

Liver cell adenoma: A case report with clonal analysis and literature review

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Abstract

We report a case of liver cell adenoma (LCA) in a 33-year-old female patient with special respect to its clonality status, pathogenic factors and differential diagnosis. The case was examined by histopathology, immunohistochemistry and a clonality assay based on X-chromosomal inactivation mosaicism in female somatic tissues and polymorphism at androgen receptor focus. The clinicopathological features of the reported cases from China and other countries were compared. The lesion was spherical, sizing 2 cm in its maximal dimension. Histologically, it was composed of cells arranged in cords, most of which were two-cell-thick and separated by sinusoids. Focal fatty change and excessive glycogen storage were observed. The tumor cells were round or polygonal in shape, resembling the surrounding parenchymal cells. Mitosis was not found. No portal tract, central vein or ductule was found within the lesion. The tumor tissue showed a positive reaction for cytokeratin (CK) 18, but not for CK19, vimentin, estrogen and progesterone receptors. Monoclonality was demonstrated for the lesion, confirming the diagnosis of an LCA. Clonality analysis is helpful for its distinction from focal nodular hyperplasia.

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Key words: Liver Cell adenoma; Clonality; Literature review

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INTRODUCTION

Liver cell adenoma (LCA) is a rare benign liver tumor. It is considered a hepatocellular neoplasm, while some authors hold that LCA originated from liver progenitor cell^[1]. A female predominance was noticed for the development of LCA based on the reports from Western countries, with a majority of cases associated with a long-term use of oral contraceptives or other steroids. Other diseases should be excluded before establishing the diagnosis of an LCA. It is indeed a difficult task to distinguish LCA from focal nodular hyperplasia (FNH) when the morphologic features are not prominent and a central scar is absent. Then some molecular approaches, including clonality analysis, may be helpful. In this article, a case of LCA in a female Chinese patient is presented, with its clonality status demonstrated.

CASE REPORT

A hepatic mass was found in the right lobe of a 33-year-old woman during her routine medical check-up. She was then admitted to Tangdu Hospital in Xi'an in January 28, 2003. She had never used oral contraceptives, and she had no history of alcohol abuse or hepatitis. No record of HCC or any hereditary disease was found among her family members. The parameters of routine clinical biochemistry, including values of aspartate aminotransferase (AST), alanine aminotransferase (ALT), lactate dehydrogenase (LDH), γ -glutamyltransferase (γ -GTP) and concentrations of α -fetoprotein (AFP) and plasma proteins, were all within normal ranges. The laboratory tests failed to show any positive signal in her serum for hepatitis B surface antigen (HBsAg) or anti-hepatitis C virus (HCV) antibody.

Ultrasonography revealed a solid mass in the posterior part of the right lobe of liver. Computed tomography (CT) scanning showed reduction of density for the lesion sizing 2.0 cm in diameter (Figure 1), indicating malignant potential. Laparotomy was then performed on February 1, 2003. Size of the liver appeared normal, with a mass found in the right lobe. Partial hepatectomy was then performed. Appearance, color and texture of the surrounding liver

were normal, without any indication of cirrhosis, pronounced fibrosis or cholestasis.

Histological and immunohistochemical procedures

The sample was fixed in 40 g/L formaldehyde solution, embedded in paraffin. Sections of 4 μm in thickness were prepared and stained by hematoxylin and eosin (HE), Masson trichrome methods and periodic acid-Schiff (PAS) reaction. Immunostaining was carried out using a streptavidin-labeled peroxidase (S-P) kit (KIT9730) as described previously^[2]. The primary antibodies used in this study included those against cytokeratin (CK) 18, CK19, vimentin, CD34, estrogen receptor (ER), progesterone receptor (PR), AFP, S-100 protein, HBsAg, hepatitis B core antigen (HBcAg), as well as an anti-HCV antibody. All of the reagents for immunostaining were supplied by Maxim Biotechnology Corporation Limited, Fuzhou, China.

