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Differentiation-dependent Requirement of Tsix long non-coding RNA in Imprinted X-chromosome Inactivation

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Abstract

Imprinted X-inactivation is a paradigm of mammalian transgenerational epigenetic regulation resulting in silencing of genes on the paternally-inherited X-chromosome. The pre-programmed fate of the X-chromosomes is thought to be controlled in *cis* by the parent-of-origin-specific expression of two long non-coding RNAs, Tsix and Xist, in mice. Exclusive expression of Tsix from the maternal-X has implicated it as the instrument through which the maternal germline prevents inactivation of the maternal-X in the offspring. Here, we show that Tsix is dispensable for inhibiting Xist and X-inactivation in the early embryo and in cultured stem cells of extra-embryonic lineages. Tsix is instead required to prevent Xist expression as trophectodermal progenitor cells differentiate. Despite induction of wild-type Xist RNA and accumulation of histone H3-K27me₃, many Tsix-mutant X-chromosomes fail to undergo ectopic X-inactivation. We propose a novel model of lncRNA function in imprinted X-inactivation that may also apply to other genomically imprinted loci.

X-chromosome inactivation results in the mitotically-stable epigenetic transcriptional silencing of genes along one of the two X-chromosomes in female mammals, thereby equalizing X-linked gene expression between males and females¹. X-inactivation is thought to be separable into three phases: initiation, establishment, and maintenance². During the initiation phase, cell autonomous epigenetic mechanisms identify the future inactive X-chromosome and trigger the formation of transcriptionally inert heterochromatin on that X. This heterochromatic configuration is then proposed to spread during the establishment phase to envelop genes across most of the inactive X-elect. Once inactivated, replicated copies of that X-chromosome are transmitted as inactive through multiple mitotic cell division cycles during the maintenance phase. While one X-chromosome is transmitted as

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AUTHOR CONTRIBUTIONS

E.M. and S.K. conceived the study and designed the experiments. E.M. performed all dissections and analysis of peri-implantation and post-implantation embryos. E.B. developed strand-specific RNA FISH probes and assisted with FISH staining of embryos. E.M., S.P., and M.H. isolated and analyzed blastocyst stage embryos. C.H., S.G., M.K.S., and E.M. derived and characterized TS cell lines. E.M., E.B., and S.K. wrote the manuscript.

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inactive during mitosis, the other X within the same nucleus is maintained in a transcriptionally active state.

The epigenetic transcriptional states of both the inactive and active X-chromosomes are controlled in *cis* by a segment on the X-chromosome that encodes long non-coding RNAs (lncRNAs) which play key roles in both X-inactivation and in forestalling inactivation of the active-X³⁻⁵. The two most prominent lncRNAs are Xist and Tsix. Xist is induced exclusively from the inactive X-chromosome and is considered a primary determinant of X-inactivation^{6,7}. Current models posit that Xist RNA transcription initiates a cascade of events that ultimately leads to X-inactivation⁸. Xist RNA physically coats the chromosome from which it is expressed, leading to the deposition of proteins that catalyze epigenetic transcriptional silencing along this X-chromosome⁹. The most notable of the Xist RNA recruits are Polycomb group proteins. Distinct Polycomb group complexes are thought to contribute to the formation of the unique facultative heterochromatic structure of the inactive-X via post-translational modification of histones¹⁰⁻¹³. While expression of the Xist RNA is required in *cis* for X-inactivation, transcription of the Xist anti-sense RNA, Tsix, is necessary to prevent inactivation of the active-X¹⁴⁻¹⁶. Tsix transcription across the *Xist* gene is posited to inhibit Xist expression, potentially by influencing chromatin modifications at the *Xist* promoter^{17,18}.

In mice, all cells of the developing zygote initially undergo imprinted inactivation of the paternal X-chromosome, beginning at around the 4–8 cell stage of zygotic development¹⁹⁻²². The pre-programmed fate of the two Xs during imprinted X-inactivation implies that the X-chromosomes are differentially marked in the parental germlines. In the pre-implantation embryo, Xist is expressed exclusively from the paternal-X and Tsix only from the maternal-X^{15,16}. The mutually exclusive expression and divergent transcriptional impact of Xist and Tsix lncRNAs represent a paradigm of how parent-of-origin specific gene regulation is executed in the offspring²³. While the paternal-X undergoes imprinted X-inactivation, evidence indicates that the epigenetic imprint itself resides on the maternal-X²⁴. This notion is supported by the observation that in early embryos that harbor two maternal X-chromosomes, neither X-chromosome undergoes X-inactivation²⁵⁻²⁷. Conversely, embryos with two paternal-Xs initially express Xist from both X-chromosomes, but then down-regulate Xist from one of the two Xs and appear to stably inactivate the other Xist-coated X-chromosome²⁸. Due to its expression exclusively from the maternal-X, its *Xist*-antagonistic function, and that embryos harboring a Tsix-mutant maternal X-chromosome die during gestation, Tsix RNA has been nominated as the factor via which the oocyte prevents inactivation of the maternal X-chromosome in the embryo^{15,16}. However, the temporal requirement of Tsix in imprinted X-inactivation, i.e., whether it functions during the initiation phase in the early embryo, which would support an oogenic imprint role, or in the maintenance phase in the later embryo, remains unclear.

At the peri-implantation (~128-cell) stage of embryogenesis, the pluripotent epiblast precursor cells in the inner cell mass compartment reactivate the inactive paternal X-chromosome^{29,30}. The epiblast lineage is the sole contributor of cells that will form the fetus; the rest of the cells from the early embryo give rise to the trophectoderm and the primitive endoderm lineages that will contribute to the extra-embryonic structures of the

placenta and yolk-sac, respectively³¹. After the embryo implants, descendants of the epiblast precursor cells undergo transcriptional reactivation of the paternal X-chromosome, followed by random X-inactivation of either the maternal or paternal X-chromosome³².

The transcriptional reactivation of the inactive paternal X-chromosome in pluripotent epiblast progenitors is characterized by loss of Xist RNA coating^{29,30,33}. Xist RNA depletion is thought to contribute to the epigenetic remodeling of the inactive paternal X-chromosome, leading to the re-expression of paternal X-linked genes²⁹. Similarly, Xist repression and the absence of or reactivation of the inactive-X are considered epigenetic hallmarks of pluripotency; female mouse embryonic stem (ES) cells and induced pluripotent stem (iPSCs) both display two active-Xs^{34–36}. As a negative regulator of Xist, Tsix is proposed to facilitate the loss of Xist RNA coating and reactivation of the inactive-X^{37–39}. To date, however, genetic evidence linking Tsix expression to Xist repression during X-chromosome reactivation is lacking.

Here, we comprehensively examine the role of Tsix RNA in imprinted X-inactivation and -reactivation *in vivo* and *in vitro*. We find that Tsix is dispensable in suppressing Xist and for preventing X-inactivation during the initiation as well as maintenance phases of imprinted X-inactivation through studies in the pre-implantation embryo and in stem cells of the trophoblast and primitive endoderm lineages. Instead, Tsix is required to prevent Xist induction during the differentiation of trophoblast cells *in vivo* and *in vitro*. Despite the induction of intact Xist RNA and accumulation of Polycomb group-catalyzed histone methylation on the Tsix-mutant X-chromosome, a substantial number of trophoblast cells do not display ectopic X-inactivation. We further find that both the repression of Xist and the reversal of imprinted X-inactivation that occur in epiblast precursor cells prior to random X-inactivation do not require Tsix RNA.

RESULTS

Role of Tsix During Initiation of Imprinted X-inactivation

If Tsix expression serves to repress Xist during the initiation phase of imprinted X-inactivation, then a maternal-X devoid of Tsix transcription should ectopically express Xist in the pre-implantation embryo. We therefore generated embryonic day (E) 3.5 blastocyst-stage (~64-cell) embryos that inherit either a wild-type (WT) or a Tsix-mutant maternal-X from Tsix-heterozygous females. The Tsix mutation, *Tsix*^{AA2 1.7}, truncates the Tsix transcript in exon 2 and deletes the critical *DXPas34* repeat sequence that controls Tsix expression (herein referred to as *X*^{Tsix}) (Fig. 1a)^{16,40–42}. Since Tsix transcription across the Xist promoter region is required for Tsix RNA to inhibit Xist expression, *X*^{Tsix} is functionally a null Tsix mutation^{16–18}. We first assayed Tsix and Xist expression by RNA fluorescence *in situ* hybridization (RNA FISH) with single-stranded riboprobes that uniquely detect Tsix or Xist. Tsix expression is detectable from the active (maternal) X-chromosome in most nuclei of approximately half of the embryos; in the remaining embryos, Tsix is undetectable in all nuclei (Fig. 1b). The active-X is additionally marked by expression of the *Atrx* gene, which is subject to X-inactivation. We therefore classified embryos with Tsix RNA FISH signals as WT XX and XY and the ones without as mutant *X*^{Tsix}X and *X*^{Tsix}Y. These embryos displayed Xist RNA coating in XX but not XY

embryos, as expected (Fig. 1b). To our surprise, $X^{Tsix}X$ blastocysts also showed Xist RNA coating of only one X-chromosome, and no inactivation of the X^{Tsix} maternal X-chromosome (Fig. 1b–c). As in $X^{Tsix}X$ females, $X^{Tsix}Y$ males also failed to display Xist induction or defective gene expression from their single, maternally-inherited X-chromosome (Fig. 1b–c).

To independently validate the RNA FISH data, we performed allele-specific RT-PCR amplification of Xist RNA in individual hybrid blastocyst stage embryos harboring polymorphic X-chromosomes. While the maternal X-chromosome is transmitted by *M. domesticus*-derived laboratory and Tsix-mutant strains (WT X^{Lab} and mutant X^{Tsix} , respectively), the paternal X-chromosome is derived from the divergent *M. molossinus* JF1 (X^{JF1}) strain; these strains contain numerous single nucleotide polymorphisms (SNPs), which permits defining the allele-specific origin of RNAs. We again assigned genotypes to the embryos by assaying Tsix RNA expression (Fig. 1d). As expected, in WT $X^{Lab}X^{JF1}$ and $X^{Lab}Y$ embryos Xist was expressed only in females and not in males (Fig. 1d). We observed a similar pattern of Xist RNA expression in $X^{Tsix}X^{JF1}$ and $X^{Tsix}Y$ mutant blastocysts (Fig. 1d). $X^{Tsix}Y$ mutant males did not exhibit Xist expression, consistent with the RNA FISH data. We exploited a SNP in the Xist RT-PCR amplicon to identify the chromosomal source of Xist RNA in $X^{Lab}X^{JF1}$ and $X^{Tsix}X^{JF1}$ embryos. Both genotypes displayed Xist expression only from the paternal X-chromosome (Fig. 1e). Taken together, the RNA FISH and RT-PCR results lead us to conclude that Tsix is dispensable in pre-implantation embryos during the initiation phase of imprinted X-inactivation, both to prevent Xist expression and to forestall inactivation of the maternally-inherited X-chromosome.

Post-implantation Role of Tsix in Suppressing Xist

Maternally-inherited Tsix mutations are typically embryonic lethal, suggesting an essential requirement for Tsix during embryonic development^{15,16}. Since $X^{Tsix}X$ and $X^{Tsix}Y$ pre-implantation embryos displayed normal imprinted X-inactivation, we investigated imprinted X-inactivation in peri- and post-implantation embryos to pinpoint when Tsix is required. At the E4.0 peri-implantation stage, $X^{Tsix}X$ and $X^{Tsix}Y$ embryos begin to exhibit Xist induction from the X^{Tsix} X-chromosome in a few nuclei, typically fewer than 3% (Supplementary Fig. 1). We next examined Xist expression and X-inactivation in XX, $X^{Tsix}X$, XY, and $X^{Tsix}Y$ E6.5 post-implantation embryos. We initially assessed Xist RNA coating and expression of the X-linked gene *Pgkl*, which is subject to X-inactivation, in whole E6.5 embryos using double-stranded probes that detect Xist and Tsix simultaneously^{21,43}. At E6.5, the extra-embryonic cell types derived from the trophectoderm and the primitive endoderm of earlier embryos maintain imprinted X-inactivation, while the epiblast cells display random X-inactivation. We observed ectopic Xist expression from the mutant X-chromosome in both $X^{Tsix}X$ and $X^{Tsix}Y$ embryos but not in WT XX and XY counterparts (Supplementary Fig. 2 a–d, Supplementary Movies 1–4). Using single-stranded riboprobes, we next quantified Xist RNA coating and *Pgkl* expression in isolated extra-embryonic tissues and found that 15% of $X^{Tsix}X$ and 12% of $X^{Tsix}Y$ cells showed ectopic Xist RNA coating and *Pgkl* silencing. These percentages represent a significant level of ectopic X-inactivation in extra-embryonic cells ($p=3.5\times 10^{-5}$ and $p=0.0003$, respectively, Fisher's exact test; Fig. 2a–b). We confirmed ectopic expression of Xist RNA from the

maternal X-chromosome in mutant extra-embryonic cells and its absence in WT cells by allele-specific RT-PCR followed by Sanger sequencing to determine the allelic origin of the transcript (Fig. 2c–d). *Tsix* deficiency, therefore, induces *Xist* expression and inactivation of the maternal X-chromosome in the extra-embryonic tissues of post-implantation embryos.

