

### rstb.royalsocietypublishing.org

## Review



**Cite this article:** Hileman LC. 2014 Trends in flower symmetry evolution revealed through phylogenetic and developmental genetic advances. *Phil. Trans. R. Soc. B* **369**: 20130348. http://dx.doi.org/10.1098/rstb.2013.0348

One contribution of 14 to a Theme Issue 'Contemporary and future studies in plant speciation, morphological/floral evolution and polyploidy: honouring the scientific contributions of Leslie D. Gottlieb to plant evolutionary biology'.

#### Subject Areas:

developmental biology, evolution, plant science

#### Keywords:

flower symmetry, *CYCLOIDEA*, parallel evolution, character evolution, evolution of development

#### Author for correspondence:

Lena C. Hileman e-mail: lhileman@ku.edu

## Trends in flower symmetry evolution revealed through phylogenetic and developmental genetic advances

### Lena C. Hileman

Ecology and Evolutionary Biology, University of Kansas, 1200 Sunnyside Avenue, Lawrence, KS 66045, USA

A striking aspect of flowering plant (angiosperm) diversity is variation in flower symmetry. From an ancestral form of radial symmetry (polysymmetry, actinomorphy), multiple evolutionary transitions have contributed to instances of non-radial forms, including bilateral symmetry (monosymmetry, zygomorphy) and asymmetry. Advances in flowering plant molecular phylogenetic research and studies of character evolution as well as detailed flower developmental genetic studies in a few model species (e.g. Antirrhinum majus, snapdragon) have provided a foundation for deep insights into flower symmetry evolution. From phylogenetic studies, we have a better understanding of where during flowering plant diversification transitions from radial to bilateral flower symmetry (and back to radial symmetry) have occurred. From developmental studies, we know that a genetic programme largely dependent on the functional action of the CYCLOIDEA gene is necessary for differentiation along the snapdragon dorsoventral flower axis. Bringing these two lines of inquiry together has provided surprising insights into both the parallel recruitment of a CYC-dependent developmental programme during independent transitions to bilateral flower symmetry, and the modifications to this programme in transitions back to radial flower symmetry, during flowering plant evolution.

### 1. Introduction

Variation in flower symmetry has attracted the attention of botanists for more than a century [1–4]. Research has centred on understanding the developmental mechanisms that establish patterns of symmetry, the ecological contexts in which alternative patterns of symmetry are favoured, and the evolutionary history of transitions between different forms. This research has provided key insights into how, when and why transitions in floral symmetry evolve.

During the diversification of flowering plants (angiosperms), there have been numerous evolutionary transitions between radial flower symmetry (polysymmetry, actinomorphy; figure 1*a*) and bilateral flower symmetry (monosymmetry, zygomorphy; figure 1*a*), or in more extreme cases, flower asymmetry (figure 1*c*) [5,6]. Bilateral symmetry is predominant in a number of species-rich lineages for example, Lamiales (mints and allies) and Fabaceae (legumes) in eudicots, and Orchidaceae in the monocots. Bilateral symmetry in these lineages is not only common, but also highly elaborate. However, a survey of flowering plant lineages demonstrates that both elaborate and subtle forms of bilateral flower symmetry have evolved from radially symmetrical ancestors many times, and that reversals from bilateral to radial, or approximately radial symmetry are not uncommon (reviewed in [6]).

It is generally accepted that these transitions in flower symmetry are associated with pollination syndromes. For example, transitions from radial to bilateral flower symmetry appear to be linked to the evolution of specialized plant–pollinator interactions. Bilateral symmetry is most often evident in the petal and stamen whorls and may promote pollinator approach and legitimate (pollen transferring) landings, and may increase the specificity of pollen deposition during pollinator visits [7–9]. In part because of the relationship between





bilateral symmetry

**Figure 1.** Flower symmetry diversity and bilateral flower symmetry developmental genetics. The range of floral symmetries include radial symmetry with multiple planes of mirror image symmetry (*a*, *Potentilla* sp.), disymmetry with two planes of mirror image symmetry (*b*, *Cardaminopsis arenosa*), asymmetry with zero planes of mirror image symmetry (*c*, *Pedicularis racemosa*) and bilateral symmetry with just a single plane of mirror image symmetry (*d*, *Antirrhinum majus*). At the developmental level, one or more genetic signals must differentiate the dorsal (adaxial) from the ventral (abaxial) domains of the developing flower, for example a genetic programme that distinguishes dorsal identity (*e*, dorsal shading in cartoon of early developing flower). In the model species *A. majus*, the genetic programme that establishes dorsoventral flower identity from early stages of development includes the dorsal identity genes and protein products CYCLOIDEA (CYC), DICHOTOMA (DICH) and RADIALIS (RAD) as well DIVARICATA (DIV) which specifies ventral flower development (*f*). DIV is excluded from the dorsal domain of the developing *A. majus* flower through post-translational negative regulation by RAD. *C. arenosa* flower is taken from image for which copyright is held by Meneerke bloem (http://commons.wikimedia.org/wiki/File:Cardaminopsis\_arenosa\_02.jpg). This image is used under a Creative Commons Attribution-Share Alike 3.0 Unported, 2.5 Generic, 2.0 Generic and 1.0 Generic licence. *P. racemosa* flower taken from image for which copyright is held by Jerry Friedman (http://commons.wikimedia.org/wiki/File:Pedicularis\_racemosa\_1.jpq). This image is used under the Creative Commons Attribution-Share Alike 3.0 Unported, wiki/File:Pedicularis\_racemosa\_1.jpq). This image is used under the Creative Commons Attribution-Share Alike 3.0 Unported licence. (Online version in colour.)

symmetry and specialized pollination biology, transitions to bilateral flower symmetry are hypothesized to represent key innovations associated with diversification of species-rich flowering plant lineages [10,11].

More recently, much attention has turned to the developmental programmes that specify bilateral flower symmetry. Now, many genes and genetic interactions necessary for the development of bilateral flowers are understood from the model species *Antirrhinum majus* (snapdragon, reviewed in [12]). Because of the historical interest in floral symmetry pollination ecology and evolution, these newer insights from *A. majus* provide the jumping board for comparative studies. The primary comparative question has been whether the developmental programme identified in *A. majus* contributes to the establishment of bilateral symmetry in other flowering plant lineages. Strikingly, current evidence suggests that a similar developmental programme, first identified in *A. majus*, has been recruited many times independently during the parallel evolution of bilateral flower symmetry (reviewed in [12–16]).