Clonal analysis

Sections of 10 μm in thickness were prepared, deparaffinized, rehydrated and HE stained. Neoplastic tissues were dissected using a syringe needle from 4 different tumor areas, sizing 0.5 cm \times 0.5 cm for each. Normal liver tissue was also isolated from the surrounding parenchyma at 3 different sites of the same size and analyzed in a parallel way as reference samples. Genomic DNA was extracted using a QIAamp kit (Qiagen, Mannheim, Germany). Polymorphism was examined at the androgen receptor (AR) and phosphoglycerokinase (PGK) loci as described previously^[3,4], with the former gene proved polymorphic at the CAG short-tandem repeat (STR) located in exon 1. Loss of X-chromosomal inactivation mosaicism was demonstrated by pretreatment of DNA with methylation-sensitive restriction enzyme *Hha* I and amplification via nested PCR. The CAG STR-polymorphic alleles were resolved on a 100 g/L denaturing polyacrylamide gel at 120 V for 4 h. A reduction of at least 50% in density of either band, as compared to that obtained using the sample not treated with *Hha* I, is regarded as loss of X-chromosomal inactivation mosaicism^[5].

RESULTS

A partial resection liver specimen, sizing 4.0 cm \times 3.0 cm \times 2.0 cm, presented with a spherical mass of 2.0 cm in its maximal dimension. The mass was beneath the hepatic capsule, yellow brown in color and soft in its texture, without any necrotic focus or fibrotic scar in its cut surface. There was a clear border, but not a fibrotic septum between the lesion and surrounding liver tissue that appeared red brown and apparently normal. Microscopically, the lesion was composed of cells arranged in two-cell-thick cords, with the cell cords separated by sinusoids (Figure 2A). Focal fatty change and excessive glycogen storage were present (Figure 2A and 2B). The tumor cells were round or polygonal, apparently resembling the surrounding liver parenchyma cells in size and shape. Mitosis was not found. There was no portal tract or hepatic venule in the tumor. Ductule or scattered ductular cells (the so-called "oval cells"^[6,7] or "liver progenitor cells"^[1]) were absent, and immunostaining for the ductular cell markers, includ-

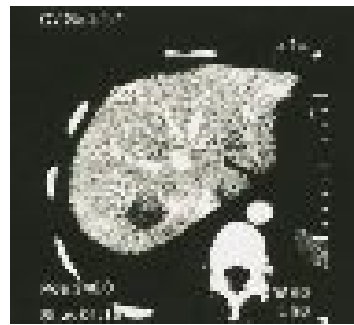


Figure 1 CT scanning showing a lesion with reduction of density in the posterior part of the right lobe of liver.

