

***Enterococcus faecium* AND *Enterococcus faecalis* IN BLOOD OF NEWBORNS WITH SUSPECTED NOSOCOMIAL INFECTION**

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SUMMARY

Enterococci are Gram-positive cocci saprophyte of the human gastrointestinal tract, diners who act as opportunistic pathogens. They can cause infections in patients hospitalized for a long time or who have received multiple antibiotic therapy. *Enterococcus faecalis* and *Enterococcus faecium* are the most common species in human infections. To evaluate the possibility of rapid detection of these species and their occurrence in the blood of newborns with suspected nosocomial infection, blood samples were collected from 50 newborns with late infections, admitted to the Neonatal Care Unit of the University Hospital Federal de Mato Grosso do Sul (UFMS-HU), from September 2010 to January 2011. The samples were subjected to conventional PCR and real time PCR (qPCR) to search for *Enterococcus faecium* and *Enterococcus faecalis*, respectively. The PCR results were compared with respective blood cultures from 40 patients. No blood cultures were positive for Enterococci, however, eight blood samples were identified as genomic DNA of *Enterococcus faecium* by qPCR and 22 blood samples were detected as genomic DNA of *Enterococcus faecalis* by conventional PCR. These findings are important because of the clinical severity of the evaluated patients who were found positive by conventional PCR and not through routine microbiological methods.

KEYWORDS: *Enterococcus faecium*; *Enterococcus faecalis*; Prematurity; PCR.

INTRODUCTION

Neonatal sepsis is the most frequent nosocomial infection in neonatal intensive care units and its incidence is increasing due to the increased survival of infants with very low birth weights^{13,15}. The greatest risk factors for late onset sepsis in neonates include low birth weight, prematurity, long periods of hospitalization and invasive procedures³. The late sepsis usually manifests as septicemia and pneumonia, occurring 72 hours after birth²⁰, caused by pathogens found in the nosocomial environment, including enterococci among the main etiological agents^{3,11}.

The genus *Enterococcus* is comprised of gram-positive cocci in pairs or short chains^{6,9}, considered saprophytes of the human gastrointestinal tract. They can survive on inanimate objects such as thermometers and stethoscopes, as well as on the hands of health professionals for long periods¹⁸.

Symptoms of neonatal infection are not specific, even for the different agents, with a need for more sensitive and specific microbiological and molecular tests¹⁰.

In a study in Spain by FERNANDEZ *et al.*, 2004², 95 episodes of

bacteremia caused by *Enterococcus faecalis* were documented, 83.2% with nosocomial origin, 85.3% associated with invasive procedures, 9.5% in neonates and 41.1% that had previously received broad spectrum antibiotics. In Brazil, a study by TITZE-DE-ALMEIDA *et al.* 2004¹⁹, with results of phenotypic and molecular analysis showed rates of around 95% in both *Enterococcus faecalis* and *Enterococcus faecium*.

Blood culture is the gold standard for diagnosis of sepsis, but has low sensitivity and the results are available in no less than 48 to 72 hours⁹. The PCR methods are shown to be fast and specific^{8,14}, but when we compare qPCR with conventional PCR, it is known that even though it consists of a more stringent and sensitive technique, in practice it demands higher costs than conventional PCR.

Thus, the objective of this study was to identify through the technique of qPCR and conventional PCR, respectively, the presence of *Enterococcus faecium* and *Enterococcus faecalis* in the blood of newborns, suspected of infection and admitted to the Intensive Care Unit.

MATERIAL AND METHODS

The study included infants who were suspected of nosocomial

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infection, admitted to the Neonatal Intensive Care Unit, University Hospital, Federal University of Mato Grosso do Sul, with at least 72 hours of admission or those with clinical worsening or a change of antibiotics, in a five month follow up, from September 2010 to January 2011.

This research was initiated only after approval by the Ethics Committee on Human Research of the Federal University of Mato Grosso do Sul (Protocol 1520).

The peripheral blood samples were collected with all accuracy aseptically as usually taken for routine under medical request. Samples for blood cultures were sent to the Central Laboratory of the HU-UFMS, where they were processed and analyzed by the automated system, BACTTEC™ FX (BD, New Jersey, USA) and the results compared with those obtained by conventional PCR.

