respectively. The purified PCR product was cloned into the corresponding restriction sites of pET28a(+) (Novagen, USA) vector and then the recombinant plasmid was transformed into E. coli BL21(DE3) (Promega). Recombinant C. senensis serpin protein (rCsproSERPIN3) was expressed by inducing with isopropyl-beta-D-thiogalactopyranoside at a final concentration of 0.2 mM at 30°C for 4 hours. The bacterial cells were harvested by centrifuging at 4°C for 15 minutes at 8000g and suspended in lysis buffer (0.5 M NaCl, 20 mM Tris-HCl, 5 mM imidazole, pH 8.0), sonicated on ice, and centrifuged at 10 000g for 15 minutes at 4°C. The recombinant fusion protein was batch-purified with His Bind Purification kit (Novagen) according to the manufacturer's instructions. The eluted fractions were pooled and dialyzed with PBS (10 mM phosphate buffer, 27 mM KCl, 137 mM NaCl, pH 7.4). The purification and purity of the recombinant protein was monitored by sodium dodecyl sulphatepolyacrylamide gel electrophoresis (SDS-PAGE, 12% gel) coloured with *Coomassie* blue, and protein concentration was estimated using Bradford assay. rCsproSERPIN was achieved as previously described.<sup>7</sup>

## Acquirement of rat anti-rCsSERPIN immune serum and detection of IgG titre

For production of polyclonal antiserum for rCspro SERPIN and rCsproSERPIN3, Sprague–Dawley rat were immunized with purified rCsproSERPIN (100  $\mu$ g) or rCsproSERPIN3 (100 µg) three times at 2-week intervals. Anti-sera were collected at two weeks after the second boost and the IgG titre was determined by enzyme-linked immunosorbent assay. Briefly, the plates were coated with 5 µg/ml purified rCsproSERPIN or rCsproSERPIN3 in coating buffer (0.05 M carbonatebicarbonate, pH 9.6) and incubated at 4°C overnight, followed by blocked with 5% skimmed milk in PBS containing 0.05% Tween 20 (pH 7.4) for 2 hours at 37°C. After washing three times, the plates were incubated with different dilutions of corresponding immune sera. Rat sera immunized with PBS was measured as negative controls. Then they were washed and incubated with horseradish peroxidase-conjugated IgG  $(1:20\ 000\ dilutions\ in\ 0.1\%\ BSA-PBST)$  for 1 hour. After washing, the enzyme reaction was developed with TMB substrate solution (BD Biosciences) and terminated by 2 M H<sub>2</sub>SO<sub>4</sub>. The absorbance was measured at 450 nm. Finally, they were incubated with the supernatant of *E. coli* BL21(DE3) containing pET28a(+) overnight at a ratio of 1:9 and centrifuged. The immune sera were stored at  $-80^{\circ}$ C for use.

## Western blotting analysis of CsSERPIN and CsSERPIN3

Crude soluble antigens (CSAs) of adult worms and metacercariae were prepared as previously described. The rCsproSERPIN (5  $\mu$ g), rCsproSERPIN3 (5  $\mu$ g), or CSA (20 µg) were subjected to SDS-PAGE (12% gel) and then electrotransferred to membrane (Millipore). The polyvinylidene difluoride membranes were probed with naive rat serum, antirCsproSERPIN and anti-rCsproSERPIN3 rat serum (1:4000 dilutions), rat serum infected with C. sinensis (1:400 dilution), normal human serum, and serum from human infected with C. sinensis, respectively. After washed three times, membranes were incubated with horseradish horseradish-conjugated anti-rat immunoglobulin G (Boster, 1:2000 dilution) for 1 hour at room temperature (RT). After washed again, membranes were visualized with diaminobenzidine showing colour reagent (Boster) as substrate.

## *Immunolocalization of* CsSERPIN at metacercaria of C. sinensis

Living metacercariae were washed three times in PBS (pH 7.4) and fixed in 4% paraformaldehyde in PBS (pH 7.2), embedded with paraffin, then sliced into section of 4-5 µm in thickness. The slides were blocked with normal goat sera containing 1% BSA for 2 hours at RT then incubated in a humid atmosphere at RT for 2 hours with serum from rat immunized in PBS (negative contrast), antirCsSERPIN rat serum and anti-rCsSERPIN3 rat serum which were diluted (0.1% BSA in PBST) to 1:200. After washed five times with PBST, the slides were incubated further with fluorescein isothiscyanate-conjugated goat anti-rat IgG (1:400 with 0.1% BSA in PBST; Invitrogen) for 1 hour at RT in dark, washed five times with PBST in dark again and subsequently observed under fluorescence microscope.