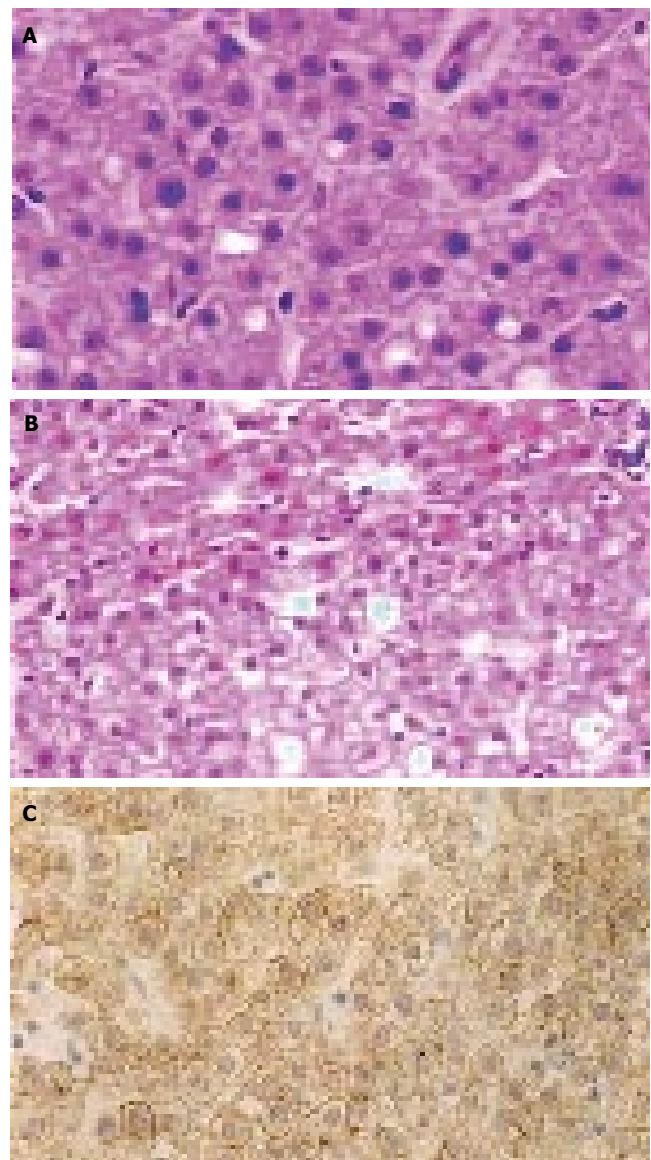


Figure 2 Liver cell adenoma (\times 400). **A:** Similar to normal hepatocytes, arranged in two-cell-thick cords separated by hepatic sinusoids (HE); **B:** Showing the border between LCA (lower) and surrounding liver parenchyma (upper) (HE); **C:** CK18 immunoreactivity (S-P).

ing CK19 and S-100 protein^[7,8], failed to show any positive cell within the lesion. The tumor cells were positive for CK18 (Figure 2C), but negative for AFP, vimentin and p53 protein. They did not show any positive signal for ER or PR. Both neoplastic and the adjacent parenchymal tissues were negative for HBsAg, HBcAg and HCV antigen.

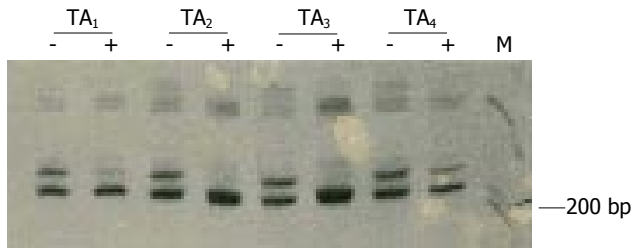


Figure 3 Clonal analysis on tissues from 4 tumor areas (TAs). The pretreatment with *Hha* I results in loss of the upper band for TA₂ and TA₃, and intensity reduction by factors of 71.8% and 57.1% for TA₁ and TA₄, respectively. +, with *Hha* I pretreatment; -, without *Hha* I treatment.

Clonality status of the lesion was determined by an assay based on the CAG-STR polymorphism at exon 1 of AR gene. Pretreatment with *Hha* I resulted in pronounced reduction or loss of the upper band for all of the tissue samples from 4 different areas of the lesion (Figure 3), demonstrating loss of the X-chromosomal inactivation mosaicism. The adjacent liver parenchyma, however, did not show the change (Figure 4). The data proved the monoclonal, neoplastic nature of the lesion, confirming the diagnosis of LCA. The patient has survived for 30 mo after the operation without any indication of recurrence.

