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Differential role of psoriasin (S100A7) in estrogen receptor α positive and negative breast cancer cells occur through actin remodeling

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Abstract

Psoriasin (S100A7) is a calcium-binding protein that has shown to be highly expressed in high-grade ductal carcinoma in situ (DCIS) and a subset of invasive breast cancers. However, its role in invasion and metastasis is not very well known. In this study, we have shown that S100A7 differentially regulates epidermal growth factor (EGF)-induced cell migration and invasion in ER α ⁻ MDA-MB-231 cells and ER α ⁺ MCF-7 and T47D breast cancer cells. Further signaling studies revealed that S100A7 enhances EGF-induced EGFR phosphorylation and actin remodeling that seems to favor lamellipodia formation in ER α ⁻ cells. In addition, S100A7 overexpression enhanced NF- κ B-mediated matrix metalloproteinase-9 (MMP-9) secretion in MDA-MB-231 cells

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Conflict of interest None.

indicating its role in enhanced invasiveness. However, S100A7 overexpression inhibited migration and invasion of MCF-7 cells by inactivating Rac-1 pathway and MMP-9 secretion. Moreover, S100A7 overexpressing MDA-MB-231 cells showed enhanced metastasis compared to vector control in in vivo nude mice as detected by bioluminescence imaging. Our tissue microarray data also revealed predominant expression of S100A7 in ER α ⁻ metastatic carcinoma, especially in lymph node regions. Overall these studies suggest that S100A7 may enhance metastasis in ER α ⁻ breast cancer cells by a novel mechanism through regulation of actin cytoskeleton and MMP-9 secretion.

Keywords

Breast cancer; S100A7; Estrogen receptor- α ; Actin remodeling; MMP-9

Introduction

Psoriasin (S100A7) is a low molecular weight S100 gene family protein, originally isolated from skin psoriatic lesions [1]. The S100 gene family consisting of ~20 members is defined by calcium binding helix-loop-helix structural EF hand motif [2]. Apart from two calcium-binding sites, S100A7 has an additional zinc-binding site [3]. S100A7 expression has been reported in epithelial malignancies such as breast, lung, bladder, skin, esophageal, gastric, and head and neck [4–7]. In breast cancer, S100A7 expression is highly associated with high-grade ductal carcinoma in situ (DCIS) and invasive carcinoma compared to normal breast tissues [4, 8, 9]. Differential expression of S100A7 in breast cancer was initially observed in primary breast cancer compared to normal tissue [10]. Previous studies have shown that S100A7 is expressed in ~50 % of ER α ⁻ and only ~20 % of ER α ⁺ cases of breast cancer [4, 8, 9, 11–13]. Interestingly, in terms of prognosis, both DCIS and invasive breast cancer forms showed consistent association of S100A7 with ER α ⁻ tumors [4]. Furthermore, S100A7 has been known to modulate tumor growth by activating several signaling pathways, including PI3K, NF- κ B, AP-1, and Jab1 [13–15]. However, recent studies have reported the tumor suppressive effects of S100A7 in ER α ⁺ breast cancer cells [16, 17].

Breast carcinoma is classified based on the expression of three receptors: EGFR, ER, and HER-2. EGFR expression is closely related to ER receptor status and has adverse association to overall patient survival with poor prognosis. One prominent feature of ER α ⁻ tumors, especially triple-negative basal-like subtype, is the expression of EGFR [18]. These basal-like tumors are associated with aggressive histological features, poor prognosis and are extremely difficult to treat. High EGFR expression is also associated with metastatic and invasive form of breast cancer [18]. EGFR activation provokes a plethora of signaling pathways that includes cell proliferation, adhesion and motility and promotes invasion and angiogenesis [19]. Recent studies in our laboratory and others have shown that S100A7 regulates epidermal growth factor (EGF)/EGFR-mediated signaling pathways [13–15]. Studies have also shown that S100A7 and EGFR are associated with ER α ⁻ tumors in a large unselected cohort of breast cancer patients [13].

Actin dynamics and remodeling have been identified as major determinant of metastasis and invasion that are the key basis of most cancer-related deaths. During cell motility, branched network of actin filaments are required to assemble beneath the plasma membrane to consistently progress the cell forward to form lamellipodia [20]. The recruitment of active Rac1, a small Rho GTPase at the leading edge, is itself sufficient for cell extension and further movement [21, 22]. Moreover, the influence of S100A7 in calcium-mediated signal transduction and cellular events through direct interactions with intermediate filaments also implies its role in modulation of the cytoskeleton [2].

Hence, present study investigated the influence of S100A7 on metastatic and invasive abilities of ER α ⁻ and ER α ⁺ breast cancer cells upon EGF stimulation. Our studies showed that S100A7 differentially regulates migration and invasion in ER α ⁻ and ER α ⁺ cells. S100A7 overexpression enhanced EGF-induced migration/invasion in ER α ⁻ cells, while its overexpression inhibited migration/invasion in ER α ⁺ cells. We also showed that S100A7 enhances metastasis in vivo and its predominant expression was observed in ER α ⁻ lymph node metastatic group of breast patient samples. In addition, actin polymerization pathway seems to play an important role in establishing the differential effect of S100A7 in ER α ⁻ and ER α ⁺ breast cancer cells.

Materials and methods

Cells, stable transfections, and antibodies

The vector information and generation of stable clones of MDA-MB-231, MCF7, and T47D breast carcinoma cells with stable vector and S100A7 overexpression used in the present study are as described earlier [16, 23]. Knockdown of p65-NF- κ B was performed using its siRNA transfection (50 nM; Dharmacon) for 72 h using lipofectamine 2000 as per manufacturer's protocol. Antibodies used were mostly from Cell Signaling, GAPDH (Santa Cruz Biotechnology) and Phalloidin-568 (Invitrogen).

Migration and invasion assay

Wound healing assay, chemotaxis assay, and invasion assay were performed and calculated as described previously [15, 16, 23, 24].

Gelatin zymography

This method was used to compare MMP-2 and MMP-9 with gelatinase activity of MDA-MB-231 and MCF-7 cells upon S100A7 overexpression (24 h). Supernatants containing secreted form of MMPs were concentrated using centrifugal filter units (Millipore) and detected using Novex gelatin zymography. Renaturing, developing, and staining steps were followed to visualize active MMP bands according to the manufacturer's instructions (Life technologies).