The DNA extraction was performed using the GE Health Care extraction Kit and the blood samples of 50 infants, identifying *Enterococcus faecium* by qPCR and *Enterococcus faecalis*, by conventional PCR.

For qPCR reaction, the reagents kit from: SYBR Green PCR Biosystems (Applied Biosystems, Warrington, UK) was used. Designed oligonucleotide *primers* selected within the PBP5 (penicillin-binding protein) genome region were FAEFOR-1 = 5'-ggt acaaccgattactctgtcccat-3' and FAEFOR-2 = 5'-ggtacaaccgattacttggccat-3'; FAEREV-1 = 5'-tctgccgtctactcttggatgt-3' and FAEREV-2 = 5'-tctgccgtctactcttgaatgt-3', to obtain a specific target fragment of 94 bp.

The thermocycling conditions for qPCR were those standardized in the ABI Prism 7000 unit (52 °C / 2 min, 95 °C / 10 min, 45 cycles of: 95 to 60 °C / 15 °C / 15 s) with a final dissociation curve for each sample. *Primers* (1.5 pmol each), DNA (50-100 ng) and Syber Green PCR Master Mix (Applied Biosystems, Warrington, UK) were combined and qPCR carried out under conditions recommended by the manufacturer. All positive and negative tests had controls included. A human s26 normalizer was used as internal control. The cumulative CT values (cycle threshold) were collected in qPCR for each positive sample amplified in triplicate.

Standard conventional PCR was used for the detection of *Enterococcus faecalis*¹². The specific oligonucleotides *primers* designed and selected to amplify an amplicon of 87 bp, within the target region EntC2 (enterocin peptide precursor) of the genome of *Enterococcus faecalis*, were FAECFOR (5'-gcaattactgtggaggacctgg-3') and FAECREV (5'-tccaattcttttgaagacctgc-3'). *Primers* were designed based on sequences selected in the Genbank (program NCBI-BLASTn).

PCR consisted of: 9.75 µL of water sterile filtered, 1.5 µL 10x PCR buffer, 1.2 µL of dNTP mix (200 mM each), 0.4 µL of 50 mM MgCl₂, 0.5 µL of each primer (10 pmol / microl), 0.15 µL (5 U / microl) of Taq DNA polymerase (Ludwig Biotec, Alvorada, Brazil) and 1 µL of genomic DNA (200 ng) in a total volume of 15 µL reaction. The reaction was carried out in a MJ Research PTC 100 Thermal Cycler with the following program: 95 °C / 5 min, followed by 05 cycles of 94 °C / 1 min, 56 °C / 1 min, 72 °C / 1 min, followed by 40 cycles: 92 °C / 1 min, 60 °C / 1 min, 72 °C / 1 min, and final extension at 72 °C / 4 min.

The amplified fragments were photographed after electrophoresis on 6% of polyacrilamide gel (Ludwig Biotec) stained with silver nitrate. ATCC 19433 was used as a positive control for *Enterococcus faecalis*, autoclaved ultrapure water was used as a negative control.

The comparison between the groups of newborns evaluated in relation to blood PCR was performed using the nonparametric chi-square contingency table with 2x2 cells and with a value greater than five. The other results of the variables evaluated in this study were presented as descriptive statistics or graphs. Statistical analysis was performed using the SigmaStat version 2.0, whereas there were significant differences for values of $p < 0.05$ ¹⁶.

RESULTS

We evaluated 50 newborns of both genus; the mean gestational age was 31 weeks, and the mean birth weight was equal to 1.745 kg.

With respect to invasive procedures, all newborns in which we detected the presence of genetic material of bacteria studied were subjected to some type of vascular catheter. The use of mechanical ventilation and parenteral nutrition represent important features among these patients.

Of the 50 patients studied, 40 had blood culture and none of the cultures observed *Enterococcus faecalis* or *Enterococcus faecium*. For the other ten blood cultures, the results were not included in the medical record.

The results showed 16% (n = 8) positive for *Enterococcus faecium* in peripheral blood of the newborn evaluated by qPCR (Fig. 1) and 44% (n = 22) were positive for *Enterococcus faecalis*, by simple conventional PCR (Fig. 2). Among all the infants evaluated, 12% (n = 6) were positive for both types of bacteria, *Enterococcus faecalis* being more frequent ($p = 0.004$).