DISCUSSION

LCA is a rare benign hepatic neoplasm, accounting for less than 2% of all hepatic tumors^[9]. A pronounced female predominance was noted mainly by authors from Western countries. It often occurs in women of 20 to 40 years during their child-bearing period^[10,11], being closely associated with long-term use of steroids, mainly oral contraceptives^[12-14]. In fact, it was rarely reported before the introduction of oral contraceptives in 1960s. Edmondson *et al.*^[15] found only two cases in 48 900 necropsies performed in Los Angeles General Hospital during the period from 1918 to 1954. In 1973, Baum *et al.*^[16] pointed out the possible link between the use of oral contraceptives and LCA development. Data from several reports has confirmed the etiologic association. Leese *et al.*^[17] reported 24 cases of HA, 16 (66.7%) of them with a history of using oral contraceptives. Tumor regression was observed in some cases after withdrawal of the hormones^[18,19], and then the tumor remained silent or grew slowly for many years, or even progressed to HCC^[20], albeit infrequently. Complete remission was observed in an LCA patient, who had used oral contraceptives for 8 years, after the hormone withdrawal for 9 months^[21].

Through literature review, the clinicopathological data of 127 cases of LCA reported from Chinese patients were collected and compared to those of 130 patients from Western countries, with the male/female ratios being 1.8/1 and 1/2.9, respectively. The ages of the Chinese patients ranged from 2 to 73 years, with their mean and average values estimated to be 31.0 and 35.8 years, respectively. For patients from Western countries, the ages ranged from 11 months to 82 years, with their mean and average values 30.0 and 31.6 years, respectively. Only 3.9% (5/127) of Chinese patients had a history of using oral contraceptives,

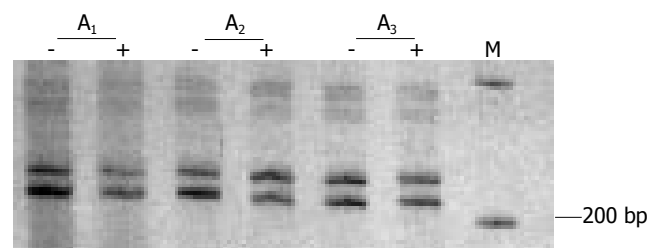


Figure 4 Data of clonality test of three areas of liver parenchymal tissues (A₁-A₃). Neither of the bands on gel shows marked intensity change for the samples pretreated with *Hha* I (+) compared to those not treated (-).

the percentage being much lower than that for the patients from Western countries (54/130, 41.5%). Difference is evident, therefore, between the LCAs occurring in China and in Western countries in their etiologic associations, indicating possibly different pathogenic pathways. This, at least partly, provides an explanation for the distinct gender distribution pattern in Chinese patients.

Other factors are also linked to LCA development, including glycogen storage diseases^[22] and administration of danazol^[23], phenobarbital^[24] and androgenic/anabolic steroids^[9,25-33]. Among the 130 cases from Western countries, 4 (3.1%) were found to have glycogen-storage diseases, but only one (0.8%), in the Chinese group, was with the association. For some LCA cases, there seemed no identifiable pathogenic factor. The majority of the cases from the Chinese group fell into this category. Among the 127 Chinese patients, 13 (10.2%) were seropositive for HBsAg, and 8 of them were with chronic hepatitis^[34-37]. Among the patients from Western countries, however, only one (0.8%) was shown to be an HBsAg-carrier. In consideration of the high prevalence (up to 10%^[38]) of HBsAg-carrier state in China, the data does not provide support for the role of persistent HBV infection in development of the solitary LCA.

Most of LCAs grow slowly and are asymptomatic or cause only mild symptoms, while rupture and hemorrhage may occur in some tumors (15%-33%^[39]), and malignant transformation was also observed in a minority of the cases (5/39, 12.8%^[20]). Ultrasonography, CT scanning and magnetic resonance imaging (MRI) are useful for detecting hepatic occupation and determining its location and size. Imaging approaches may be helpful for identifying LCA from other hepatic lesions, and a peritumorous halo, demonstrated by CT scanning, was considered indicative of LCA^[40,41]. These features, however, are not specific^[40], and the accurate preoperative diagnosis of LCA, particularly the distinction from HCC, is frequently problematic. This is even more serious as a surgical consideration in China where incidence of HCC is overwhelmingly high compared to that of LCA. For the pathologists, well differentiated HCC and FNH are among the hepatic lesions that should be excluded before making a diagnosis of LCA.

