



Published in final edited form as:

*Diabetologia*. 2014 May ; 57(5): 1027–1036. doi:10.1007/s00125-014-3180-5.

## Adiponectin reduces thermogenesis by inhibiting brown adipose tissue activation in mice

**Liping Qiao,**

Department of Pediatrics, University of California San Diego, 9500 Gilman Drive, MC 0983, La Jolla, CA 92093, USA

**Hyung sun Yoo,**

Department of Pediatrics, University of California San Diego, 9500 Gilman Drive, MC 0983, La Jolla, CA 92093, USA

**Chris Bosco,**

Department of Pediatrics, University of California San Diego, 9500 Gilman Drive, MC 0983, La Jolla, CA 92093, USA

**Bonggi Lee,**

Department of Pediatrics, University of California San Diego, 9500 Gilman Drive, MC 0983, La Jolla, CA 92093, USA

**Gen-Sheng Feng,**

Department of Pathology, University of California San Diego, La Jolla, CA, USA

**Jerome Schaack,**

Department of Microbiology, University of Colorado at Denver and Health Sciences Center, Aurora, CO, USA

**Nai-Wen Chi,** and

Veterans Affairs San Diego Healthcare System, and Department of Medicine, University of California San Diego, La Jolla, CA, USA

**Jianhua Shao**

Department of Pediatrics, University of California San Diego, 9500 Gilman Drive, MC 0983, La Jolla, CA 92093, USA

### Abstract

---

© Springer-Verlag Berlin Heidelberg 2014

J. Shao jishao@ucsd.edu.

**Electronic supplementary material** The online version of this article (doi:10.1007/s00125-014-3180-5) contains peer-reviewed but unedited supplementary material, which is available to authorised users.

**Duality of interest** The authors declare that there is no duality of interest associated with this manuscript.

**Contribution statement** LQ, HY, CB and BL performed experiments, analysed data and revised the article. GF, N-WC and JSc contributed study design, reviewing and editing of this manuscript. JSh is responsible for the integrity of this work as a whole and conceived and supervised the study and wrote the manuscript. All authors approved the final version of this manuscript.

**Aims/hypothesis**—Adiponectin is an adipocyte-derived hormone that plays an important role in energy homeostasis. The main objective of this study was to investigate whether or not adiponectin regulates brown adipose tissue (BAT) activation and thermogenesis.

**Methods**—Core body temperatures (CBTs) of genetic mouse models were monitored at room temperature and during cold exposure. Cultured brown adipocytes and viral vector-mediated gene transduction were used to study the regulatory effects of adiponectin on *Ucp1* gene expression and the underlying mechanisms.

**Results**—The CBTs of adiponectin knockout mice (*Adipoq*<sup>-/-</sup>) were significantly higher than those of wild type (WT) mice both at room temperature and during the cold (4°C) challenge. Conversely, reconstitution of adiponectin in *Adipoq*<sup>-/-</sup> mice significantly blunted β adrenergic receptor agonist-induced thermogenesis of interscapular BAT. After 10 days of intermittent cold exposure, *Adipoq*<sup>-/-</sup> mice exhibited higher UCP1 expression and more brown-like structure in inguinal fat than WT mice. Paradoxically, we found that the anti-thermogenic effect of adiponectin requires neither AdipoR1 nor AdipoR2, two well-known adiponectin receptors. In sharp contrast to the anti-thermogenic effects of adiponectin, AdipoR1 and especially AdipoR2 promote BAT activation. Mechanistically, adiponectin was found to inhibit *Ucp1* gene expression by suppressing β<sub>3</sub>-adrenergic receptor expression in brown adipocytes.

**Conclusions/interpretation**—This study demonstrates that adiponectin suppresses thermogenesis, which is likely to be a mechanism whereby adiponectin reduces energy expenditure.

### Keywords

Adiponectin; Brown adipose tissue; Energy expenditure; Thermogenesis

### Introduction

Fat depots have been generally classified as white adipose tissue (WAT) and brown adipose tissue (BAT). In humans and most other animals, WAT is the primary depot that stores chemical energy as triacylglycerol-enriched lipid droplets. By contrast, BAT is smaller in mass and specialised for thermogenesis. In humans, BAT was previously believed to regress after birth and vanish in adults. However, this dogma has been convincingly refuted by studies using [<sup>18</sup>F]fluorodeoxyglucose positron emission tomography scans [1–4]. These studies uncovered metabolically active BAT depots in adult humans that are quickly activated by cold exposure [2–4]. More importantly, human BAT mass and activity are inversely correlated with BMI, suggesting that BAT is an important player in energy metabolism in humans and may be harnessed to increase energy expenditure [2, 3, 5].

BAT is enriched in mitochondria that use uncoupling protein 1 (UCP1)-mediated uncoupling to convert significant amounts of chemical energy to heat [6]. UCP1 is exclusively expressed in brown adipocytes. Two types of brown adipocytes have been distinguished: (1) classical brown adipocytes that develop from myogenic factor 5 (Myf5)-positive, myoblast-like precursors and reside in the interscapular and perirenal regions [7]; (2) brown-like adipocytes that develop from a Myf5-negative precursors and are embedded in WAT [8, 9]. Differentiation into these brown-like adipocytes, generally referred to as

'browning', can be induced by prolonged cold exposure and  $\beta$ -adrenergic receptor ( $\beta$ AR) agonists [9–11]. In contrast to white adipocytes, brown-like adipocytes express UCP1 and harbour high levels of mitochondria and multilocular lipid droplets [8].

Adiponectin (also known as Adipoq or adipocyte complement-related protein of 30 kDa [Acrp30]) is an adipocyte-secreted hormone. It exhibits many characteristics of a starvation hormone in that it suppresses energy expenditure and enhances energy conservation [12–15]. Adiponectin overexpression increases fat mass in wild type (WT) mice and leads to a super-obese phenotype in the *ob/ob* background [12, 16]. Conversely, adiponectin ablation leads to a lean phenotype in two independent lines of *Adipoq* gene knockout (*Adipoq*<sup>-/-</sup>) mice as well as in transgenic mice expressing antisense mRNA against adiponectin [13, 14, 17]. Adiponectin suppresses basal and catecholamine-induced lipolysis in white adipocytes [17, 18]. We have reported that protein kinase A (PKA), an essential mediator of  $\beta$ AR signalling in BAT, is downregulated in WAT by adiponectin treatment [17]. Although an inhibitory effect of adiponectin on *Ucp1* gene expression has been observed in mouse BAT [13, 15, 19], it is not clear whether or not, or indeed how, adiponectin regulates BAT activation and thermogenesis.

Using genetic mouse models and cold challenge tests, we found that *Adipoq*<sup>-/-</sup> mice maintained a higher core body temperature (CBT) than WT mice, and were protected from hypothermia during acute cold exposure. After prolonged intermittent cold exposure, these mice harbour a greater number of brown-like adipocytes in the inguinal fat than WT control mice. Interestingly, the present study revealed that the anti-thermogenic effect of adiponectin is independent of two of its receptors, AdipoR1 and AdipoR2. In addition, we found that adiponectin inhibits UCP1 expression by reducing  $\beta_3$  adrenergic receptor (also known as *Adrb3*) expression in brown adipocytes. Therefore, this study demonstrated that adiponectin reduces BAT thermogenesis in mice.