## Results

#### Sequence analysis of CsSERPIN and CsSERPIN3

The inserted sequence of 133f12 was 1149 bp in length and 84g02 was 1161 bp. They encoded proteins of 382 amino acids (aa) and 386 aa, respectively, which dominated as *Cs*SERPIN and *Cs*SERPIN3. The calculated molecular weight of them is 42.2 and 43.5 kDa. Neither amino-acid sequence of *Cs*SERPIN and *Cs*SERPIN3 has Nterminal signal peptide. Both sequences have SERPIN domain and the greatest difference between the two domains is the reactive centre loop. Transmembrane region was found in *Cs*SERPIN3 while not in *Cs*SERPIN. The deduced amino-acid sequence of *Cs*SERPIN3 was highly homologous to serpins from other species. It has been deposited to GenBank database with accession number KJ489357 (Fig. 1).

## Expression, purification, and characterization of rCsproSERPIN and rCsproSERPIN3

The two r*Cs*proSERPINs could express as soluble fusion protein in *E. coli* BL21/DE with  $6 \times$  His-tag at the N-terminals. The yield of purified r*Cs*proSERPIN in soluble cell was 7.5 mg/l of *E. coli*, and that of r*Cs*proSERPIN3 was 2 mg/l. The concentration of purified r*Cs*proSERPIN3 was 2 mg/l. The concentration of the two rat anti-r*Cs*SERPIN sera is above 51 200. The two kinds of r*Cs*proSERPIN could be probed by mouse anti-His-tag monoclonal antibody at 44.5 and 45.8 kDa, respectively, which could also be recognized by the corresponding rat anti-r*Cs*SERPIN serum, rat and human serum infected with *C. sinensis*. A pale band in the normal human sera could be found upon closer examination<sup>7</sup> (Fig. 3).

## *Expression of* CsSERPIN and CsSERPIN<sub>3</sub> at life stage of metacercaria and adult of C. sinensis

The transcription of the two *Cs*SERPINs could be detected at both metacercaria and adult stage. Compared with the mRNA level at adult stage, the transcription at metacercaria stage of *Cs*SERPIN and *Cs*SERPIN3 was 3.249 and 11.314 times higher with *C. sinensis* beta-*actin* as internal standard. Furthermore, the expression of *Cs*SERPIN was 4.32-fold of that of *Cs*SERPIN3 at metacercaria stage (Fig. 4). By Western-blot analysis, CSA of metacercariae can be obviously probed by anti-r*Cs*proSERPIN and anti*rCs*proSERPIN3 rat serum at about 45 kDa, while corresponding band could hardly be observed with CSA of adult probed by the immune serum (Fig. 5).

#### *Immunolocalization of* CsSERPIN and CsSERPIN3 *in metacercaria of* C. sinensis

By immunofluorescence localization with anti-CsSERPIN and anti-CsSERPIN3 rat serum, the deposition of CsSERPIN and CsSERPIN3 in metacercariae was detected. The results showed that CsERPIN was dispersed at subtegument and oral sucker of metacercaria, while CsSERPIN3 localized mostly in the tegument of metacercaria of C. sinensis inside of the cyst wall (Fig. 6).

## Discussion

In our previous study, a serpin of *C. sinensis* named *Cs*SERPIN was acquired and characterized. It was highly expressed at the stage of metacercaria, which suggested that it played an important role in the stage of *C. sinensis* metacercaria. Another novel sequence encoding serpin named *Cs*SERPIN3 was identified and characterized in this study. The cDNA sequence is 1161 bp in length which encoded a protein of 386 aa. It shares typical serpin motif, reactive centre loop,



Figure 1 Multiple sequence alignment. The deduced amino-acid sequence of *Cs*SERPIN3 (GenBank accession number: KJ489357) from *C. sinensis* was aligned with those of presently known serpin. Gaps are introduced into the sequences for optimal alignment. The serpin motif is indicated as a single bold line, the reactive site loop shown as a dotted line, and the serpin signature indicated as a double thin line, and the transmembrane region of *Cs*SERPIN3 presented as a single thin line. The scissile bond is marked with a red arrow.

and serpin signature, which is well conserved in the serpin superfamily. Bioinformatics analysis identified that the greatest difference between *CsSERPIN* and *CsSERPIN3* is the reactive centre loop. Neither of the two sequences was found containing characteristics N-terminal signal peptide cleavage sites. Transmembrane region was found in *CsSERPIN3* while not in *CsSERPIN*. The transcription and expression of both *CsSERPINs* was much higher at the stage of metacercariae than that at adult. *CsSERPIN* was mostly deposited in oral sucker and some were dispersed at subtegument, while *CsSERP* 

PIN3 localized in the tegument of metacercaria of *C. sinensis* inside of the cyst wall. Besides, soluble r*Cs*proSERPIN and r*Cs*proSERPIN3 were obtained and the recombinant protein could be probed by sera from infected rats or patients.