HCC is diagnosed usually at ages between 40 and 60 years, with a pronounced male predominance. In China, including Hong Kong and Taiwan, about 80% of HCCs were found in patients with chronic hepatitis B and cirrhosis or advanced liver fibrosis. An elevated level of circulating AFP is indicative of HCC, but this change may

not be evident in patients with an early-stage, often well differentiated, HCC. Liver-cell plates more than three cells thick, acinar structures, increased nuclear/cytoplasmic ratio, prominent nucleoli, mitoses, increased cytoplasmic basophilia, loss of the reticulin fibers, absence of Kupffer cells, presence of vascular invasion or immunoreactivity for AFP, all indicate HCC. However, none of these features can be relied upon with certainty, as some are also seen in the hepatic lesions with high-grade SCC, including the premalignant nodules of altered hepatocytes^[2], adenomatous hyperplasia^[42-46] and LCA. The most helpful parameters, in our consideration, are thickness of the liver-cell plates (more than three cells), cell density (an increase of two folds compared to the surrounding liver parenchyma) and vascular invasion^[38]. It should be noted that male, cirrhosis, and chronic HBV infection strongly indicate that a hepatic neoplasm is malignant. Conversely, female and a history of oral contraceptive use support the diagnosis of LCA. Exceptional difficulties may be encountered in some of the LCAs associated with long-term use of anabolic steroids or metabolic disorders. Some of them may show pronounced architectural disturbance and cellular atypia, which make their distinction impossible from well-differentiated HCCs based on histological grounds alone. Malignant transformation is proposed for such cases^[13,20,47,48], but they often show more favorable clinical courses, or even regression after withdrawal of the steroid^[21]. Such lesions may represent the borderline hepatocellular neoplasm, and their behaviors should be determined by careful postoperative observations.

FNH occurs most commonly in young women, having similar etiological associations and clinical manifestations to LCA^[49-51]. It is a localized lesion, frequently solitary, within an otherwise normal or nearly normal liver. The lesion is similar to cirrhosis by its histology, and a central stellate fibrous region containing large vessels can often be found. Its development has been attributed to the vascular malformation^[52,53]. Usually, FNH is readily distinguished from LCA by its central scar, multinodularity and presence of proliferating bile ductules in the fibrous septa. It may become a diagnostic problem, however, when the central fibrous region is not evident. Data from different laboratories have demonstrated the polyclonal cell composition and indicated non-neoplastic nature for the lesion^[54,55]. In contrast, an LCA was shown to be monoclonal^[56], and our data confirm the conclusion that LCA is a neoplastic lesion. The clonality assays, therefore, are helpful for the differential diagnosis between LCA and some FNH lesions without an identifiable central scar.