Western blotting

Western blot analysis was done as previously described [23, 24].

Rac1 activation assay

Activation of Rac1 was determined using the Rac/Cdc42 activation assay kit as per manufacturer's protocol (Millipore). Briefly, cell lysates were incubated with 10 µg/mL p21-activated kinase 1 agarose beads for 60 min at 4 °C. Agarose beads were collected by centrifugation followed by heat denaturation of samples and Rac1 activation was evaluated by immunoblotting by anti-human Rac1 antibody.

G-actin/F-actin in vivo biochemical assay

This quantitative assay was performed to determine the relative effect of EGF on filamentous actin (G-actin) versus free globular actin (F-actin) content. Briefly, cells were suspended in F-actin stabilizing buffer and separated from G-actin by ultra-centrifugation as per manufacturer's directions (Cytoskeleton Inc.). The difference in G-actin and F-actin content was examined by western blot using G-actin antibody.

Confocal microscopy

Briefly, treated cells were fixed with 4 % paraformaldehyde at room temperature. Cells were washed with 1× PBS, blocked with 5 % BSA in 1× PBS for 60 min and incubated with Phalloidin-568 overnight at 4 °C. Cells were washed with 1× PBS and mounted using vectashield mounting medium containing DAPI and examined under Olympus FV1000 Filter confocal microscope. Images were acquired with 40× objective and modified using FV10-ASW2.0 software.

Bioluminescent imaging (BLI) and analysis

Nude mice obtained from Charles River Laboratories, were maintained at Ohio State University animal facility under IACUC rules and regulations. Nude mice ($n = 10$) were injected intracardially with MDA-MB-231-luc-D3H2LN-S100A7-luciferase or vector control ($1 \times 10^5/100 \mu\text{L}$) and were weekly assessed for tumor burden (IVIS System 200, Xenogen Corporation). Mice were anesthetized intraperitoneally with 0.15 mg/mL of D-luciferin (PBS) and bioluminescent images were collected between 2 and 5 min post-injection. The light intensity was detected by IVIS camera system, integrated, digitalized, and displayed for relative photon flux as calculated per mouse.

Tissue microarrays (TMA) and immunohistochemical analysis

TMA were obtained from Imgenex (San Diego, CA) and immunohistochemistry (IHC) analysis was performed on paraffin-embedded formalin fixed breast tissue specimens. TMAs were de-paraffinized according to manufacturer's recommendation and immunostained with S100A7 antibody at 1:50 dilution (Imgenex). Vectastain Elite ABC reagents (Vector Laboratories) using avidin DH:biotinylated horseradish peroxidase H complex, 3,3'-diaminobenzidine (Polysciences), and Mayer's hematoxylin (Fisher Scientific) were used for detection of the bound antibodies.

Statistical analysis

All the experiments were performed at least three to four times to confirm the results. The results were then expressed as mean \pm SD of data obtained from these three or four

experiments. The statistical significance was determined by the Student's *t* test and value of $p < 0.05$ was considered significant as denoted by asterisks.

Results

S100A7 overexpression differentially activates EGFR in ER α ⁻ and ER α ⁺ breast cancer cells

It has been shown that S100A7 downregulation inhibits EGFR-mediated signaling in ER α ⁻ cells [15]. Here, we have analyzed the effect of S100A7 overexpression on EGF-induced receptor activation in ER α ⁻ (MDA-MB-231) and ER α ⁺ (MCF-7 and T47D) cells by EGFR phosphorylation. We observed an increase in EGFR phosphorylation in S100A7 overexpressing MDA-MB-231 cells upon EGF treatment (Fig. 1a). However, S100A7 overexpression reduced EGF-induced EGFR phosphorylation in MCF7 cells compared to vector (Fig. 1b). In another ER α ⁺ cell line, T47D, we observed similar results of time-dependent inhibition of EGFR phosphorylation upon EGF stimulation (Fig. 1c). The quantitative analysis of all immunoblots showed consistent increase and decrease in EGFR phosphorylation of S100A7 overexpressing ER α ⁻ and ER α ⁺ cells, respectively (Fig. 1d-f). Therefore, differential EGFR phosphorylation might play an important role in S100A7 overexpressing ER α ⁻ and ER α ⁺ breast cancer cells.

S100A7 overexpression affects cell motility of ER α ⁻ and ER α ⁺ cells

The motile ability of tumor cells determines their meta-static phenotype. In the present study, EGF-induced cell migration was performed to analyze the cell motility of ER α ⁻ and ER α ⁺ cells upon S100A7 overexpression. The wound healing assay revealed the effect of S100A7 in directional cell migration of ER α ⁻ and ER α ⁺ cells. The assay showed S100A7 to significantly increase EGF-mediated migratory abilities of S100A7 overexpressing MDA-MB-231 cells (Fig. 2a). We observed significant increase in wound closure of S100A7 overexpressing MDA-MB-231 cells compared to vector control. In contrast, S100A7 inhibited the directional cell migration of ER α ⁺ MCF-7 cells by relatively slowing down their wound closure compared to vector cells (Fig. 2b). Moreover, cell migration assay using transwell chambers showed ~five fold increase in EGF-induced migration of S100A7 overexpressing MDA-MB-231 cells compared to its vector control (Fig. 2c). SCP6, a single cell progeny of MDA-MB-231 cells, which has previously been characterized as a low metastatic cell line, was also analyzed to evaluate the effect of S100A7 overexpression on cell migration (Supplementary Fig. 1a) [25]. Importantly, S100A7 was able to promote EGF-induced cell migration in SCP6 as well. However, S100A7 overexpression in MCF-7 cells inhibited EGF-induced cell migration by ~eight fold (Fig. 2d). Similar results were seen on S100A7 overexpression in ER α ⁺ T47D cell line, which showed lesser inhibition in cell migration compared to MCF-7 cells (Supplementary Fig. 1b). Taken together, these data reveal the differential consequence of S100A7 on migratory response of breast cancer cells depending on their ER α status.

