

RAPID COMMUNICATION

Blockade of high mobility group box-1 protein attenuates experimental severe acute pancreatitis

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Supported by Grants-in-Aid for Scientific Research from the Ministry of Education, Science, Sports and Culture of Japan and from the Ministry of Health, Labor and Welfare of Japan

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Received: 2006-10-17 Accepted: 2006-11-20

Nakajima T, Kuroda Y. Blockade of high mobility group box-1 protein attenuates experimental severe acute pancreatitis. *World J Gastroenterol* 2006; 12(47): 7666-7670

<http://www.wjgnet.com/1007-9327/12/7666.asp>

Abstract

AIM: To examine the effects of anti-high mobility group box 1 (HMGB1) neutralizing antibody in experimental severe acute pancreatitis (SAP).

METHODS: SAP was induced by creating closed duodenal loop in C3H/HeN mice. SAP was induced immediately after intraperitoneal injection of anti-HMGB1 neutralizing antibody (200 µg). Severity of pancreatitis, organ injury (liver, kidney and lung), and bacterial translocation to pancreas was examined 12 h after induction of SAP.

RESULTS: Anti-HMGB1 neutralizing antibody significantly improved the elevation of the serum amylase level and the histological alterations of pancreas and lung in SAP. Anti-HMGB1 antibody also significantly ameliorated the elevations of serum alanine aminotransferase and creatinine in SAP. However, anti-HMGB1 antibody worsened the bacterial translocation to pancreas.

CONCLUSION: Blockade of HMGB1 attenuated the development of SAP and associated organ dysfunction, suggesting that HMGB1 may act as a key mediator for inflammatory response and organ injury in SAP.

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Key words: Severe acute pancreatitis; High mobility group box-1; Neutralizing antibody; Inflammatory response; Organ dysfunction; Bacterial translocation

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INTRODUCTION

In severe acute pancreatitis (SAP), multiple organ dysfunction syndrome (MODS) in the early phase^[1,2] and complications of infection (infected pancreatic necrosis and sepsis) in the late phase are contributors to high mortality in SAP^[3,4]. MODS is a consequence of the systemic inflammatory response syndrome, and it is conceivable that release of humoral mediators from the excessive activated macrophages/monocytes and neutrophils may lead to the remote organ injury. Complications of infection are thought to be a result of bacterial translocation from the gastrointestinal tract, and breakdown of intestinal integrity is considered to be implicated in the mechanism^[5-7].

High mobility group box 1 (HMGB1) protein, originally discovered 30 years ago as a nuclear DNA binding protein^[8-10], was recently identified as a late-acting mediator of endotoxin lethality^[11]. Injection of HMGB1 itself was lethal, and serum levels of HMGB1 increased from 8 to 32 h after the administration of endotoxin, when the tumor necrosis factor (TNF) peak had already occurred^[11]. Antibodies to HMGB1 attenuated the mortality associated with endotoxemia, even when the antibodies were administered 2 h after the onset of endotoxemia^[11]. HMGB1 was also found to have the capacity to induce cytokines and activate inflammatory cells when it was applied extracellularly^[11-13]. This implicates HMGB1 as a proinflammatory mediator. Recent investigations reported that serum HMGB1 levels increased in patients with sepsis/endotoxemia^[11,14,15], hemorrhagic shock^[16], acute lung injury^[17,18], rheumatoid arthritis^[19] and disseminated intravascular coagulation^[20]. It has been demonstrated that HMGB1 is secreted actively by living inflammatory cells such as stimulated macrophages/monocytes, and is released passively from necrotic or damaged cells^[21-23]. Therefore, HMGB1 may be related to inflammation and necrosis in SAP, and may be an important mediator for multiple organ failure.

In a recent study, we have first demonstrated that

serum HMGB1 levels were significantly elevated in patients with SAP on admission, and were correlated with the severity^[24,25] of the disease. The HMGB1 levels were higher in patients with organ dysfunction and infection during the clinical course. The HMGB1 levels in non-survivors were higher than those in survivors. These results suggest that HMGB1 may play a pivotal role in the pathogenesis of SAP, and that HMGB1 may act as a key mediator for inflammation and organ failure in this disease. In the present study, to clarify the role of HMGB1 in the pathophysiology of SAP, effects of anti-HMGB1 neutralizing antibody were investigated in SAP in mice.

