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Correction

# Correction: Nuclear Motility in Glioma Cells Reveals a Cell-Line Dependent Role of Various Cytoskeletal Components

**The *PLOS ONE* Staff**

There is an error in the first sentence of the legend for Figure 2. This error is only present in the XML version of the article and not the PDF version. Please view the correct Figure 2 and legend below.

**Citation:** The *PLOS ONE* Staff (2014) Correction: Nuclear Motility in Glioma Cells Reveals a Cell-Line Dependent Role of Various Cytoskeletal Components. PLoS ONE 9(7): e101593. doi:10.1371/journal.pone.0101593

**Published** July 2, 2014

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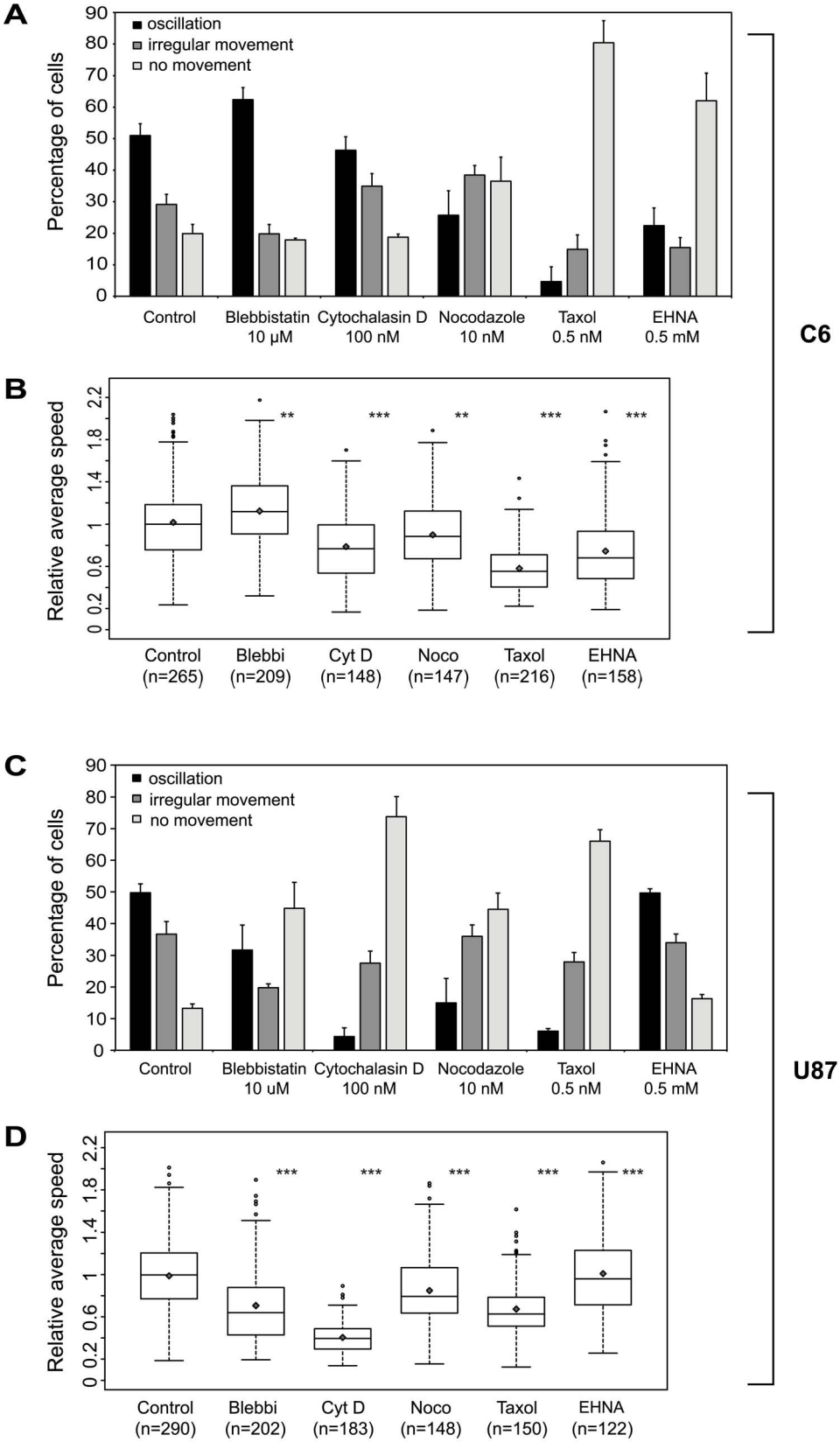
A
oscillation
irregular movement
no movement
Percentage of cells
Control
Blebbistatin 10 μM
Cytochalasin D 100 nM
Nocodazole 10 nM
Taxol 0.5 nM
EHNA 0.5 mM
B
Relative average speed
Control (n=265)
Blebbi (n=209)
Cyt D (n=148)
Noco (n=147)
Taxol (n=216)
EHNA (n=158)
C6
C
oscillation
irregular movement
no movement
Percentage of cells
Control
Blebbistatin 10 uM
Cytochalasin D 100 nM
Nocodazole 10 nM
Taxol 0.5 nM
EHNA 0.5 mM
D
Relative average speed
Control (n=290)
Blebbi (n=202)
Cyt D (n=183)
Noco (n=148)
Taxol (n=150)
EHNA (n=122)
U87