DISCUSSION

Prematurity and low birth weights are described as neonatal factors relevant to the development of late onset sepsis, since it is the main cause of admission of neonates in intensive care units⁴. We also observed prematurity (46%) as an important characteristic of this population.

In this study, among the blood samples in which at least one of the microorganisms described here was detected, the use of central venous catheters has been the procedure with the highest correlation with the presence of bacteria (54%), followed by mechanical ventilation (32%) and parenteral nutrition (32%).

This data agrees with HERRMANN *et al.*⁴, who in 2008 also found that the use of invasive procedures such as venous catheters, parenteral nutrition and mechanical ventilation had an important relationship with the development of late neonatal sepsis.

Some practices may be adopted as mitigation measures for late sepsis. Knowing that the use of vascular catheters and mechanical ventilation are closely related to the presence of infection, time of use must be reduced with safe and hygienic measures in the handling of equipment, and proper

Setup		Instrument		Results		
Plate	Software	Component	Amplification Plot	Standard Curve	Dissociation	Report
Well	Sample Name	Detector	Task	Ct	StdDev Ct	Qty
A11	11	Sybr Green	Unknown	17.81		Undet.
C1	25	Sybr Green	Unknown	38.34		Undet.
C4	28	Sybr Green	Unknown	18.96		Undet.
C5	29	Sybr Green	Unknown	17.24		Undet.
C6	30	Sybr Green	Unknown	38.48		Undet.
C7	31	Sybr Green	Unknown	36.64		Undet.
D2	38	Sybr Green	Unknown	Undet.		Undet.
D6	42	Sybr Green	Unknown	Undet.		Undet.
D10	46	Sybr Green	Unknown	37.15		Undet.
E2	50	Sybr Green	Unknown	39.26		Undet.
F8	e. faecium pasteo	Sybr Green	Standard	20.95		Undet.

Fig. 1 - Samples positive for *Enterococcus faecium* by Real-Time PCR.

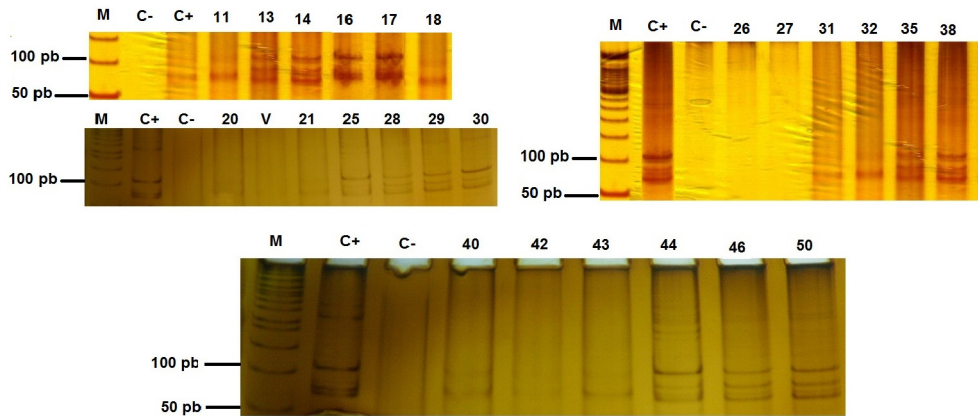


Fig. 2 - Conventional PCR for *Enterococcus faecalis* present in blood of newborns with suspected nosocomial infection. (M) 50 bp ladder marker (1µg); (C+) positive control for *E. faecalis*, ATCC 19433. (C-) negative control; 22 positive samples = (11,13,14,16,17,18,20,21,25,28,29,30,31,32,35,38,40,42,43,44,46,50).

hand washing are important measures in preventing infection nosocomial⁴.

The use of parenteral nutrition has been associated with late infection and delaying the start of enteral feeding. Other authors suggest that the enteral route should be initiated as early as possible to reduce the time of parenteral nutrition, which is associated with a high risk of infection^{4,7,11}.