MATERIALS AND METHODS

Animals

Female C3H/HeN mice (weighing 20–22 g, 9 weeks old) were purchased from CLEA Japan (Tokyo, Japan). The protocol for this animal experiment was approved by the Institutional Animal Committee of Kobe University Graduate School of Medical Sciences.

Anti-HMGB1 neutralizing antibody

Anti-HMGB1 neutralizing antibody (chicken anti-HMGB1 polyclonal antibody) was obtained from Shino-Test Corporation (Sagamihara, Japan). This antibody recognizes mouse HMGB1. The specificity and neutralizing activity of this antibody was confirmed by western blot analysis.

Model for acute necrotizing pancreatitis (closed duodenal loop-induced pancreatitis)

Under general anesthesia with a subcutaneous injection of carbamic acid ethyl ester (urethane) at a dosage of 1.5 g/kg, a midline laparotomy was performed, and a closed loop (2 cm in length) was created by ligating the duodenum at 1 cm proximal and distal sides to the biliopancreatic duct outlet. Only laparotomy was performed in sham-operated mice.

Experimental design

Saline (0.2 mL) or anti-HMGB1 neutralizing antibody (200 µg, 0.2 mL) was injected intraperitoneally, and immediately closed duodenal loop-induced pancreatitis was made. Mice were divided into three groups as follows. Group A: Sham, laparotomy with saline injection. Group B: SAP, severe acute pancreatitis with saline injection. Group C: HMGB1 Ab + SAP, severe acute pancreatitis with anti-HMGB1 antibody injection. Mice were sacrificed 12 h after induction of SAP. Pancreas and lung tissue was removed, fixed in 10% formalin, and stained with hematoxylin and eosin for light microscopic analysis. Blind analysis was carried out for all histological studies. Blood sample was drawn from heart. Serum amylase, aspartate aminotransferase (AST), alanine aminotransferase (ALT), lactate dehydrogenase (LDH), blood urea nitrogen (BUN), and creatinine (Cr) levels were measured using standard clinical automated analyzer.

Bacterial culture of pancreas

The skin was cleaned with 10% povidone iodine. Pancreas

Table 1 Blood biochemical parameters

Parameter	Sham (n = 6)	SAP (n = 20)	HMGB1 Ab + SAP (n = 12)
Amylase (IU/L)	2220 ± 707	52155 ± 14449 ^a	8994 ± 1623 ^c
AST (IU/L)	1499 ± 335	5056 ± 545 ^a	4193 ± 561
ALT (IU/L)	395 ± 288	1215 ± 118 ^a	848 ± 145 ^c
LDH (IU/L)	6743 ± 1206	16403 ± 1072 ^a	15220 ± 4687
BUN (mg/dL)	32 ± 3	93 ± 6 ^a	94 ± 7
Cr (mg/dL)	0.10 ± 0.00	0.42 ± 0.10 ^a	0.16 ± 0.03 ^c

Blood samples were obtained 12 h after induction of SAP. AST: Aspartate aminotransferase; ALT: Alanine aminotransferase; LDH: Lactate dehydrogenase; BUN: Blood urea nitrogen; Cr: Creatinine. ^a*P* < 0.05 vs Sham group; ^c*P* < 0.05 vs SAP group.

was obtained 0, 4, 8, and 12 h after induction of SAP under sterile conditions, and processed for culture of aerobic and anaerobic organisms using a standardized method. Specimens were inoculated onto agar plates including BTB agar, sheep blood agar, chocolate agar (Nippon Becton Dickinson Co. Ltd., Tokyo, Japan), brucella HK agar (Kyokuto Pharmaceutical Co. Ltd., Tokyo, Japan), and GAM (Gifu Anaerobic Medium) agar (Nissui Pharmaceutical Co. Ltd., Tokyo, Japan). BTB agar was incubated in the aerobic chamber at 37°C, sheep blood agar and chocolate agar were incubated in the O₂/CO₂ incubator, and brucella HK agar was incubated in the anaerobic chamber for 48 h, respectively. GAM agar was incubated in the ambient chamber at 37°C for 72 h. When the colony forming was detected, it was considered to be positive for bacterial translocation.