Differentiation-dependent Function of *Tsix* in TS Cells

Whole-mount RNA FISH stains of *Tsix*-mutant E6.5 embryos suggested that ectopic *Xist* induction primarily characterized the trophoctoderm-derived extra-embryonic ectoderm or its differentiated derivatives (Supplementary Fig. 2a–d, Supplementary Movies 1–4). To further examine the role of *Tsix* in imprinted X-inactivation, we generated $X^{\text{Lab}}X^{\text{JF1}}$, $X^{\text{Tsix}}X^{\text{JF1}}$, $X^{\text{Lab}}Y$, and $X^{\text{Tsix}}Y$ trophoblast stem (TS) cells. TS cells arise from trophoctoderm cells of the early embryo, and provide an *in vitro* model of early imprinted X-inactivation^{13,43,44}. Strand-specific RNA FISH analysis of the TS cells showed that *Xist* RNA is expressed from and coats a single X-chromosome in undifferentiated WT and X^{Tsix} -mutant female TS cells, and is not expressed in male TS cells of either genotype (Fig. 3a). We confirmed that *Xist* RNA is restricted to female cells despite the absence of *Tsix* by allele-specific RT-PCR (Fig. 3b). These data not only reinforce the conclusion from embryos that X-inactivation is unperturbed in the absence of *Tsix* transcription from the maternal X-chromosome in trophoctoderm cells, but also demonstrate that *Tsix* is not required to stably maintain *Xist* repression in undifferentiated TS cells in culture.

To reconcile why the extra-embryonic ectoderm in $X^{\text{Tsix}}X^{\text{JF1}}$ and $X^{\text{Tsix}}Y$ post-implantation embryos but not TS cells displayed ectopic *Xist* expression and X-inactivation, we hypothesized that *Tsix* is required in a differentiation-dependent manner, rather than a temporally-specific manner, in the trophoctoderm lineage. We therefore tested if differentiation of $X^{\text{Tsix}}X^{\text{JF1}}$ and $X^{\text{Tsix}}Y$ TS cells would cause *Xist* induction from the *Tsix*-mutant X-chromosome. TS cell differentiation indeed led to ectopic *Xist* induction from the *Tsix*-mutant but not the WT maternal X-chromosome in both male and female cells by RNA FISH (Fig. 3c–e). We confirmed ectopic *Xist* induction from the maternal X^{Tsix} in differentiated TS cells by allele-specific RT-PCR followed by Sanger sequencing (Fig. 3f–g).

Next, we characterized when during differentiation TS cells induce *Xist* from the X^{Tsix} . Only cells devoid of CDX2, a marker of trophoblast progenitor cells, displayed ectopic *Xist* induction (Fig. 4a–d). Moreover, *Xist* induction from the maternal X^{Tsix} coincided with a failure of the mutant TS cells to differentiate to completion. Whereas WT TS cells of both sexes are able to terminally differentiate into trophoblast giant cells, $X^{\text{Tsix}}X^{\text{JF1}}$ and $X^{\text{Tsix}}Y$ TS cells displayed a significant reduction in the percentage of giant cells (Fig. 4e–f). Together, these results demonstrate that *Tsix* prevents *Xist* induction from the maternal X-chromosome during the differentiation of trophoctodermal progenitor cells. Ectopic *Xist* induction and the ensuing inactivation of both Xs in females or of a single X in males results in a paucity of X-linked gene expression, which in turn is expected to cause reduced or stalled cell proliferation and prevent terminal differentiation. A block in TS cell differentiation is also consistent with the variable ectopic *Xist* induction and X-inactivation

in the trophectoderm lineage of $X^{TsixX^{JF1}}$ and X^{TsixY} post-implantation embryos (Fig. 2, Supplementary Fig. 2, Supplementary Movies 1–4).

In addition to TS cells, XEN cells undergo imprinted X-inactivation of the paternal X-chromosome^{43,45}. XEN cells are derived from the primitive endoderm layer of blastocysts that generates the extra-embryonic yolk-sac in later-stage embryos. To test if *Tsix*-mutant XEN cells can stably undergo imprinted X-inactivation, we derived $X^{LabX^{JF1}}$, $X^{TsixX^{JF1}}$, X^{LabY} , and X^{TsixY} XEN cells. By both RNA FISH and allele-specific RT-PCR, we found that, just like in TS cells, XEN cells can stably repress *Xist* from the maternal X-chromosome independently of *Tsix* (Fig. 5).

Disassociation of H3-K27me3 Enrichment and X-inactivation

Xist RNA coating is postulated to lead to X-inactivation in *cis*^{8,46,47}. We noticed, however, that in 5% of the E6.5 X^{TsixX} extra-embryonic cells *Xist* RNA coated both X-chromosomes, but one *Xist* coated X-chromosome remained active, as indicated by expression of *Pgk1* from one of the two Xs in these cells (Fig. 2a–b). In E6.5 X^{TsixY} extra-embryonic nuclei, 17% of cells displayed both ectopic *Xist* RNA coating and *Pgk1* expression from the single X-chromosome (Fig. 2a–b). To determine if *Pgk1* expression from the ectopically *Xist* RNA-coated X-chromosome agreed with expression of other X-linked genes subject to X-inactivation, we performed pair-wise comparisons of expression of *Pgk1* with *Atrx* and *Atrx* with *Rnfl2* (Supplementary Fig. 3). We observed a high level of concordant expression of both sets of X-linked genes, leading to the conclusion that the ectopically *Xist* RNA-coated X-chromosome remained transcriptionally active in a subset of nuclei.

To investigate the uncoupling of *Xist* RNA coating and X-linked gene silencing, we tested whether ectopic *Xist* RNA expression and coating led to the functional enrichment of the Polycomb group proteins on that X-chromosome. *Xist* RNA is thought to induce X-inactivation via the recruitment of protein complexes, most notably the Polycomb repressive complex 2 (PRC2), that inhibit transcription on the inactive-X^{10,12,48}. PRC2 catalyzes methylation of lysine at amino acid position 27 in the tail of histone H3 (H3-K27me3), which is associated with transcriptional repression and is required for imprinted X-inactivation^{10,12,43,49}. Moreover, both PRC2 components and H3-K27me3 accumulate on the inactive-X early during X-inactivation, suggesting an intimate role for PRC2 and H3-K27me3 in the formation of the inactive-X heterochromatin^{10,12}. We therefore tested ectopic accumulation of PRC2-catalyzed H3-K27me3 in E6.5 XX, X^{TsixX} , XY, and X^{TsixY} extra-embryonic cells. We found discordance between *Xist* RNA coating and H3-K27me3 accumulation; nuclei with ectopic *Xist* RNA coating in the *Tsix*-mutant cells did not always display H3-K27me3 enrichment (Fig. 6a–c). In cells with ectopic *Xist* RNA coating and concomitant H3-K27me3 accumulation, however, a substantial percentage (20% of female and 16% of male nuclei) harbored a transcriptionally-competent X-chromosome, as reflected by *Pgk1* expression (Fig. 6b–c). Together, these data demonstrate that *Xist* RNA induction often, but not always, leads to H3-K27me3 accumulation on the X^{Tsix} maternal X-chromosome, and that X-inactivation does not necessarily follow.

Tsix is Dispensable in X-chromosome Reactivation

In addition to preventing Xist RNA expression and X-inactivation, Tsix is also implicated in Xist repression in pluripotent embryonic stem (ES) cells, an *in vitro* analog of the pluripotent epiblast precursor cells within the inner cell mass (ICM) of hatched blastocysts^{37–39}. Loss of Xist RNA coating is a hallmark of the chromosome-wide epigenetic remodeling that accompanies reactivation of the inactive-X in epiblast precursors^{29,30,33}. To test whether Tsix is required to repress Xist in the epiblast precursors, we isolated ICMs from XX and XX^{Tsix} E4.0 embryos. In both XX and XX^{Tsix} ICMs, we observed loss of Xist RNA coating and biallelic expression of *Atrx*, a gene subject to X-inactivation, indicating that reactivation of the paternal-X had occurred in spite of its lacking functional Tsix (Fig. 7a–c). We independently validated reactivation of the X^{Tsix} paternal X-chromosome by allele-specific RT-PCR amplification of the X-linked genes *Pdhal*, *Rnf12*, and *Utx* in E5.0 X^{JF1}X^{Lab} and X^{JF1}X^{Tsix} epiblasts (Fig. 7d). Whereas *Pdhal* and *Rnf12* are subject to X-inactivation, *Utx* escapes X-inactivation and serves as a control for the assay to gauge biallelic X-linked gene expression. If the paternal X-chromosome is reactivated, then transcription of all three genes should be apparent from both X-chromosomes. In agreement, all three genes displayed biallelic expression in both genotypes in E5.0 epiblasts. Based on these data, we conclude that the paternal X-chromosome is reactivated in pluripotent stem cells independently of Tsix.

DISCUSSION

Tsix transcription across the Xist promoter is thought to inhibit Xist expression, and thereby prevent X-inactivation^{17,18}. We show instead that Tsix is not required to repress Xist and prevent X-inactivation in the early embryo and in stem cells of the trophectoderm and primitive endoderm lineages. Earlier studies implicating Tsix in imprinted X-inactivation did not profile the onset of X-inactivation in Tsix-mutant preimplantation embryos or in TS and XEN cells^{15,16}. We find that Tsix expression in *cis* is required to forestall inactivation of the maternal X-chromosome as the trophectoderm cells differentiate. By the post-implantation stage, the Tsix-mutant maternal X-chromosome displayed ectopic Xist RNA coating in 25% and 20% of extra-embryonic cells isolated from E6.5 X^{Tsix}X and X^{Tsix}Y male and female embryos, respectively. The variable induction of Xist reflects a requirement for Tsix in differentiating but not undifferentiated trophoblast cells to prevent Xist induction and X-inactivation.

Xist expression from and coating of the X^{Tsix} maternal X-chromosome coincided with enrichment of the histone modification H3-K27me3 on the mutant-X in many, but not all, X^{Tsix}X (54%) and X^{Tsix}Y (70%) extra-embryonic nuclei. Despite Xist RNA coating and H3-K27me3 accumulation, however, we unexpectedly found that 20% of the X^{Tsix} maternal X-chromosomes in females and 16% in males did not undergo inactivation. This finding is consistent with and extends previous work showing that the recruitment of PRC2 and the catalysis of H3-K27me3 are insufficient to trigger gene silencing by ectopically integrated Xist transgenes or by a mutant Xist RNA expressed from the endogenous locus^{10,12,50}. In these studies, however, the site of integration of the Xist transgene or the specific mutation in Xist can not be excluded as the cause of defective silencing. We find

that ectopic coating of the X-chromosome by an unmodified and endogenous Xist RNA followed by robust H3-K27me3 enrichment nevertheless results in active transcription of endogenous X-linked genes.