Linking model system findings (e.g. in *A. majus* flower development) to comparative developmental questions is not a new concept. Leslie Gottlieb (1936–2012) was an early and strong proponent of the idea that developmental genetic studies in model plant species can inform our understanding of natural variation in flower form [17,18]. For example, he furthered the hypothesis that induced mutations, identified early in the establishment of *A. majus* as a model species [19], may provide genetic information about floral traits that distinguish species or genera. With respect to floral symmetry, he recognized that *A. majus* mutants with

increased petal and/or stamen number and radial flower symmetry may be significant for understanding *Verbascum* flower development. Likewise, he pointed out that *A. majus* mutants with a reduced corolla limb and tubular configuration show similarities to *Rhinanthus* flowers [18]. Similar to Gottlieb's early examples, many current comparative flower developmental genetic (evo-devo) studies aim to test how variation in genetic pathways, identified largely through loss or gain of function mutations in model plant species, may explain natural variation in flower form.

Extrapolating from model system studies of flower development to the genetic basis of interspecific variation in flower form is best approached in a phylogenetic framework. A wellresolved hypothesis of phylogenetic relationships among flowering plant species allows assessment of ancestral character states, and pinpoints evolutionary transitions towards or away from character states of interest. The past 40 plus years of molecular phylogenetic studies in flowering plants have provided and continue to provide this critical framework (reviewed in [20]). Early on, Gottlieb embraced molecular tools for plant phylogenetic and evolutionary studies [21-25]. His contributions made an impact that provided momentum to the field, and this momentum has not waned. We now have a clearer understanding of relationships among major lineages of flowering plants (figure 2a and see [26-28]); likewise, molecular phylogenetic studies have contributed to resolution of relationships within many key lineages (e.g. figure 2b and [29-32]). With respect to flower symmetry development and evolution, the products of molecular phylogenetic studies allow researchers to determine



**Figure 2.** Evolutionary transitions in floral symmetry in a phylogenetic framework. (*a*) Phylogeny of major angiosperm lineages (from [26]). Lineages in which elaborate bilateral flower symmetry can be found (from [6]) are in red text (grey text in print version). Stars indicate lineages containing species for which *CYCLOI-DEA* homologues have been implicated in transitions to bilateral flower symmetry. (*b*) Phylogeny of representative Lamiales lineages (from [31]). One possible parsimonious history of floral symmetry evolution is shown suggesting multiple transitions from radial to bilateral flower symmetry early in Lamiales diversification, followed by multiple transitions from bilateral to radial (or approximately radial) flower symmetry. Lineages with radial flower symmetry are in black/bold; those with bilateral flower symmetry are in red or grey/not bold. Taxa were scored at the species level (see Schaferhoff *et al.* [31] for complete taxon list). Species exhibiting elaborate bilateral symmetry in the corolla and/or androecium were scored as having bilaterally symmetrical flowers. (Online version in colour.)

how often and in which lineages transition from radial flower symmetry to bilateral symmetry (and back to radial flower symmetry) have occurred [16,31,33–37], thus providing the framework for informed choice of species when addressing comparative developmental questions. Here, I review some recent advances in understanding flower symmetry evolution. I address multiple important contributions of molecular phylogenies to the field. Additionally, I demonstrate how the past 10 years of linking model system findings to comparative developmental questions

has shed light on the extensive developmental parallelism in independent transitions between flower symmetry forms.

## 2. Diversity in floral symmetry

Although the focus of this review is evolutionary transitions between radial and bilateral flower symmetry (figure 1a,d), it is important to recognize that these two symmetry forms represent only part of the diversity in symmetry found across flowering plants. Flower symmetry is generally assessed via the face-on view of a flower at the time of anthesis, and is usually expressed most strongly in the petal and stamen whorls of the flower. Radially symmetrical flowers (figure 1a) display several planes of symmetry that bisect the flower into mirror images, and bilaterally symmetrical flowers (figure 1d) display just a single plane of mirror image symmetry. However, flowers may be disymmetrical (figure 1b), having just two planes of mirror image symmetry, or asymmetrical (figure 1c), lacking altogether a plane of symmetry that bisects the flower into mirror images. Interestingly, bilateral symmetry may often be an intermediate state between radial symmetry and asymmetry. For example, asymmetric *Pedicularis* (figure 1c) and asymmetric Phaseolus and Lathyrus are nested within Lamiales (figure 2b) and Faboideae, respectively, two flowering plant lineages in which bilateral flower symmetry is predominant. Likewise, multiple forms of bilateral flower symmetry are derived from disymmetry. For example, bilaterally symmetrical Iberis (Brassicaceae) and Corydalis (Papaveraceae) are derived from ancestors with disymmetrical flowers [38-41].

Bilateral flower symmetry itself can range from elaborate to subtle patterns of low complexity (reviewed in [6]). Most familiar forms of complex bilateral flower symmetry are the bilabiate (lipped and keeled) flower forms. In bilabiate flowers, reproductive organs (stamens and carpels) are found inside an elaborate corolla that is differentiated along the dorsal/ventral (adaxial/abaxial) floral axis. In lip flowers, the reproductive organs are held in the upper side of the corolla resulting in pollen transfer on the backs of visiting pollinators; in keel flowers, the reproductive organs are held in the lower (keel) side of the corolla resulting commonly in pollen transfer on the underside of pollinators. Bilaterally symmetrical flowers of the lip form are extremely prevalent in Lamiales (e.g. A. majus, figure 1d), but are found in other lineages, including Campanulales and Orchidaceae. Those of the keel form are well known from Fabaceae, but can also be found in Polygalaceae. Less elaborate forms of bilateral flower symmetry also result from organ differentiation primarily in the petal and/or stamen whorls, and may be due to displacement of organ initiation, size or shape variation in organs along the dorsoventral axis of the flower, or sigmoidal curvature of organs (reviewed in [6,16]).

## 3. A phylogenetic context for floral symmetry evolution

From assessments of taxonomic distribution of bilateral flower symmetry [1], and variation in the form of bilaterally symmetrical flower (e.g. lip versus keel bilabiate flowers), it has historically been quite clear that transitions from radial to bilateral flower symmetry were probably frequent during flowering plant diversification. However, it is only in the context of robust phylogenetic hypotheses for the relationships among flowering plant lineages that we can determine along which lineages evolutionary transitions from radial to bilateral (and back to radial) flower symmetry have occurred [42]. And it is primarily advances in molecular phylogenetics that provide the context for studies of floral symmetry character evolution.

Studies that have used molecular phylogenies to reconstruct the ancestral flower conclude that it was radially symmetrical [43]. A clear understanding of the ancestral form of symmetry is an excellent starting point for determining where bilateral flower symmetry has been gained or lost in flowering plants. A number of recent molecular phylogenies that sample taxa at approximately the family level are now being used to assess patterns of floral character evolution, including symmetry [26-28]. Figure 2a shows the ordinal-level backbone phylogeny from Soltis et al. [26] on which orders containing species with more or less elaborate bilateral flower symmetry [6] are indicated. This is by no means a critical evaluation of floral symmetry evolution, but illustrates the widely dispersed nature of transitions to bilateral flower symmetry. Citerne et al. [16] undertook an excellent analysis of floral symmetry evolution on the estimate of flowering plant family relationships presented in Bremer et al. [27]. Using a parsimony approach, and scoring for flower symmetry at the family level (which is likely to underestimate the number of transitions to bilateral flower symmetry), they identified a single transition to bilateral flower symmetry among the basal angiosperms, 23 transitions in monocots, and 46 independent transitions in the eudicots. Therefore, using a well resolved and densely sampled (at the family level) estimate of flowering plant phylogeny, Citerne et al. [16] suggest at least 70 transitions to bilateral flower symmetry-twice as many as previously reported.

