

## High prevalences of infection with *Giardia intestinalis* genotype B among children in urban and rural areas of Argentina

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The protozoan parasite most frequently associated with diarrhoea worldwide is *Giardia intestinalis*. In 2005, a study was initiated to identify the genotypes of this parasite infecting children in the Argentinian provinces of Buenos Aires, Mendoza and Chaco, and to explore the associations between the genotype detected in a child, the characteristics of the child's household and the child's clinical presentation. Overall, 998 children (504 boys and 494 girls) aged between 2–14 years, with or without symptoms, were enrolled. The *G. intestinalis* in 94 of the 117 stool samples found positive for the parasite by microscopy were successfully genotyped by PCR. Seventy-seven of the children were found to be infected with genotype B only and 14 with genotype AII only, three children being found to have mixed (AII and B) infections.

Only genotype B was detected in children from rural areas ( $P < 0.05$ ) and most *Giardia* detected in children from households with a piped water supply were also of this genotype ( $P < 0.05$ ). The other household characteristics investigated (quality of building, history of flooding, type of sanitation, level of overcrowding, and presence/absence of pet dogs) had no significant effect on the genotype distribution. Children infected with genotype AII were significantly younger than those infected with genotype B ( $P < 0.05$ ) and there was a significant positive association between infection with genotype B and abdominal pain ( $P < 0.05$ ). Diarrhoea was not, however, found to be significantly associated with genotype-AII or genotype-B infection.

This is the first published report on the *Giardia* genotypes circulating in the provinces of Mendoza and Chaco. The results indicate the importance of asymptomatic children in the transmission of *Giardia* among the young.

The protozoan parasite that causes most cases of diarrhoea worldwide is *Giardia intestinalis* (*G. duodenalis*, *G. lamblia*), this parasite being responsible for an estimated 2.5 million cases of diarrhoea each year among children in developing countries (Mahdy *et al.*, 2009). *Giardia* affects mainly children and has a wide range of clinical manifestations, from asymptomatic carriage

to acute diarrhoea with nutritional malabsorption (Thompson, 2004). The World Health Organization (WHO) recently estimated that about 200 million people in Asia, Africa and Latin America have symptomatic giardiasis (Mahdy *et al.*, 2009).

Human giardiasis is a cosmopolitan (sometimes zoonotic) infection whose prevalence varies from 2% (in some developed countries) to 70% (in some developing countries) (Eligio-García *et al.*, 2008). In Argentina, prevalences of giardial infection

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have been found to vary from 6% to 36% and to depend largely on the local levels of sanitation and personal hygiene (Minvielle *et al.*, 2004; López Santorno *et al.*, 2006; Basualdo *et al.*, 2007; Milano *et al.*, 2007; Salomón *et al.*, 2007; Gamboa *et al.*, 2011).

Although seven main genotypes of *G. intestinalis* have been identified (A–G), only genotypes A and B have been associated with human infection (Thompson, 2004). According to Cacciò and Ryan (2008), most human infections involve genotype B, although the relative commonness of genotypes A and B varies geographically. Among Ugandans sharing gorilla habitats (Graczyk *et al.*, 2002), symptomatic patients in Mexico (Ponce-Macotela *et al.*, 2002) and Canada (Van Keulen *et al.*, 2002) and children in Brazilian day-care centres (Volotao *et al.*, 2007), for example, genotype A has been found to be predominant. Among symptomatic and asymptomatic young adults in India (Paintlia *et al.*, 1998), English patients with diarrhoea (Amar *et al.*, 2002), children in Australian day-care centres (Read *et al.*, 2002), symptomatic patients in France (Bertrand *et al.*, 2005) and young children (aged 1 month–9 years) in Peru (Pérez-Cordón *et al.*, 2008), however, B was found to be the most common genotype. The distribution of *Giardia* genotypes among communities in Argentina has been the subject of just two previous studies: Molina *et al.* (2007) found genotype B in a rural population in Buenos Aires province, and Minvielle *et al.* (2008) identified genotypes AII and B in the same province.

The factors behind the variability seen in the morbidity of giardial infection in humans are not clear and there is certainly no consensus on any links between the infecting genotype (A or B) and any clinical manifestation (Homan and Mank, 2001; Read *et al.*, 2002; Cacciò *et al.*, 2005; Haque *et al.*, 2005; Robertson *et al.*, 2009).

The main aims of the present study were to identify the genotypes of *G. intestinalis* circulating among children living in the

Argentinian provinces of Buenos Aires, Mendoza and Chaco, and to explore the associations, if any, between the infecting genotypes, epidemiological characteristics and clinical presentations.