Our work shows that the oocyte-derived imprint that prevents inactivation of the maternal X-chromosome in the early embryo does not act through Tsix RNA. We instead propose a model where the oocyte marks the maternal-X with chromosome-wide histone modifications, ensuring that genes along the maternal X-chromosome remain transcriptionally competent during early embryogenesis (Fig. 8). This chromatin profile is sufficient to repress Xist during the initiation of imprinted X-inactivation in the early embryo independently of Tsix RNA. Tsix RNA is also not needed to maintain Xist repression in undifferentiated trophoblast stem cells. During trophoblast differentiation, by contrast, Tsix absence leads to Xist induction from and X-inactivation of the maternal-X. These data are consistent with previous findings that cellular differentiation can trigger X-inactivation defects, for example through large-scale chromatin changes that are inherent to differentiation^{43,51}. The insufficiency of the germline-derived chromatin imprint to prevent inactivation of the maternal X-chromosome is also highlighted by the observations that the maternal-X in parthenogenetic embryos harboring two maternal-Xs is subject to inactivation in extra-embryonic tissues of post-implantation embryos but not in pre-implantation embryos^{25,52}. We further propose that Tsix expression from the maternal X-chromosome is not directly programmed by the oocyte, but is simply a byproduct of the absence of Xist transcription from that chromosome. Indeed, Tsix RNA is induced when Xist expression is absent^{52,53}. Tsix transcription then contributes to the chromatin structure at the Xist promoter region, and these Tsix-induced modifications may assume the Xist-inhibitory role in differentiating trophoblast cells^{17,18}. This mode of lncRNA function in epigenetic transcriptional regulation may also apply to other loci subject to imprinted, parent-of-origin specific gene expression, where opposing sense-antisense lncRNA functions are invoked⁵⁴. The reciprocal regulation by Tsix of Xist, in turn, is the cause of the Tsix-mutant maternal-Xs ectopically accumulating Xist RNA and H3-K27me3 in trophoblast cells. But, despite the enrichment of both Xist RNA and H3-K27me3 a significant percentage of X^{Tsix} maternal X-chromosomes do not undergo inactivation. These findings are consistent with the hypothesis that factors in addition to or other than Xist RNA and H3-K27me3 contribute to the initiation of X-inactivation^{21,43,55}.

Our analysis additionally uncouples Tsix expression from the reactivation of the inactive paternal X-chromosome, a process that characterizes pluripotent epiblast precursor cells in the developing embryo^{29,30}. X-reactivation is also a prominent epigenetic feature of pluripotent ES and iPSC cells³⁴⁻³⁶. Tsix induction from the inactive-X is posited to contribute to Xist repression and to the transcriptional equality of the two Xs in pluripotent cells³⁷⁻³⁹. We find, however, that despite Tsix-absence, Xist RNA is repressed and the paternal X-chromosome is efficiently reactivated in epiblast precursor cells. In addition to by Tsix, Xist repression is postulated to occur via the pluripotency factors NANOG, OCT4, and SOX2, which are expressed in pluripotent epiblast progenitors and have been shown to bind within intron 1 of *Xist* in undifferentiated ES cells^{37,39,56,57}. Thus, it is conceivable that these pluripotency factors may function to repress Xist and induce reactivation of the X^{Tsix}

paternal X-chromosome in the embryo. Countering this argument, however, Xist RNA upregulation and coating during random X-inactivation in the epiblast occurs despite high levels of NANOG, OCT4, and SOX2 expression, suggesting the involvement of additional factors in both Xist repression and X-reactivation⁵⁸.

METHODS

Ethics Statement

This study was performed in strict accordance with the recommendations in the Guide for the Care and Use of Laboratory Animals of the National Institutes of Health. All animals were handled according to protocols approved by the University Committee on Use and Care of Animals (UCUCA) at the University of Michigan (Protocol #PRO00004007).

Mice

Mice harboring the *Tsix*^{AA2 1.7} mutation were generated from targeted ES cells that were a kind gift of Takashi Sado and have been described elsewhere^{16,55}. *Tsix* mutant mice were maintained on a CD1 strain background. The mice recapitulate the published transmission frequency and phenotype¹⁶. The X-linked *GFP* transgenic (*X-GFP*) strain is available via Jackson Labs [Tg(CAG-EGFP)D4Nagy/J] and have been described previously^{21,43,55,59}. The *M. molossinus* JF1 strain was sourced from Jackson Laboratories (JF1/ms).

Embryo Dissections and Processing

E3.5-E4.0 embryos were flushed from the uterine limbs in 1X PBS (Invitrogen, #14200075) containing 6% bovine serum albumin (BSA; Invitrogen, #15260037). Zona pellucidas surrounding E3.5 embryos were removed through incubation in cold acidic Tyrode's Solution (Sigma, #T1788), immediately followed by neutralization through several transfers of cold M2 medium (Sigma, #M7167). GFP fluorescence conferred by the paternal transmission of the *X-GFP* transgene was used to distinguish female from male embryos, since only females inherit the paternal-X. Embryos were rinsed in 1X PBS with 6 mg/ml BSA, plated on gelatin-coated glass coverslips, excess solution was aspirated, and the embryos air-dried for 15 min. After drying, embryos were permeabilized and fixed with 50 mL of 1X PBS containing 0.05% Tergitol (Sigma, #NP407) together with 1% paraformaldehyde for 10 min. Excess solution was tapped off, and coverslips were rinsed 3X with 70% ethanol and stored in 70% ethanol at -20°C prior to RNA fluorescence *in situ* hybridization (RNA FISH) staining.

For isolation of E5.0-E6.5 embryos, dissections were carried out in 1X PBS containing 6% BSA. Individual implantation sites were cut from the uterine limbs, and decidua were removed with forceps. Embryos were dissected from the decidua, and the Reichert's membranes surrounding post-implantation embryos were removed using fine forceps. For separation of extra-embryonic and epiblast portions of E6.5 embryos, fine forceps were used to physically bisect the embryos at the junction of the extra-embryonic ectoderm and epiblast. Immunofluorescence (IF) and/or RNA FISH staining were performed as described below.

Trophoblast Stem Cells

E3.5 embryos were flushed out from the uterus with MEM α (Invitrogen, #12561) with 10% fetal bovine serum (FBS; Invitrogen, #10439-024) and plated on mouse embryonic fibroblast (MEF) cells in medium consisting of RPMI (Invitrogen, #21870076) with 20% FBS, 1 mM sodium pyruvate (Invitrogen, #11360-070), 100 μ M β -mercaptoethanol (Sigma, #M7522), 2 mM L-glutamine (Invitrogen, #25030), 37.5 ng/mL FGF4 (R&D Systems, #235-F4-025), and 1.5 μ g/mL heparin (Sigma, #H3149-10KU). Following five days of growth at 37°C with 5% CO₂ blastocyst outgrowths were dissociated with 0.05% trypsin (Invitrogen, #25300-054). Dissociated cells were plated on MEFs and cultured at 37°C with 5% CO₂. RNA was harvested from TS cells using Trizol (Invitrogen, #15596-018) and RT-PCR was performed as described below. For RNA FISH and/or IF, TS cells were split onto gelatin-coated glass coverslips and allowed to grow for 3–6 days. The cells were then permeabilized through sequential treatment with ice-cold cytoskeletal extraction buffer (CSK; 100 mM NaCl, 300 mM sucrose, 3 mM MgCl₂, and 10 mM PIPES buffer, pH 6.8) for 30 sec, ice-cold CSK buffer containing 0.4% Triton X-100 (Fisher Scientific, #EP151) for 30 sec, followed twice with ice-cold CSK for 30 sec each. After permeabilization, cells were fixed by incubation in 4% paraformaldehyde for 10 min. Cells were then rinsed 3X in 70% ethanol and stored in 70% ethanol at –20°C prior to IF and/or RNA FISH. For differentiation of TS cells, cells were split onto gelatinized dishes or coverslips and cultured for 6 days (d6) in media without FGF4 or heparin. On d6 of differentiation, RNA was harvested or cells were processed as described above for IF and RNA FISH.

RNA Fluorescence *in situ* Hybridization (RNA FISH)

Double-stranded RNA FISH (dsRNA FISH) probes were created by randomly priming DNA templates using BioPrime DNA Labeling System (Invitrogen, #18094011). Probes were labeled with Fluorescein-12-dUTP (Invitrogen), Cy3-dCTP (GE Healthcare, #PA53021), or Cy5-dCTP (GE Healthcare, #PA55031). Labeled probes for multiple genes were precipitated in a 3M sodium acetate (Teknova, #S0298) solution along with 300 mg of yeast tRNA (Invitrogen, #15401-029), 15 μ g of mouse COT-1 DNA (Invitrogen, #18440-016) and 150 mg of sheared, boiled salmon sperm DNA (Invitrogen, #15632-011). The solution was then spun at 15,000 rpm for 20 min at 4°C. The resulting pellet was washed in 70% ethanol, then washed in 100% ethanol, dried, and re-suspended in deionized formamide (ISC Bioexpress, #0606-500ML). The re-suspended probe was denatured via incubation at 90°C for 10 min followed by an immediate 5 min incubation on ice. A 2X hybridization solution consisting of 4X SSC, 20% Dextran sulfate (Millipore, #S4030), and 2.5 mg/ml purified BSA (New England Biolabs, #B9001S) was added to the denatured solution. The probe was then pre-annealed by incubation at 37°C for 1 hr to minimize probe hybridization to repetitive sequences. Probes were stored at –20°C until use. Strand-specific RNA FISH (ssRNA FISH) probes were labeled with Fluorescein-12-UTP (Roche, #11427857910) or Cy3 CTP (GE Healthcare, #25801086) using the Invitrogen MAXIscript Kit (Invitrogen, #AM-1324). Labeled probes were column purified (Roche, #11814427001). The labeled probes were then precipitated in an 0.25M ammonium acetate solution essentially as described above for dsRNA FISH probes, but without the addition of COT-1 DNA. Probes were resuspended as described for dsRNA FISH probes and stored at –20 without preannealing. Embryos, embryo fragments, or TS cells mounted on coverslips were

dehydrated through 2 min incubations in 70%, 85%, 95%, and 100% ethanol solutions and subsequently air-dried. The coverslips were then hybridized to the probe overnight in a humid chamber at 37°C. The samples were then washed 3X for 7 min each while shaking at 39°C with 2XSSC/50% formamide, 2X with 2X SSC, and 2X with 1X SSC. A 1:250,000 dilution of DAPI (Invitrogen, #D21490) was added to the third 2X SSC wash. The embryos were then mounted in Vectashield (Vector Labs, #H-1200). A total of 64 E3.5 embryos (18 XX, 16 X^{Tsix}X, 17 XY, and 13 X^{Tsix}Y) from eight litters were analyzed by RNA FISH. For assessment of E4.0 embryos, 13 embryos from 2 litters were analyzed by RNA FISH.

Whole-mount RNA FISH

E6.5 embryos were permeabilized through sequential transfers into ice-cold cytoskeletal extraction buffer (CSK; 100 mM NaCl, 300 mM sucrose, 3 mM MgCl₂, and 10 mM PIPES buffer, pH 6.8) for 1 min, ice-cold CSK buffer containing 0.4% Triton X-100 (Fisher Scientific, #EP151) for 10 min, followed twice with ice-cold CSK for 1 min each. After permeabilization, the embryos were rinsed 3X in 70% ethanol and stored in 70% ethanol at -20°C prior to RNA FISH. Embryos were rehydrated by incubating in decreasing concentrations of ethanol diluted in 2X SSC for 3 min each. Embryos were then placed in a droplet of 2X SSC in a depression well slide (Fisher Scientific, #S175201). Embryos were incubated in 10 µL of probe (probe generation described in Experimental Procedures) overnight in a depression-well slide (Fisher Scientific, #S175201) sealed with a glass coverslip in a humid chamber at 37°C. Embryos were then rinsed with pre-warmed (50°C) 2X SSC/50% deionized formamide (Amresco, #NC9473844) and washed with 50% formamide/2X SSC solution 3X for 15 min each at 50°C, with periodic agitation via pipetting. Embryos were next washed with pre-warmed 2X SSC (50°C) 3X for 15 min each. In the first two 2X SSC washes, 4',6-diamidino-2-phenylindole (DAPI; Invitrogen, #D21490) was included at a dilution of 1:200,000. Embryos were next washed in pre-warmed 1X SSC solution (50°C) 2X for 15 min each. After washing, embryos were processed through sequential incubations in PBS with 25%, 50%, 75%, and 100% Vectashield (Vector Labs, #H-1000) and mounted in depression-well slides with Vectashield.