Studies of character evolution on large-scale phylogenies, such as the one undertaken by Citerne et al., represent important advances in our understanding of floral evolution. Ideally, as advances are made in molecular phylogenetics, we will have at our disposal estimates of the flowering plant phylogeny that are densely sampled at the genus (or even species) level, and for which phylogenetic branch length estimates are available. It will be in this context that floral symmetry evolution will be most critically evaluated using statistical methods for ancestral state reconstruction [44]. That bilateral flower symmetry is a key innovation leading to increased diversification rates has been hypothesized, and to a limited extent tested [10,11]. As with studies of character state evolution, it will be in this context of densely sampled phylogenies that the relationships between shifts in flower symmetry and clade diversification will best be investigated [45]. Excitingly, researchers are anticipating these large datasets. For example, both the National Evolutionary Synthesis Center (NESCent)-supported working group 'Floral assembly: quantifying the composition of a complex adaptive structure' (http:// www.nescent.org/science/awards\_summary.php?id=90) and eFLOWER (http://eflower.myspecies.info/) are developing massive data matrices of floral traits, including floral symmetry, scored at the species level.

If we move our focus from the entire clade to specific lineages within flowering plants, then we find that more fully resolved assessments of floral symmetry evolution are possible. This more focused view will certainly suggest additional transitions to and from bilateral symmetry to those that would be seen on an ordinal- or family-level sampled phylogeny of flowering plants. For example, Schaferhoff et al. [31] generated a densely sampled, well-resolved phylogeny of Lamiales. Scoring for corolla symmetry at just the family level of their backbone phylogeny, they recover one transition from radial to bilateral symmetry, and one transition back to radial symmetry. Using the same phylogeny, but scoring for corolla and stamen whorl symmetry at the species level, based on the species sampled in the Schaferhoff et al. [31] phylogeny, I recover possibly two transitions from radial to bilateral flower symmetry early in Lamiales diversification (considering the bilateral symmetry in the stamen whorl of many Oleaceae species), and multiple transitions from bilateral to radial flower symmetry (figure 2b). Others have undertaken similar analyses of floral symmetry evolution in large flowering plant lineages, scoring symmetry for genera or species. Some key findings are multiple transitions from radial to bilateral symmetry inferred during Solanaceae [29,35], Brassicaceae [38] and Ranunculales [46] diversification. By contrast, in Malpighiales, a single transition to bilateral flower symmetry is recovered, followed by multiple transitions from bilateral to radial symmetry [37,47].

## 4. Developmental genetics of floral symmetry

As described above, bilateral flower symmetry has evolved multiple times and its form varies in complexity. Research in the model species *A. majus*, with its elaborate bilabiate form (figure 1*d*), provided the first ground-breaking insights into the genetic control of bilateral flower symmetry. At the foundation of this control is a programme that differentiates dorsal (adaxial) and ventral (abaxial) flower identity from very early stages of floral organ initiation and differentiation (figure 1*e*).

Two recently duplicated TCP (Teosinte branched 1/Cycloidea/proliferating cell factors) family transcription factors [48-51], CYCLOIDEA (CYC) and DICHOTOMA (DICH), function partially redundantly to specify dorsal flower identity (figure 1f) [52,53]. These paralogues represent the upstream extent of our knowledge of dorsal flower specification. In other words, we do not yet know what gene products control the regulation of CYC and DICH. Expression of CYC and DICH corresponds with their function in specifying dorsal flower identity. Both are expressed in the dorsal region of the floral meristem from initiation, and their dorsal-restricted expression is maintained throughout petal and stamen development [52-54]. CYC and DICH expression and function in the dorsal flower domain are necessary for establishing the distinct shape of dorsal petals (figure 1d), abortion of the dorsal (medial) stamen, as well as petal and stamen merosity. In an A. majus cyc;dich double mutant background, flowers completely lack dorsal identity, are radially symmetrical and develop with ventral identity in the ventral, lateral and dorsal domains [52,53]. CYC and DICH appear to determine the distinct shape of dorsal petals and the formation of the dorsal staminode by affecting patterns of cell growth and proliferation. This is in line with the widely recognized function of TCP transcription factors in promoting and/or repressing tissue growth (reviewed in [12,51]).

While *CYC* and *DICH* are necessary to differentiate dorsal floral identity, a single MYB family transcription factor, *DIVARICATA* (*DIV*), functions to specify ventral identity (figure 1*f*) [54,55]. *DIV* expression and function in the ventral flower domain are necessary for establishing the shape of the

ventral (medial) petal, which distinctly contributes to the lower lip of the bilabiate *A. majus* flower (figure 1*d*). Interestingly, in early flower development, *DIV* is expressed in both the dorsal and ventral domains of the flower, but its expression becomes somewhat restricted to the developing ventral petal at later stages of development [54]. The effects of *CYC* and *DICH* on dorsal flower development and of *DIV* on ventral flower development are in part mediated through an additional MYB transcription factor, *RADIALIS (RAD). RAD* expression is positively regulated by *CYC* and *DICH*. Therefore, *RAD* expression and function are primarily restricted to the dorsal domain of developing flowers (figure 1*f*) [56,57]. It is RAD protein in the dorsal flower domain that posttranslationally restricts DIV function to the ventral domain (figure 1*f*) [55,56,58,59].

# 5. Parallel recruitment of a *CYC*-dependent pathway in bilateral symmetry evolution

From extensive molecular phylogenetic work and studies of character evolution in flowering plants, we have a clearer understanding of the history of flower symmetry evolution. Additionally, from research on flower development in A. majus, we know at least one way by which flower symmetry can be established at the molecular level. Together, these provide a foundation for comparative developmental studies. Bilateral flower symmetry evolved early in the diversification of Lamiales (figure 2b); therefore, bilateral symmetry in A. majus is homologous to bilateral flower symmetry found in other Lamiid lineages (with the possible exception of Oleaceae; figure 2b). A reasonable, testable hypothesis is that the A. majus CYC/RAD/DIV developmental programme (figure 1f) evolved early in Lamiales and is conserved among relatives of A. majus with bilateral flower symmetry. In addition, either similar or divergent genetic programmes may have been recruited to specify independent origins of bilateral flower symmetry elsewhere in flowering plants (figure 2a). Possibilities include independent recruitment of a CYC-dependent programme to specify dorsal or ventral identity (figure 3a,b), or novel recruitment of a CYC-independent developmental programme to specify either dorsal or ventral flower identity (figure  $3c_{d}$ ). Results from many comparative studies now demonstrate that there is striking parallelism in the independent evolution of bilateral symmetry with a CYC-dependent programme frequently recruited to specify dorsal identity, and in some cases ventral identity (reviewed in [13,15,16]).