## Methods

### Mouse colonies and cold exposure

All mice were in the C57BL/6 background. *AdipoR2*<sup>+/-</sup> mice were obtained from the Jackson Laboratory (Bar Harbor, ME, USA). *Adipoq*<sup>-/-</sup> mice were generated by P. Scherer's laboratory (The University of Texas Southwestern Medical Center, Dallas, TX, USA) and had been backcrossed to C57BL/6 background for nine generations. WT littermates of the last round of backcrossing served as controls. *AdipoR1*<sup>-/-</sup> mice were obtained from the National Institutes of Health-supported Mutant Mouse Regional Resource Centers (Chapel Hill, NC, USA). *AdipoR1* and *AdipoR2* gene knockout (*AdipoR1*<sup>-/-</sup> and *AdipoR2*<sup>-/-</sup>) mice were generated by crossing heterozygous breeders using their respective *AdipoR1*<sup>+/+</sup> and *AdipoR2*<sup>+/+</sup> littermates as controls. All mice were housed under standard conditions and handled according to the Association for Assessment and Accreditation of Laboratory Animal Care guidelines with approval from the University of California San Diego Animal Care and Use Committee.

Intraperitoneally implanted emitters, ER4000 receiver and the Vital View data acquisition system (Mini Mitter, Bend, OR, USA) were used to wirelessly monitor CBT. CBTs were

recorded at 1 min intervals. During fasting, CBTs were recorded from 20:00 to 07:30. Mice were singly caged for all cold exposure studies, which started at 08:00 or 08:30. For acute cold exposure, mice were transferred to a 4°C room. A Bio-Thermo Lifechip (Destron Fearing, South St Paul, MN, USA) was implanted underneath the interscapular BAT (iBAT) to measure iBAT temperature at 5 min intervals after i.p. injection of isoprenaline (ISO, 0.1 µmol/g of body weight) or saline control. For intermittent cold exposure, the exposure to 4°C was progressively extended as follows: 3 h on the first day and one additional hour each day up to 6 h/day for 7 days.

### **In vivo gene transduction**

Adenovirus-encoded adiponectin (Ad-Acrp30) was used to overexpress or reconstitute adiponectin in mice [20, 21]. Purified adenovirus vector ( $1 \times 10^9$  pfu per mouse) was injected through the tail vein. Experiments and tissue sample collection were performed 3 days later when blood adiponectin levels were increased 10–12-fold without overt signs of adenovirus-induced hepatitis [21, 22]. As we previously reported [21, 22], transduction with Ad-Acrp30 significantly reduced blood FFA and triacylglycerol levels without affecting body weight or composition (data not shown).

### **Mitochondrial respiration assay**

Mitochondria were purified from freshly collected tissues [23], and oxygen consumption was measured in a Mitocell Miniature respirometer (model 782, Strathkelvin Instruments Limited, Motherwell, UK) using pyruvate or palmitoyl-DL-carnitine (Sigma-Aldrich, St Louis, MO, USA) as substrates [24, 25].

### **Citrate synthase activity assay**

Citrate synthase activity was measured spectrophotometrically using tissue homogenates [21]. After an initial 2 min absorbance reading taken at 412 nm, the reaction was initiated with the addition of 3.0 mmol/L acetyl-CoA, and the change in absorbance was measured every 10 s for 2 min.

### **Immunoblotting and quantitative real-time PCR**

Previously described procedures were used [21, 22]. Briefly, protein samples (50 µg) were separated using NuPAGE 4–12% Bis-Tris gels (Life Technologies, Carlsbad, CA, USA). Proteins were transferred to polyvinylidene difluoride membranes, which were then blotted with primary antibodies overnight and visualised using ECL Plus substrate (Thermo Scientific, Waltham, MA, USA). The density of the bands was quantified using Quantity One 1-D Analysis Software (Bio-Rad Laboratories, Hercules, CA, USA). Trizol (Life Technologies) was used to isolate total mRNA. Relative PCR quantification was performed using a Stratagene Mx3000P (Agilent Technologies, Santa Clara, CA, USA). Expression data were normalised to the amount of 18S rRNA.

### **Brown adipocyte culture**

Brown adipocytes were differentiated from either immortalised mouse brown pre-adipocytes (obtained from Y.-H. Tseng's laboratory, Joslin Diabetes Center, Harvard Medical School,

Boston, MA, USA) or the stromal vascular fraction of mouse iBAT [26]. Specifically, cells were cultured in a differentiation medium (DMEM with 10% FBS, 20 nmol/l insulin and 1 nmol/l T3) for 3 days and then switched to induction medium (differentiation medium plus 0.125 mmol/l indometacin, 5  $\mu$ mol/l dexamethasone and 0.5 mmol/l IBMX). Two days later, the cells were switched back to differentiation medium for 5 days. The co-culture system was used for in vitro adiponectin treatment [17, 21]. For in vitro lipolysis assays, a co-culture system was used to treat brown adipocytes (derived from the stromal vascular fraction of iBAT from *Adipoq*<sup>-/-</sup> mice) with adiponectin for 24 h [17, 21]. The cells were then treated with the selective  $\beta$ 3-adrenergic agonist BRL37344 (10 ng/ml) or PBS for 1 h. Glycerol levels in culture medium were measured as previously described [17].

### Statistical analysis

Data are expressed as mean  $\pm$  SEM. Statistical analyses were performed using the Student's *t* test or ANOVA, followed by Bonferroni post-hoc tests using GraphPad Prism version 6.00 for Mac, GraphPad Software, La Jolla, CA, USA. A *p* value of <0.05 was considered to be statistically significant.

## Results

### Adiponectin inhibits thermogenesis and energy expenditure in mice

Thermogenesis is the main function of BAT, especially in rodents. To study the effects of adiponectin on BAT activation, CBTs were compared between *Adipoq*<sup>-/-</sup> and WT mice using intraperitoneally implanted wireless sensors. As shown in Fig. 1a, the CBTs of *Adipoq*<sup>-/-</sup> mice housed at room temperature (RT) were significantly higher than those of WT mice. During acute cold exposure (4°C air), *Adipoq*<sup>-/-</sup> mice were remarkably more resistant to hypothermia than WT mice (Fig. 1b) and their average CBTs significantly exceeded those of WT control mice (Fig. 1b inset). The resistance of *Adipoq*<sup>-/-</sup> mice to hypothermia was abolished when their iBAT was excised (Electronic supplementary material [ESM] Fig. 1a). Conversely, using adenoviral vectors to express adiponectin in the liver of *Adipoq*<sup>-/-</sup> mice, we found significantly lower CBTs in adiponectin-reconstituted mice than adenovirus-encoded green fluorescent protein-transduced control mice during acute cold exposure (ESM Fig. 1b).

The sympathetic nervous system (SNS) plays a critical role in BAT activation, a process mediated primarily by  $\beta$ ARs, especially  $\beta_3$  [7, 27]. We measured the  $\beta$  agonist ISO-induced changes of iBAT temperature to assess the effect of adiponectin on thermogenesis in *Adipoq*<sup>-/-</sup> mice. Adiponectin reconstitution significantly attenuated ISO-induced increase of iBAT temperature (Fig. 1c), an effect consistent with the cold exposure findings (ESM Fig. 1b). Together, these results suggest that adiponectin inhibits BAT activation and thermogenesis in mice.