Some studies showed a serine proteinase-mediated encystation of *Acanthamoeba* and chemically synthesized serine proteinase inhibitors significantly reduced the encystation efficiency of *Acanthamoeba*.<sup>11</sup> Both encystment and excystment of *Acanthamoeba castellanii* were found to be dependent on serine protease function.<sup>12</sup> However, the function of serine protease



Figure 2 Expression and purification of r*Cs*proSERPIN3 by 12% SDS–PAGE stained with *Coomassie* brilliant blue. Protein molecular weight markers (M), lysate of *E. coli* with pET28a(+) before induction (lane1) and after induction (lane 2), lysate of *E. coli* with pET28a(+)-*Cs*SERPIN3 before induction (lane 3) and after induction (lane 4), supernatant (lane 5), and precipitant (lane 6) of lysate of *E. coli* with pET28a(+)-*Cs*SERPIN3 after induction; lane 7, r*Cs*proSERPIN3 after purification.

should be tightly regulated by its inhibitor. In fact, the concept that parasites may utilize proteinase inhibitors to survive within the host which has been with us for many years.<sup>13,14</sup> Many evidence demonstrated that parasite could encode serpins to inhibit the hosts' serine proteinases and block their immune systems. A number of serpins have now been identified in parasitic helminths with putative involvement in immune regulation and in parasite survival through interference with the host immune response.<sup>15</sup> A novel serpin expressed by blood-borne microfilariae of the parasitic nematode *Brugia malayi* 



Figure 3 Western blot analysis of r*Cs*proSERPIN3. Protein molecular weight markers; r*Cs*proSERPIN3 probed with naive rat serum (lane 1), with anti-r*Cs*proSERPIN3 rat serum (lane 2), with serum from rat infected with *C. sinensis* (lane 3), with serum from normal human (lane 4), with serum from human infected with *C. sinensis* (lane 5); lysate of *E. coli* with pET28a(+)-*Cs*SERPIN3 before induction (lane 6) and after induction (lane 7) probed with anti-r*Cs*proSERPIN3 rat serum; lysate of *E. coli* with pET28a(+)-*Cs*SERPIN3 before induction (lane 8) and after induction (lane 9) probed with His monoclonal antibody of mouse; r*Cs*proSERPIN3 after purification probed with His monoclonal antibody of mouse (lane 10).



Figure 4 Transcript analysis of *Cs*SERPIN and *Cs*SERPIN3 at the stage of adult and metacercaria of *Clonorchis sinensis*. Beta-*actin* of *C. sinensis* (accession number: EU109284) was used as the transcription control. The  $\Delta C_t$ values of each adult and metacercaria were calculated by deducting the values of beta-*actin*, and the  $-\Delta\Delta C_t$  values were figured out according to the  $2^{-\Delta\Delta C_t}$  method. The amplification of *Cs*SERPIN3 of adult worms was employed as the calibrator to evaluate relative expression levels of the others. Each bar represented the mean value from three experiments with standard deviation (\**P*<0.05).

inhibited human neutrophil serine proteinases.<sup>16,17</sup> Serpins isolated from adult *Schistosoma mansoni* and *Schistosoma haematobium* possibly play a dual role of both in parasite physiology and in interactions with their hosts.<sup>18,19</sup>