Statistical analysis

The results are expressed as mean ± SE. The Mann-Whitney *U* test and Chi-square test were used to evaluate differences between two groups. A *P* value < 0.05 was considered statistically significant.

RESULTS

Serum amylase level

Twelve hours after induction of SAP, serum amylase levels were significantly elevated in SAP group, and anti-HMGB1 neutralizing antibody significantly reduced its elevation (Table 1).

Morphology of pancreas and lung

Twelve hours after induction of SAP, HE staining of the pancreas showed edema, hemorrhage, leukocyte infiltration, and necrosis. Anti-HMGB1 neutralizing antibody improved the histological alterations of pancreas (Figure 1A). Twelve hours after induction of SAP, HE staining of the lung showed edema, inflammatory infiltration, hemorrhage and thickening of the alveolar membrane. In contrast, anti-HMGB1 neutralizing antibody ameliorated the histological changes in the lungs (Figure 1B).

Hepatic and renal dysfunction

Twelve hours after induction of SAP, serum AST, ALT,

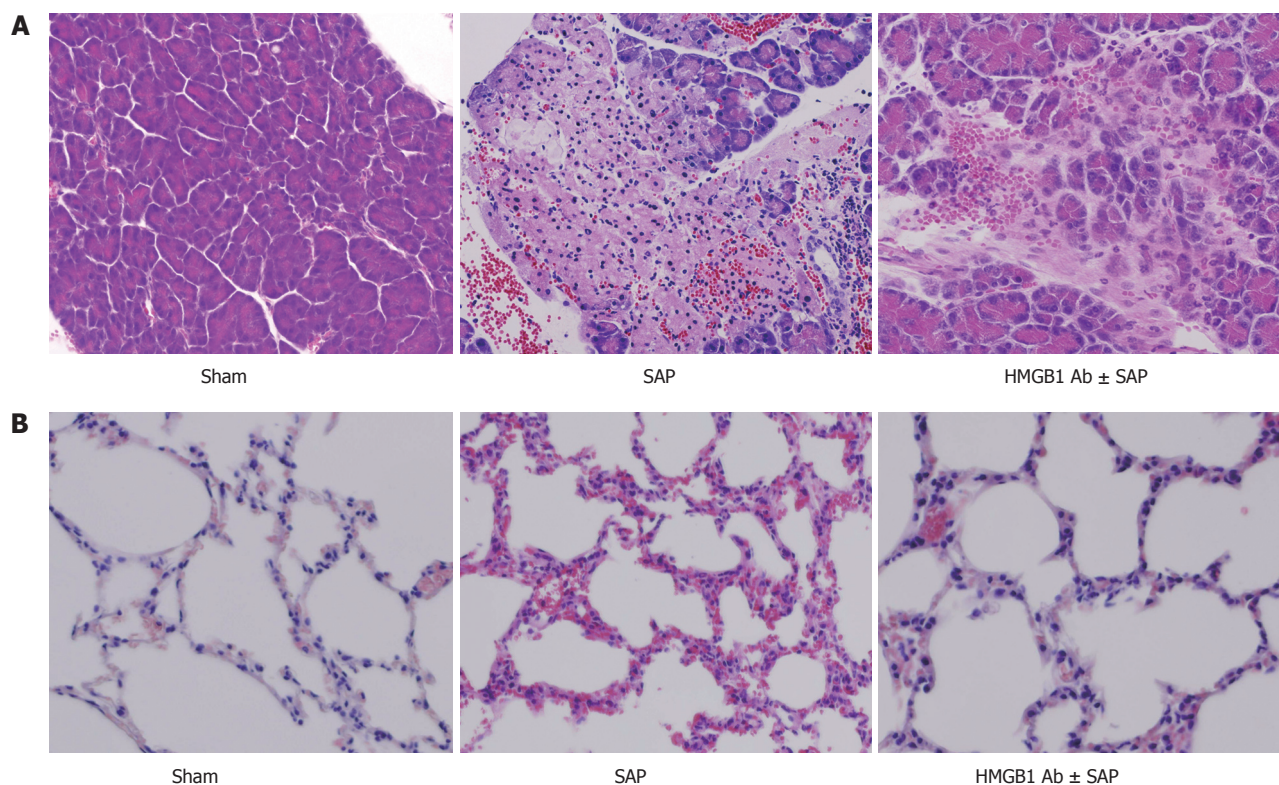


Figure 1 Effect of anti-HMGB1 antibody on the morphology of pancreas and lung in mouse severe acute pancreatitis (SAP). Tissue samples were obtained 12 h after induction of closed duodenal loop pancreatitis. **A:** HE staining of pancreas ($\times 200$); **B:** HE staining of lung ($\times 200$).