Given the importance of BAT in energy metabolism, we used metabolism cages to investigate the effects of adiponectin on energy expenditure. Consistent with previous reports [12–15], *Adipoq*<sup>-/-</sup> mice exhibited significantly higher energy expenditures than WT

mice, particularly at the onset of the dark cycle (ESM Fig. 1c). These results further confirm the inhibitory effects of adiponectin on energy expenditure.

### Adiponectin reduces Ucp1 expression in iBAT

iBAT is the largest BAT in mice. Although CBTs were significantly increased in *Adipoq*<sup>-/-</sup> mice, their iBAT mass was indistinguishable from that of WT mice ( $n=8$ ,  $101.56\pm 6.5$  vs  $94.29\pm 6.2$  mg;  $p=0.44$ ). In addition, there was no obvious change in their iBAT morphology as assessed by hematoxylin and eosin staining (ESM Fig. 2).

UCP1 is essential for thermogenesis in BAT. Its expression in iBAT was compared between *Adipoq*<sup>-/-</sup> and WT mice. Similar to previous reports [13, 15], we found significantly more UCP1 protein in the iBAT of *Adipoq*<sup>-/-</sup> mice than WT controls housed at RT (Fig. 2a). The difference became striking after 4 h of cold air exposure (Fig. 2b). In addition, cold exposure significantly increased *PGC-1 $\alpha$*  gene expression in the iBAT of *Adipoq*<sup>-/-</sup> mice compared with WT control mice (Fig. 2b, c).

Citrate synthase is the first enzyme of the tricarboxylic acid cycle. Its activity is often used to assess the oxidative status of mitochondria, which is key to thermogenesis. We found significantly higher citrate synthase activity in the iBAT of *Adipoq*<sup>-/-</sup> mice than that of WT control mice (Fig. 2d).

Taking an alternative approach, we used an adenoviral vector to overexpress adiponectin in WT mice to investigate the effect of adiponectin on UCP1 expression and citrate synthase activity in iBAT. As shown in Fig. 2e, f, adiponectin overexpression significantly decreased UCP1 and peroxisome proliferator-activated receptor  $\gamma$  co-activator 1  $\alpha$  (PGC-1 $\alpha$ ) protein levels, as well as citrate synthase activity in iBAT. These in vivo effects were recapitulated in vitro using cultured brown adipocytes, showing that adiponectin treatment significantly reduced ISO-induced UCP1 expression (Fig. 2g). Together, these results indicate that adiponectin reduces *Ucp1* gene expression and citrate synthase activity in mouse iBAT.

Similar to our previous observation in WAT [17], phosphorylation of hormone-sensitive lipase was robustly elevated in iBAT of *Adipoq*<sup>-/-</sup> mice at RT and after cold exposure (Fig. 2a, b). Interestingly, remarkably increased protein levels of adipose triacylglycerol lipase were also found in the iBAT of *Adipoq*<sup>-/-</sup> mice (Fig. 2a, b), but not in WAT [17]. Furthermore, by using cultured brown adipocytes from *Adipoq*<sup>-/-</sup> mice, we found that glycerol release induced by the selective  $\beta_3$  agonist BRL37344 was significantly decreased by adiponectin treatment (Fig. 2h). Together, these results indicate that adiponectin inhibits lipolysis in brown adipocytes similar to its effect on white adipocytes [17, 18]. Lipolysis provides fatty acids to fuel thermogenesis in BAT. Therefore, we postulated that increased lipolysis and fuel supply contributed to the cold-resistant phenotype of *Adipoq*<sup>-/-</sup> mice.

### Adiponectin reduces mitochondrial respiration in iBAT without overtly altering mitochondrial biogenesis

Mitochondrial enrichment is a hallmark of brown adipocytes. We and others have reported that adiponectin enhances mitochondrial biogenesis in skeletal muscle [21, 28, 29]. However, the current study shows that in BAT, adiponectin reduces expression of PGC-1 $\alpha$



(Fig. 2b, c, e), a key regulator of mitochondrial biogenesis. We, therefore, set out to investigate the effects of adiponectin on mitochondrial biogenesis in brown adipocytes. We found no significant difference in mitochondrial DNA copy number or the mitochondrial marker *Cox5a* between *Adipoq*<sup>-/-</sup> and WT or between adiponectin-overexpressing and control mice (ESM Fig. 3a, b). In addition, by comparing the ultrastructure of iBAT, we found no significant difference in mitochondrial structure or density between *Adipoq*<sup>-/-</sup> and WT mice (ESM Fig. 3c, d). These results indicate that despite inhibiting PGC-1 $\alpha$  expression in iBAT, adiponectin does not significantly affect mitochondrial biogenesis in brown adipocytes. Given the important role of PGC-1 $\alpha$  in mitochondrial biogenesis, there was an apparent discordance between the effects of adiponectin on PGC-1 $\alpha$  expression, mitochondrial biogenesis and citrate synthase activity in brown adipocytes. This discordance is reminiscent of mice with *Lkb1* deficiency in skeletal muscle, where exercise increases mitochondrial protein expression and citrate synthase activity without changing PGC-1 $\alpha$  or mitochondrial DNA [30]. Moreover, PGC-1 $\alpha$  deficiency fails to abolish exercise-induced adaptive mitochondrial protein expression in muscle [31].

Oxygen consumption is a readout for mitochondrial metabolic state. We compared the respiration rates of mitochondria purified from iBAT of *Adipoq*<sup>-/-</sup> and adiponectin-overexpressing mice in comparison with their respective controls. We found no difference in any of the four states of respiration when pyruvate was used as substrate (ESM Fig. 3e). However, in the presence of the lipid substrate palmitoyl carnitine, the respiration rate of *Adipoq*<sup>-/-</sup> mitochondria was significantly higher than that in WT controls (Fig. 3a). Conversely, the rate was decreased by adiponectin reconstitution of *Adipoq*<sup>-/-</sup> mice (Fig. 3b). These results suggest that despite the lack of a measurable effect on iBAT mitochondrial biogenesis, adiponectin suppresses fatty acid combustion in brown adipocytes.

### Adiponectin decreases browning of subcutaneous fat

Browning has been observed in rodent models [8, 9]. Hyperplasia of beige cells in WAT has been suggested to enhance thermogenesis and energy expenditure [8–11]. By comparing inguinal fat from mice that were housed under regular conditions, we found no significant difference in UCP1 expression or histology structure between WT and *Adipoq*<sup>-/-</sup> mice (data not shown). We then exposed *Adipoq*<sup>-/-</sup> and WT mice to cold air intermittently for 10 days prior to comparing the histology of and gene expression in inguinal fat. As expected, cold exposure significantly increased brown adipocyte-like structure in inguinal fat of both WT and *Adipoq*<sup>-/-</sup> mice (data not shown). Figure 4 shows that *Adipoq*<sup>-/-</sup> mice contained significantly more brown adipocyte-like regions (Fig. 4a) and higher UCP1 protein levels (Fig. 4b) than WT control mice. These results indicate that the inguinal fat of *Adipoq*<sup>-/-</sup> mice is more prone to cold-induced browning, suggesting that adiponectin normally inhibits cold-induced browning of subcutaneous fat.

### Adiponectin inhibits thermogenesis independently of adiponectin receptors 1 and 2

AdipoR1 and AdipoR2 have been reported to be the main cell-surface receptors that mediate adiponectin signalling [32]. To investigate the roles of these receptors in the inhibition of BAT activation by adiponectin, we challenged *AdipoR1*<sup>-/-</sup> and *AdipoR2*<sup>-/-</sup> mice to cold air.