## SUBJECTS AND METHODS

### Subjects and Study Areas

Between August 2005 and July 2007, 1035 children (533 boys and 502 girls) aged 2–14 years and with or without symptoms were enrolled at health centres or in public schools or soup kitchens. All the subjects were permanent residents in the provinces of Buenos Aires, Mendoza or Chaco, in Argentina. Enrollment was voluntary.

Buenos Aires province, in central–eastern Argentina, has a humid and moderate climate, with a mean annual rainfall of 1100 mm. The subjects from this province lived in Hipólito Vieytes (35°16'S, 57°34'E), Atalaya (35°01'S, 57°31'E), Barrio Obrero (34°56'S, 57°52'E), El Paligüe (34°58'S, 57°53'E), or 4 de Junio (30°50'S, 58°06'E), which lie 40, 45, 7, 3 and 35 km, respectively, from the provincial capital of La Plata.

The subjects from Mendoza province (which lies in central–western Argentina and has a moderate and arid climate, with a mean annual rainfall of 250 mm) were enrolled in Barrio Jardín Aeroparque (32°53'S, 68°41'E), which lies 4 km from the provincial capital of Mendoza (and 1090 km from La Plata).

The subjects from the north–eastern province of Chaco province (which has a warm and humid subtropical climate, with a mean annual rainfall of 1200 mm) were enrolled in the city of Resistencia, which lies at 27°27'S and 58°59'E (and 1070 km from La Plata).

### Parasitology and Data Collection

A medical team conducted a copro–parasitological examination of each child and collected relevant clinical–epidemiological

data on each child, from the child's parents, who were asked to complete a standardized questionnaire.

#### DATA COLLECTION

The questionnaire recorded details of the child's house — setting (rural or urban), quality ['precarious' (built of wood/cardboard/metal sheet) or 'substantial' (built of cement and masonry)], sanitation (sewerage, latrine or cesspool), source of water (piped or pump), history of flooding (whether or not the house was flooded more than twice/year), pet dogs (present or absent), level of overcrowding (whether or not there were more than three people/room) — and whether or not the child had any symptoms (presence/absence of diarrhoea in the last week, anorexia, vomiting, asthenia and/or abdominal pain).

#### FAECAL SAMPLES

Faecal material was collected, by each child's parents or tutors, on a daily basis for 5 consecutive days, in the same disposable container with 5% formaldehyde. These samples were concentrated using a modification of the Telemann method. Briefly, 10 g of homogenized sample were macerated in 15 ml Telemann solution [made by dissolving 5 g NaCl in 50 ml of 40% (w/v) formaldehyde and 950 ml distilled water]. After filtering the sample through a double layer of gauze, 1 ml of ether was added and then the suspension was centrifuged at  $1000 \times g$  for 3 min. Three wet smears of each resultant pellet, each smear treated with a drop of Lugol's solution, were then checked for *Giardia* and other intestinal parasites under a light microscope, at  $\times 100$  and  $\times 400$  (Minvielle *et al.*, 2008).

#### *Giardia* GENOTYPING

All faecal samples found smear-positive for *Giardia* cysts were processed further, to concentrate and clean the cysts and extract their DNA, so that the genotype could be

identified (Molina *et al.*, 2007). The cysts were first isolated using sucrose-gradient concentration, counted in a Neubauer chamber and preserved at 4°C until the DNA could be isolated.

Prior to the DNA extraction, the cysts were broken open, by three cycles of freezing (at -80°C for 30 min) and thawing (at +80°C for 30 min), and then incubated at 60°C for 24 h with lysis E buffer [100 mM Tris-HCl, 100 mM EDTA, 2% (w/v) SDS, 0.2 M NaCl and 1 mM mercaptoethanol, containing 1 mg proteinase K/ml] before being preserved at -20°C.

The DNA in the suspensions of broken cysts was purified using the QIAamp DNA stool mini kit (QIAGEN, Hilden, Germany) according to the manufacturer's instructions and then preserved at -20°C. These DNA samples, once thawed, were used as the templates in a hemi-nested PCR designed to amplify a fragment of the *Giardia tpi* (triosephosphate isomerase) gene (Amar *et al.*, 2002). In the first round, 3 µl of purified DNA were used in a 30-µl reaction mixture of 1 × PCR buffer containing 3 mM MgCl<sub>2</sub>, 0.25 mM of each deoxyribonucleotide triphosphate (dNTP), 0.25 µM of each primer (TPIAF, TPIAR, TPIBF and TPIBR), 0.1 µg bovine serum albumin (BSA)/µl and 0.1 U *Taq* DNA polymerase/µl. The thermocycler used was set to give an initial denaturation at 94°C for 4 min, then 30 cycles, each of 30 s at 94°C, 30 s at 52°C and 1 min at 72°C, before a final extension for 10 min at 72°C.