### (a) Asterids

Limited data support the hypothesis that the *CYC/RAD/DIV* programme is conserved across Lamiales. In bilaterally symmetrical flowers of *Veronica* and *Gratiola* (belonging to the same family as *A. majus*, Plantaginaceae), *CYC* and *RAD* homologues are expressed in the dorsal regions of flowers and with nearly identical spatial distributions, suggesting conservation of positive regulation of the *RAD* gene by CYC protein [60]. This is also true in *Chirita* and *Bournea* from the early diverging Lamiales lineage Gesneriaceae [61,62]. Whether post-translational negative regulation of DIV by RAD protein is conserved in Lamiales is not clear and difficult to test, because analyses of *DIV* transcript localization will not reflect where in the flower DIV protein is present and



localization of a *CYC*-independent developmental programme specifying radial flower symmetry derived from *CYC*-dependent bilateral flower symmetry

**Figure 3.** Hypothesized *CYC*-dependent and *CYC*-independent pathways for recurrent evolutionary transitions from radial to bilateral, and bilateral to radial flower symmetry. (a,b) A *CYC*-dependent programme is necessary for the development of bilateral flower symmetry through the specification of dorsal or ventral identity, respectively. (c,d) A *CYC*-independent programme is necessary for the development of bilateral flower symmetry through the specification of dorsal or ventral identity, respectively. (c,d) A *CYC*-independent programme is necessary for the development of bilateral flower symmetry through the specification of dorsal or ventral identity, respectively. (e,f) Radial flower symmetry is derived from *CYC*-dependent bilateral symmetry through loss of the dorsoventral restricted *CYC*-dependent programme. (g,h) Radial flower symmetry is derived from *CYC*-dependent bilateral symmetry through an independent programme that compensates for the effects of the *CYC*-dependent programme. (Online version in colour.)

functional (although see [62]). In addition to limited information on the conservation of the *CYC/RAD/DIV* programme within Lamiales, few studies have investigated the regulatory interactions among these genes/gene products in other asterid lineages (but see [63,64] summarized below), especially those most closely related to Lamiales (e.g. Boraginaceae, Solanales, Gentianales; figure 2*a*). In the distantly related model species *Arabidopsis thaliana* (rosid lineage), *CYC-* and *RAD-*like genes and gene products do not seem to be directly regulated by one another [57,65], but the phylogenetic distance makes it difficult to draw conclusions about when the *CYC/RAD* network interactions evolved.

Elsewhere within asterids, the role of *CYC*-like genes in independent transitions to bilateral flower symmetry has been investigated in Dipsacales and Asterales (figures 2*a* and 4). In the bilaterally symmetrical flower of *Lonicera* (Caprifoliaceae, Dipsicales), duplicate *CYC*-like genes are expressed in the dorsal or dorsal plus lateral petals (figure 4). This is in striking contrast to the radially symmetrical flowers of related *Viburnum* (Adoxaceae, Dipsacales) where these *CYC*-like orthologues show no pattern of differential expression across the floral axis. Interestingly, a *Lonicera RAD* orthologue is expressed similarly to one of the *Lonicera CYC* paralogues, providing some indication that the *CYC/RAD* regulatory interaction may have been established early in asterid evolution and retained in both Dipsacales and Lamiales, but this hypothesis requires extensive further testing.

Multiple lines of evidence demonstrate that *CYC*-like genes play a role in establishing the developmental differentiation of ray flowers (bilaterally symmetrical) from disc flowers (radially symmetrical) in Asteraceae inflorescences (capitula). *CYC* homologues in *Helianthus, Senecio* and *Gerbera* are preferentially expressed around the periphery of the capitulum where ray flowers are expected to develop, but either at low levels, or not at all in the region of disc flower development (figure 4) [66–68]. In *Helianthus* (sunflower), naturally occurring mutations transform disc flowers to ray flowers (double-flowered mutants), and ray flowers to disc-like tube flowers (tubular-rayed mutants). Double-flowered mutants are due to mutations that cause overexpression of a *CYC*-like gene in the region of disc flower development, thus causing their transformation to ray identity [66]. Similarly, transgenic overexpression of a *CYC* homologue in *Gerbera* leads to transformation of disc flowers to ray identity [68]. Tubular-rayed mutants are due to loss-of-function mutations in a *CYC*-like gene resulting in conversion of ray flowers to disc-like tubular flowers [66,69]. Interestingly, it appears to be different *CYC* paralogues in different Asteraceae lineages that are responsible for differentiation of ray flowers [66]. Although this is somewhat surprising, this is consistent with ray flowers having evolved multiple times in the family [70].

### (b) Rosids

Similar to asterids, bilateral flower symmetry has evolved multiple times in rosids, and in at least three instances is associated with independent recruitment of a CYC-dependent developmental programme (Fabaceae, Brassicaceae and Malpighiaceae; figure 4). Developmental genetic studies of bilateral symmetry in Lotus and Pisum (Fabaceae) are extensive and second only to the work on floral symmetry in A. majus. In both Lotus and Pisum, there are three CYC-like paralogues. Through analysis of naturally occurring mutants, as well as gene silencing and overexpression transgenic studies, it is clear that two of these paralogues, CYC1 and CYC2 (LST in Pisum, and SQU1 in Lotus) function redundantly to establish dorsal petal identity [71-74]. Unlike in A. majus where the ground state for development seems to be lateral petal identity (cyc;dich;div mutant background [75]), in Pisum and Lotus the ground state appears to be ventral petal identity because in addition to dorsal identity controlled by CYC1 and CYC2, the

bilateral symmetry



**Figure 4.** Summarized expression of *CYC*-like genes from comparative developmental genetic studies. Blue (grey in print version) shading indicates approximate pattern of *CYC*-like gene expression in the corolla of representative taxa. Phylogeny as in figure 2*a*, but for only the subset of orders with bilaterally symmetrical species. (Online version in colour.)

third paralogue, CYC3 (K in *Pisum*, KEW in *Lotus*), directs the development of lateral petals (figure 4) [71–74].