To our surprise, both groups of mice were more susceptible than WT to cold-induced hypothermia (Fig. 5a, b). In addition, ISO-induced iBAT hyperthermia was significantly attenuated in *AdipoR1*<sup>-/-</sup> mice and, more strikingly, abolished in *AdipoR2*<sup>-/-</sup> mice (Fig. 5c). These results indicate that in contrast to the cold resistance of *Adipoq*<sup>-/-</sup> mice, both *AdipoR1*<sup>-/-</sup> and *AdipoR2*<sup>-/-</sup> mice were cold intolerant.

Receptor deletion could conceivably lead to compensatory increases in ligand levels. We determined adiponectin levels in serum and iBAT to investigate why *AdipoR1*<sup>-/-</sup> and *AdipoR2*<sup>-/-</sup> mice were impaired in thermogenesis. Remarkably, serum and iBAT adiponectin levels were decreased in *AdipoR1*<sup>-/-</sup> mice (ESM Fig. 4a, b) but remained unaffected in *AdipoR2*<sup>-/-</sup> mice (ESM Fig. 4c, d). Given the anti-thermogenic effect of adiponectin, these results suggest that the cold intolerance of *AdipoR1*<sup>-/-</sup> and *AdipoR2*<sup>-/-</sup> mice is not due to changes in adiponectin levels.

Given that thermogenesis is increased in *Adipoq*<sup>-/-</sup> mice but decreased in both *AdipoR1*<sup>-/-</sup> and *AdipoR2*<sup>-/-</sup> mice, we created double knockouts to further explore this paradox. Upon cold challenge, both *AdipoR1*<sup>-/-</sup> and *AdipoR2*<sup>-/-</sup> mice became hypothermic faster than WT mice (ESM Fig. 4e). Remarkably, the cold intolerance of both groups of knockout mice was alleviated by concomitant deletion of adiponectin (ESM Fig. 4e). These results suggest that adiponectin regulates thermogenesis through a mechanism distinct from AdipoR1 and AdipoR2.

In line with their cold intolerant phenotype, the iBAT of *AdipoR1*<sup>-/-</sup> and *AdipoR2*<sup>-/-</sup> mice expressed less UCP1 protein and *Ucp1* mRNA than WT controls (Fig. 5d–g). In cultured brown adipocytes, we found that AdipoR2 overexpression robustly increased UCP1 protein levels regardless of ISO treatment (Fig. 5h). These results indicate that AdipoR2 enhances UCP1 expression in brown adipocytes.

AMP-activated protein kinase (AMPK) is an effector of AdipoR1 and AdipoR2 [32]. However, we detected no significant alteration in protein levels or phosphorylation of AMPK in iBAT of *AdipoR1*<sup>-/-</sup> or *AdipoR2*<sup>-/-</sup> mice (Fig. 5d, f), suggesting that AMPK may not be responsible for AdipoR1- or AdipoR2-dependent thermogenesis.

### **Adiponectin inhibits Ucp1 expression by downregulating $\beta_3$ adrenergic receptor in adipocytes**

To study the effects of adiponectin on *Ucp1* gene expression and its underlying mechanism, in vitro differentiated brown adipocytes were treated with adiponectin using a co-culture system [21]. Interestingly, despite a minor effect on basal *Ucp1* gene expression, adiponectin treatment robustly diminished UCP1 induction by the  $\beta_3$ -specific agonist BRL37344 (Fig. 6a) and the nonselective  $\beta$  agonist ISO (Fig. 2g). Given the critical role of the SNS in BAT development and activation, the results of this in vitro study suggest that the SNS may mediate the inhibitory effect of adiponectin on UCP1 expression and BAT thermogenesis. To test this hypothesis, sympathetic denervation of iBAT was performed by injecting 6-hydroxydopamine into the iBAT of both *Adipoq*<sup>-/-</sup> and adiponectin-reconstituted mice [33, 34]. A significant decrease of tyrosine hydroxylase mRNA in iBAT was apparent (ESM Fig. 5a). Despite the denervation, adiponectin robustly inhibited UCP1 expression in



iBAT (ESM Fig. 5b). We, therefore, concluded that adiponectin acted directly on iBAT to secondarily modulate the SNS effect.

The  $\beta_3$ -adrenergic receptor *Adrb3* is highly expressed in rodent adipocytes and is pivotal to SNS-mediated lipolysis and thermogenesis in BAT. We found a remarkable upregulation of *Adrb3* mRNA levels in iBAT (Fig. 6b) and inguinal fat (ESM Fig. 6a) of *Adipoq*<sup>-/-</sup> mice compared with WT controls. In brown adipocytes differentiated from iBAT stromal vascular fraction, adiponectin treatment significantly reduced *Adrb3* mRNA and protein expression (Fig. 6c, d). In line with these in vitro results, adiponectin reconstitution in *Adipoq*<sup>-/-</sup> mice also robustly reduced *Adrb3* mRNA and protein levels in iBAT (Fig. 6e, f). PKA is a main effector of *Adrb3* in adipocytes. Consistent with the changes in *Adrb3* expression, phosphorylation of PKA substrates was significantly reduced in adiponectin-treated brown adipocytes and iBAT of adiponectin-reconstituted *Adipoq*<sup>-/-</sup> mice (ESM Fig. 6b, c). These results clearly indicate that adiponectin inhibits *Adrb3* gene expression in mouse brown adipocytes.

To verify whether or not *Adrb3* downregulation mediates the inhibitory effects of adiponectin on UCP1 expression, *Adrb3* was overexpressed in cultured brown adipocytes. The overexpression prevented adiponectin from inhibiting UCP1 expression in ISO-treated brown adipocytes (Fig. 6g), supporting the hypothesis that adiponectin inhibits UCP1 expression in brown adipocytes by suppressing *Adrb3*.

## Discussion

Emerging evidence suggests that adiponectin plays an important role in energy homeostasis beyond its insulin-sensitising effect. The current study shows that adiponectin inhibits BAT activation and thermogenesis by suppressing both *Ucp1* gene expression in brown adipocytes and browning of subcutaneous fat. This study also reveals that adiponectin suppresses BAT activation by inhibiting  $\beta$ AR signalling in that tissue. To our knowledge, this is the first study to date that systematically demonstrates the regulatory effect of adiponectin on BAT activation and thermogenesis. Identification of the inhibitory effects of adiponectin on BAT provides mechanistic insights into how this fat-secreted starvation hormone reduces energy expenditure and modulates energy metabolism.

Thermogenesis is the main function of BAT. By monitoring the CBT of genetically manipulated mouse models with and without acute cold stress, our study demonstrated that adiponectin inhibits thermogenesis in mice. This anti-thermogenic effect likely has two components: the extent of mitochondrial uncoupling and the availability of fuels. UCP1 is expressed only in brown adipocytes, where it promotes thermogenesis by uncoupling fuel oxidation from ATP formation. Although iBAT in our *Adipoq*<sup>-/-</sup> mice showed no obvious alterations in tissue mass or histology, it expresses more UCP1 protein than in WT mice both in the basal state and after cold exposure. Conversely, *Ucp1* gene expression is down-regulated in adiponectin-overexpressing mice. These results confirm previous reports that adiponectin inhibits *Ucp1* gene expression in brown adipocytes [13, 15, 19]. Fatty acids are an important fuel for thermogenesis. In response to cold exposure, the SNS turns on both lipolysis and thermogenesis. Lipolysis releases fatty acids for mitochondrial oxidation and

thermogenesis. Previous studies have demonstrated that adiponectin inhibits lipolysis in white adipocytes [17, 18]. The current study shows that adiponectin also inhibits lipolysis in brown adipocytes. In both types of adipocyte, the inhibition of lipolysis should reduce the supply of fatty acids as thermogenic substrates. Additionally, we found that adiponectin suppresses browning of subcutaneous fat in mice repetitively exposed to cold. Although beige cells can contribute to thermogenesis, their limited presence in WAT under physiological conditions suggests a minor role in adiponectin-inhibited thermogenesis.