Role of MMP-9 activation in invasiveness of S100A7 overexpressing ER α ⁻ and ER α ⁺ cells

One of the hallmarks of tumor metastasis is its ability to degrade extracellular matrix to invade distant organs. Matrigel invasion assay was performed to analyze the EGF-induced

invasion of S100A7 overexpressing ER α ⁻ and ER α ⁺ cells. We found that S100A7 overexpression has significantly increased EGF-induced invasive ability of MDA-MB-231 cells compared to vector control by approximately twofold (Fig. 3a). However, there was a considerable decrease in invaded population of S100A7 overexpressing MCF7 cells upon EGF stimulation (Fig. 3b). Similar EGF-mediated inhibition of cell invasion was observed in S100A7 overexpressing T47D cells compared to control cells (Supplementary Fig. 2). The difference in invasion of vector and S100A7 overexpressing cells has been expressed as percentage, which was statistically significant. It is known that cancer cells require matrix metalloproteinases (MMPs) to invade the extracellular matrix underlying their basement membrane and stroma. Hence, we have analyzed the presence of S100A7 expression on activation status of MMP-2 and -9 that have implications in the process of tumor invasion [26]. We observed an increase in active form of MMP-9 secretion in S100A7 overexpressing MDA-MB-231 cells, while its activity was decreased in S100A7 overexpressing MCF7 cells (Fig. 3c, d). However, the MMP-2 activation was not affected in both vector and S100A7 overexpressing MDA-MB-231 and MCF-7 cells. Therefore, diverse MMP-9 activation in S100A7 overexpressing MDA-MB-231 and MCF7 cells suggests the significance of MMP-9 in S100A7 associated invasiveness.

Role of NF- κ B in S100A7-mediated MMP-9 secretion in ER α ⁻ cells

It is known that NF- κ B binding on MMP-9 gene regulates TNF- α -mediated MMP-9 secretion [27]. It has also been reported that S100A7 promotes pro-survival pathways in ER α ⁻ cells through Akt-mediated NF- κ B activation [14]. Since, S100A7 regulates NF- κ B activation in ER α ⁻ cells, we sought to see the effect of NF- κ B knockdown on MMP-9 secretion in S100A7 overexpressing MDA-MB-231 cells. We observed a significant NF- κ B downregulation in S100A7 overexpressing MDA-MB-231 cells (Fig. 3e, f). Active MMP-9 secretion was significantly reduced in NF- κ B-knocked-down cells compared to scramble siRNA control indicating the direct role of NF- κ B in S100A7-mediated MMP-9 secretion (Fig. 3g).

In vivo metastatic potential of S100A7 overexpressing MDA-MB-231 cells and clinicopathological S100A7 expression analysis in breast carcinomas

Since, S100A7 overexpression in breast cancer cell lines has been shown to enhance tumor growth [12], we investigated its significance in metastasis in vivo. In this study, we have used IVIS imaging system to analyze the meta-static potential of S100A7 overexpressing MDA-MB-231 with luciferase reporter gene (Fig. 4a). Higher metastatic progression with elevated radiation flux was observed in intracardially injected nude mice with S100A7 overexpressing MDA-MB-231 cells compared to vector control (Fig. 4b). This demonstrates that S100A7 plays an important role in promoting metastatic phenotype of ER α ⁻ breast cancer cells. Furthermore, we have analyzed the S100A7 expression in a cohort of breast tissue specimens using tissue microarray ($n = 59$). The TMA and IHC data has been summarized as in Supplementary Table 1. Our study revealed that S100A7 expression was highly prevalent in metastatic tumors, especially at lymph node region (Fig. 4c). Ninety percent of metastatic tumors showed good expression of S100A7, while normal breast tissues ($n = 9$) were devoid of S100A7 protein. Interestingly, S100A7 expressing lymph node metastatic group were mostly ER α negative. In addition, we observed predominant S100A7 expression

in ER α ⁻ (~54 %) and PR⁻ (~50 %) compared to ER α ⁺ type (~36 %) and PR⁺ (~44 %) among 35 infiltrating ductal carcinoma carcinomas (Fig. 4d). Hence, the considerable role of S100A7 as a regulator of breast cancer metastasis seems to be directly linked to ER α status.

Role of S100A7 overexpression on actin polymerization

Actin polymerization is a well-known process that drives cell migration with the most evident feature of lamellipodia formation at leading edges of motile cells. Our immunofluorescence studies showed increased actin accumulation at the leading edges of S100A7 over-expressing MDA-MB-231 cells compared to vector upon EGF stimulation (Fig. 5a). However, there was comparatively lesser actin accumulation at the leading edges of S100A7 overexpressing MCF7 cells compared to vector control. Instead, actin seems to be distributed as small actin-filament structures that became more prominent in vector compared to S100A7 overexpressing MCF-7 cells on EGF treatment (Fig. 5b). Actin accumulation at leading edges might be responsible for differential migratory response of S100A7 in ER α ⁻ and ER α ⁺ breast cancer cells. Hence, a direct relationship between activity of actin polymerization and formation of migratory structures could be possible.

S100A7 overexpression affect EGF-induced Rac1 activation and associated signaling

In order to investigate the role of actin polymerization pathway in S100A7-mediated differential effect in ER α ⁻ and ER α ⁺ cells, we have analyzed the activity status of Rac1, which is a key molecule in actin polymerization. Rac1 pathway is downstream of LIMK1/2 and mediates its activation through intermediate kinases like PAK1 [28]. We observed that Rac1 activity was enhanced along with LIMK1/2 expression in S100A7 overexpressing MDA-MB-231 cells compared to vector control cells at all time points (Fig. 6a, c). However, Rac1 activation was inhibited in MCF-7 cells on S100A7 overexpression and even EGF stimulation did not affect its activity suggesting the importance of Rac1 pathway (Fig. 6e, f). We also observed the downregulated expression of LIMK1/2 upon EGF treatment in S100A7 overexpressing MCF7 cells compared to vector control cells (Fig. 6g, h). Consistently enhanced Rac1 activity and LIMK1/2 expression reveal Rac-1 pathway as a modulator of increased migration and metastasis in S100A7 overexpressing MDA-MB-231 cells. However, Rac1 appears to act as a negative regulator in cell migration of S100A7 overexpressing MCF-7 cells.