For genomic detection of bacteria *Enterococcus faecium* and *Enterococcus faecalis* in the blood of newborns with risk factors for infection, we used qPCR and conventional PCR, respectively, for being rapid and specific methods. We also compared these results with blood culture.

There was no positive result for the bacteria studied by means of blood cultures. Other studies have reported lower sensitivity of blood cultures^{1,5,21}, despite being the gold standard for diagnosis of sepsis, is a time consuming technique, whose results are ready in no less time than 48 to 72 hours and several authors have questioned its credibility⁵.

Several methods for the molecular detection of *Enterococcus faecium* and *Enterococcus faecalis* have been reported as a powerful tool in the investigation of outbreaks of nosocomial infections¹². Such techniques

are distinguished by the rapidity and specificity of their results, which are essential for the prevention and control of transmission of infection, being all PCR based methods.

Further studies are needed to reveal in more detail the late onset of sepsis and its risk factors among newborns. Even though the presence of genomic DNA by conventional PCR and qPCR does not necessarily mean sepsis, possibly just colonization, adding to the patients symptoms and clinical observations, especially prematurity conditions, together lead the physician to take proper action or medical procedures in favor of children's treatment, avoiding deadly sepsis. High prematurity, associated with *Enterococcus faecium* and *Enterococcus faecalis* in the blood of newborns, reinforces the usage of conventional PCR and qPCR as additional tools for clinics.

CONCLUSIONS

The present finding is important due to the clinical severity of the evaluated patients who were positive by methods PCR and were not detected in routine microbiological methods. These methods were more effective compared to blood cultures that did not show any positive case for *enterococci*.

RESUMO

***Enterococcus faecium* e *Enterococcus faecalis* no sangue de recém-nascidos com suspeita de infecção nosocomial**

Os enterococos são cocos Gram-positivos saprófitas do trato gastrointestinal humano, atuam como patógenos oportunistas. Podem causar infecções em pacientes: hospitalizados por um longo tempo ou que receberam antibioticoterapia múltipla. *Enterococcus faecalis* e *Enterococcus faecium* são as espécies mais comuns em infecções humanas. Para avaliar a possibilidade de detecção rápida dessas espécies e sua ocorrência no sangue de recém-nascidos com suspeita de infecção hospitalar, foram coletadas amostras de sangue de 50 recém-nascidos, com infecção tardia, internados na Unidade de Terapia Neonatal do Hospital Universitário da Universidade Federal de Mato Grosso do Sul (UFMS-HU), no período de setembro de 2010 a janeiro de 2011. As amostras foram submetidas a PCR convencional e PCR em tempo real (qPCR) para pesquisa de *Enterococcus faecium* e *Enterococcus faecalis*, respectivamente. Os resultados da PCR foram comparados com culturas de sangue respectivos de 40. Nenhuma hemocultura foi positiva para enterococos, no entanto, em oito amostras de sangue foi identificado DNA genômico de *Enterococcus faecium* através da técnica de reação em cadeia da polimerase em tempo real, e em 22 amostras de sangue, foram detectados DNA genômico de *Enterococcus faecalis*, através de PCR convencional. A descoberta é importante por causa da gravidade clínica dos pacientes avaliados que foram positivos por PCR convencional e não foram detectados na rotina por métodos microbiológicos.