In contrast to leptin, adiponectin exhibits several characteristics of a starvation hormone. For example, it inhibits energy expenditure and enhances fat accumulation in several mouse studies [12–15]. Although other mechanisms may be involved, our discovery of adiponectin-mediated BAT inhibition provides mechanistic insights into how this hormone suppresses energy expenditure and enhances fat accumulation. We realise that the effects of adiponectin on BAT activation and fat accumulation are modest under our experimental conditions. However, the effects could be more pronounced in different settings since the importance of UCP1 is highly context dependent. Specifically, *Ucp1* knockout mice are obese at thermoneutrality but not at RT [6]. Our mice were housed at RT, which is below thermoneutrality for mice. Therefore, the magnitude of adiponectin-inhibited energy expenditure may be attenuated by counter-regulatory factors in energy metabolism and BAT activation due to the constant cold stress of RT.

Several plasma membrane proteins have been described as adiponectin receptors [32, 35]. Although detailed signalling pathways have not been elucidated, AdipoR1 and AdipoR2 have been reported to transduce the effects of adiponectin on both glucose and lipid metabolism [32]. We were surprised to find that neither AdipoR1 nor AdipoR2 is necessary for adiponectin to inhibit BAT activation. Our study showed that AdipoR2 knockout dramatically impaired thermogenesis (Fig. 5a–c), while concomitant deletion of adiponectin in these mice attenuated the phenotype (ESM Fig. 4e), suggesting that adiponectin and AdipoR2 regulate thermogenesis through different pathways. Lending support to the notion that AdipoR2 modulates thermogenesis independently of adiponectin is the nematode *Caenorhabditis elegans*. Despite the absence of an adiponectin homologue in this worm, deletion of its AdipoR2 homologue impairs both cold adaptation and lipid metabolism [36]. While we were preparing this paper, Kajimura et al reported that the regulatory effect of adiponectin on bone mass is independent of AdipoR1 and AdipoR2 [15]. Therefore, we propose that AdipoR1 and AdipoR2 may also serve as receptors for other ligands or may have ligand-independent activities that control BAT activation and energy metabolism. Further investigations are warranted to determine the signalling pathways that mediate the inhibitory effects of adiponectin on thermogenesis.

The SNS plays a critical role in BAT development and thermogenesis. *Adrb3*, a member of the  $\beta$ AR family, is predominantly expressed in adipose tissues in rodents. Our study found that sympathetic denervation of iBAT did not prevent the inhibitory effect of adiponectin on UCP1 expression in that tissue (ESM Fig. 5b). This is in contrast to cultured brown adipocytes, where the inhibitory effect of adiponectin on UCP1 requires the presences of adrenergic agonists (Figs. 2g and 6a). These results suggest that adiponectin inhibits UCP1 and thermogenesis through a pathway within adipocytes and not through the SNS. Indeed,

overexpression of *Adrb3* in brown adipocytes abolished this inhibitory effect (Fig. 6g), supporting the hypothesis that *Adrb3* downregulation mediates inhibition of BAT activation and thermogenesis in mice by adiponectin. Our study does not address mechanistically how adiponectin downregulates *Adrb3* in adipocytes. Given that the phenotype of *Adipoq*<sup>-/-</sup> mice is the opposite to that of *AdipoR1*<sup>-/-</sup> and *AdipoR2*<sup>-/-</sup> mice, it seems unlikely that adiponectin inhibits *Adrb3* through *AdipoR1* or *AdipoR2*. It is noteworthy that *Adrb3* expression and function in humans, unlike rodents, is not restricted to adipocytes and is uncertain. Caution should be taken in extrapolating the anti-thermogenic effect of adiponectin in mice to that in humans.

In summary, this study demonstrates that adiponectin inhibits BAT activation and thermogenesis in mice by suppressing UCP1 expression, lipolysis and brown adipocyte recruitment. The findings expand our understanding of how adiponectin regulates energy homeostasis.

## Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

## Acknowledgments

We thank Y.-H. Tseng (Joslin Diabetes Center, Harvard Medical School, Boston, MA, USA) and P. Scherer (The University of Texas Southwestern Medical Center, Dallas, TX, USA) for providing brown pre-adipocytes and *Adipoq*<sup>-/-</sup> mice, respectively.

**Funding** This work was supported by grants DK080418 (JS), HD069634 (JS), DK095132 (JS) and R01DK075916 (GF) from the National Institutes of Health and 5-I01-BX000702 (N-WC) from the Department of Veterans Affairs.

## Abbreviations

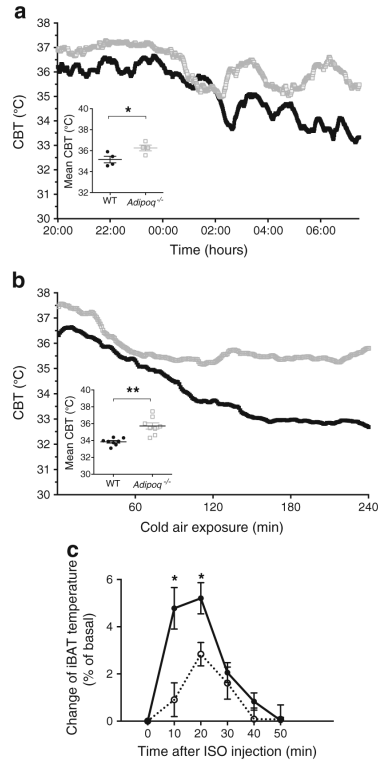
<b>Acrp30</b>	Adipocyte complement-related protein of 30 kDa (adiponectin)
<b>Ad-Acrp30</b>	Adenovirus-encoded adipocyte complement-related protein of 30 kDa (adiponectin)
<b>Ad-GFP</b>	Adenovirus-encoded green fluorescent protein
<b>AMPK</b>	AMP-activated protein kinase
<b>ATGL</b>	Adipose triacylglycerol lipase
<b>βAR</b>	β-adrenergic receptor
<b>BAT</b>	Brown adipose tissue
<b>CBT</b>	Core body temperature
<b>GAPDH</b>	Glyceraldehyde 3-phosphate dehydrogenase
<b>HSL</b>	Hormone-sensitive lipase
<b>iBAT</b>	Interscapular brown adipose tissue
<b>ISO</b>	Isoprenaline

<b>Myf5</b>	Myogenic factor 5
<b>PGC-1<math>\alpha</math></b>	Peroxisome proliferator-activated receptor $\gamma$ co-activator 1 $\alpha$
<b>PKA</b>	Protein kinase A
<b>RT</b>	Room temperature
<b>SNS</b>	Sympathetic nervous system
<b>UCP1</b>	Uncoupling protein 1
<b>WAT</b>	White adipose tissue
<b>WT</b>	Wild type