S100A7 overexpression affects cofilin

Cofilin is one of the downstream molecules of actin polymerization that directly has impact on cell motility through activation of actin-filament dynamics. Cofilin is de-phosphorylated upon Rac1 activation and leads to the polymerization of the F-actin filaments and lamellipodium formation [29]. Our results showed time-dependent decrease in phosphorylation of cofilin in S100A7 overexpressing MDA-MB-231 cells on EGF stimulation compared to vector control cells (Fig. 7a, b). Since p-cofilin is an inactive form of cofilin, its declined level leads to increased actin polymerization–depolymerization and enhanced lamellipodium formation as migratory structures in S100A7 overexpressing MDA-MB-231 cells seen in our immunofluorescence studies. This effect was not seen on cofilin phosphorylation in MCF7 cells upon S100A7 expression suggesting the effect of inactive Rac1 pathway and cofilin on slow mobility of S100A7 overexpressing ER α ⁺ cells

(Fig. 7c, d). Hence, actin-associated regulatory pathway appears to play an important role in migratory potential of S100A7 in ER α ⁻ and ER α ⁺ breast cancer cells.

S100A7 regulated EGF-induced actin turnover

So far, our studies suggest that S100A7-mediated cell motility is controlled by actin polymerization and associated downstream signaling. However, actin-filament subunits (F-actin) need to be recycled back to monomeric forms (G-actin) to maintain further polymerization and motility [30]. Hence, the influence of S100A7 associated actin polymerization on G-actin/F-actin content upon EGF stimulation was examined by an in vivo assay kit. The assay showed more G-actin content with decreased F-actin quantity in S100A7 overexpressing MDA-MB-231 cells on EGF stimulation (Fig. 7e). The decrease in F-actin and simultaneous increase in G-actin content might be responsible for enhancing ATP based actin recycling during actin polymerization, thus significantly enhancing actin turnover with time. Moreover, there was no effect of EGF on F-actin and G-actin content on S100A7 overexpression in MCF7 cells (Fig. 7f). The diverse influence of EGF on G-actin/F-actin content and actin turnover in S100A7 overexpressing ER α ⁻ and ER α ⁺ cells significantly correlated with the activation of actin polymerization pathway.

Discussion

In this study, we report for the first time that S100A7 differentially regulates EGF-induced EGFR phosphorylation and migration of ER α ⁻ and ER α ⁺ breast cancer cells. We observed an increase in EGFR phosphorylation in S100A7 overexpressing ER α ⁻, whereas reduced EGFR phosphorylation was seen in ER α ⁺ cells. The estrogen receptor pathway is known to crosstalk with EGFR pathway and since S100A7 negatively regulates ER, it could be possible that S100A7 may likely inhibit EGF activity in ER α ⁺ cells [13, 16]. However, EGF-induced downregulation of EGFR phosphorylation in S100A7 overexpressing ER α ⁺ cells could also be due to the downregulation of β -catenin/TCF4 pathway shown in previous studies [16, 31]. Therefore, it is reasonable to mention that S100A7-mediated differential EGF receptor activation appears to be regulated through different pathways in ER α ⁻ and ER α ⁺ cells.

Increased invasive and migratory properties are important characteristics of metastatic breast cancer cells. Our previous studies on MVT-1 orthotopic syngeneic bi-transgenic mS100a7a15 mouse model showed enhanced metastasis through M2-macrophage recruitment [23]. EGF/EGFR-axis is known to regulate cell spreading, motility and invasion through extracellular matrix (ECM). In this study, we demonstrate EGF-induced increase in migration and invasion of S100A7 overexpressing MDA-MB-231 cells. In addition, our in vivo nude mouse model study revealed that S100A7 is associated with increased meta-static capacity of ER α ⁻ cells. Furthermore, our studies revealed that S100A7 enhanced NF- κ B-mediated MMP-9 secretion in MDA-MB-231 cells. MMP-9 has been shown to play an important role in breast cancer invasion and metastasis [32]. Consistent with our studies, other S100 gene family proteins also promote tumor metastasis through MMP's activation [33, 34]. Our patient sample data also suggest that S100A7 is widely expressed in metastatic carcinoma, especially in lymph node regions. Interestingly, all S100A7 expressing

metastatic samples were ER α negative providing further evidence of S100A7 involvement in ER α ⁻ tumor metastasis. However, S100A7-mediated effects on cell migration and invasion were inhibited in ER α ⁺ breast cancer cells with decreased MMP-9 activity.

Metastasis is a multi-step process which can be driven by several ways such as actin polymerization, cell adhesion, and acto-myosin contraction [35]. Hence, we have analyzed the influence of S100A7 in ER α ⁻ and ER α ⁺ cells on actin polymerization pathway, which has been extensively studied in cancer metastasis. We revealed an increase in actin polymerization in MDA-MB-231 cells and decrease in MCF7 cells on S100A7 overexpression compared to vector control. Furthermore, we have shown that increased actin polymerization in S100A7 over-expressing ER α ⁻ cells is due to more lamellipodia formation at leading edges that is regulated by Rac1 pathway. Rac1 has been shown to control cofilin phosphorylation through the activity of class II PAKs that is regulated through LIM kinases and other downstream effectors of the Rho family of GTPases, Cdc42, Rac, and Rho [36]. Our results suggest that S100A7 overexpression in ER α ⁻ cells downregulates cofilin phosphorylation with increased LIMK1/2 expression. This can increase the number of barbed ends available during directional cell movement [28, 36]. Therefore, increased LIMK1/2 and cofilin de-phosphorylation mediates enhanced directional migration of S100A7 overexpressing ER α cells. However, downregulated LIMK1/2 expression and presence of inactive phosphorylated form of cofilin appears to inhibit local actin polymerization at leading edges and reduced EGF-induced migration in S100A7-overexpressing ER α ⁺ cells. Interestingly, prominent cytoplasmic staining of actin filaments in S100A7 overexpressing MDA-MB-231 cells on EGF stimulation could be explained by increased actin turnover. Our in vivo G-actin/F-actin assay demonstrates that G-actin/F-actin ratio regulates actin turnover, which is maintained by S100A7-mediated activation of Rac1 pathway on EGF stimulation in MDA-MB-231 cells. However, ineffective actin turnover in MCF-7 cells could be due to inhibited actin regulatory Rac1 pathway.