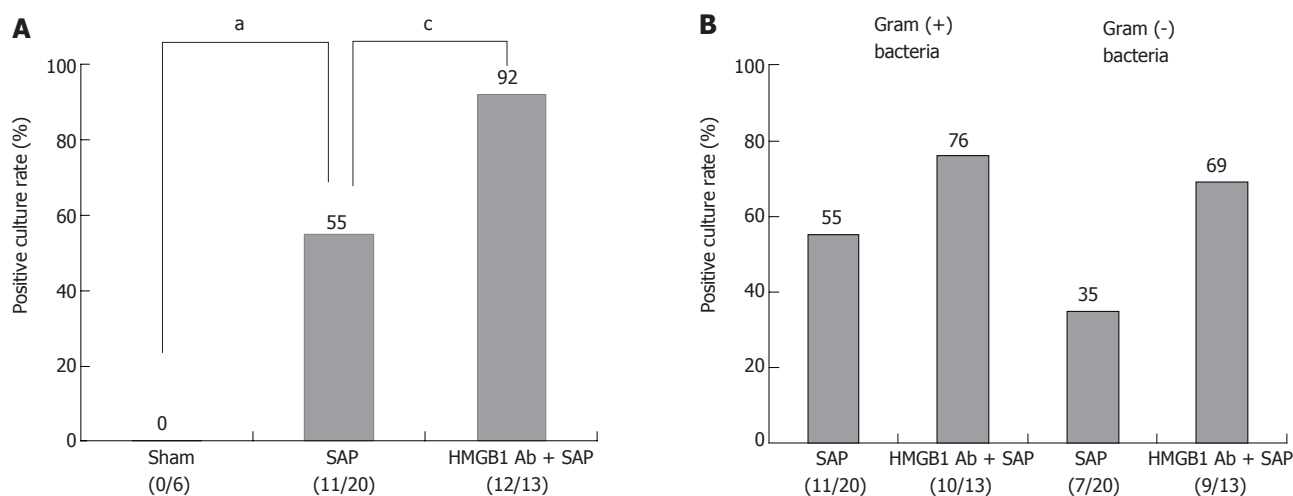


Figure 2 Effect of anti-HMGB1 antibody on the bacterial translocation to pancreas in mouse severe acute pancreatitis (SAP). Bacterial culture of pancreas was examined 12 h after induction of SAP. **A:** Positive rate of bacterial culture of pancreas (overall). ^a $P < 0.05$ vs Sham group; ^b $P < 0.05$ vs SAP group; **B:** Positive rate of bacterial culture of pancreas (Gram-positive and Gram-negative bacteria).

LDH, BUN, and Cr levels were significantly elevated in SAP group, and anti-HMGB1 neutralizing antibody significantly improved the elevated ALT and Cr (Table 1).

Bacterial translocation to pancreas

Bacterial translocation to pancreas was not observed in Sham group (Figure 2A), but could be detected 12 h after the induction of SAP. In earlier periods (0, 4, and 8 h), it was not detected. In SAP group, 55% of mice (11/20) exhibited positive bacterial culture. Anti-HMGB1 antibody significantly increased the positive culture rate to

92% (12/13) (Figure 2A). Positive rate of gram-positive and gram-negative bacterial culture in SAP group was 55% (11/20) and 35% (7/20), respectively. Anti-HMGB1 antibody increased them to 76% (10/13) and 69% (9/13), respectively, but no significant difference was observed (Figure 2B).