## References

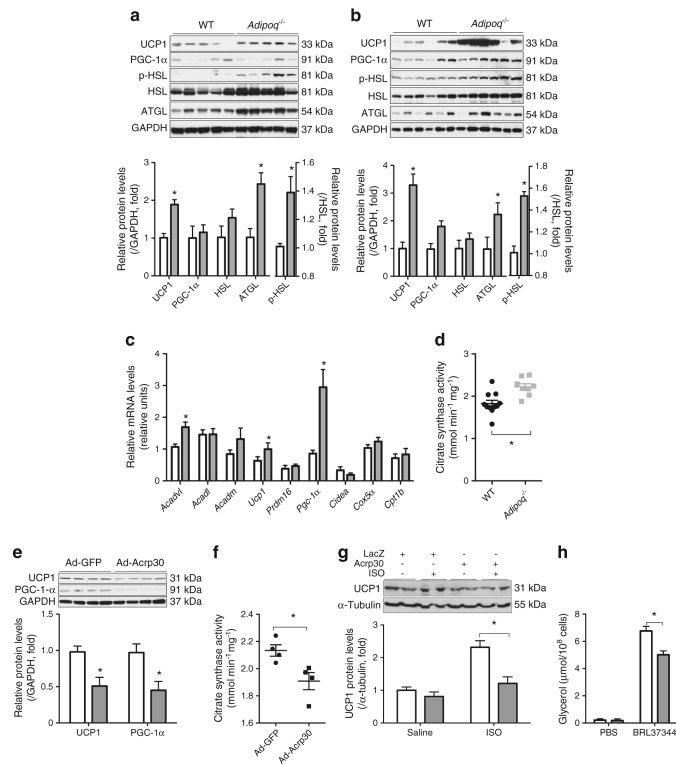
1. Nedergaard J, Bengtsson T, Cannon B. Unexpected evidence for active brown adipose tissue in adult humans. *Am J Physiol Endocrinol Metab.* 2007; 293:E444–E452. [PubMed: 17473055]
2. Cypess AM, Lehman S, Williams G, et al. Identification and importance of brown adipose tissue in adult humans. *N Engl J Med.* 2009; 360:1509–1517. [PubMed: 19357406]
3. Virtanen KA, Lidell ME, Orava J, et al. Functional brown adipose tissue in healthy adults. *N Engl J Med.* 2009; 360:1518–1525. [PubMed: 19357407]
4. van Marken Lichtenbelt WD, Vanhommerig JW, Smulders NM, et al. Cold-activated brown adipose tissue in healthy men. *N Engl J Med.* 2009; 360:1500–1508. [PubMed: 19357405]
5. Saito M, Okamatsu-Ogura Y, Matsushita M. High incidence of metabolically active brown adipose tissue in healthy adult humans: effects of cold exposure and adiposity. *Diabetes.* 2009; 58:1526–1531. [PubMed: 19401428]
6. Feldmann HM, Golozoubova V, Cannon B, Nedergaard J. UCP1 ablation induces obesity and abolishes diet-induced thermogenesis in mice exempt from thermal stress by living at thermoneutrality. *Cell Metab.* 2009; 9:203–209. [PubMed: 19187776]
7. Virtanen KA, Nuutila P. Brown adipose tissue in humans. *Curr Opin Lipidol.* 2011; 22:49–54. [PubMed: 21157334]
8. Boström P, Wu J, Jedrychowski MP, et al. A PGC1- $\alpha$ -dependent myokine that drives brown-fat-like development of white fat and thermogenesis. *Nature.* 2012; 481:463–468. [PubMed: 22237023]
9. Cinti S. Adipocyte differentiation and transdifferentiation: plasticity of the adipose organ. *J Endocrinol Invest.* 2002; 25:823–835. [PubMed: 12508945]
10. Fisher FM, Kleiner S, Douris N, et al. FGF21 regulates PGC-1 $\alpha$  and browning of white adipose tissues in adaptive thermogenesis. *Genes Dev.* 2012; 26:271–281. [PubMed: 22302939]
11. Ohno H, Shinoda K, Spiegelman BM, Kajimura S. PPAR $\gamma$  agonists induce a white-to-brown fat conversion through stabilization of PRDM16 protein. *Cell Metab.* 2012; 15:395–404. [PubMed: 22405074]
12. Kim J-Y, van de Wall E, Laplante M, et al. Obesity-associated improvements in metabolic profile through expansion of adipose tissue. *J Clin Invest.* 2007; 117:2621–2637. [PubMed: 17717599]
13. Kubota N, Yano W, Kubota T, et al. Adiponectin stimulates AMP-activated protein kinase in the hypothalamus and increases food intake. *Cell Metab.* 2007; 6:55–68. [PubMed: 17618856]
14. Saito K, Arata S, Hosono T, et al. Adiponectin plays an important role in efficient energy usage under energy shortage. *Biochim Biophys Acta.* 2006; 1761:709–716. [PubMed: 16815093]
15. Kajimura D, Lee HW, Riley KJ, et al. Adiponectin regulates bone mass via opposite central and peripheral mechanisms through FoxO1. *Cell Metab.* 2013; 17:901–915. [PubMed: 23684624]
16. Combs TP, Pajvani UB, Berg AH, et al. A transgenic mouse with a deletion in the collagenous domain of adiponectin displays elevated circulating adiponectin and improved insulin sensitivity. *Endocrinology.* 2004; 145:367–383. [PubMed: 14576179]