In summary, our proposed model describes the EGF-induced differential role of S100A7-mediated actin remodeling and MMP-9 in ER α ⁺ and ER α ⁻ breast cancer cells (Supplementary Fig. 3). S100A7 was expressed predominantly in lymph node ER α ⁻ metastatic tumors. Its overexpression enhanced in vivo metastasis of ER α ⁻ cells. Furthermore, our studies suggest that S100A7 regulates breast cancer metastasis through a novel pathway by modulating actin cytoskeleton and MMP-9 activation. Since metastasis is the leading cause of cancer-related deaths, S100A7 could be a novel therapeutic target in ER α ⁻ metastatic breast cancer.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

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Abbreviations

S100A7	Psoriasin
ERα	Estrogen receptor α
EGF	Epidermal growth factor
DCIS	Ductal carcinoma in situ
IHC	Immunohistochemistry

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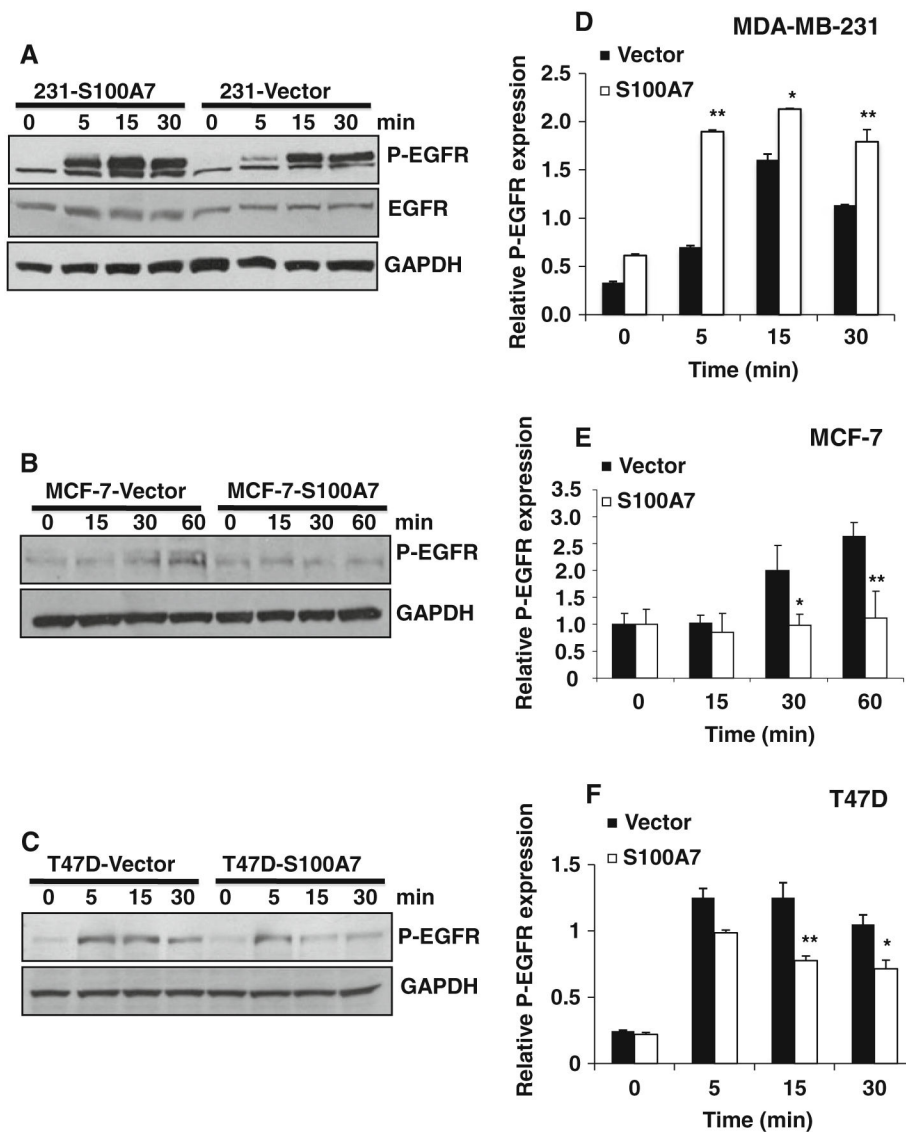


Fig. 1. EGF-induced differential phosphorylation of EGFR in ER α ⁻ and ER α ⁺ breast cancer cells with S100A7 overexpression. EGFR phosphorylation status was analyzed in S100A7 overexpressing ER α ⁻ MDA-MB-231 cells (a) and ER α ⁺ MCF-7 (b) and T47D cells (c) on EGF stimulation (100ng/mL) by western blotting. The relative EGFR phosphorylation levels in all cells are representative of three independent experiments, calculated with respect to GAPDH for MDA-MB-231, MCF-7, and T47D (d-f) at indicated time. The statistically significant *p* values were indicated as * <0.05 and ** <0.005