In Brassicaceae, the role of *CYC*-like genes for directing development of bilateral flower symmetry has been investigated in *Iberis*, a close relative to the model species *Arabidopsis*. In *Iberis*, the two ventral petals are expanded relative to the two dorsal petals (figure 4). This difference is established late during *Iberis* flower development, and is associated with relatively late expression of *IaTCP1*, a *CYC* homologue, in the smaller dorsal petals (figure 4) [38,39]. Because *Iberis* is closely related to *Arabidopsis*, heterologous functional studies of *IaTCP1* in *Arabidopsis* provided meaningful assessment of *IaTCP1* function. Overexpression of *IaTCP1* in *Arabidopsis* resulted in reduced cell proliferation in both vegetative organs and petals [39], consistent with reduced dorsal petal size where *IaTCP1* is expressed in *Iberis*. Lastly for the rosids, and similar to Fabaceae and Brassicaceae, *CYC*-like genes have been implicated in the evolution of bilateral symmetry in Malpighiaceae, with expression of *CYC*-like genes restricted to dorsal and dorsal/lateral petals (figure 4) [47].

### (c) Early diverging eudicots and monocots

While most of the comparative work of flower symmetry developmental genetics has been undertaken in core eudicot lineages,

radial symmetry

a handful of studies have tested whether the extensive parallel recruitment of a *CYC*-dependent programme for bilateral flower symmetry extends to non-core eudicot taxa. And again, we find evidence supporting a role for *CYC*-like genes in the development of bilaterally symmetrical flowers from early diverging eudicot and monocot lineages. Bilaterally symmetrical flowers of *Capnoides* in the Fumariodeae lineage of Papaveraceae (Ranunculales) are derived from disymmetric flowers [40,41]. The plane of bilateral symmetry in Fumariodeae flowers is transverse (figure 4), although partial resupination ultimately brings the transverse plane into dorsoventral orientation. In *Capnoides*, expression of two *CYC*-lineage paralogues [40,41] is asymmetric, with slightly stronger expression at the base of the outer petal that forms a nectary [76].

In monocots, transitions from radial to bilateral flower symmetry are pervasive [16], yet are quite under studied. In bilaterally symmetrical flowers of Costus and Heliconia (Zingerberales), as well as bilaterally symmetrical flowers of Commelina (Commelinales), expression of at least one CYC-like gene is asymmetric across the dorsoventral flower axis. In both monocot lineages studied, asymmetric CYC-like gene expression in the perianth is restricted to the ventral side of flowers (figure 4) [77,78]. This is in striking contrast to the general pattern of a CYC-dependent programme independently recruited to specify dorsal flower development across eudicots (figure 4). It is interesting to note, however, that a CYC-like gene from rice, RETARDED PALEA1, functions to specify palea development [79], an organ that develops on the dorsal side of grass florets. Whether the emerging pattern of dorsal flower expression in eudicots and ventral expression in monocots is a general pattern, perhaps reflecting developmental constraints, awaits further comparative work in monocots, as well as a clearer understanding of how CYC homologue expression is regulated during monocot and eudicot flower development.

## 6. Evolutionary transitions from bilateral to radial flower symmetry

Given the frequent association of bilateral symmetry with restricted expression of CYC-like genes to either the dorsal (most dicots), or ventral (most monocots) side of developing flowers, it is expected that reversals from CYC-dependent bilateral symmetry to radial symmetry will involve functional or regulatory changes to CYC homologues or their upstream regulators. There are, however, multiple hypothesized ways by which CYC-dependent bilateral flower symmetry might be lost in derived species with radial flower symmetry. One possibility is complete loss of CYC-like gene expression in flowers, through either regulatory evolution or gene loss (figure 3e). By contrast, regulatory evolution may result in expansion of CYClike gene expression across the dorsoventral axis of developing flowers (figure 4f). Alternatively, the evolution of radial symmetry from CYC-dependent bilateral symmetry could arise through mechanisms independent of functional or regulatory evolution of CYC-like genes. For instance, compensatory changes might evolve in genes/genetic pathways downstream of CYC, or in developmental pathways non-overlapping with a CYC-like programme (figure 4g,h). Results from multiple comparative studies suggest that evolutionary changes at or upstream of CYC-like genes frequently underlie transitions from CYC-dependent bilateral to radial flower symmetry. However, results from some studies are not inconsistent with a hypothesis of compensatory evolution.

Examples of derived radial symmetry (from CYC-dependent bilateral symmetry) for which the expression of CYC homologues has been studied include *Plantago* (Plantaginaceae, Lamiales), *Cadia* (Fabaceae, Fabales), two independent transitions to radial from bilateral symmetry in Gesneriaceae (Lamiales)—*Bournea* and *Tengia*, and four independent transitions to radial from bilateral symmetry in Malpighiaceae (Malpighiales)—*Psychopterys*, *Sphedamnocarpus*, *Microsteria* and *Lasiiocarpus* (figure 4) [62,80–83]. For each of these, two or more paralogous *CYC*-like genes are dorsally expressed in close relatives. Therefore, expression of all paralogues was investigated in these derived radially symmetrical lineages.

The most common pattern observed is a paralogue-specific combination of CYC loss of expression (figure 3e) with expanded CYC expression (figure 3f). In Plantago, Tengia, Cadia and Microsteria, one CYC-like paralogue (or set of closely related paralogues in the case of Tengia) is expressed across the dorsoventral flower axis, owing to regulatory evolution either at or upstream of that paralogue. The other CYC-like paralogue has been lost (Plantago and Microsteria), or is no longer expressed in flowers (Cadia and Tengia) [80-83]. Alternatively, both CYC-like paralogues are expressed across the dorsoventral flower axis (Psychopterys), or neither is expressed in flowers (Sphedamnocarpus) [82]. For two studied lineages with derived radially symmetrical flowers, one CYC-like paralogue has either expanded or lost floral expression, but the other paralogue retains dorsal-specific expression (Lasiocarpus and Bournea, respectively) [81,82]. In these cases, dorsalspecific CYC-like gene expression should be interpreted with caution. Dorsally restricted expression may be transient, or only occur early in development, and therefore may not specify a dorsal-specific developmental programme. Alternatively, there may indeed be functional consequences to retention of dorsal-specific expression, and these developmental consequences may be compensated by evolutionary changes in downstream or independent developmental programmes (figure  $3g_{,h}$ ).

### 7. Prospects

Our current knowledge of the repeated recruitment of a CYC-dependent developmental programme during independent transitions to bilateral flower symmetry is staggering. These insights are possible through a combination of advances in flowering plant molecular phylogenetic research and studies of character evolution, as well as detailed flower developmental genetic studies in a few model species (namely A. majus and L. japonica). Likewise, it is daunting to imagine how little we would know about these evolutionary developmental processes had evolution not proceeded with such extensive parallelism. I believe we are now justified in stating that parallel recruitment of a CYC-dependent developmental programme for bilateral flower symmetry is extensive. However, we have yet to determine the depth of this parallelism: does independent recruitment occur through regulatory changes at CYC-like loci, or through evolutionary changes to one or multiple upstream regulators of CYC, or through a combination of these possibilities that is taxon specific? Additionally, paralogues belonging to different CYC-like gene lineages are implicated in the evolution of bilateral

flower symmetry in the core eudicots, early diverging eudicots and monocots. Also, CYC-like genes are generally regulators of cell proliferation [51]. Other than this broad recruitment from different CYC paralogue groups, and a possible general role in regulating cell proliferation, we know little about the specific function or regulation of CYC- dependent genetic pathways that might shed light on why they have so frequently been recruited to flower symmetry developmental programmes.