**Figure 2: Cell-line dependent effects of actin and microtubule inhibitors on nuclear motility in C6 and U87 cells.** Cells were imaged for at least 14 hours in the presence of DMSO (solvent control) or various cell-permeable cytoskeletal drugs. A–B: C6 cells, C–D: U87 cells (A and C) Proportion of cells within the different motility subgroups† upon inhibitor treatments. Error bars indicate mean+SE. (B and D) Average nuclear speeds in the two cell lines. Diamonds mark mean values, empty circles represent outliers. Data of at least three independent experiments is shown. Statistical analysis was performed using Kruskal-Wallis test. Significance codes: *: $p<0.05$, **: $p<0.01$, ***: $p<0.001$ †: explanation of the three migration types is discussed in Figure S3. The images and legends for Figures S2 and S3 are incorrectly switched. Please view the correct Figure S2 and Figure S3 below.
doi:10.1371/journal.pone.0093431.g002

The images and legends for Figures S2 and S3 are incorrectly switched. Please view the correct Figure S2 and Figure S3 below.

## Supporting Information

**Figure S2. Coupling between nuclear migration and cellular movements.** Cell extensions and nuclei of C6 and U87 cells seeded on patterns were manually tracked (n = 15). Representative example of an oscillating C6 (A) and U87 cell (B). Top panels: Positions of the cell center, the nucleus and the cell edges projected along the pattern over time. Middle panels: Relative position of the nucleus within the cell, normalized to the cell edges*. Allows visualizing the nuclear movements inside the cell. Lower panels: Related cross-correlation plots indicate no coupling between the movement of the nucleus and the cell centroid in C6 cells, and a strong correlation between their movements in U87 cells. Red vertical lines mark the lag at 0, red dashed lines indicate 95% confidence intervals. *Cell edges are defined at the start of tracking process, thus the "leading" or "trailing" edge terms are arbitrary.
(TIF)

**Figure S3. Categorization of nuclear movements in U87 cells.** Based on the coordinates of nuclei projected to the movement axis (i.e. along the pattern) and visual inspection of their corresponding trajectories, we have established the following categories: (A) Oscillatory movement: nuclei display a periodic movement along the pattern in at least 80% of the measured time. (B) Irregular movement: nuclei move without recurrent periodicity. (C) No movement: nuclei show no significant positional change over most of the time. This means that the cumulative nuclear displacement within 14 hours was below 200 μm for C6 cells, or below 300 μm in the case of U87 cells.
(TIF)

## Reference

1. Kiss A, Horvath P, Rothballer A, Kutay U, Csucs G (2014) Nuclear Motility in Glioma Cells Reveals a Cell-Line Dependent Role of Various Cytoskeletal Components. PLoS ONE 9(4): e93431. doi:10.1371/journal.pone.0093431.