PCR and Allele-specific Reverse Transcriptase Polymerase Chain Reaction (RT-PCR)

Embryos and embryo fragments were lysed in 100 µL of lysis/binding buffer (Dynabeads mRNA DIRECT Micro Kit; Invitrogen, #610.21). Messenger RNA was isolated by following manufacturer's instructions. SuperScript III One-Step RT-PCR Platinum *Taq* enzyme mixture (Invitrogen, #12574-035) was used to prepare and amplify the complementary DNA (cDNA). Strand-specific reverse transcription of *Xist* was performed using the XR-9816 primer, which spans bp 9815-9775 of *Xist* (CTCCACCTAGGGATCGTCAA). For PCR amplification, the forward primer XF-9229, which spans bp 9229-9248 of *Xist* (GACAACAATGGGAGCTGGTT) was added upon completion of the RT reaction and prior to the PCR step. The *Xist* amplicon spans two introns, thus permits distinguishing genomic DNA sequence amplification by size. Genomic PCR for the *Xist* locus was performed the XR-271 primer, which spans 721-740 of *Xist* (CGGGGCTTGGTGGATGGAAAT), and XF-1083, which spans 1083-1064 of *Xist* (GCACAACCCCGCAAATGCTA). RT-PCR amplification of *Tsix* exon 4 in in TS cells

and E3.5 embryos was performed using the RT primer TR-4224, which spans base pairs 4224–4205 of *Tsix* (TCGGATCCCCTACAGATGA), and the forward PCR primer TF-3796, which spans base pairs 3796–3815 of *Tsix* (CTAAGAGCACCTGGCTCCAC). For E3.5 blastocysts, an additional round of nested PCR was performed to detect *Tsix* using TR-4224 and TF-3987, which spans base pairs 3987–4006 of *Tsix* (TCCCAATTCTTGCAAACCTC). RT-PCR for the *Tsix* amplicon spanning exons 2–4 was performed using the RT primer TR-732, which spans base pairs 732–713 of *Tsix* (GGAGAGCGCATGCTTGCAAT) and the forward PCR primer TF-350, which spans base pairs 350–369 of *Tsix* (CCTGCAAGCGCTACACACTT). RT-PCR for β -Actin was performed using the RT primer β A-R, which spans base pairs 673–655 of *Actb* (GTAGCCACGCTCGGTCAGG), and the forward primer β A-F, which spans base pairs 142–159 of *Actb* (CGCGGGCGACGATGCTCC). Amplified cDNAs were run on agarose gels and purified using the Clontech NucleoSpin Kit (Clontech, #740609). The purified cDNAs were then sequenced and sequencing traces were examined for single nucleotide polymorphisms (SNPs) characteristic of the *M. musculus*-derived X^{JF1} chromosome and the *M. domesticus*-derived wild-type X^{Lab} and mutant X^{Tsix} chromosomes. The SNP within the *Xist* amplicon localizes to bp 9399 of *Xist*. The *M. domesticus*-derived wild-type X^{Lab} and mutant X^{Tsix} SNP is an adenosine while the *M. musculus* derived X^{JF1} SNP is a guanosine. The SNP within the *Xist* genomic PCR amplicon localizes to bp 804 of *Xist*. The *M. domesticus*-derived wild-type X^{Lab} and mutant X^{Tsix} SNP is a thymidine while the *M. musculus* derived X^{JF1} SNP is an adenosine. SNPs within X-linked genes *Rnf12* (bp 860, NM_011276), *Pdha1* (bp 969, NM_008810.2), and *Utx* (bp 1383, NM_009483.1) have been described previously²¹. RT and PCR primers for these genes have also been described previously^{16,21}.

Immunofluorescence

Embryo fragments mounted on glass coverslips were washed 3X in PBS for 3 min each while shaking. The fragments were then incubated in blocking buffer (0.5 mg/mL BSA (New England Biolabs, #B9001S), 50 μ g/mL yeast tRNA, 80 units/mL RNaseOUT (Invitrogen, #10777–019), and 0.2% Tween 20, in PBS) in a humid chamber for 30 min at 37°C. The samples were next incubated with primary antibody, diluted in blocking buffer, for 1 hr in the humid chamber at 37°C. The H3-K27me3 (EMD Millipore, #ABE44) was used at 1:2500 dilution in blocking buffer. The CDX2 antibody (BioGenex, #MU328A–UC) was used at a 1:75 dilution in blocking buffer. The p57^{Kip2} antibody (Thermo Scientific, #RB-1637) was used at a 1:150 dilution in blocking buffer. Following three washes in PBS/0.2% Tween-20 for 3 min each while shaking, the embryos were incubated in blocking buffer for 5 min at 37°C in the humid chamber. The embryos were then incubated in blocking buffer containing a 1:300 dilution of fluorescently-conjugated secondary antibody (Alexa Fluor, Invitrogen) for 30 min in a humid chamber, followed by three washes in PBS/0.2% Tween-20 while shaking for 3 min each. The samples were then processed for RNA FISH as described above.

Immunosurgery

To isolate the inner cell mass (ICM) of E4.0 embryos, embryos were incubated in pre-warmed rabbit anti-mouse serum (Rockland Immunochemicals, #110–4101) at a 1:5 dilution

in M2 medium for 1 hr in a humid chamber at 37°C. After briefly rinsing in M2 medium, embryos were incubated in pre-warmed guinea pig complement (Sigma, cat No. S1639) at a 1:5 dilution in M2 medium for 45 min to 1 hr in a humid chamber at 37°C. The embryos were then repeatedly pipetted using a finely-pulled glass Pasteur pipette to remove trophectoderm cells. Isolated ICMs were treated with 0.05% trypsin for 10 min for mild dissociation, then incubated in M2 media with 10% fetal bovine serum (FBS) for 10 min to neutralize the trypsin. ICMs were then rinsed in 1X PBS and permeabilized through sequential transfers into ice-cold CSK for 1 min, ice-cold CSK containing 0.4% Triton X-100 buffer for 5 min, followed twice with ice-cold CSK for 1 min each. ICMs were mounted on a glass coverslip coated with 1X Denhardt's solution in a small drop of ice-cold solution of 1X PBS containing 1% paraformaldehyde and 20% CSK buffer. Excess solution was aspirated off and the coverslip air-dried for 15 min. The ICMs were then fixed in cold 3% paraformaldehyde for 10 min. After fixation, the coverslips were rinsed 3X in 70% ethanol and stored in 70% ethanol at -20°C prior to use.

Microscopy

Stained samples were imaged using a Nikon Eclipse TiE inverted microscope with a Photometrics CCD camera. The images were deconvolved and uniformly processed using NIS-Elements software.

Statistics

Comparisons between gene expression patterns were performed using a two-tailed Fisher's exact test. $p=0.01$ was used as the cutoff for statistical significance.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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REFERENCES

1. Morey C, Avner P. The demoiselle of X-inactivation: 50 years old and as trendy and mesmerising as ever. *PLoS Genet.* 2011; 7:e1002212. [PubMed: 21811421]
2. Plath K, Mlynarczyk-Evans S, Nusinow DA, Panning B. Xist RNA and the mechanism of X chromosome inactivation. *Annu Rev Genet.* 2002; 36:233-78. [PubMed: 12429693]
3. Lee JT. Gracefully ageing at 50, X-chromosome inactivation becomes a paradigm for RNA and chromatin control. *Nat Rev Mol Cell Biol.* 2011; 12:815-26. [PubMed: 22108600]
4. Rastan S. Non-random X-chromosome inactivation in mouse X-autosome translocation embryos--location of the inactivation centre. *J Embryol Exp Morphol.* 1983; 78:1-22. [PubMed: 6198418]

5. Rastan S, Robertson EJ. X-chromosome deletions in embryo-derived (EK) cell lines associated with lack of X-chromosome inactivation. *J Embryol Exp Morphol.* 1985; 90:379–88. [PubMed: 3834036]
6. Marahrens Y, Loring J, Jaenisch R. Role of the Xist gene in X chromosome choosing. *Cell.* 1998; 92:657–64. [PubMed: 9506520]
7. Penny GD, Kay GF, Sheardown SA, Rastan S, Brockdorff N. Requirement for Xist in X chromosome inactivation. *Nature.* 1996; 379:131–7. [PubMed: 8538762]
8. Payer B, Lee JT. X chromosome dosage compensation: how mammals keep the balance. *Annu Rev Genet.* 2008; 42:733–72. [PubMed: 18729722]
9. Kalantry S. Recent advances in X-chromosome inactivation. *J Cell Physiol.* 2011; 226:1714–8. [PubMed: 21344379]
10. Plath K, et al. Role of histone H3 lysine 27 methylation in X inactivation. *Science.* 2003; 300:131–5. [PubMed: 12649488]
11. Plath K, et al. Developmentally regulated alterations in Polycomb repressive complex 1 proteins on the inactive X chromosome. *J Cell Biol.* 2004; 167:1025–35. [PubMed: 15596546]
12. Silva J, et al. Establishment of histone h3 methylation on the inactive X chromosome requires transient recruitment of Eed-Enx1 polycomb group complexes. *Dev Cell.* 2003; 4:481–95. [PubMed: 12689588]
13. Mak W, et al. Mitotically stable association of polycomb group proteins eed and enx1 with the inactive x chromosome in trophoblast stem cells. *Curr Biol.* 2002; 12:1016–20. [PubMed: 12123576]
14. Lee JT, Lu N. Targeted mutagenesis of Tsix leads to nonrandom X inactivation. *Cell.* 1999; 99:47–57. [PubMed: 10520993]
15. Lee JT. Disruption of imprinted X inactivation by parent-of-origin effects at Tsix. *Cell.* 2000; 103:17–27. [PubMed: 11051544]
16. Sado T, Wang Z, Sasaki H, Li E. Regulation of imprinted X-chromosome inactivation in mice by Tsix. *Development.* 2001; 128:1275–86. [PubMed: 11262229]
17. Sado T, Hoki Y, Sasaki H. Tsix silences Xist through modification of chromatin structure. *Dev Cell.* 2005; 9:159–65. [PubMed: 15992549]
18. Navarro P, Pichard S, Ciaudo C, Avner P, Rougeulle C. Tsix transcription across the Xist gene alters chromatin conformation without affecting Xist transcription: implications for X-chromosome inactivation. *Genes Dev.* 2005; 19:1474–84. [PubMed: 15964997]
19. Okamoto I, Otte AP, Allis CD, Reinberg D, Heard E. Epigenetic dynamics of imprinted X inactivation during early mouse development. *Science.* 2004; 303:644–9. [PubMed: 14671313]
20. Patrat C, et al. Dynamic changes in paternal X-chromosome activity during imprinted X-chromosome inactivation in mice. *Proc Natl Acad Sci U S A.* 2009; 106:5198–5203. [PubMed: 19273861]
21. Kalantry S, Purushothaman S, Bowen RB, Starmer J, Magnuson T. Evidence of Xist RNA-independent initiation of mouse imprinted X-chromosome inactivation. *Nature.* 2009; 460:647–651. [PubMed: 19571810]
22. Namekawa SH, Payer B, Huynh KD, Jaenisch R, Lee JT. Two-step imprinted X inactivation: repeat versus genic silencing in the mouse. *Mol Cell Biol.* 2010; 30:3187–205. [PubMed: 20404085]
23. Lee JT, Bartolomei MS. X-inactivation, imprinting, and long noncoding RNAs in health and disease. *Cell.* 2013; 152:1308–23. [PubMed: 23498939]
24. Lyon MF, Rastan S. Parental source of chromosome imprinting and its relevance for X chromosome inactivation. *Differentiation.* 1984; 26:63–7. [PubMed: 6714546]
25. Goto Y, Takagi N. Maternally inherited X chromosome is not inactivated in mouse blastocysts due to parental imprinting. *Chromosome Res.* 2000; 8:101–9. [PubMed: 10780698]
26. Tada T, et al. Imprint switching for non-random X-chromosome inactivation during mouse oocyte growth. *Development.* 2000; 127:3101–5. [PubMed: 10862747]