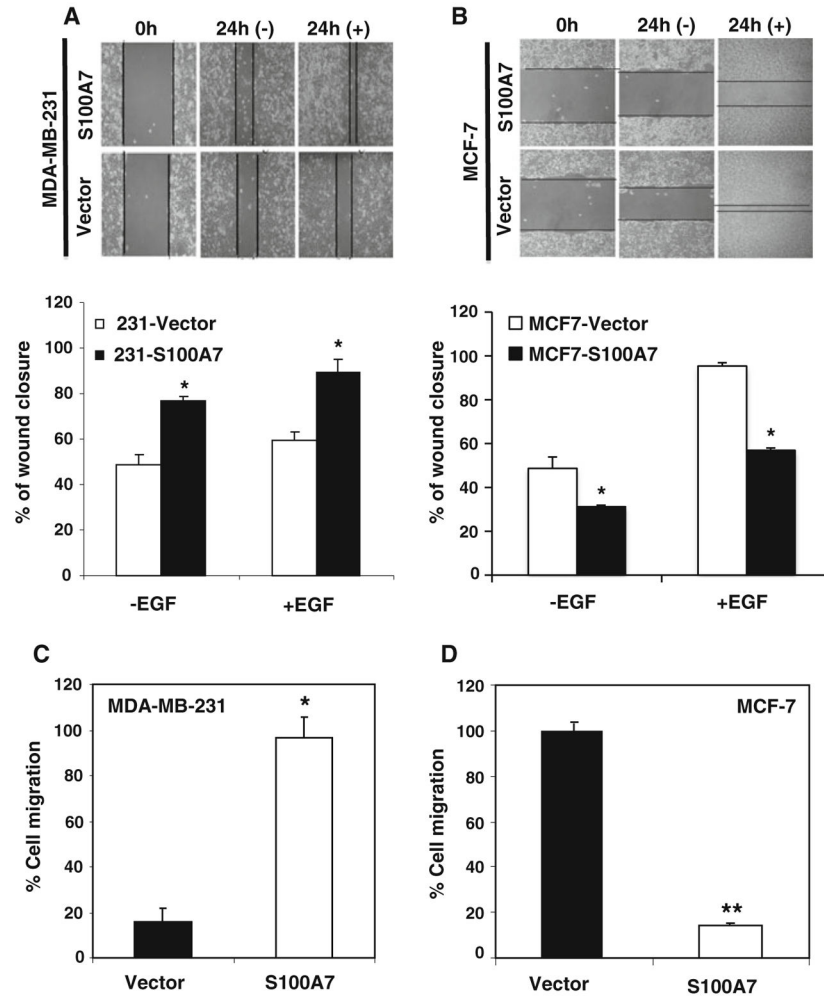
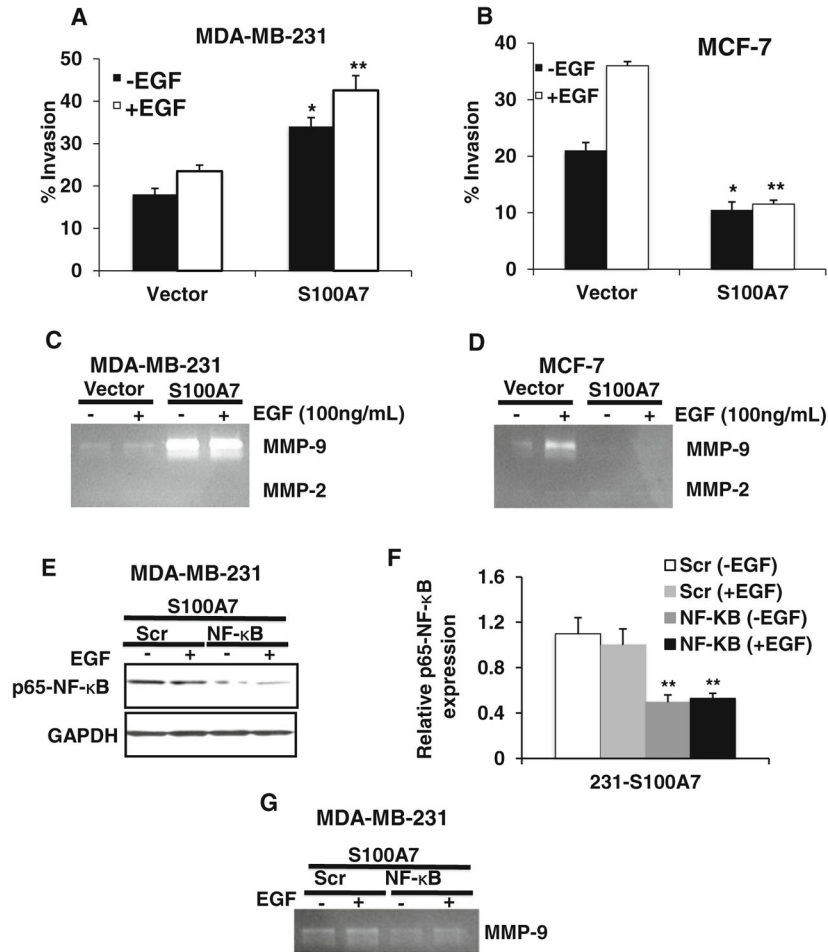


Fig. 2. Effect of S100A7 on EGF-induced cell motility, migration, and invasion of ER α ⁻ and ER α ⁺ cells. Wound healing assay images represent EGF-induced effect on cell motility of S100A7 overexpressing MDA-MB-231 and MCF-7 cells compared to their vector control (**a**, **b**). Their quantitative analysis represent one of the three independent experiments performed with p value denoted as $* < 0.05$. Relative migratory potential of S100A7 overexpression in MDA-MB-231 (**c**) and MCF-7 cells (**d**) was compared to vector by transwell migration assay, represented as percent cell migration with P value as $* < 0.05$ and $** < 0.005$. EGF-induced effects were analyzed at 100ng/mL concentration for cell motility assays

**Fig. 3.**

Role of NF- κ B in S100A7-mediated MMP-9 secretion in ER α^- and ER α^+ cells. The effect of EGF on relative invasiveness of MDA-MB-231 (a) and MCF-7 cells (b) on S100A7 overexpression was analyzed by matrigel invasion assay and represented as percent invasion with p value <0.05 as * and <0.005 as **. Gelatin zymography revealing the level of MMP-9 secretion in MDA-MB-231 cells (c) and MCF-7 cells (d) upon S100A7 overexpression without any effect on MMP-2 levels. Images represent one of the three independently performed experiments. Knockdown of p65-NF- κ B subunit (e, f) markedly inhibited the MMP-9 secretion of MDA-MB-231 cells analyzed by gelatin zymography (g). Images represent one of the three experiments and their quantitative analyses were statistically significant with p value $* < 0.05$ and $** < 0.005$. Where, EGF used was 100ng/ mL and *Scr* stands for scramble siRNA