17. Qiao L, Kinney B, Schaack J, Shao J. Adiponectin inhibits lipolysis in mouse adipocytes. *Diabetes*. 2011; 60:1519–1527. [PubMed: 21430087]
18. Wedellová Z, Dietrich J, Siklová-Vítková M, et al. Adiponectin inhibits spontaneous and catecholamine-induced lipolysis in human adipocytes of non-obese subjects through AMPK-dependent mechanisms. *Physiol Res*. 2011; 60:139–148. [PubMed: 20945960]
19. Dong M, Yang X, Lim S, et al. Cold exposure promotes atherosclerotic plaque growth and instability via UCPI-dependent lipolysis. *Cell Metab*. 2013; 18:118–129. [PubMed: 23823482]
20. Qiao L, MacLean PS, You H, et al. Knocking down liver ccaat/enhancer-binding protein alpha by adenovirus-transduced silent interfering ribonucleic acid improves hepatic gluconeogenesis and lipid homeostasis in db/db mice. *Endocrinology*. 2006; 147:3060–3069. [PubMed: 16543372]
21. Qiao L, Kinney B, Yoo HS, et al. Adiponectin increases skeletal muscle mitochondrial biogenesis by suppressing mitogen-activated protein kinase phosphatase-1. *Diabetes*. 2012; 61:1463–1470. [PubMed: 22415879]
22. Qiao L, Zou C, van der Westhuyzen DR, Shao J. Adiponectin reduces plasma triglyceride by increasing VLDL triglyceride catabolism. *Diabetes*. 2008; 57:1824–1833. [PubMed: 18375436]
23. Frezza C, Cipolat S, Scorrano L. Organelle isolation: functional mitochondria from mouse liver, muscle and cultured fibroblasts. *Nat Protoc*. 2007; 2:287–295. [PubMed: 17406588]
24. Yeh T-YJ, Beiswenger KK, Li P, et al. Hypermetabolism, hyperphagia, and reduced adiposity in tankyrase-deficient mice. *Diabetes*. 2009; 58:2476–2485. [PubMed: 19651815]
25. Zechner C, Lai L, Zechner JF, et al. Total skeletal muscle PGC-1 deficiency uncouples mitochondrial derangements from fiber type determination and insulin sensitivity. *Cell Metab*. 2010; 12:633–642. [PubMed: 21109195]
26. Tseng Y-H, Kokkotou E, Schulz TJ, et al. New role of bone morphogenetic protein 7 in brown adipogenesis and energy expenditure. *Nature*. 2008; 454:1000–1004. [PubMed: 18719589]
27. Blaak EE, van Baak MA, Kempen KP, Saris WH. Role of alpha- and beta-adrenoceptors in sympathetically mediated thermo-genesis. *Am J Physiol*. 1993; 264:E11–E17. [PubMed: 8094272]
28. Iwabu M, Yamauchi T, Okada-Iwabu M, et al. Adiponectin and AdipoR1 regulate PGC-1alpha and mitochondria by Ca(2+) and AMPK/SIRT1. *Nature*. 2010; 464:1313–1319. [PubMed: 20357764]
29. Civitarese AE, Ukropcova B, Carling S, et al. Role of adiponectin in human skeletal muscle bioenergetics. *Cell Metab*. 2006; 4:75–87. [PubMed: 16814734]
30. Tanner CB, Madsen SR, Hallowell DM, et al. Mitochondrial and performance adaptations to exercise training in mice lacking skeletal muscle LKB1. *Am J Physiol Endocrinol Metab*. 2013; 305:E1018–E1029. [PubMed: 23982155]
31. Leick L, Wojtaszewski JFP, Johansen ST, et al. PGC-1alpha is not mandatory for exercise- and training-induced adaptive gene responses in mouse skeletal muscle. *Am J Physiol Endocrinol Metab*. 2008; 294:E463–E474. [PubMed: 18073319]
32. Yamauchi T, Kamon J, Ito Y, et al. Cloning of adiponectin receptors that mediate antidiabetic metabolic effects. *Nature*. 2003; 423:762–769. [PubMed: 12802337]
33. Chao P-T, Yang L, Aja S, et al. Knockdown of NPY expression in the dorsomedial hypothalamus promotes development of brown adipocytes and prevents diet-induced obesity. *Cell Metab*. 2011; 13:573–583. [PubMed: 21531339]
34. Thureson-Klein A, Mill-Hyde B, Barnard T, Lagercrantz H. Ultrastructural effects of chemical sympathectomy on brown adipose tissue. *J Neurocytol*. 1976; 5:677–690. [PubMed: 1003260]
35. Hug C, Wang J, Ahmad NS, et al. T-cadherin is a receptor for hexameric and high-molecular-weight forms of Acrp30/adiponectin. *Proc Natl Acad Sci U S A*. 2004; 101:10308–10313. [PubMed: 15210937]
36. Svensson E, Olsen L, Mörck C, et al. The adiponectin receptor homologs in *C. elegans* promote energy utilization and homeostasis. *PLoS ONE*. 2011; 6:e21343. [PubMed: 21712952]

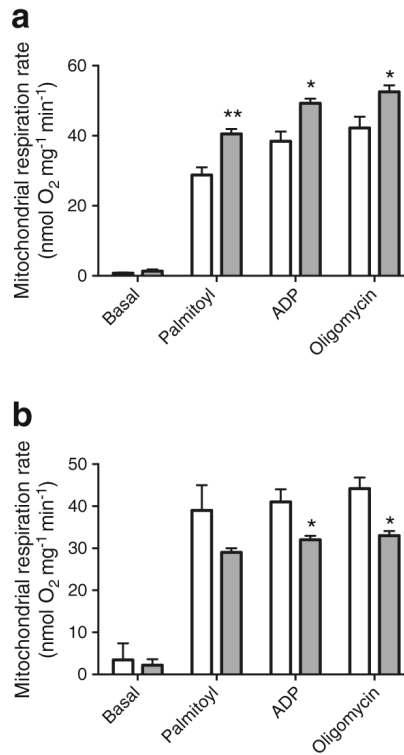


**Fig. 1.** Adiponectin inhibits thermogenesis in mice. **(a, b)** An emitter was implanted into the peritoneal cavity of 2-month-old male WT (black squares) and *Adipoq*<sup>-/-</sup> mice (grey outlined squares). After 1 week of recovery, CBTs were monitored during **(a)** overnight fasting ( $n=4$  per group) or **(b)** 4°C cold exposure ( $n=7-9$  per group). Average CBT ( $\pm$ SEM) is displayed in the insets. \* $p<0.05$ , \*\* $p<0.01$  vs WT. **(c)** male *Adipoq*<sup>-/-</sup> mice implanted with a Bio-Thermo Lifechip (Destron Fearing, South St Paul, MN, USA) underneath iBAT were injected i.v. with purified Ad-GFP (black circles) or Ad-Acrp30 (white circles) virus. Three days later, iBAT temperature was remotely measured before and after ISO injection ( $n=5$  per group). Increase of iBAT temperature (% of basal  $\pm$  SEM) is displayed. \* $p<0.05$  vs Ad-Acrp30-injected mice