27. Kay GF, Barton SC, Surani MA, Rastan S. Imprinting and X chromosome counting mechanisms determine Xist expression in early mouse development. *Cell*. 1994; 77:639–50. [PubMed: 8205614]
28. Okamoto I, Tan S, Takagi N. X-chromosome inactivation in XX androgenetic mouse embryos surviving implantation. *Development*. 2000; 127:4137–45. [PubMed: 10976046]
29. Mak W, et al. Reactivation of the paternal X chromosome in early mouse embryos. *Science*. 2004; 303:666–9. [PubMed: 14752160]
30. Williams LH, Kalantry S, Starmer J, Magnuson T. Transcription precedes loss of Xist coating and depletion of H3K27me3 during X-chromosome reprogramming in the mouse inner cell mass. *Development*. 2011; 138:2049–57. [PubMed: 21471155]
31. Hogan, B.; Beddington, R.; Costantini, F.; Lacy, E. *Manipulating the Mouse Embryo*. Cold Spring Harbor Press; 1994.
32. Rastan S. Timing of X-chromosome inactivation in postimplantation mouse embryos. *J Embryol Exp Morphol*. 1982; 71:11–24. [PubMed: 6185603]
33. Sheardown SA, et al. Stabilization of Xist RNA mediates initiation of X chromosome inactivation. *Cell*. 1997; 91:99–107. [PubMed: 9335338]
34. Payer B, Lee JT, Namekawa SH. X-inactivation and X-reactivation: epigenetic hallmarks of mammalian reproduction and pluripotent stem cells. *Hum Genet*. 2011; 130:265–80. [PubMed: 21667284]
35. Minkovsky A, Patel S, Plath K. Concise review: Pluripotency and the transcriptional inactivation of the female Mammalian X chromosome. *Stem Cells*. 2011; 30:48–54. [PubMed: 21997775]
36. Plath K, Lowry WE. Progress in understanding reprogramming to the induced pluripotent state. *Nat Rev Genet*. 2011; 12:253–65. [PubMed: 21415849]
37. Navarro P, et al. Molecular coupling of Tsix regulation and pluripotency. *Nature*. 2010; 468:457–60. [PubMed: 21085182]
38. Navarro P, et al. A role for non-coding Tsix transcription in partitioning chromatin domains within the mouse X-inactivation centre. *Epigenetics Chromatin*. 2009; 2:8. [PubMed: 19615107]
39. Nesterova TB, et al. Pluripotency factor binding and Tsix expression act synergistically to repress Xist in undifferentiated embryonic stem cells. *Epigenetics Chromatin*. 2011; 4:17. [PubMed: 21982142]
40. Vigneau S, Augui S, Navarro P, Avner P, Clerc P. An essential role for the DXPas34 tandem repeat and Tsix transcription in the counting process of X chromosome inactivation. *Proc Natl Acad Sci U S A*. 2006; 103:7390–5. [PubMed: 16648248]
41. Stavropoulos N, Rowntree RK, Lee JT. Identification of developmentally specific enhancers for Tsix in the regulation of X chromosome inactivation. *Mol Cell Biol*. 2005; 25:2757–69. [PubMed: 15767680]
42. Cohen DE, et al. The DXPas34 repeat regulates random and imprinted X inactivation. *Dev Cell*. 2007; 12:57–71. [PubMed: 17199041]
43. Kalantry S, et al. The Polycomb group protein Eed protects the inactive X-chromosome from differentiation-induced reactivation. *Nat Cell Biol*. 2006; 8:195–202. [PubMed: 16415857]
44. Tanaka S, Kunath T, Hadjantonakis AK, Nagy A, Rossant J. Promotion of trophoblast stem cell proliferation by FGF4. *Science*. 1998; 282:2072–5. [PubMed: 9851926]
45. Kunath T, et al. Imprinted X-inactivation in extra-embryonic endoderm cell lines from mouse blastocysts. *Development*. 2005; 132:1649–61. [PubMed: 15753215]
46. Schulz EG, Heard E. Role and control of X chromosome dosage in mammalian development. *Curr Opin Genet Dev*. 2013
47. Brockdorff N. Chromosome silencing mechanisms in X-chromosome inactivation: unknown unknowns. *Development*. 2012; 138:5057–65. [PubMed: 22069184]
48. Schoeftner S, et al. Recruitment of PRC1 function at the initiation of X inactivation independent of PRC2 and silencing. *EMBO J*. 2006; 25:3110–22. [PubMed: 16763550]
49. Wang J, et al. Imprinted X inactivation maintained by a mouse Polycomb group gene. *Nat Genet*. 2001; 28:371–5. [PubMed: 11479595]

50. Wutz A, Rasmussen TP, Jaenisch R. Chromosomal silencing and localization are mediated by different domains of Xist RNA. *Nat Genet.* 2002; 30:167–74. [PubMed: 11780141]
51. Corbel C, Diabangouaya P, Gendrel AV, Chow JC, Heard E. Unusual chromatin status and organization of the inactive X chromosome in murine trophoblast giant cells. *Development.* 2013; 140:861–72. [PubMed: 23362347]
52. Rastan S, Kaufman MH, Handyside AH, Lyon MF. X-chromosome inactivation in extra-embryonic membranes of diploid parthenogenetic mouse embryos demonstrated by differential staining. *Nature.* 1980; 288:172–3. [PubMed: 7432515]
53. Hoki Y, et al. A proximal conserved repeat in the Xist gene is essential as a genomic element for X-inactivation in mouse. *Development.* 2009; 136:139–46. [PubMed: 19036803]
54. Barlow DP. Genomic imprinting: a mammalian epigenetic discovery model. *Annu Rev Genet.* 2011; 45:379–403. [PubMed: 21942369]
55. Kalantry S, Magnuson T. The Polycomb group protein EED is dispensable for the initiation of random X-chromosome inactivation. *PLoS Genet.* 2006; 2:e66. [PubMed: 16680199]
56. Navarro P, et al. Molecular coupling of Xist regulation and pluripotency. *Science.* 2008; 321:1693–5. [PubMed: 18802003]
57. Donohoe ME, Silva SS, Pinter SF, Xu N, Lee JT. The pluripotency factor Oct4 interacts with Ctf and also controls X-chromosome pairing and counting. *Nature.* 2009; 460:128–32. [PubMed: 19536159]
58. Pfister S, Steiner KA, Tam PP. Gene expression pattern and progression of embryogenesis in the immediate post-implantation period of mouse development. *Gene Expr Patterns.* 2007; 7:558–73. [PubMed: 17331809]
59. Hadjantonakis AK, Gertsenstein M, Ikawa M, Okabe M, Nagy A. Non-invasive sexing of preimplantation stage mammalian embryos. *Nat Genet.* 1998; 19:220–2. [PubMed: 9662390]

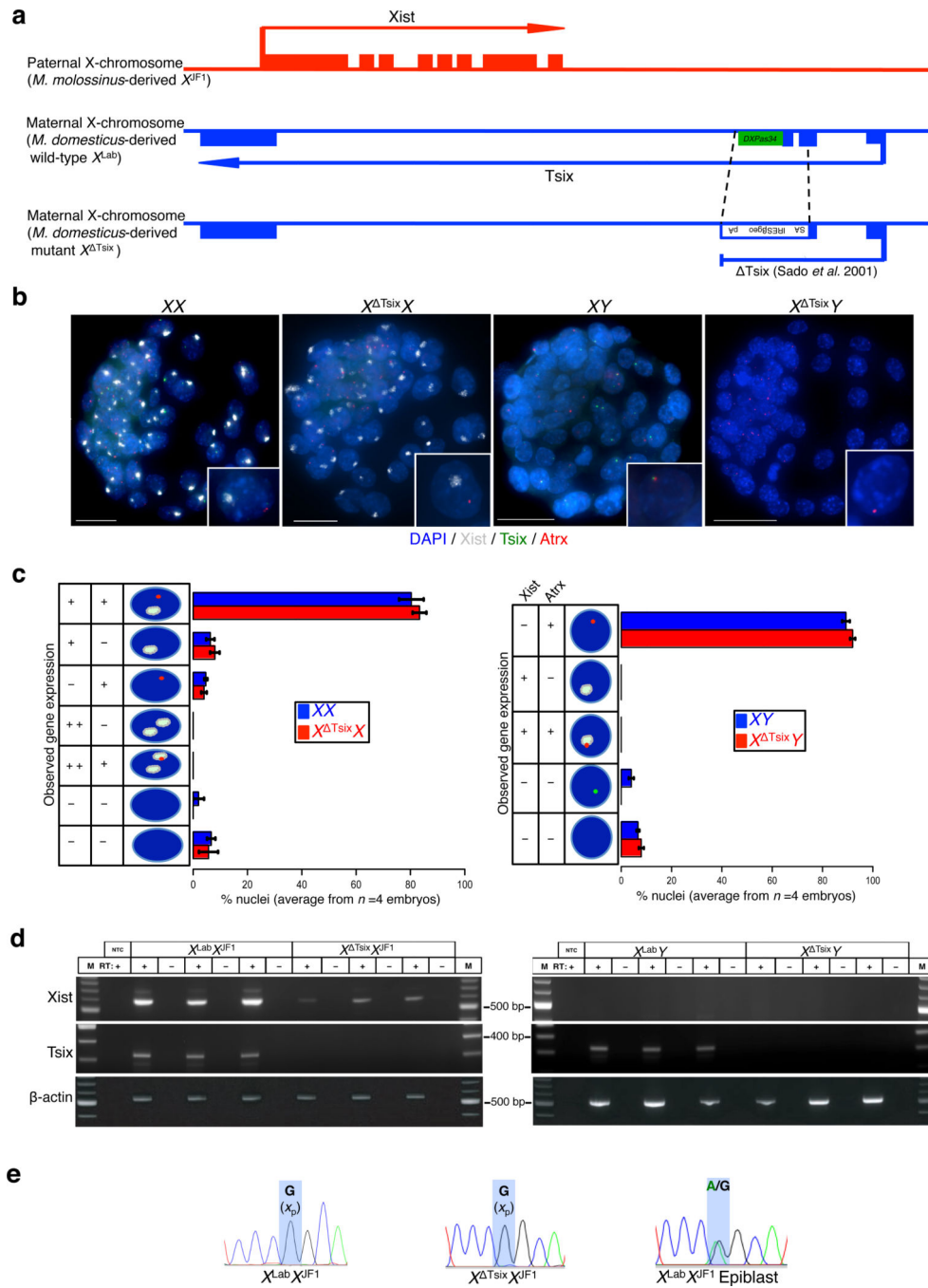


Figure 1. Absence of ectopic Xist induction from the X^{Tsix} maternal X-chromosome in embryonic day (E) 3.5 blastocyst embryos
(a) Schematic representation of the genomic structure of *Xist*, *Tsix*, and the *Tsix* RNA truncation mutant X^{Tsix}
(b) RNA FISH detection of Xist (white), *Tsix* (green) and Atrx (red) RNAs in representative E3.5 embryos. Nuclei are stained blue with DAPI. Insets show representative nuclei. Scale bar, 25 μ m.
(c) Quantification of Xist, *Tsix*, and Atrx RNA expression patterns in blastocyst nuclei. The X-axis of each graph represents the average % nuclei observed in each class for each genotype. $n=4$ embryos per genotype. Diagrams along

the Y-axis depict all observed expression patterns. +, RNA expression detected from a single X-chromosome; ++, RNA expression detected from both X-chromosomes; -, absence of RNA detection. Gene expression pattern does not differ significantly between wild-type and Tsix mutant blastocysts (Fisher's exact test). Error bars, S.D. **(d)** RT-PCR detection of Xist, Tsix, and control b-actin RNAs. Three individual embryos are shown for each genotype. M, marker; NTC, no template control; +, reaction with reverse transcriptase (RT); -, no RT control lane. **(e)** Sanger sequencing chromatograms of representative Xist RT-PCR products. Highlights mark a single nucleotide polymorphism that differs between the maternal X^{Lab} / X^{Tsix} alleles and the paternal X^{JF1} allele (see Methods). Both $X^{Lab}X^{JF1}$ and $X^{Tsix}X^{JF1}$ females express Xist only from the paternally-inherited X-chromosome (Xp). The $X^{Lab}X^{JF1}$ epiblast is a control sample displaying expression from both parental alleles.