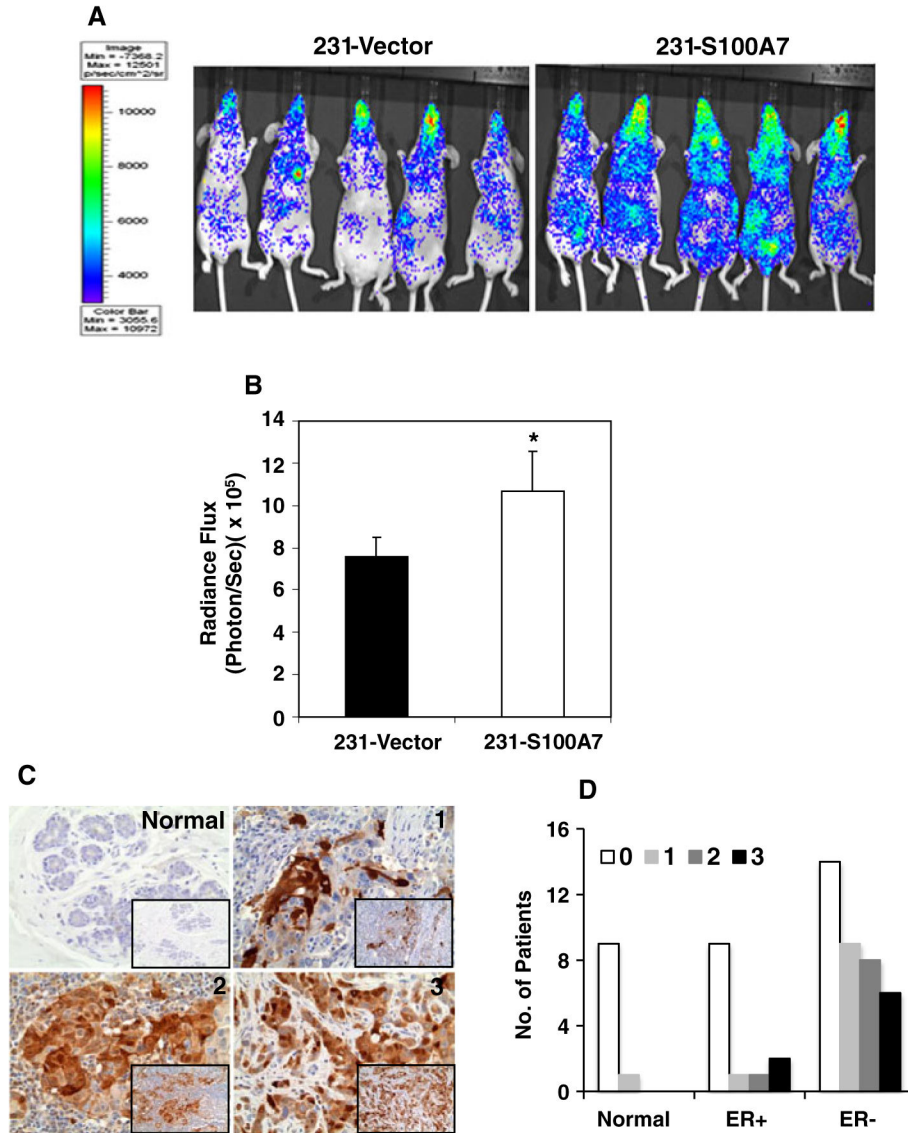


Fig. 4. In vivo metastatic potential of S100A7 in ER α ⁻ cells and its association with ER α status. Representative BLI images show comparative metastases of control and S100A7 overexpressing MDA-MB231-luc-D3H2LN cells in nude mouse model (a), while the statistically significant high radiance flux was observed in the presence of S100A7 with *p* value <0.05 (b). Representative immunohistochemistry images show different levels of S100A7 staining pattern in lymph node region of metastatic tissue specimens compared to normal tissue, as a negative control (c). Based on clinico-pathologist analysis, the overall S100A7 expression levels were quantitatively represented in ER α ⁻ and ER α ⁺ cells compared to normal breast tissues (d). Where 0, 1, 2, and 3 represent different S100A7 expression levels

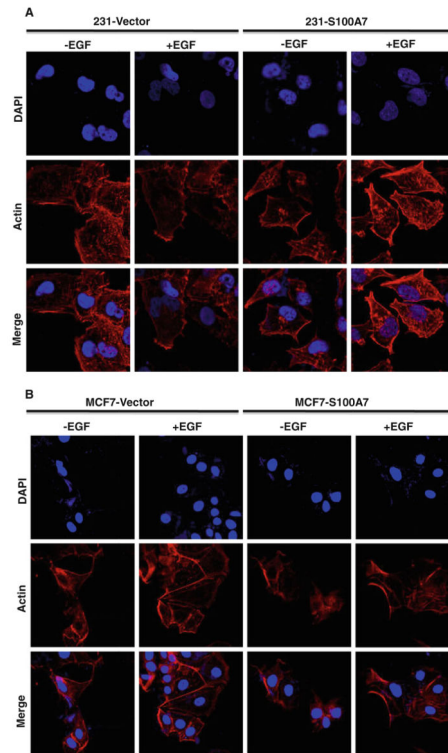


Fig. 5. Effect of EGF on actin polymerization of S100A7 overexpressing ER α ⁻ and ER α ⁺ cells. Immunofluorescence images showing EGF-induced consequences on actin-based cell protrusions of MDA-MB-231 (**a**) and MCF-7 cells (**b**) on S100A7 expression. Images represent one of three independent experiments with phalloidin-568 and DAPI staining as indicated at 100ng/ mL of EGF treatment