Figure 5 Western blot analysis of r*Cs*proSERPIN and r*Cs*proSERPIN3 in adult and metacercaria. Lane 1, soluble antigen of *C. sinensis* adult probed with naive rat serum; lane 2, soluble antigen of *C. sinensis* metacercaria probed with naive rat serum; lane 3, soluble antigen of *C. sinensis* adult probed with anti-r*Cs*proSERPIN rat serum; lane 4, soluble antigen of *C. sinensis* metacercaria probed with anti-r*Cs*proSERPIN rat serum; lane 5, soluble antigen of *C. sinensis* adult probed with anti-r*Cs*proSERPIN rat serum; lane 6, soluble antigen of *C. sinensis* metacercaria probed with anti-r*Cs*proSERPIN3 rat serum; lane 6, soluble antigen of *C. sinensis* metacercaria probed with anti-r*Cs*proSERPIN3 rat serum; lane 6, soluble antigen of *C. sinensis* metacercaria probed with anti-r*Cs*proSERPIN3 rat serum; lane 6, soluble antigen of *C. sinensis* metacercaria probed with anti-r*Cs*proSERPIN3 rat serum; lane 6, soluble antigen of *C. sinensis* metacercaria probed with anti-r*Cs*proSERPIN3 rat serum; lane 6, soluble antigen of *C. sinensis* metacercaria probed with anti-r*Cs*proSERPIN3 rat serum; lane 6, soluble antigen of *C. sinensis* metacercaria probed with anti-r*Cs*proSERPIN3 rat serum; lane 6, soluble antigen of *C. sinensis* metacercaria probed with anti-r*Cs*proSERPIN3 rat serum.



Figure 6 Immunolocalization of *Cs*SERPIN and *Cs*SERPIN3 in metacercariae of *C. sinensis*. A–C were probed with serum from rats immunized with PBS (negative control); D–F were probed with serum from rats immunized with anti-r*Cs*proSERPIN; G–I were probed with serum from rats immunized with anti-r*Cs*proSERPIN3. B, E, and H are fluorescence microscopy images; A, D, and G are corresponding optical images of B, E, and H. C, F and I are superposed images (C, F, and I) of the previous two. OS, oral sucker; EB, excretory bladder; CW, cyst wall.

The roles of serpins in parasites are usually closely related with their locations. The intracellular serpins are believed to be the protection of cells from superfluous proteolytic damage and the maintenance of cellular homeostasis.<sup>20</sup> *Schistosoma japonicum* (*S. japonicum*) serpin is located on the tegument in *S. japonicum* adult worms and is speculated to be a vaccine candidate or a novel target for anti-schistosome drugs.<sup>19</sup> Parasitederived serpins with signal peptide are suggested to play essential roles in the survival of the parasites in the hosts via host immune evasion and protection from exogenous host proteases.<sup>13</sup>

The expression of the *Cs*SERPIN and *Cs*SERPIN3 at metacercariae stage was significantly high than that

at adult stage (see supplementary material online for this article www.maneyonline.com/doi/suppl/10.1179/ 2047773214Y.0000000138), which gives us a clue that the two CsSERPINs play an important role in metacercariae. Although neither of them has signal peptide, there was great difference between CsSERPIN and CsSERPIN3 at the reactive centre loop. Transmembrane region was found in CsSERPIN3 and immunofluerence also showed that CsSERPIN3 localized in the tegument of metacercaria inside of the cyst wall, indicating that it should be a tegument-anchored protein. Moreover, rCspro SERPIN3 can be probed by the sera from patients infected with C. sinensis. The most likely reason for the

band in the nomal human sera may be that the human was infected subclinical previously and there was a little cross-reactivity. All these manifested that CsSERPIN3 may be a candidate of vaccine and drug target for clonorchiasis. CsSERPIN did not contain any transmembrane region and it was mostly deposited in oral sucker and some were dispersed at subtegument, which implied that CsSERPIN was an intracellular serpin and not a secreted or membranebound protein. In view that it was highly expressed at the stage of metacercaria and the expression was 4.32 times of that of CsSERPIN3, the role of CsSERPIN may be the regulation the activities of intracellular serine proteinase of the parasite. Studies also displayed that rCsproSERPIN could be recognized by the sera of rabbits immunized with excretory/secretory products and may be involved in immunoregulation of host by inducing Th1-biased type cytokines in rats.<sup>8</sup>

Taken together, two different serpins which were both highly expressed at the stage of metacercaria were compared in this study. Their different locations may result in different functions. *CsSERPIN* probably plays an important role in regulation of endogenous serine proteinase and *CsSERPIN3* is a potential candidate for vaccine or drug target. Further studies will be assessing the specific immune responses against *CsSERPINs* in intermediate host.

#### **Disclaimer Statements**

#### Contributors None.

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**Ethics approval** This study was prospectively performed and approved by the institutional Ethics Committees of Sun Yat-Sen University and conducted in accordance with the ethical guidelines of the Declaration of Helsinki.

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