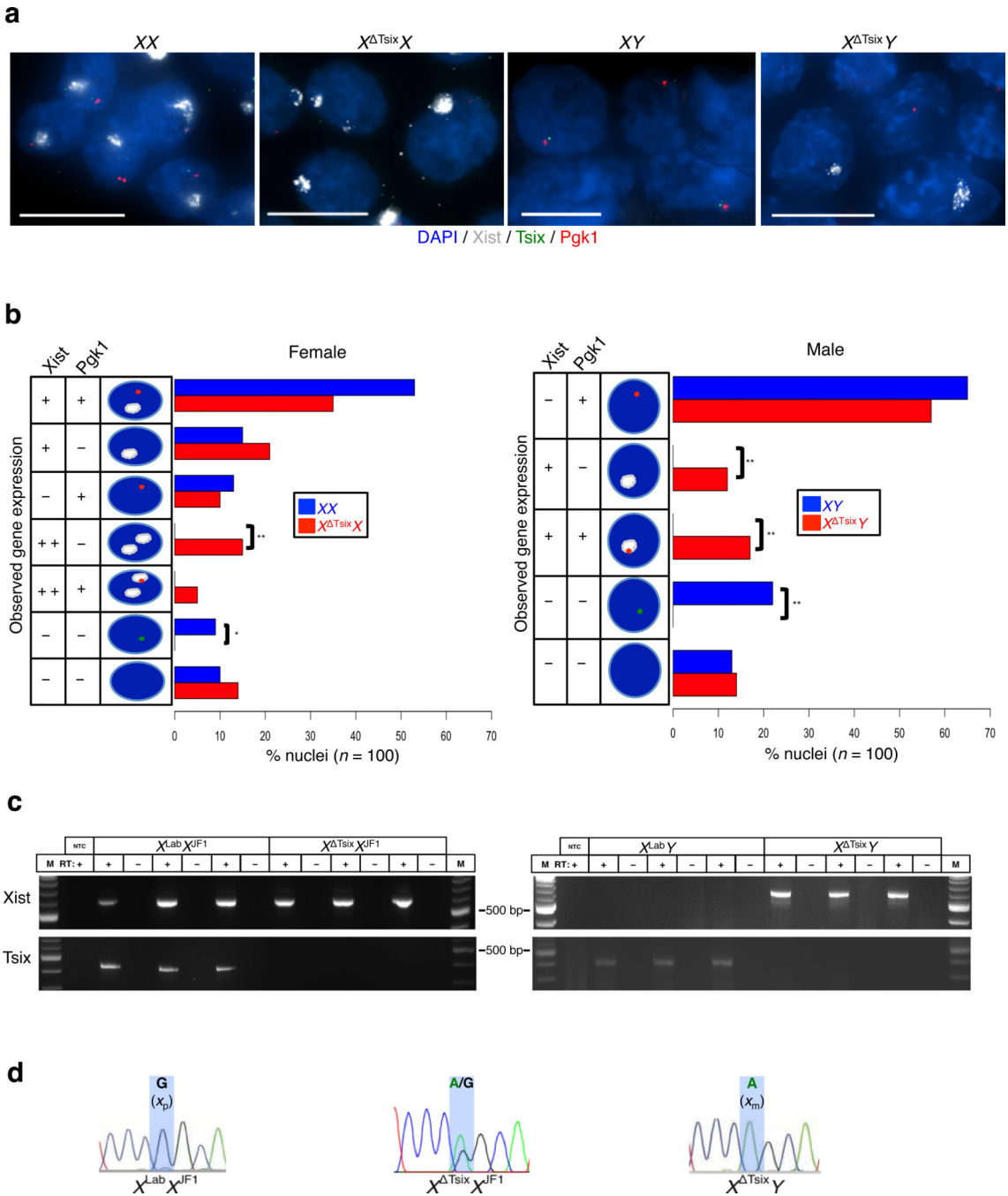


Figure 2. Xist induction from the X^{Tsix} maternal X-chromosome in E6.5 extra-embryonic cells
(a) RNA FISH detection of Xist (white), Tsix (green), and Pgl1 (red) RNAs in E6.5 extra-embryonic cells. Nuclei are stained blue with DAPI. Dashed boxes mark representative nuclei. Scale bar, 10 μ m. **(b)** Quantification of Xist, Tsix, and Pgl1 RNA expression patterns. The X-axis of each graph represents the % nuclei in each class out of 100 total nuclei counted per genotype (from $n > 3$ embryos per genotype). Diagrams along the Y-axis depict all observed expression patterns. +, RNA expression detected from a single X-chromosome; ++, RNA expression detected from both X-chromosomes; -, absence of RNA

detection. Pairwise comparisons of the frequency of individual gene expression patterns between wild-type and X^{Tsix} mutant embryos were performed using Fisher's exact test. *, $0.001 < p < 0.01$; **, $p < 0.001$. Extra-embryonic cells show significantly increased level of inactivation of the X^{Tsix} X-chromosome ($p=0.0003$ for males; $p=3.5 \times 10^{-5}$ for females). **(c)** RT-PCR detection of Xist and Tsix RNAs in extra-embryonic tissues from individual E6.5 embryos. Results from three individual embryos of each genotype are shown. M, marker; NTC, no template control; +, RT; -, no RT control lane. **(d)** Sanger sequencing chromatograms of Xist RT-PCR products. Highlights mark a single nucleotide polymorphism that differs between the maternal X^{Lab} / X^{Tsix} alleles and the paternal X^{JF1} allele. $X^{Lab}X^{JF1}$ females express Xist only from the paternally-inherited X-chromosome, while $X^{Tsix}X^{JF1}$ females express Xist biallelically in extra-embryonic tissues. $X^{Tsix}Y$ embryos variably express Xist from the maternally-inherited X-chromosome.

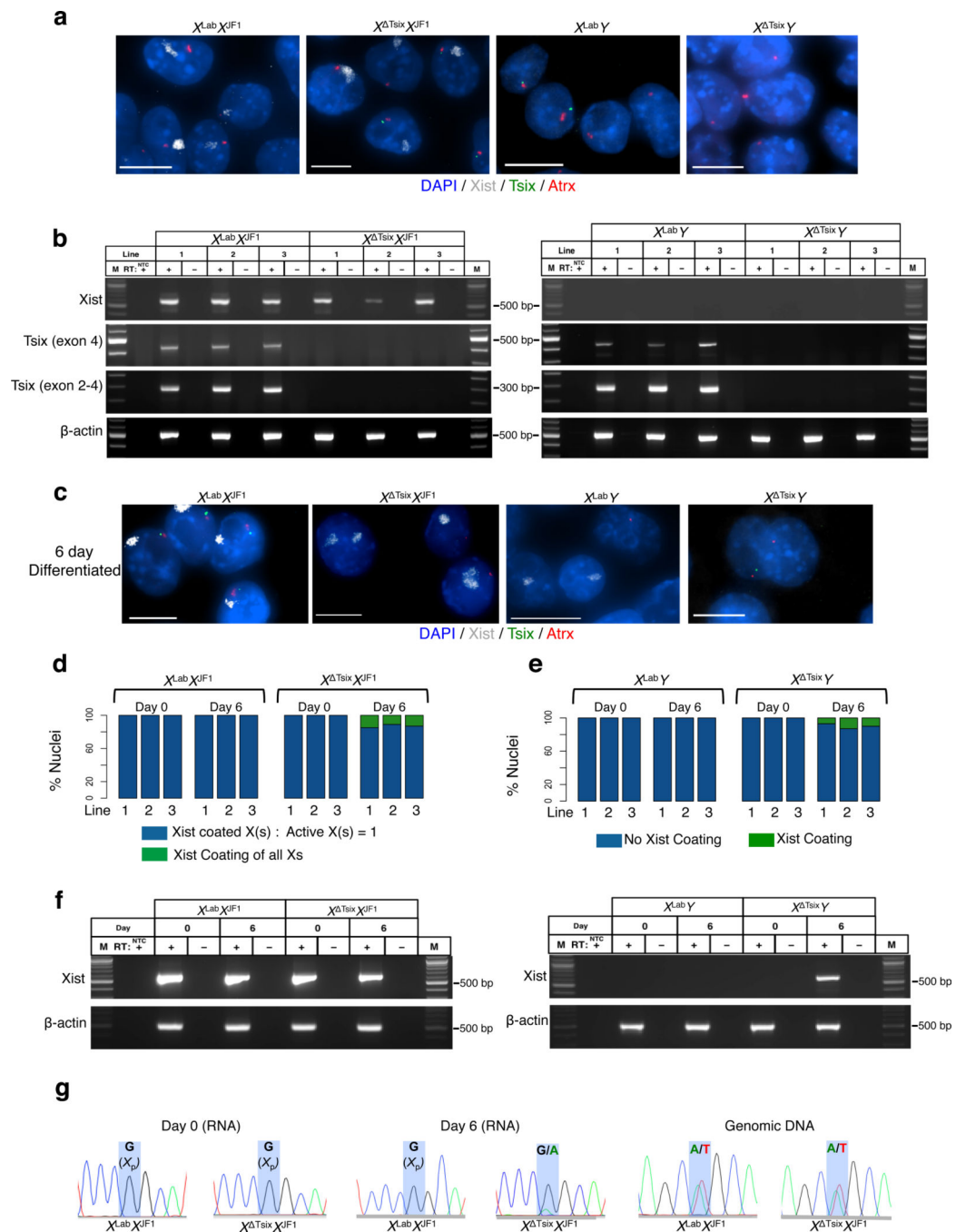


Figure 3. The X^{Tsix} maternal X-chromosome displays ectopic Xist induction only upon differentiation in trophoblast stem (TS) cells
 RNA FISH detection of Xist (white), Tsix (green), and Atrx (red) RNAs in representative TS cell lines. Nuclei are stained blue with DAPI. Scale bar, 10 μm. Three cell lines of each genotype were analyzed. **(b)** RT-PCR detection of Xist, Tsix (two different amplicons), and control b-actin RNAs in wild-type (WT) and Tsix-mutant TS cells. Three TS cell lines of each genotype were analyzed. M, marker; NTC, no template control; +, RT; -, no RT control lane. **(c)** RNA FISH detection of Xist (white), Tsix (green), and Atrx (red) RNAs in 6-day

(d6) differentiated TS cell lines. Three cell lines of each genotype were analyzed. Scale bar, 10 μm . **(d)** Quantification of Xist induction from the X^{Tsix} X-chromosome in females. Aberrant Xist RNA coating (defined as two Xist RNA coats in a diploid cell or Xist coating of all chromosomes in polyploid giant cells) is observed in mutant but not WT d6 differentiated TS cells. For giant cells, the number of X-chromosomes was identified based on distinct Xist and Atrx RNA FISH signals. $n=100$ nuclei counted for each cell line per day of differentiation. **(e)** Quantification of Xist induction from the X^{Tsix} X-chromosome in males. Aberrant Xist RNA coating is observed in mutant but not WT d6 differentiated TS cells. $n=100$ nuclei counted for each cell line per day of differentiation. **(f)** RT-PCR detection of Xist and control b-actin RNA in undifferentiated (d0) and d6 differentiated wild-type (WT) and Tsix-mutant TS cells. A single representative TS cell line from each genotype is shown. M, marker; NTC, no template control; +, RT; -, no RT control lane. **(g)** Sanger sequencing chromatograms of representative $X^{\text{Lab}}X^{\text{JF1}}$ and $X^{\text{Tsix}}X^{\text{JF1}}$ Xist RT-PCR products (RNA), and an Xist genomic DNA amplicon (gDNA) within exon 1. Highlights mark a single nucleotide polymorphism that differs between the maternal $X^{\text{Lab}} / X^{\text{Tsix}}$ alleles and the paternal X^{JF1} Xist allele.