DISCUSSION

Extracellular HMGB1 was recently identified as a novel proinflammatory cytokine. In a previous study, we dem-

onstrated that serum HMGB1 levels were significantly elevated in patients with SAP, and were correlated with disease severity^[24]. In this study, we have for the first time demonstrated that blockade of HMGB1 attenuated the development of SAP and associated organ dysfunction, suggesting that HMGB1 may act as a key mediator for inflammatory response and organ injury in SAP. We think that raised HMGB1 may represent a cause of aggravation of SAP (progression to SAP) and associated organ dysfunction as well as a consequence of SAP. On the other hand, HMGB1 can promote alterations in gut barrier function by increasing the permeability in enterocytic monolayers and increasing bacterial translocation in mice^[26]. Similar contributions of HMGB1 to SAP were supposed, but blockade of HMGB1 adversely worsened the bacterial translocation against our expectation.

There have been several reports concerning effects of anti-HMGB1 neutralizing antibody in other pathological conditions. It has been demonstrated that anti-HMGB1 antibody protected against organ injury and improved survival in murine sepsis^[14] and rat sepsis^[27]. Tsung *et al.*^[28] clarified that inhibition of HMGB1 with neutralizing antibody significantly decreased liver damage after ischemia/reperfusion, whereas administration of recombinant HMGB1 worsened it. It has been reported that anti-HMGB1 antibody improved lipopolysaccharide (LPS)-induced acute lung injury in mice^[18], and ventilator-induced lung injury in rabbits^[29]. These observations together with our results in this study indicate that HMGB1 is one of the deteriorating factors in the development of organ injury.

Concerning the elevation of serum HMGB1 levels in SAP, two possible mechanisms can be assumed^[24]. First, HMGB1 may be produced and released by macrophages/monocytes in response to inflammatory mediators. In SAP, it is conceivable that release of humoral mediators from the excessive activated macrophages/monocytes may lead to the remote organ injury. Thus, release of HMGB1 from activated macrophages/monocytes may participate in tissue injury and organ failure in SAP. Second, HMGB1 may be produced and released by injured pancreas or other damaged organs. Recent investigations demonstrated that HMGB1 mRNA expression was significantly increased in liver and lung after rat thermal injury^[30], that HMGB1 concentration was increased in lung epithelial lining fluid of patients with acute lung injury^[18], and that HMGB1 expression was up-regulated in the liver after murine liver ischemia-reperfusion^[28]. Therefore, it is likely that HMGB1 is produced and released by damaged organs in SAP. Change of HMGB1 expression in various tissues should be investigated in SAP.

It was recently clarified that extracellular HMGB1 acts as a cytokine by signaling *via* the receptor for advanced glycosylated end-products (RAGE)^[31,32] and/or *via* members of the toll-like receptor (TLR) family (TLR2 and 4)^[33]. Activation of RAGE and TLR leads to the induction of inflammatory responses *via* NF- κ B. Tsung *et al.*^[28] demonstrated that anti-HMGB1 antibody failed to provide protection in TLR4-defective mice, but successfully reduced liver damage after ischemia/reperfusion in wild-type mice, suggesting that TLR4 is involved in the process as one of the receptors. As TLR4 recognizes LPS of gram-negative

bacilli^[34,35], interactions of HMGB1 with TLR4 may provide an explanation for the ability of HMGB1 to generate inflammatory responses that are similar to those initiated by LPS. Moreover, TLR is involved in not only inflammatory response but also host defense mechanism, and it is postulated that TLR may function defensively against infection. In our result, anti-HMGB1 antibody worsened the bacterial translocation (especially gram-negative bacteria) in SAP, suggesting that HMGB1 may function at least partially *via* TLR (especially TLR4).

Results obtained here raise the possibility that blockade of HMGB1 in the early phase is useful as a new therapeutic option against the inflammatory response and MODS in patients with SAP. Further investigations should be performed to elucidate the role of HMGB1 and HMGB1 signaling in the mechanism of inflammatory response, organ injury, and infection in SAP.

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S- Editor Wang GP L- Editor Ma JY E- Editor Bi L