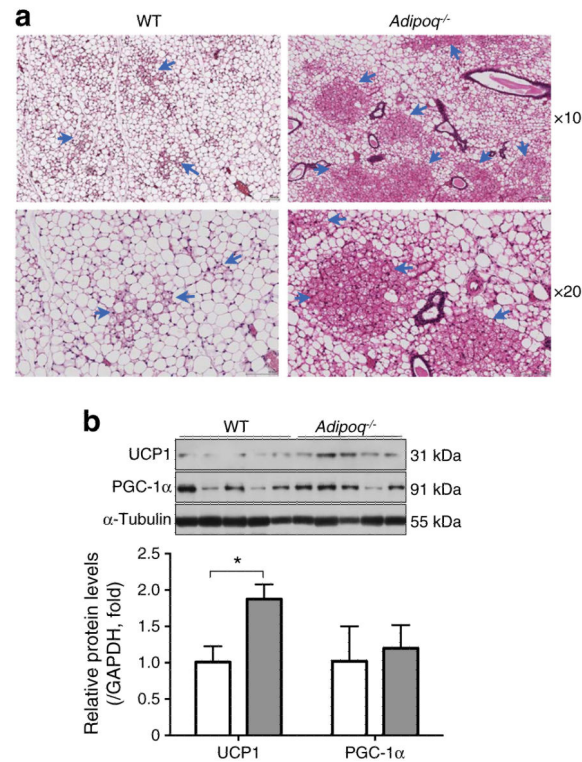


**Fig. 2.**

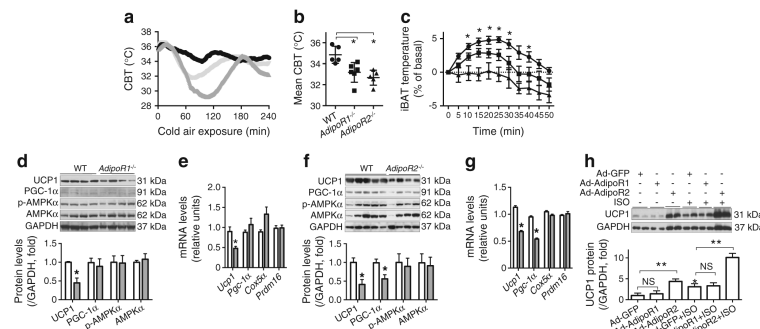
Adiponectin reduces UCP1 expression in iBAT. iBATs were collected from 8-week-old WT and *Adipoq*<sup>-/-</sup> mice at (a) RT or (b–d) after 4 h of cold exposure. (e, f) WT mice were injected i.v. with Ad-GFP or Ad-Acrp30 3 days before iBAT collection. (d, f) Citrate synthase activity in iBAT homogenates was determined based on the acetylation of oxaloacetic acid. (g, h) brown adipocytes differentiated from stromal vascular fraction of the iBAT of *Adipoq*<sup>-/-</sup> mice were treated overnight with Acrp30 (grey bar) using a co-culture system. Protein or medium samples were collected after treatment with ISO (10 μmol/l, 4 h) or BRL37344 (10 ng/ml, 1 h). Levels of (g) UCP1 protein and (h) glycerol in medium were determined. In (a–c): white bar, WT, grey bar, *Adipoq*<sup>-/-</sup>; (e): white bar, Ad-GFP, grey bar, Ad-Acrp30. *n*=6–8, \**p*<0.05 vs control mice



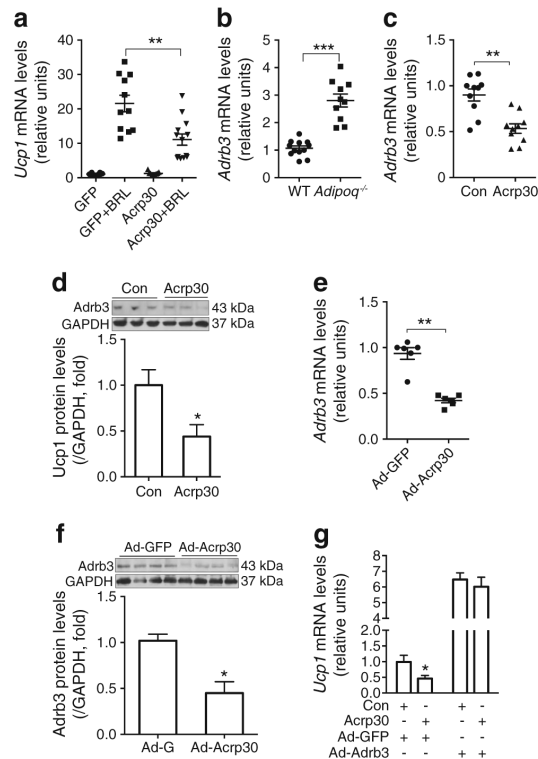
**Fig. 3.** Adiponectin reduces mitochondrial respiration rate in iBAT. Mitochondria were purified from (a) freshly isolated iBAT of *Adipoq*<sup>-/-</sup> (grey bars) and WT mice (white bars) or (b) adiponectin-reconstituted mice (Ad-GFP, white bars; Ad-Acrp30, grey bars), all housed at RT. Respiration rates were determined using palmitoyl carnitine (10 μmol/l) as substrate. Data are presented as mean ± SEM. *n*=6–8 per group of mice. \**p*<0.05, \*\**p*<0.01 vs control mice



**Fig. 4.** Enhanced browning of inguinal fat of *Adipoq*<sup>-/-</sup> mice. *Adipoq*<sup>-/-</sup> and WT mice were intermittently exposed to cold air for 10 days. **(a)** inguinal fat pads were collected for haematoxylin and eosin staining, and brown adipocyte-like structures are marked (blue arrows). **(b)** UCP1 and PGC-1α protein levels were determined by western blotting. White bars, WT; grey bars, *Adipoq*<sup>-/-</sup>. The data shown are mean ± SEM, *n*=5 per group. \**p*<0.05



**Fig. 5.** *AdipoR1*<sup>-/-</sup> and *AdipoR2*<sup>-/-</sup> mice are cold intolerant. **(a, b)** Mice of three genotypes were exposed to 4°C 1 week after emitter implantation and CBTs were monitored. The data are presented as mean or mean ± SEM ( $n=5-6$  per group). **(a)** Black circles, WT, light grey triangles; *AdipoR1*<sup>-/-</sup>; dark grey triangles, *AdipoR2*<sup>-/-</sup>. **(c)** temperatures of iBAT in mice kept at RT were remotely recorded before and after ISO injection. Black circles, WT; black squares, *AdipoR1*<sup>-/-</sup>; black triangles, *AdipoR2*<sup>-/-</sup>. The basal ± SEM percentage temperature changes are presented. \* $p<0.05$  vs *AdipoR1*<sup>-/-</sup> or *AdipoR2*<sup>-/-</sup> mice. **(d-g)** iBATs were collected from *AdipoR1*<sup>-/-</sup> or *AdipoR2*<sup>-/-</sup> mice after 4 h of cold air exposure. Levels of protein and mRNA **(d, e** grey bars, *AdipoR1*<sup>-/-</sup>; **f, g** grey bars, *AdipoR2*<sup>-/-</sup>) were measured by immunoblotting or real-time PCR ( $n=5-8$  per genotype). \* $p<0.05$  vs WT. **(h)** Differentiated brown adipocytes from immortalised pre-adipocytes were transduced for 24 h with adenovirus vectors encoding AdipoR1 or AdipoR2 (Ad-AdipoR1 and Ad-AdipoR2). Four hours after ISO treatment, protein levels of UCP1 were measured by immunoblotting ( $n=8$ ). \* $p<0.05$  vs Ad-GFP-transduced cells without ISO treatment. \*\* $p<0.01$  vs indicated group

**Fig. 6.**

Adiponectin inhibits UCP1 expression in brown adipocytes through downregulating Adrb3.

(a) Brown adipocytes differentiated from the stromal vascular fraction of iBAT from *Adipoq*<sup>-/-</sup> mice were treated overnight with Acrp30 using a co-culture system. BRL (50 ng/ml) was added to the medium 4 h before sample collection. Expression levels of *Ucp1* were determined by real-time PCR ( $n=8-13$  per group). (b) *Adrb3* mRNA levels of the iBAT of *Adipoq*<sup>-/-</sup> and WT mice housed at RT ( $n=10-12$  per group). (c, d) differentiated brown adipocytes were treated overnight with Acrp30 using a co-culture system. Levels of (c) *Adrb3* mRNA and (d) Adrb3 protein were determined by real-time PCR and immunoblotting ( $n=10$  per group). (e, f) *Adipoq*<sup>-/-</sup> mice were transduced with Acrp30-expressing adenovirus to reconstitute adiponectin, with Ad-GFP as control. iBATs were collected 3 days later to quantify (e) *Adrb3* mRNA by real-time PCR and (f) Adrb3 protein by immunoblotting ( $n=8$  per group). (g) Adipocytes differentiated from immortalised brown preadipocytes were transduced with an adenovirus vector overexpressing Adrb3 for 24 h. ISO was added 4 h before sample collection. A co-culture system was used for adiponectin treatment. Data are presented as mean  $\pm$  SEM,  $n=8$  per group. \* $p<0.05$  vs cells treated with control medium (d) or cells transduced with Ad-GFP without Ad-Adrb3. \*\* $p<0.01$  vs indicated group. Con, control