Acknowledgements. L.C.H.'s research is supported by the University of Kansas and NSF grant no. IOS-1255808.

### References

- 1. Stebbins GL. 1974 Flowering plants: evolution above the species level. Cambridge, MA: Harvard University Press.
- 2. Leppik EE. 1972 Origin and evolution of bilateral symmetry in flowers. In Evolutionary biology (eds T Dobzhansky, MK Hecht, WC Steere). New York, NY: Appleton-Century-Crofts.
- 3. Delpino F. 1887 Zigomorfia florale e sue cause. *Malpighia* **1**, 245–262.
- 4. Church AH. 1908 Types of floral mechanism. Oxford, UK: Clarendon.
- 5. Endress PK. 1999 Symmetry in flowers: diversity and evolution. Int. J. Plant Sci. 160, S3-S23. (doi:10. 1086/314211)
- 6. Endress PK. 2012 The immense diversity of floral monosymmetry and asymmetry across angiosperms. Bot. *Rev.* **78**, 345–397. (doi:10.1007/s12229-012-9106-3)
- 7. Fenster CB, Armbruster WS, Dudash MR. 2009 Specialization of flowers: is floral orientation an overlooked first step? New Phytol. 183, 502-506. (doi:10.1111/j.1469-8137.2009.02852.x)
- 8. Ushimaru A, Dohzono I, Takami Y, Hyodo F. 2009 Flower orientation enhances pollen transfer in bilaterally symmetrical flowers. Oecologia 160, 667-674. (doi:10.1007/s00442-009-1334-9)
- Kampny CM. 1995 Pollination and flower diversity 9. in Scrophulariaceae. Bot. Rev. 61, 350-366. (doi:10.1007/BF02912622)
- 10. Sargent RD. 2004 Floral symmetry affects speciation rates in angiosperms. Proc. R. Soc. Lond. B 271, 603-608. (doi:10.1098/rspb.2003.2644)
- 11. Vamosi JC, Vamosi SM. 2010 Key innovations within a geographical context in flowering plants: towards resolving Darwin's abominable mystery. Ecol. Lett. 13, 1270-1279. (doi:10.1111/j.1461-0248.2010.01521.x)
- 12. Preston JC, Hileman LC. 2009 Developmental genetics of floral symmetry evolution. Trends Plant Sci. 14, 147-154. (doi:10.1016/j.tplants. 2008.12.005)
- 13. Preston JC, Hileman LC, Cubas P. 2011 Reduce, reuse, and recycle: developmental evolution of trait diversification. Am. J. Bot. 98, 397-403. (doi:10. 3732/ajb.1000279)
- 14. Busch A, Zachgo S. 2009 Flower symmetry evolution: towards understanding the abominable mystery of angiosperm radiation. Bioessays 31, 1181-1190. (doi:10.1002/bies.200900081)
- 15. Rosin FM, Kramer EM. 2009 Old dogs, new tricks: regulatory evolution in conserved genetic modules leads to novel morphologies in plants. Dev. Biol. **332**, 25–35. (doi:10.1016/j.ydbio.2009.05.542)
- 16. Citerne H, Jabbour F, Nadot S, Damerval C. 2010 The evolution of floral symmetry. In Advances in

botanical research (eds JC Kader, M Delseny), pp. 85-137, vol. 54. London, UK: Academic Press Ltd-Elsevier Science Ltd.

- 17. Ford VS, Gottlieb LD. 1992 Bicalyx is a natural homeotic floral variant. Nature 358, 671-673. (doi:10.1038/358671a0)
- 18. Gottlieb LD. 1984 Genetics and morphological evolution in plants. Am. Nat. 123, 681-709. (doi:10.1086/284231)
- 19. Stubbe H. 1959 Considerations on the genetical and evolutionary aspects of some mutants of Hordeum, Glycine, Lycopersicon, and Antirrhinum. Cold Spring Harbor. Symp. Quant. Biol. 24, 31-40. (doi:10. 1101/SQB.1959.024.01.005)
- 20. Zimmer EA, Wen J. 2012 Using nuclear gene data for plant phylogenetics: progress and prospects. Mol. Phylogenet. Evol. 65, 774-785. (doi:10.1016/j. ympev.2012.07.015)
- 21. Gottlieb LD. 1973 Enzyme differentiation and phylogeny in Clarkia fransciscana, C. rubicunda and C. amoena. Evolution 27, 205-214. (doi:10.2307/ 2406961)
- 22. Gottlieb LD. 1977 Electrophoretic evidence and plant systematics. Ann. Missouri Bot. Garden 64, 161-180. (doi:10.2307/2395330)
- 23. Gottlieb LD, Ford VS. 1996 Phylogenetic relationships among the sections of Clarkia (Onagraceae) inferred from the nucleotide sequences of PgiC. Syst. Bot. 21, 45-62. (doi:10. 2307/2419562)
- 24. Sytsma KJ, Gottlieb LD. 1986 Chloroplast DNA evolution and phylogenetic relationships in Clarkia sect. peripetasma (Onagraceae). Evolution 40, 1248-1261.
- 25. Sytsma KJ, Smith JF, Gottlieb LD. 1990 Phylogenetics in Clarkia (Onagraceae): restriction site mapping of chloroplast DNA. Syst. Bot. 15, 280-295. (doi:10.2307/2419184)
- 26. Soltis DE et al. 2011 Angiosperm phylogeny: 17 genes, 640 taxa. Am. J. Bot. 98, 704-730. (doi:10.3732/ajb.1000404)
- 27. Bremer B et al. 2009 An update of the angiosperm phylogeny group classification for the orders and families of flowering plants: APG III. Bot. J. Linn. Soc. 161, 105-121. (doi:10.1111/j.1095-8339.2009.00996.x)
- 28. Burleigh JG, Hilu KW, Soltis DE. 2009 Inferring phylogenies with incomplete data sets: a 5-gene, 567-taxon analysis of angiosperms. BMC Evol. *Biol.* **9**, 61. (doi:10.1186/1471-2148-9-61)
- 29. Olmstead RG, Bohs L, Migid HA, Santiago-Valentin E, Garcia VF, Collier SM. 2008 A molecular phylogeny of the Solanaceae. Taxon 57, 1159-1181.