The amplicons from the first round were used as templates for two single PCR (PCR II A and PCR II B). The 30-µl reaction mixture for each of these PCR consisted of 1 × PCR buffer containing 1.5 mM MgCl<sub>2</sub>, 0.25 mM of each dNTP, 1 µM of each primer (TPIAIF and TPIAR in PCR II A, TPIBIF and TPIBR in PCR II B), 0.1 µg BSA/µl and 0.05 U *Taq* DNA polymerase/µl. For both reactions, the thermocycler was set to give an initial denaturation at 94°C for 4 min, then 33 cycles, each of 30 s at 94°C, 30 s at 54°C and 1 min at 72°C, before a

final extension for 10 min at 72°C. In each PCR, samples of DNA from *G. intestinalis* of genotypes A and B (1 ng/μl; kindly donated by Dr H. van Keulen from Cleveland University, OH) were used as positive controls and double-distilled water was used as the negative control.

The amplicons from PCR II A (5 μl) were incubated with 5 U *Rsa*I, in a final volume of 30 μl, at 37°C for 3 h, so that they could be investigated by restriction-fragment-length-polymorphism (RFLP) analysis (Amar *et al.*, 2002). The amplicons (from PCR II A and PCR II B) and the PCR II-A digestion products were investigated via electrophoresis in 1.5% and 3% (w/v) agarose gel, respectively, so that genotypes could be identified (Amar *et al.*, 2002).

### Ethics

Stool samples were collected only after parents or tutors gave their informed consent.

The study protocol, which was approved by the Ethical Review Committees of La Plata National University, Roemmers Foundation and the Argentinian Ministry of Education, met the requirements of the Declaration of Helsinki (1964), the Nuremberg Code (1947) and the Argentinian National Act 25.326 (on the protection of personal information).

### Data Analysis

Pearson's  $\chi^2$  and Fisher's exact tests were used to explore the relationships between each of the categorical variables investigated in the present study and the relative frequency of each *Giardia* genotype. Version 14.0 of the SPSS software package (SPSS Inc, Chicago, IL) was used to store, edit and analyse the data. A *P*-value of <0.05 was considered indicative of a statistically significant difference or association.

## RESULTS

Although stool samples were collected from 1035 children, the samples and data from only

998 children (504 boys and 494 girls) were investigated, the other 37 children being excluded because their stool samples had not been collected correctly (19) or because their questionnaires were incomplete (18).

The microscopical examination of the stool samples from the 998 children revealed infections with *Blastocystis hominis* (in 39.1%), *Giardia intestinalis* (11.7%), *Entamoeba coli* (13.8%), *Endolimax nana* (10.1%), *Enterobius vermicularis* (5.3%), *Ascaris lumbricoides* (2.2%), *Hymenolepis nana* (1.2%), *Chilomastix mesnili* (1.1%) and *Trichuris trichiura* (0.5%).

Although *Giardia* cysts were found in the faeces of 117 children, the *Giardia* in only 94 (80.3%) of the 117 cyst-positive samples were successfully genotyped. Seventy-seven children were found to be infected with *Giardia* only of genotype B, and 14 with *Giardia* only of genotype AII, while three children (from the same school) were each found positive for both genotype AII and genotype B.

Faeces that apparently contained *Giardia* cysts of genotype B only, genotype AII only and both genotype AII and genotype B contained similar mean (S.D.) numbers of giardial cysts: 153 (184), 123 (146) and 147 (119) cysts/μl, respectively (*P*>0.05).

Of the 52 boys with genotyped giardial infection, 39 (75%) appeared to be infected with genotype B only, 11 (21%) with AII only, and two (4%) with both genotypes AII and B. Although PCR was more often successful with the cyst-positive faecal samples from boys than with similar samples from girls, the difference was not statistically significant (*P*>0.05).

The mean (S.D.) age of the 94 children with genotyped giardial infection was 6.0 (3.4) years, 49 (52.1%) being aged between 2 and 5 years (preschool) and the other 45 (47.9%) being schoolchildren aged 6–14 years. A child found infected with genotype B only was, on average, significantly older than a child found infected with genotype AII only (Fig. 1; *P*<0.05).