REFERENCES

- 1 Libbrecht L, De Vos R, Cassiman D, Desmet V, Aerts R, Roskams T. Hepatic progenitor cells in hepatocellular adenomas. *Am J Surg Pathol* 2001; **25**: 1388-1396
- 2 Su Q, Benner A, Hofmann WJ, Otto G, Pichlmayr R, Bannasch P. Human hepatic preneoplasia: phenotypes and proliferation kinetics of foci and nodules of altered hepatocytes and their relationship to liver cell dysplasia. *Virchows Arch* 1997; **431**: 391-406
- 3 Wang S, Su Q, Zhu S, Liu J, Hu L, Li D. Clonality of multiple uterine leiomyomas. *Zhonghua Binglixue Zazhi* 2002; **31**: 107-111
- 4 Diao XL, Su Q, Wang SF, Feng YM, Liu J. Non-isotopic clonality analysis on uterine leiomyomas based on AR gene polymorphism. *Disi Junyi Daxue Xuebao* 2002; **23**: 1969-1973
- 5 Wang SF, Liu Q, Zhang W, Liu J, Su Q. Clonality of uterine leiomyomas, an assay using X chromosome polymorphism at the phosphoglycerate kinase locus. *Disi Junyi Daxue Xuebao* 2001; **22**: 1576-1582
- 6 Hsia CC, Evarts RP, Nakatsukasa H, Marsden ER, Thorgeirsson SS. Occurrence of oval-type cells in hepatitis B virus-associated human hepatocarcinogenesis. *Hepatology* 1992; **16**: 1327-1333
- 7 Su Q, Liu YF, Zhang JF, Zhang SX, Li DF, Yang JJ. Expression of insulin-like growth factor II in hepatitis B, cirrhosis and hepatocellular carcinoma: its relationship with hepatitis B virus antigen expression. *Hepatology* 1994; **20**: 788-799
- 8 Su Q, Zerban H, Otto G, Bannasch P. Cytokeratin expression is reduced in glycogenotic clear hepatocytes but increased in ground-glass cells in chronic human and woodchuck hepatocellular infection. *Hepatology* 1998; **28**: 347-359
- 9 Ishak KG. Hepatic lesions caused by anabolic and contraceptive steroids. *Semin Liver Dis* 1981; **1**: 116-128
- 10 Hytioglou P, Theise ND. Differential diagnosis of hepatocellular nodular lesions. *Semin Diagn Pathol* 1998; **15**: 285-299
- 11 Li LG, Cui XJ, Li HN. Hepatocellular Adenoma. *Zhonghua Waikexue Zazhi* 1995; **10**: 613
- 12 Edmondson HA, Henderson B, Benton B. Liver-cell adenomas associated with use of oral contraceptives. *N Engl J Med* 1976; **294**: 470-472
- 13 Kerlin P, Davis GL, McGill DB, Weiland LH, Adson MA, Sheedy PF 2nd. Hepatic adenoma and focal nodular hyperplasia: clinical, pathologic, and radiologic features. *Gastroenterology* 1983; **84**: 994-1002
- 14 Shortell CK, Schwartz SI. Hepatic adenoma and focal nodular hyperplasia. *Surg Gynecol Obstet* 1991; **173**: 426-431
- 15 Edmondson HA, Steiner PE. Primary carcinoma of the liver a study of 100 cases among 48,900 necropsies. *Cancer* 1954; **7**: 462-503
- 16 Baum JK, Bookstein JJ, Holtz F, Klein EW. Possible association between benign hepatomas and oral contraceptives. *Lancet* 1973; **2**: 926-929
- 17 Leese T, Farges O, Bismuth H. Liver cell adenomas. A 12-year surgical experience from a specialist hepato-biliary unit. *Ann Surg* 1988; **208**: 558-564
- 18 Edmondson HA, Reynolds TB, Henderson B, Benton B. Regression of liver cell adenomas associated with oral contraceptives. *Ann Intern Med* 1977; **86**: 180-182
- 19 Tao LC. Oral contraceptive-associated liver cell adenoma and hepatocellular carcinoma: Cytomorphology and mechanism of malignant transformation. *Cancer* 1991; **68**: 341-347
- 20 Foster JH, Berman MM. The malignant transformation of liver cell adenomas. *Arch Surg* 1994; **129**: 712-717
- 21 Aseni P, Sansalone CV, Sammartino C, Benedetto FD, Carrafiello G, Giacomoni A, Osio C, Vertemati M, Forti D. Rapid disappearance of hepatic adenoma after contraceptive withdrawal. *J Clin Gastroenterol* 2001; **33**: 234-236
- 22 Labruno P, Trioche P, Duvaltier I, Chevalier P, Odièvre M. Hepatocellular adenomas in glycogen storage disease type I and III: a series of 43 patients and review of the literature. *J Pediatr Gastroenterol Nutr* 1997; **24**: 276-279
- 23 Bartley J, Lodenkemper C, Lange J, Mechsner S, Radke C, Neuhaus P, Ebert AD. Hepatocellular adenoma and focal nodular hyperplasia after long-term use of danazol for endometriosis: a case report. *Arch Gynecol Obstet* 2004; **269**: 290-293
- 24 Ferko A, Bedrna J, Nozicka J. [Pigmented hepatocellular adenoma of the liver caused by long-term use of phenobarbital]. *Rozhl Chir* 2003; **82**: 192-195
- 25 Paradinas FJ, Bull TB, Westaby D, Murray-Lyon IM. Hyperplasia and prolapse of hepatocytes into hepatic veins during longterm methyltestosterone therapy: possible relationships of these changes to the development of peliosis hepatis and liver tumours. *Histopathology* 1977; **1**: 225-246
- 26 Westaby D, Portmann B, Williams R. Androgen related primary hepatic tumors in non-Fanconi patients. *Cancer* 1983; **51**: 1947-1952