- 30. Bruneau A et al. 2013 Legume phylogeny and classification in the 21st century: progress, prospects and lessons for other species-rich clades. Taxon 62, 217-248. (doi:10.12705/622.8)
- 31. Schaferhoff B, Fleischmann A, Fischer E, Albach DC, Borsch T, Heubl G, Muller KF. 2010 Towards resolving Lamiales relationships: insights from rapidly evolving chloroplast sequences. BMC Evol. Biol. 10, 352. (doi:10.1186/1471-2148-10-352)
- 32. Aliscioni S et al. 2012 New grass phylogeny resolves deep evolutionary relationships and discovers C4 origins. New Phytol. 193, 304-312. (doi:10.1111/j. 1469-8137.2011.03972.x)
- 33. Ree RH, Donoghue MJ. 1999 Inferring rates of change in flower symmetry in asterid angiosperms. Syst. Biol. 48, 633-641. (doi:10.1080/ 106351599260201)
- 34. Jabbour F, Damerval C, Nadot S. 2008 Evolutionary trends in the flowers of Asteridae: is polyandry an alternative to zygomorphy? Ann. Bot. 102, 153-165. (doi:10.1093/aob/mcn082)
- 35. Knapp S. 2010 On 'various contrivances': pollination, phylogeny and flower form in the Solanaceae. Phil. Trans. R. Soc. B. 365, 449-460. (doi:10.1098/rstb. 2009.0236)
- 36. Wang YZ et al. 2010 Origin and phylogenetic relationships of the Old World Gesneriaceae with actinomorphic flowers inferred from ITS and trnLtrnF sequences. Taxon 59, 1044-1052.
- 37. Davis CC, Anderson WR. 2010 A complete generic phylogeny of Malpighiaceae inferred from nucleotide sequence data and morphology. Am. J. Bot. 97, 2031-2048. (doi:10.3732/ajb.1000146)
- 38. Busch A, Horn S, Muhlhausen A, Mummenhoff K, Zachgo S. 2012 Corolla monosymmetry: evolution of a morphological novelty in the Brassicaceae family. Mol. Biol. Evol. 29, 1241-1254. (doi:10.1093/ molbev/msr297)
- 39. Busch A, Zachgo S. 2007 Control of corolla monosymmetry in the Brassicaceae Iberis amara. *Proc. Natl Acad. Sci. USA* **104**, 16714–16719. (doi:10.1073/pnas.0705338104)
- 40. Damerval C, Le Guilloux M, Jager M, Charon C. 2007 Diversity and evolution of CYCLOIDEA-like TCP genes in relation to flower development in Papaveraceae. Plant *Physiol.* **143**, 759–772. (doi:10.1104/pp.106.090324)
- 41. Kolsch A, Gleissberg S. 2006 Diversification of CYCLOIDEA-like TCP genes in the basal eudicot families Fumariaceae and Papaveraceae s.str. Plant Biol. 8, 680-687. (doi:10.1055/s-2006-924286)
- 42. Huelsenbeck JP, Nielsen R, Bollback JP. 2003 Stochastic mapping of morphological characters.

*Syst. Biol.* **52**, 131–158. (doi:10.1080/106351 50390192780)

- Endress PK, Doyle JA. 2009 Reconstructing the ancestral angiosperm flower and its initial specializations. *Am. J. Bot.* 96, 22–66. (doi:10. 3732/ajb.0800047)
- Pagel M, Meade A, Barker D. 2004 Bayesian estimation of ancestral character states on phylogenies. *Syst. Biol.* 53, 673–684. (doi:10.1080/ 10635150490522232)
- Maddison WP, Midford PE, Otto SP. 2007 Estimating a binary character's effect on speciation and extinction. Syst. Biol. 56, 701–710. (doi:10.1080/ 10635150701607033)
- Damerval C, Nadot S. 2007 Evolution of perianth and stamen characteristics with respect to floral symmetry in Ranunculales. *Ann. Bot.* **100**, 631–640. (doi:10.1093/aob/mcm041)
- Zhang WH, Kramer EM, Davis CC. 2010 Floral symmetry genes and the origin and maintenance of zygomorphy in a plant – pollinator mutualism. *Proc. Natl Acad. Sci. USA* **107**, 6388–6393. (doi:10.1073/ pnas.0910155107)
- Hileman LC, Baum DA. 2003 Why do paralogs persist? Molecular evolution of *CYCLOIDEA* and related floral symmetry genes in Antirrhineae (Veronicaceae). *Mol. Biol. Evol.* **20**, 591–600. (doi:10.1093/molbev/msq063)
- Gubitz T, Caldwell A, Hudson A. 2003 Rapid molecular evolution of *CYCLOIDEA*-like genes in *Antirrhinum* and its relatives. *Mol. Biol. Evol.* 20, 1537–1544. (doi:10.1093/molbev/msg166)
- Cubas P, Lauter N, Doebley J, Coen E. 1999 The TCP domain: a motif found in proteins regulating plant growth and development. *Plant J.* 18, 215–222. (doi:10.1046/j.1365-313X.1999.00444.x)
- Martin-Trillo M, Cubas P. 2010 TCP genes: a family snapshot ten years later. *Trends Plant Sci.* 15, 31–39. (doi:10.1016/j.tplants.2009.11.003)
- Luo D, Carpenter R, Copsey L, Vincent C, Clark J, Coen E. 1999 Control of organ asymmetry in flowers of *Antirrhinum. Cell* 99, 367–376. (doi:10.1016/ S0092-8674(00)81523-8)
- Luo D, Carpenter R, Vincent C, Copsey L, Coen E. 1996 Origin of floral asymmetry in *Antirrhinum*. *Nature* 383, 794–799. (doi:10.1038/383794a0)
- Almeida J, Rocheta M, Galego L. 1997 Genetic control of flower shape in *Antirrhinum majus*. *Development* **124**, 1387–1392.
- Galego L, Almeida J. 2002 Role of *DIVARICATA* in the control of dorsoventral asymmetry in *Antirrhinum* flowers. *Genes Dev.* **16**, 880–891. (doi:10.1101/gad. 221002)
- Corley SB, Carpenter R, Copsey L, Coen E. 2005 Floral asymmetry involves an interplay between TCP and MYB transcription factors in *Antirrhinum. Proc. Natl Acad. Sci. USA* **102**, 5068–5073. (doi:10.1073/ pnas.0501340102)
- Costa MMR, Fox S, Hanna AI, Baxter C, Coen E. 2005 Evolution of regulatory interactions controlling floral asymmetry. *Development* 132, 5093 – 5101. (doi:10. 1242/dev.02085)