Both *Giardia* genotypes detected in the present study (B and AII) were found in

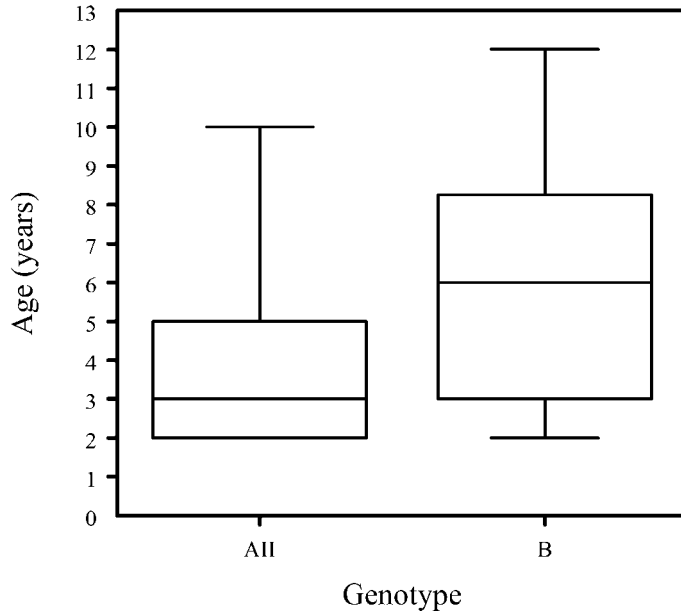


FIG. 1. A 'box-and-whisker' plot of the ages of the Argentinian children found infected with *Giardia intestinalis* of genotype AII only or B only. In each plot, the horizontal bar indicates the median, the box the range between the 25th and 75th percentiles, and the 'whiskers' the range between the fifth and 95th percentiles.

each of the three provinces under study (Fig. 2). Of the 94 children with genotyped giardial infection, 72 (76.5%) lived in urban areas [55 of them infected with genotype B, 14 with AII and three with mixed (AII+B) infections] and 22 (23.5%), all infected with genotype B, lived in rural areas. Household setting (rural *v.* urban) had a statistically

significant effect on the recorded genotype distribution, with a giardial infection in a child from a rural household more likely to be identified as genotype B than such an infection in an urban child ( $P < 0.05$ ).

The Table shows the relationships between the recorded distribution of *Giardia* genotypes and the variables investigated via the

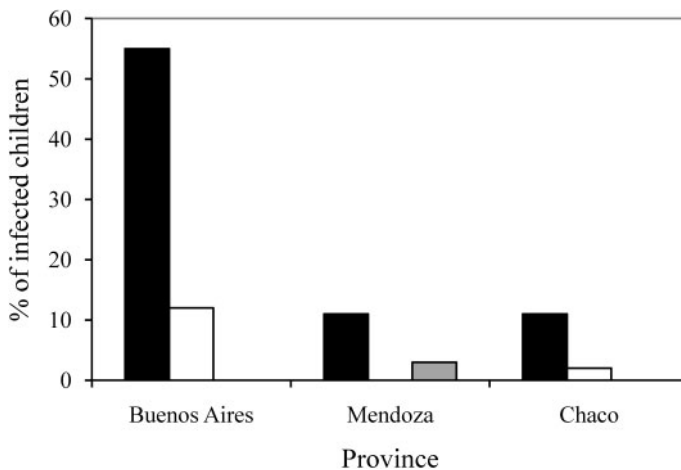


FIG. 2. The numbers of children from Buenos Aires, Mendoza and Chaco provinces found infected with *Giardia intestinalis* of genotype B only (■), genotype AII only (□) or both genotypes B and AII (▒).

epidemiological questionnaire. Only one such relationship — that between the type of water-supply system and infecting genotype — appeared statistically significant ( $P < 0.05$ ); the *Giardia*-infected study children from households with a piped water supply were less likely to be infected with genotype B than their counterparts from households that depended on wells for their water supply.

Fifty (53.2%) of the 94 children with genotyped giardial infection (42 apparently infected with genotype B only and eight apparently infected with genotype AII only) appeared to be suffering from symptomatic giardiasis (Fig. 3). Twenty (47.6%) of the symptomatic children found infected with genotype B and seven (87.5%) of the symptomatic children found infected with genotype AII were aged 2–5 years. All three children found to have mixed (AII+B) infection were asymptomatic. Genotype-B infection was significantly associated with the presence of abdominal pain ( $P < 0.05$ ).