- 27 **Chandra RS**, Kapur SP, Kelleher J Jr, Luban N, Patterson K. Benign hepatocellular tumors in the young: A clinicopathologic spectrum. *Arch Pathol Lab Med* 1984; **108**: 168-171
- 28 **Carrasco D**, Prieto M, Pallardó L, Moll JL, Cruz JM, Muñoz C, Berenguer J. Multiple hepatic adenomas after long-term therapy with testosterone enanthate, Review of the literature. *J Hepatol* 1985; **1**: 573-578
- 29 **Grangé JD**, Guéchet J, Legendre C, Giboudeau J, Darnis F, Poupon R. Liver adenoma and focal nodular hyperplasia in a man with high endogenous sex steroids. *Gastroenterology* 1987; **93**: 1409-1413
- 30 **Ishak KG**, Zimmerman HJ. Hepatotoxic effects of the anabolic/androgenic steroids. *Semin Liver Dis* 1987; **7**: 230-236
- 31 **Søe KL**, Søe M, Gluud C. Liver pathology associated with the use of anabolic-androgenic steroids. *Liver* 1992; **12**: 73-79
- 32 **Creagh TM**, Rubin A, Evans DJ. Hepatic tumours induced by anabolic steroids in an athlete. *J Clin Pathol* 1988; **41**: 441-443
- 33 **Klava A**, Super P, Aldridge M, Horner J, Guillou P. Body builder's liver. *J R Soc Med* 1994; **87**: 43-44
- 34 **Liu M**, Chen LZ, Li XH. Six cases of hepatocellular adenoma. *Zhonghua Gandan Waike Zazhi* 2003; **9**: 142-147
- 35 **Guan CN**. Liver cell adenoma. *Zhongliu Fangzhi Yanjiu* 2001; **28**: 156
- 36 **Xiao KY**, Li LQ, Peng MH, Chen B, Shang LM. Misdiagnosis analysis of hepatocellular adenoma in older man. *Linchuang Wuzhen Wuzhi* 2004; **17**: 191
- 37 **Pan SB**, Ye GR, Che SY. Analysis of nine cases of hepatocellular adenoma misdiagnosed as HCC. *Ling Nan Xiandai Linchuang Waike* 2004; **4**: 96
- 38 **Su Q**. Preneoplastic lesions in human liver. *Zhenduan Binglixue Zazhi* 2003; **10**: 112-115
- 39 **Tan M**, Di Carlo A, Robinson P, Tchervenkov JL, Barkun JS, Metrakos P. Successful outcome after transplantation of a donor liver with focal nodular hyperplasia. *Liver Transpl* 2001; **7**: 652-655
- 40 **Mathieu D**, Bruneton JN, Drouillard J, Pointreau CC, Vasile N. Hepatic adenomas and focal nodular hyperplasia: dynamic CT study. *Radiology* 1986; **160**: 53-58
- 41 **Welch TJ**, Sheedy PF 2nd, Johnson CM, Stephens DH, Charboneau JW, Brown ML, May GR, Adson MA, McGill DB. Focal nodular hyperplasia and hepatic adenoma: comparison of angiography, CT, US, and scintigraphy. *Radiology* 1985; **156**: 593-595
- 42 **Nakashima T**, Okuda K, Kojiro M, Jimi A, Yamaguchi R, Sakamoto K, Ikari T. Pathology of hepatocellular carcinoma in Japan. 232 Consecutive cases autopsied in ten years. *Cancer* 1983; **51**: 863-877
- 43 **Ferrell LD**, Crawford JM, Dhillon AP, Scheuer PJ, Nakanuma Y. Proposal for standardized criteria for the diagnosis of benign, borderline, and malignant hepatocellular lesions arising in chronic advanced liver disease. *Am J Surg Pathol* 1993; **17**: 1113-1123