- Rose A, Meier I, Wienand U. 1999 The tomato I-box binding factor *LeMYBI* is a member of a novel class of Myb-like proteins. *Plant J.* 20, 641–652. (doi:10. 1046/j.1365-313X.1999.00638.x)
- Raimundo J, Sobral R, Bailey P, Azevedo H, Galego L, Almeida J, Coen E, Costa MMR. 2013 A subcellular tug of war involving three MYB-like proteins underlies a molecular antagonism in Antirrhinum flower asymmetry. *Plant J.* **75**, 527 – 538. (doi:10.1111/tpj.12225)
- Preston JC, Kost MA, Hileman LC. 2009 Conservation and diversification of the symmetry developmental program among close relatives of snapdragon with divergent floral morphologies. *New Phytol.* 182, 751–762. (doi:10.1111/j.1469-8137.2009.02794.x)
- Gao Q, Tao JH, Yan D, Wang YZ, Li ZY. 2008 Expression differentiation of *CYC*-like floral symmetry genes correlated with their protein sequence divergence in *Chirita heterotricha* (Gesneriaceae). *Dev. Genes Evol.* **218**, 341–351. (doi:10.1007/s00427-008-0227-y)
- 62. Zhou XR, Wang YZ, Smith JF, Chen RJ. 2008 Altered expression patterns of TCP and MYB genes relating to the floral developmental transition from initial zygomorphy to actinomorphy in *Bournea* (Gesneriaceae). *New Phytol.* **178**, 532–543. (doi:10. 1111/j.1469-8137.2008.02384.x)
- Boyden GS, Donoghue MJ, Howarth DG. 2012 Duplications and expression of *RADIALIS*-like genes in Dipsacales. *Int. J. Plant Sci.* **173**, 971–983. (doi:10.1086/667626)
- Howarth DG, Martins T, Chimney E, Donoghue MJ. 2011 Diversification of *CYCLOIDEA* expression in the evolution of bilateral flower symmetry in Caprifoliaceae and *Lonicera* (Dipsacales). *Ann. Bot.* **107**, 1521–1532. (doi:10.1093/aob/mcr049)
- Baxter CEL, Costa MMR, Coen ES. 2007 Diversification and co-option of *RAD*-like genes in the evolution of floral asymmetry. *Plant J.* 52, 105–113. (doi:10.1111/j.1365-313X.2007.03222.x)
- 66. Chapman MA, Tang SX, Draeger D, Nambeesan S, Shaffer H, Barb JG, Knapp SJ, Burke JM. 2012 Genetic analysis of floral symmetry in Van Gogh's sunflowers reveals independent recruitment of *CYCLOIDEA* genes in the Asteraceae. *PLoS Genet.* 8, e1002628. (doi:10.1371/journal.pgen.1002628)
- Kim M, Cui ML, Cubas P, Gillies A, Lee K, Chapman MA, Abbott RJ, Coen E. 2008 Regulatory genes control a key morphological and ecological trait transferred between species. *Science* **322**, 1116–1119. (doi:10.1126/science.1164371)
- Broholm SK, Tahtiharju S, Laitinen RAE, Albert VA, Teeri TH, Elomaa P. 2008 A TCP domain transcription factor controls flower type specification along the radial axis of the *Gerbera* (Asteraceae) inflorescence. *Proc. Natl Acad. Sci. USA* **105**, 9117–9122. (doi:10. 1073/pnas.0801359105)
- Fambrini M, Salvini M, Pugliesi C. 2011 A transposon-mediate inactivation of a *CYCLOIDEA*-like gene originates polysymmetric and androgynous ray flowers in *Helianthus annuus*. *Genetica* **139**, 1521 – 1529. (doi:10.1007/s10709-012-9652-y)

- Panero JL, Funk VA. 2008 The value of sampling anomalous taxa in phylogenetic studies: major dades of the Asteraceae revealed. *Mol. Phylogenet. Evol.* 47, 757–782. (doi:10.1016/j.ympev.2008.02.011)
- Feng XZ *et al.* 2006 Control of petal shape and floral zygomorphy in *Lotus japonicus*. *Proc. Natl Acad. Sci. USA* **103**, 4970–4975. (doi:10.1073/pnas. 0600681103)
- Wang JC, Wang YM, Luo D. 2010 LJCYC genes constitute floral dorsoventral asymmetry in *Lotus japonicus. J. Integr. Plant Biol.* 52, 959–970. (doi:10.1111/j.1744-7909.2010.00926.x)
- Xu SL, Luo YH, Cai ZG, Cao XL, Hu XH, Yang J, Luo D. 2013 Functional diversity of *CYCLOIDEA*-like TCP genes in the control of zygomorphic flower development in *Lotus japonicus*. J. Integr. Plant Biol. 55, 221–231. (doi:10.1111/j.1744-7909.2012.01169.x)
- Wang Z et al. 2008 Genetic control of floral zygomorphy in pea (*Pisum sativum* L.). Proc. Natl Acad. Sci. USA **105**, 10 414–10 419. (doi:10.1073/ pnas.0803291105)
- Almeida J, Galego L. 2005 Flower symmetry and shape in *Antirrhinum. Int. J. Dev. Biol.* 49, 527–537. (doi:10.1387/ijdb.041967ja)
- Damerval C *et al.* 2013 Asymmetric morphogenetic cues along the transverse plane: shift from disymmetry to zygomorphy in the flower of Fumarioiceae. *Am. J. Bot.* **100**, 391–402. (doi:10. 3732/ajb.1200376)
- Preston JC, Hileman LC. 2012 Parallel evolution of TCP and B-class genes in Commelinaceae flower bilateral symmetry. *EvoDevo* 3, 6. (doi:10.1186/ 2041-9139-3-6)
- Bartlett ME, Specht CD. 2011 Changes in expression pattern of the *TEOSINTE BRANCHED1*-like genes in the Zingiberales provide a mechanisms for evolutionary shifts in symmetry across the order. *Am. J. Bot.* **98**, 227–243. (doi:10.3732/ajb. 1000246)
- Yuan Z et al. 2009 RETARDED PALEA1 controls palea development and floral zygomorphy in rice. Plant Physiol. 149, 235–244. (doi:10.1104/pp.108.128231)
- Preston JC, Martinez CC, Hileman LC. 2011 Gradual disintegration of the floral symmetry gene network is implicated in the evolution of a wind-pollination syndrome. *Proc. Natl Acad. Sci. USA* **108**, 2343 – 2348. (doi:10.1073/pnas.1011361108)
- Citerne HL, Pennington RT, Cronk QCB. 2006 An apparent reversal in floral symmetry in the legume *Cadia* is a homeotic transformation. *Proc. Natl Acad. Sci. USA* **103**, 12 017–12 020. (doi:10.1073/pnas. 0600986103)
- Zhang WH, Steinmann VW, Nikolov L, Kramer EM, Davis CC. 2013 Divergent genetic mechanisms underlie reversals to radial floral symmetry from diverse zygomorphic flowered ancestors. *Front. Plant Sci.* 4, 302. (doi:10.3389/fpls.2013.00302)
- Pang HB, Sun QW, He SZ, Wang YZ. 2010 Expression pattern of *CYC*-like genes relating to a dorsalized actinomorphic flower in *Tengia* (Gesneriaceae). *J. Syst. Evol.* 48, 309–317. (doi:10.1111/j.1759-6831.2010.00091.x)