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REFERENCES

1. Brozanski BS, Jones JG, Krohn MJ, Jordan JA. Use of polymerase chain reaction as a diagnostic tool for neonatal sepsis can result in a decrease in use of antibiotics and total neonatal intensive care unit length of stay. *J Perinatol*. 2006;26:688-92.
2. Fernández Fernández FJ, de la Fuente Aguado J, Rubianes González M, Pérez Fernández S, Alvarez Fernández M, Nodar Germañas A, et al. *Enterococcus faecalis* bacteremia. *Rev Clin Esp*. 2004;204:244-50.
3. Haque KN. Neonatal sepsis in the very low birth weight preterm infants: part 1. Review of patho-physiology. *J Med Sci*. 2010;3:1-10.
4. Herrmann DMML, Amaral LMB, Almeida SC. Fatores de risco para o desenvolvimento de sepsé neonatal tardia em uma unidade de terapia intensiva. *Pediatria(São Paulo)*. 2008;30:228-36.
5. Honest H, Sharma S, Khan KS. Rapid tests for group B. *Streptococcus* colonization in laboring women: a systematic review. *Pediatrics*. 2006;117:1055-66.
6. Koneman EW, Allen SD, Janda WM, Schreckenberger PC, Winn WC Jr, Cury AE. Diagnóstico microbiológico: texto e atlas colorido. 5. ed. Rio de Janeiro: MEDSI; 2001. p. 589-659.
7. Leal YA, Álvarez-Nemegyei J, Velázquez JR, Rosado-Quiab U, Diego-Rodríguez N, Paz-Baeza E, et al. Risk factors and prognosis for neonatal sepsis in southeastern Mexico: analysis of a four-year historic cohort follow-up. *BMC Pregnancy Childbirth*. 2012;12:48.
8. Miglioli AMD. DNA genômico de *Streptococcus* e *Escherichia coli* em sangue e aspirado traqueal e gástrico de recém-nascidos intubados imediatamente após o nascimento [tese]. Campo Grande: Universidade Federal do Mato Grosso do Sul; 2009.
9. Murray PR, Resenthal KS, Kobayashi GS, Pfaller MA. Microbiologia médica. 4. ed. Rio de Janeiro: Guanabara Koogan; 2004. p. 220-3.
10. Nourse C, Byrne C, Murphy H, Kaufmann ME, Clarke A, Butler K. Eradication of vancomycin resistant *Enterococcus faecium* from a paediatric oncology unit and prevalence of colonization in hospitalized and community-based children. *Epidemiol Infect*. 2000;124:53-9.
11. Opilla M. Epidemiology of bloodstream infection associated with parenteral nutrition. *Am J Infect Control*. 2008;36:S173-8.
12. Sader HS, Pignatari AC, Hollis RJ, Jones RN. Evaluation of interhospital spread of methicillin-resistant *Staphylococcus aureus* in São Paulo, Brazil, using pulsed field gel electrophoresis of chromosomal DNA. *Infect Control Hosp Epidemiol*. 1994;15:320-3.
13. Sader HS, Sampaio JLM, Zocolli C, Jones RN. Results of 1997 SENTRY antimicrobial surveillance program in three Brazilian medical centers. *Braz J Infect Dis*. 1999;3:63-79.
14. Saiki RK, Gelfand DH, Stoffel S, Scharf SJ, Higuchi R, Horn GT, et al. Primer directed enzymatic amplification of DNA with a thermostable DNA polymerase. *Science*. 1988;239(4839):487-91.
15. Schrag SJ, Cutland CL, Zell ER, Kuwanda L, Buchmann EJ, Velaphi SC, et al. Risk factors for neonatal sepsis and perinatal death among infants enrolled in the prevention of perinatal sepsis trial, Soweto, South Africa. *Pediatr Infect Dis J*. 2012;31:821-6.
16. Shott S. Statistics for health professionals. London: W.B. Saunders Company; 1990.
17. Stoll BJ, Gordon T, Korones SB, Shankaran S, Tyson JE, Bauer CR, et al. Late-onset sepsis in very low birth weight neonates: a report from the National Institute of Child Health and Human Development Neonatal Research Network. *J Pediatr*. 1996;129:63-71.
18. Teixeira LM, Facklam RR. Enterococcus. In: Murray PR, Baron EJ, Jorgensen JH, Pfaller MA, Tenover FC, White O, editors. *Manual of clinical microbiology*. 8. ed. Washington: American Society for Microbiology; 2003. p. 422-33.
19. Titze-de-Almeida R, Rollo Filho M, Nogueira CA, Rodrigues IP, Eudes Filho J, Nascimento RS, et al. Molecular epidemiology and antimicrobial susceptibility of Enterococci recovered from Brazilian intensive care units. *Braz J Infect Dis*. 2004;8:197-205.
20. Vergnano S, Sharland M, Kazembe P, Mwansa C, Health PT. Neonatal sepsis: an international perspective. *Arch Dis Child Fetal Neonatal Ed*. 2005;90:220-4.
21. Yadav AK, Wilson CG, Prasad PL, Menon PK. Polymerase chain reaction in rapid diagnosis of neonatal sepsis. *Indian Pediatr*. 2005;42:681-5.

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