- 44 **Le Bail B**, Belleannée G, Bernard PH, Saric J, Balabaud C, Bioulac-Sage P. Adenomatous hyperplasia in cirrhotic livers: histological evaluation, cellular density, and proliferative activity of 35 macronodular lesions in the cirrhotic explants of 10 adult French patients. *Hum Pathol* 1995; **26**: 897-906
- 45 **Hytioglou P**, Theise ND, Schwartz M, Mor E, Miller C, Thung SN. Macroregenerative nodules in a series of adult cirrhotic liver explants: issues of classification and nomenclature. *Hepatology* 1995; **21**: 703-708
- 46 Terminology of nodular hepatocellular lesions. International Working Party. *Hepatology* 1995; **22**: 983-993
- 47 **Ferrell LD**. Hepatocellular carcinoma arising in a focus of multilobular adenoma. A case report. *Am J Surg Pathol* 1993; **17**: 525-529
- 48 **Scott FR**, el-Refaie A, More L, Scheuer PJ, Dhillon AP. Hepatocellular carcinoma arising in an adenoma: value of QBend 10 immunostaining in diagnosis of liver cell carcinoma. *Histopathology* 1996; **28**: 472-474
- 49 **Stocker JT**, Ishak KG. Focal nodular hyperplasia of the liver: a study of 21 pediatric cases. *Cancer* 1981; **48**: 336-345
- 50 **Brady MS**, Coit DG. Focal nodular hyperplasia of the liver. *Surg Gynecol Obstet* 1990; **171**: 377-381
- 51 **Pain JA**, Gimson AE, Williams R, Howard ER. Focal nodular hyperplasia of the liver: results of treatment and options in management. *Gut* 1991; **32**: 524-527
- 52 **Wanless IR**, Mawdsley C, Adams R. On the pathogenesis of focal nodular hyperplasia of the liver. *Hepatology* 1985; **5**: 1194-1200
- 53 **Kondo F**, Nagao T, Sato T, Tomizawa M, Kondo Y, Matsuzaki O, Wada K, Wakatsuki S, Nagao K, Tsubouchi H, Kobayashi H, Yasumi K, Tsukayama C, Suzuki M. Etiological analysis of focal nodular hyperplasia of the liver, with emphasis on similar abnormal vasculatures to nodular regenerative hyperplasia and idiopathic portal hypertension. *Pathol Res Pract* 1998; **194**: 487-495
- 54 **Paradis V**, Laurent A, Flejou JF, Vidaud M, Bedossa P. Evidence for the polyclonal nature of focal nodular hyperplasia of the liver by the study of X-chromosome inactivation. *Hepatology* 1997; **26**: 891-895
- 55 **Zhang SH**, Cong WM, Wu MC. Focal nodular hyperplasia with concomitant hepatocellular carcinoma: a case report and clonal analysis. *J Clin Pathol* 2004; **57**: 556-559
- 56 **Paradis V**, Benzekri A, Dargère D, Bièche I, Laurendeau I, Vilgrain V, Belghiti J, Vidaud M, Degott C, Bedossa P. Telangiectatic focal nodular hyperplasia: a variant of hepatocellular adenoma. *Gastroenterology* 2004; **126**: 1323-1329

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