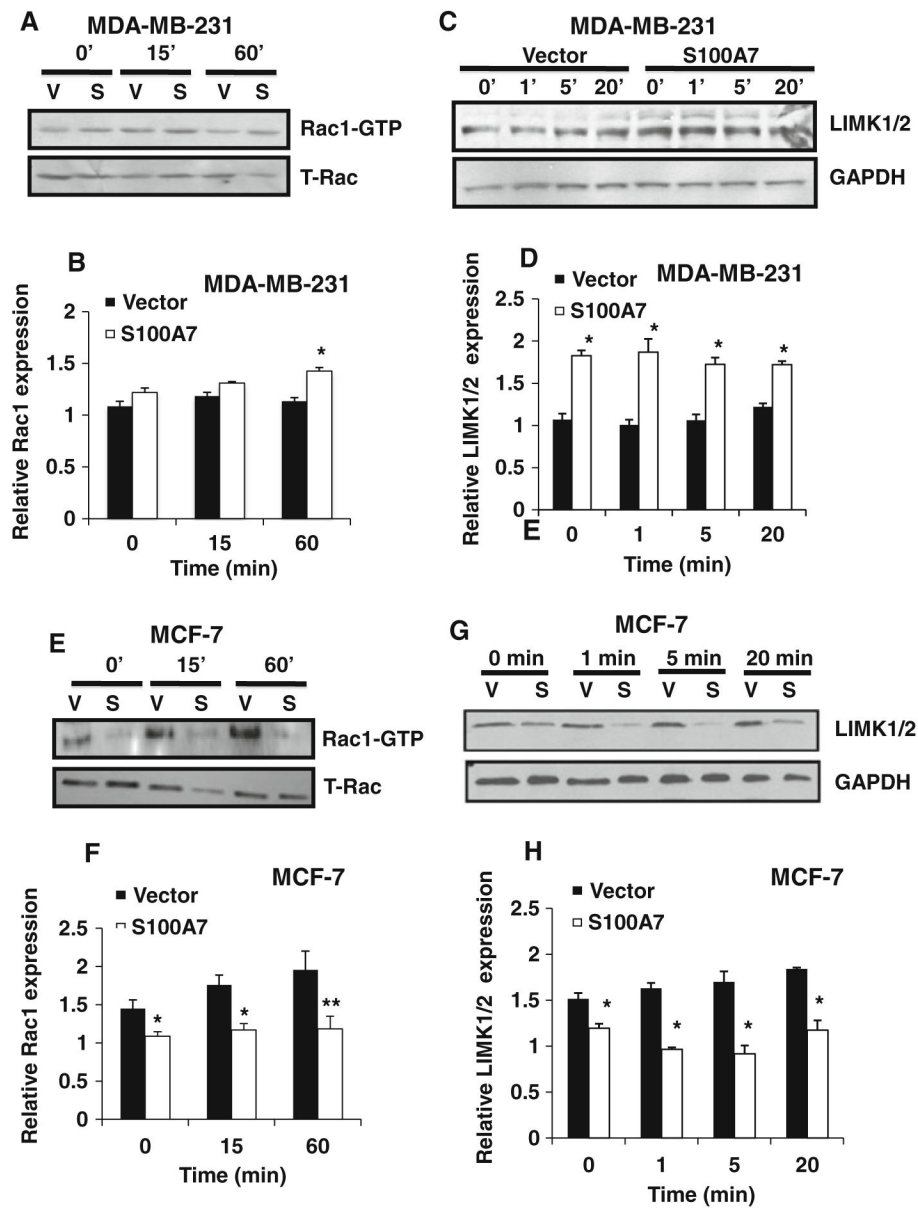
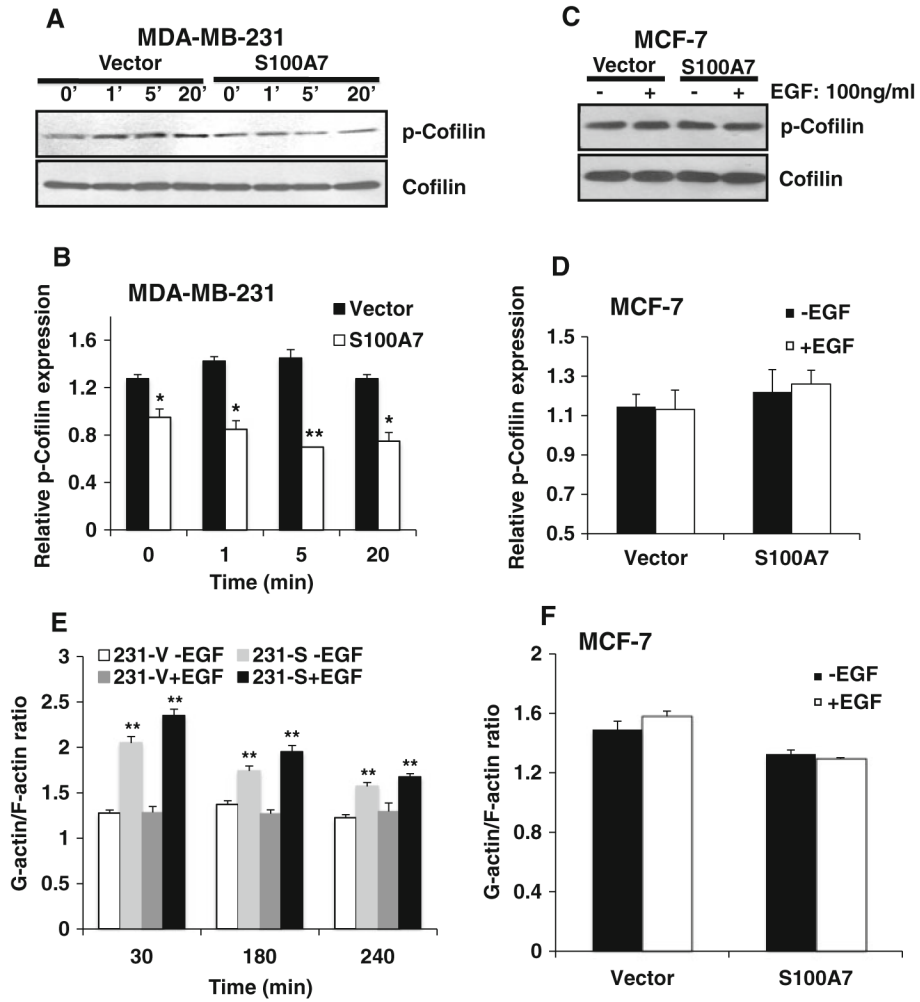


Fig. 6. Rac-1 signaling in EGF-induced cell motility of S100A7 overexpressing ER α^- and ER α^+ cells. Actin regulators, Rac-1 and LIMK1/2 activation status were evaluated on EGF stimulation (100ng/mL) in S100A7 overexpressing MDA-MB-231 (**a, c**) and MCF-7 cells (**e, g**) by western blot. Their quantitative analysis corresponds to three independent repeats as indicated by *asterisks* * and ** with significant *p* value <0.05 and <0.005, respectively (**b, d, f, h**). Where, V stands for vector and S stands for S100A7

**Fig. 7.**

Role of Cofilin signaling on actin remodeling of S100A7 overexpressing ER α ⁻ and ER α ⁺ cells. Phosphorylation status of cofilin as a negative actin regulator was assessed to show effect of EGF on S100A7 overexpressing MDA-MB-231 (a) and MCF-7 cells (c). The relative phosphorylation levels appear to be significant with p value $* < 0.05$ and $** < 0.005$ (b, d). Effect of EGF-induced actin regulation in S100A7 overexpressing MDA-MB-231 (e) and MCF-7 (f) cells were quantitatively evaluated by in vivo actin turnover assay as a change in G-actin/F-actin ratio over the indicated time. Where EGF used was 100 ng/mL