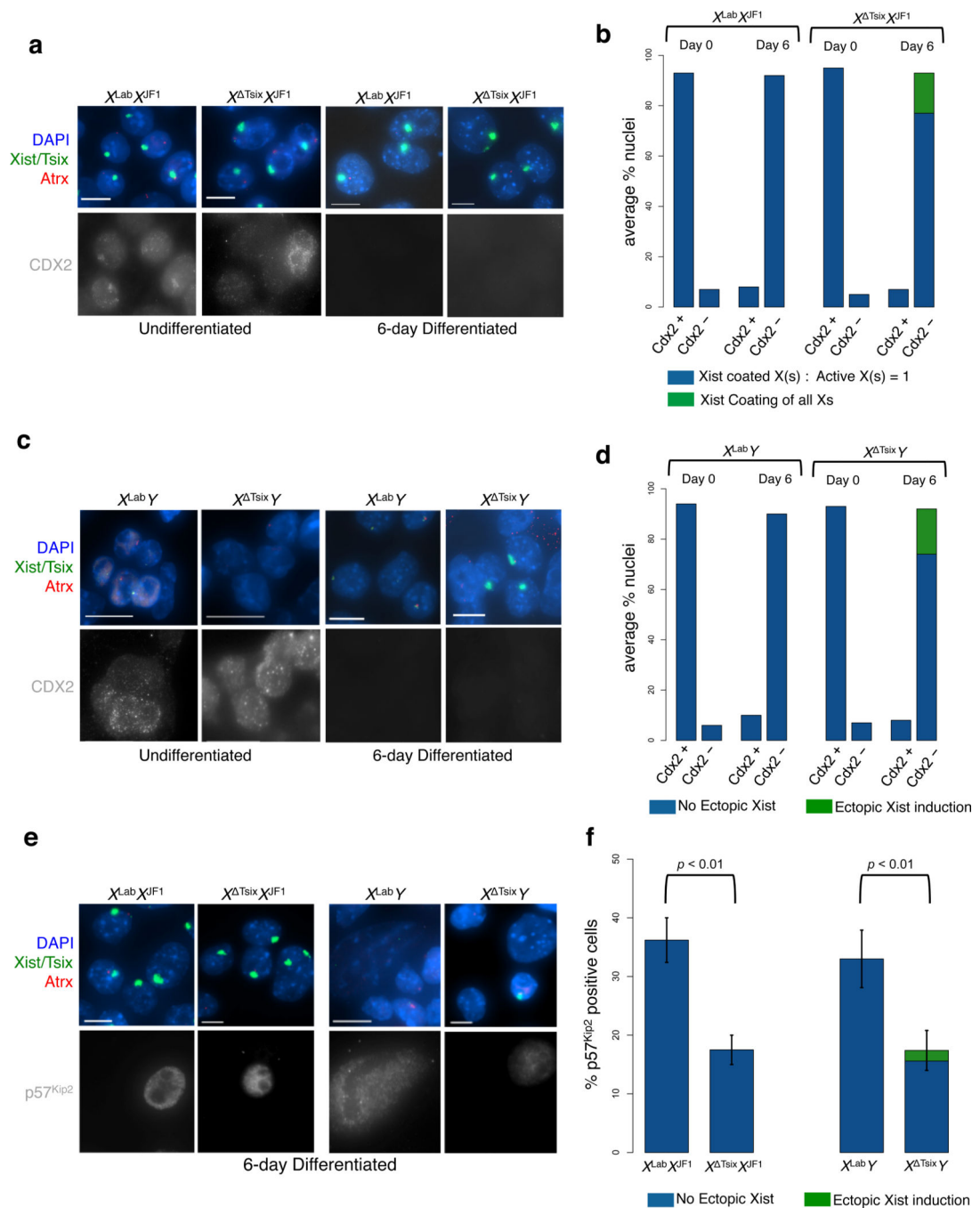


Figure 4. Characterization of differentiation-dependent Xist RNA induction from the X^{Tsix} maternal X-chromosome

(a) RNA FISH detection of Xist, Tsix, and the X-linked gene Atrx in undifferentiated and 6-day (d6) differentiated trophoblast stem (TS) cells. Immunofluorescence (IF) staining of the same cells detects CDX2, a marker of undifferentiated trophoctodermal cells. Scale bar, 10 μ m. (b) Quantification of Xist induction in CDX2 positive and negative in undifferentiated and differentiated female TS cells. 100 nuclei were counted per cell line at each time point ($n = 3$ cell lines per genotype). No aberrant Xist induction is observed from the X^{Tsix} in

undifferentiated cells. In d6 differentiated $X^{Tsix}X^{JF1}$ TS cells, ectopic Xist induction is restricted to cells that lack CDX2 staining. A subset of differentiated nuclei show both multiple Xist-coated inactive X-chromosomes and multiple active X-chromosomes, due to endoreduplication. **(c)** IF/RNA FISH analysis of male TS cells, as in **(a)**. Scale bar, 10 μm . **(d)** Quantification of Xist induction in undifferentiated and d6 differentiated male TS cells. 100 nuclei were counted per cell line at each time point ($n = 3$ cell lines per genotype). **(e)** IF/RNA FISH detection of Xist, Tsix, and the X-linked gene Atrx, in differentiated TS cells. p57^{Kip2}, a marker of trophoblast giant cells, is detected in the same cells by IF. Scale bar, 10 μm . **(f)** Quantification p57^{Kip2} positive cells and aberrant Xist induction in the TS cells. 100 nuclei were counted per cell line ($n = 3$ cell lines per genotype). $X^{Tsix}X^{JF1}$ and $X^{Tsix}Y$ TS cells show significantly reduced levels of p57^{Kip2} staining, suggesting failure of these genotypes to terminally differentiate. Error bars, S.D.

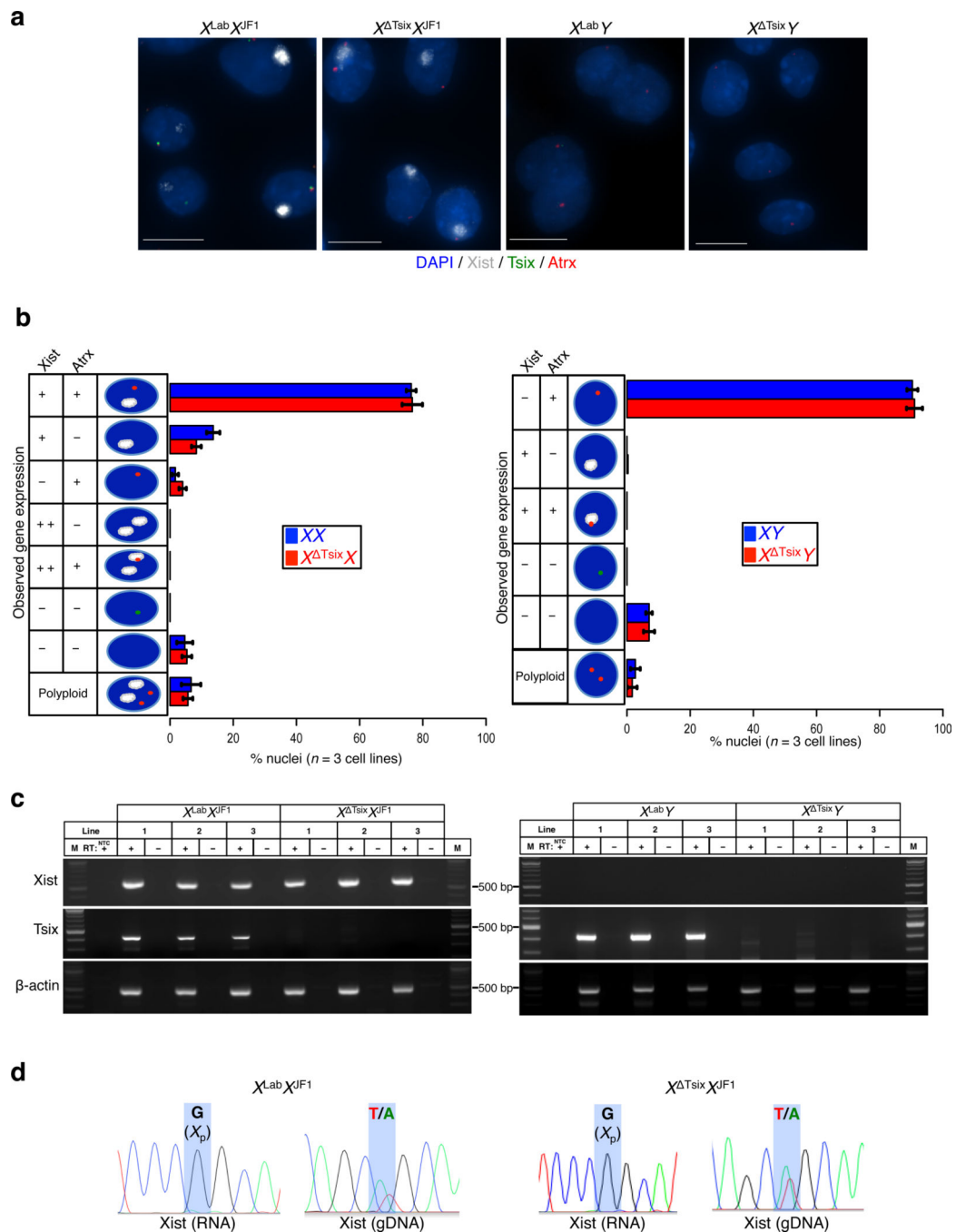


Figure 5. Lack of Xist induction from the X^{Tsix} maternal X-chromosome in cultured extra-embryonic endoderm (XEN) cells

(a) RNA FISH detection of Xist (white), Tsix (green) and Atrx (red) RNAs in representative XEN cell lines. Nuclei are stained blue with DAPI. Three cell lines of each genotype were analyzed. Scale bar, 10 μm. (b) Quantification of Xist RNA coating and X-linked gene expression in the XEN cells. The X-axis of each graph represents average % nuclei in each class from 100 cells counted per cell line (n = 3 cell lines per genotype). Diagrams along the Y-axis depict all observed expression patterns. +, RNA expression detected from a single X-

chromosome; + +, RNA expression detected from both X-chromosomes; -, absence of RNA detection. A subset of tetraploid XEN nuclei show two Xist-coated inactive X-chromosomes and two active X-chromosomes, due to endoreduplication⁴⁵. Gene expression patterns do not differ significantly between wild-type and Tsix mutant XEN cells (Fisher's exact test). Error bars, S.D. **(c)** RT-PCR detection of Xist, Tsix, and control b-actin RNAs in three individual XEN cell lines of each genotype. M, marker; NTC, no template control; +, reaction with reverse transcriptase (RT); -, no RT control lane. **(d)** Sanger sequencing chromatograms of representative $X^{\text{Lab}}X^{\text{JF1}}$ and $X^{\text{Tsix}}X^{\text{JF1}}$ RT-PCR products spanning Xist exons 1–4 (RNA), and an Xist genomic DNA amplicon (gDNA) within exon 1. Highlights mark a single nucleotide polymorphism that differs between the maternal $X^{\text{Lab}} / X^{\text{Tsix}}$ alleles and the paternal X^{JF1} allele.

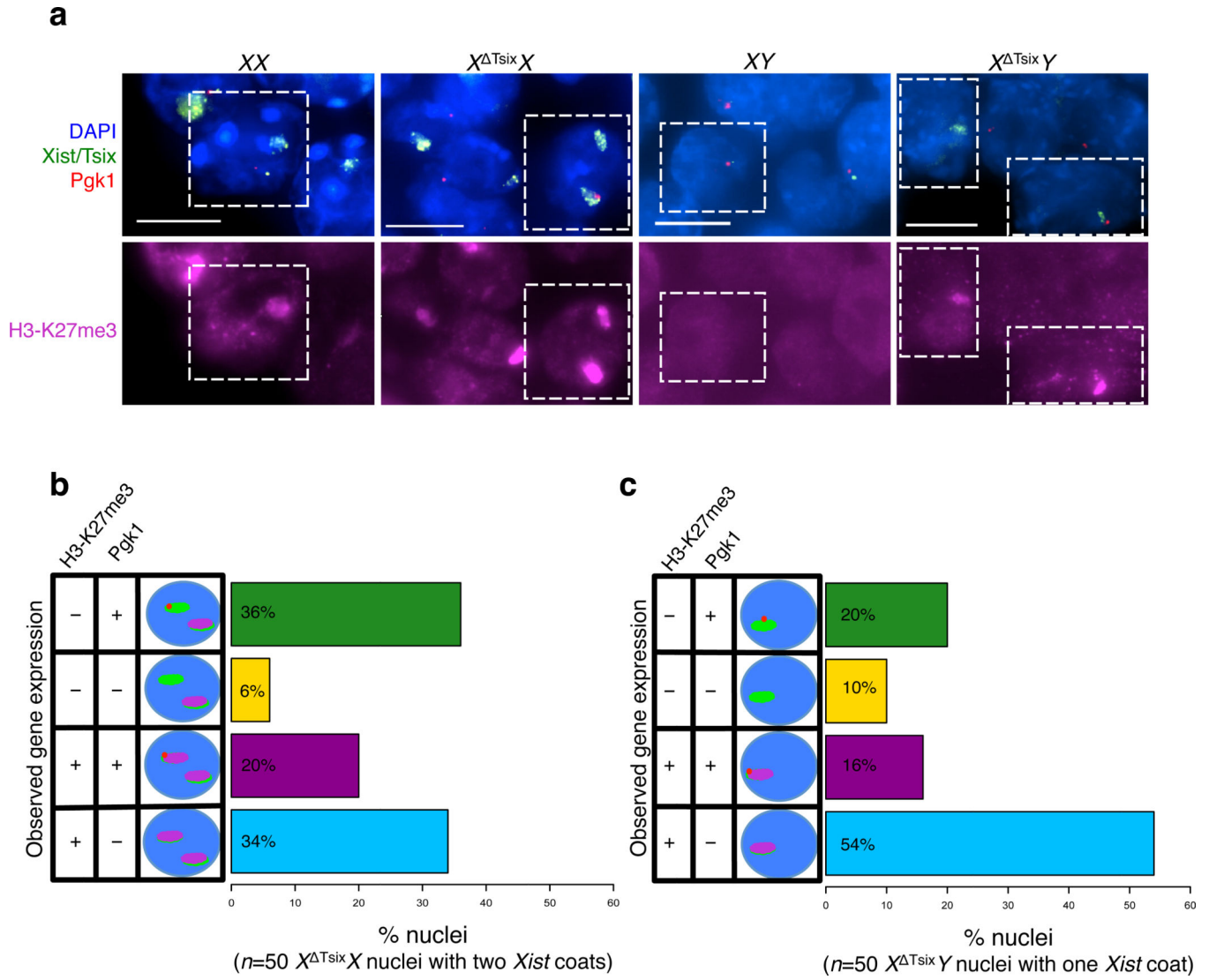


Figure 6. Disassociation of Xist induction, H3-K27me3 enrichment, and inactivation of the $X^{\Delta Tsix}$ maternal X-chromosome in E6.5 extra-embryonic cells

(a) RNA FISH detection of Xist, Tsix, and Pgf1 RNAs coupled with immunofluorescence (IF) detection of H3-K27me3 in extra-embryonic cells of E6.5 embryos. Dashed boxes mark representative nuclei. Scale bar, 10 μ m. (b) Quantification of H3-K27me3 enrichment and Pgf1 expression in nuclei displaying Xist RNA coating of both X-chromosomes in $X^{\Delta Tsix}X$ extra-embryonic cells (50 nuclei with Xist RNA coating of both X-chromosomes were analyzed [$n=5 X^{\Delta Tsix}X$ embryos]). Wild-type (WT) XX embryos show Xist RNA coating and enrichment of H3-K27me3 on a single X-chromosome ($n=5$ embryos). (c) Quantification of H3-K27me3 enrichment and Pgf1 expression in nuclei displaying Xist RNA coating of the X-chromosome in $X^{\Delta Tsix}Y$ extra-embryonic cells (50 nuclei with Xist RNA coating of the single X-chromosome [$n=4 X^{\Delta Tsix}Y$ embryos] were analyzed). WT XY cells show neither Xist RNA coating nor H3-K27me3 enrichment ($n=4$ embryos).

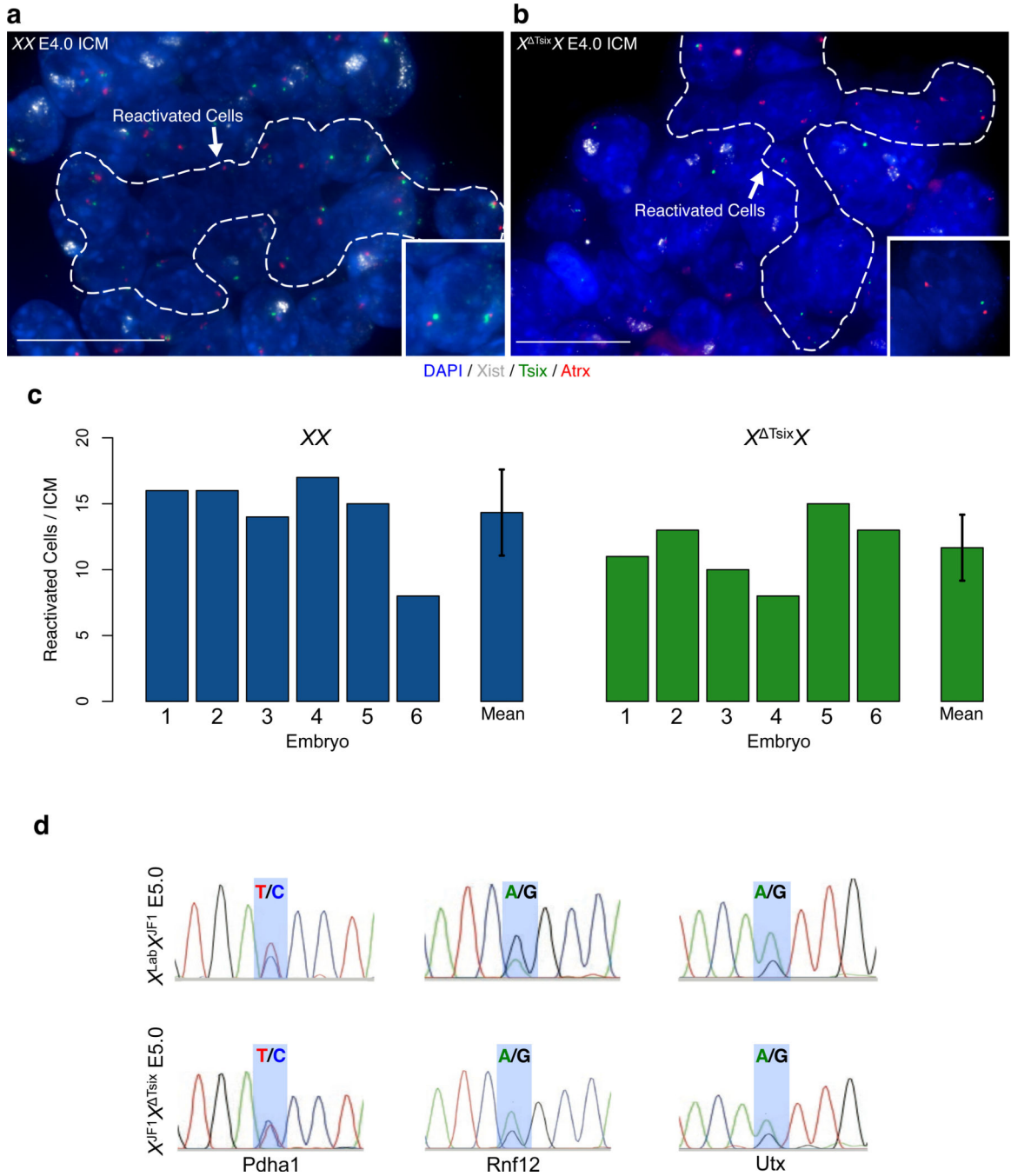


Figure 7. Reactivation of the inactive X^{Tsix} paternal X-chromosome in the inner cell mass (ICM)

RNA FISH detection of Xist (white), Tsix (green), and Atrx (red) RNAs in E4.0 ICMs. Nuclei are stained blue with DAPI. Insets show representative reactivated nuclei. Scale bar, 20 μm. (c) Quantification of the number of reactivated nuclei, as characterized by loss of Xist RNA coating and biallelic Atrx expression, in individual ICMs (n=6 ICMs per genotype). The mean number of reactivated cells per ICM does not differ significantly between XX and XX^{Tsix} ICMs (p = 0.23, two-tailed T-test). Error bars, S.D. (d) Allele-

specific X-linked gene expression analysis in E5.0 epiblast cells. Representative chromatograms of sequenced cDNAs show biallelic expression of the X-linked genes *Pdha1*, *Rnfl2*, and *Utx*, regardless of genotype.

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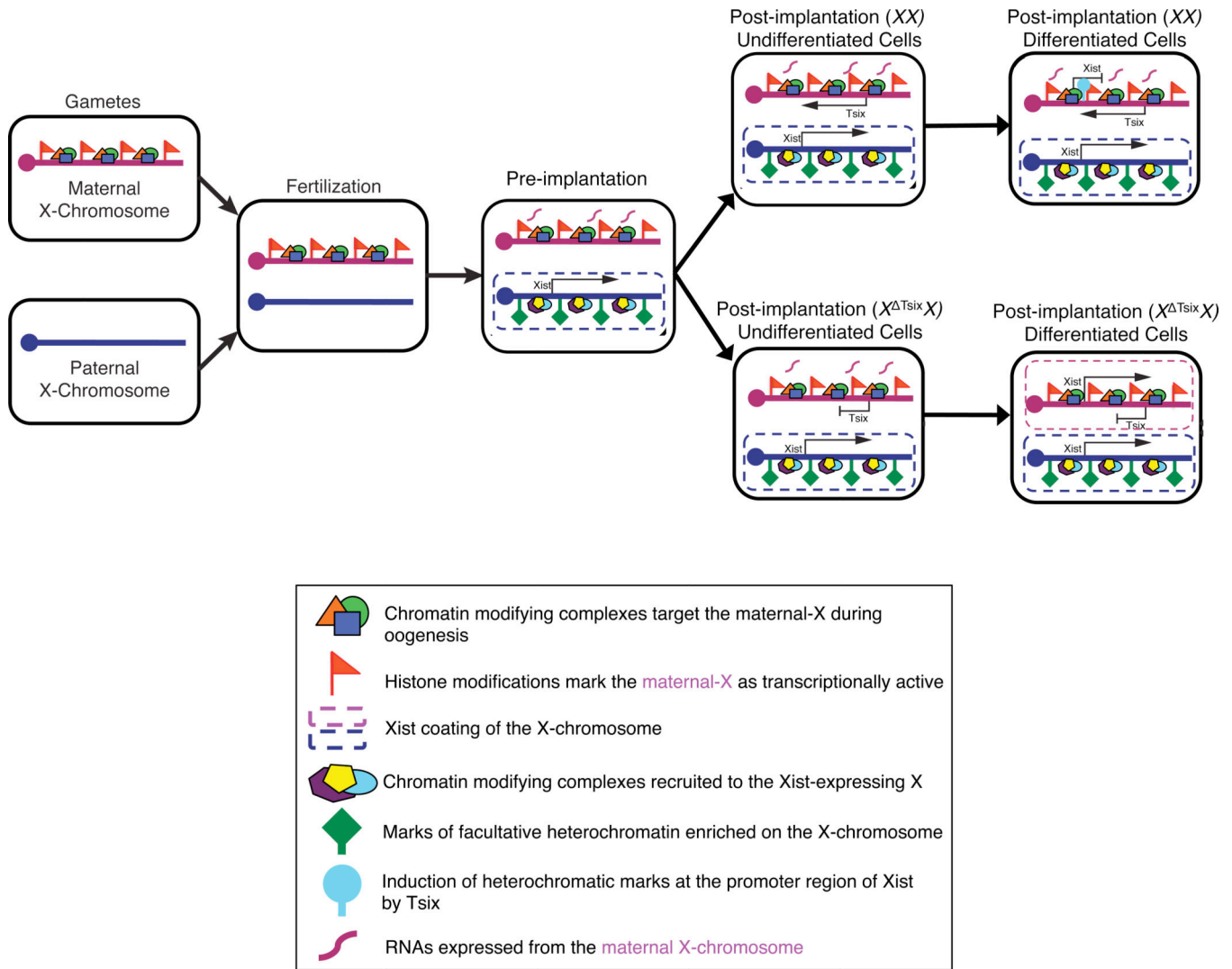


Figure 8. A model for the role of Tsix in imprinted X-Inactivation

The maternal X-chromosome, but not the paternal-X, is marked by histone modifications during gametogenesis that are transmitted to the offspring upon fertilization. In the pre-implantation embryo, these histone modifications prevent inactivation of the maternal-X, while the paternal X-chromosome is subject to inactivation. Xist is induced from the paternally-derived X-chromosome in the pre-implantation embryo, and helps recruit protein complexes that catalyze histone marks characteristic of facultative heterochromatin on the paternal-X. The oocyte-configured chromatin of the maternal-X, conversely, prevents Xist induction from the maternal X-chromosome during the initiation phase of X-inactivation (pre-implantation) and does not require Tsix. The maternal-X then remains active during the maintenance phase of imprinted X-inactivation in undifferentiated extra-embryonic nuclei (post-implantation, undifferentiated cells), independently of Tsix expression. Tsix is induced from the maternal-X due to the absence of Xist expression from this X-chromosome. Upon differentiation, Tsix transcription across the Xist promoter region is required to induce heterochromatinization of the Xist promoter to keep Xist silenced in the extra-embryonic trophoblast lineage. In XX differentiated extra-embryonic cells, the wild-type maternal-

X remains transcriptionally competent while the paternal-X is maintained as transcriptionally inactive in imprinted X-inactivated cells. Upon differentiation of $X^{Tsix}X$ trophoctoderm cells, the Tsix-mutant maternal X-chromosome induces Xist.

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