## DISCUSSION

Given its frequently high prevalence among children, giardiasis is an important infection

from an epidemiological viewpoint. It is therefore surprising that, although there have been several previous studies on the prevalence of giardial infection in Argentina (López Santorno *et al.*, 2006; Milano *et al.*, 2007; Salomón *et al.*, 2007; Gamboa *et al.*, 2011), there were no attempts before 2007 to identify the *G. intestinalis* genotypes circulating in this country. A few years ago, genotype B was detected in two cities in Buenos Aires province (Molina *et al.*, 2007; Minvielle *et al.*, 2008). One aim of the present study was to identify the genotypes circulating, among children, in other Argentinean provinces, and this article represents the first report of *G. intestinalis* genotypes in the provinces of Mendoza and Chaco. Genotype B appeared to be the predominant form of *G. intestinalis* infecting children in all three provinces under study. This genotype has also been found to predominate in recent studies on schoolchildren in the Philippines (Yason and Rivera, 2007), Thailand (Ratanapo *et al.*, 2008) and India (Ajampur *et al.*, 2009).

In the present study, as in an earlier study in Nicaragua (Lebbad *et al.*, 2008) and the previous studies in Argentina (Molina *et al.*, 2007; Minvielle *et al.*, 2008), genotype AII appeared rarer in humans than genotype

TABLE. The recorded epidemiological variables and their links with the *Giardia* genotypes detected in children in the Argentinian provinces of Buenos Aires, Mendoza and Chaco, in 2005–2007

Variable	No. and (%) of the children found infected with genotype(s):		
	AII only	B only	Both AII and B
BUILD QUALITY OF HOUSE			
Precarious	7 (50)	26 (34)	0 (0)
Substantial	7 (50)	51 (66)	3 (100)
SANITATION			
Latrine	7 (50)	19 (25)	0 (0)
Cesspool	6 (43)	42 (55)	0 (0)
Sewerage	1 (7)	16 (20)	3 (100)
WATER SUPPLY			
Piped	12 (86)	24 (31)	3 (100)
Well	2 (14)	53 (69)	0 (0)
House flooded at least twice/year	8 (57)	33 (43)	0 (0)
Household owns pet dog	9 (64)	60 (78)	3 (100)
Household overcrowded	5 (36)	17 (22)	0 (0)

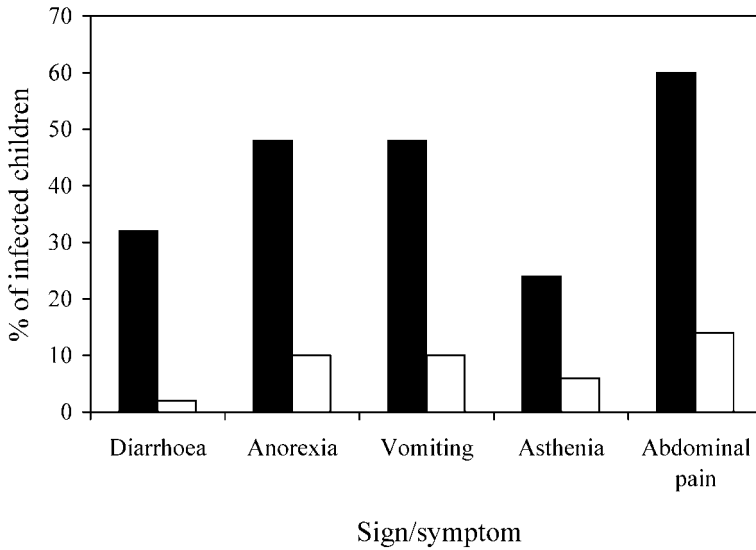


FIG. 3. The percentages of the children found infected with *Giardia intestinalis* of genotype B only (■) or genotype AII only (□) who were suffering from diarrhoea, anorexia, vomiting, asthenia and/or abdominal pain when they were enrolled.

B. Elsewhere in Latin America, however, in Cuba (Pelayo *et al.*, 2008) and Peru (Pérez-Cordón *et al.*, 2008), AII has been found more common than genotype B, while the predominant *G. intestinalis* genotype infecting humans in Brazil appears to vary according to the region (Souza *et al.*, 2007; Volotao *et al.*, 2007). The predominance of one *G. intestinalis* genotype over another in a particular area has been attributed to biological as well as geographical factors and, in certain endemic areas, all the giardial infections in humans appear to involve just one genotype (Volotao *et al.*, 2007). Despite the geographical differences, the predominant genotype in the present study was the same in each study province, perhaps indicating that historical immigration and other mass movements of humans (Ponce-Macotela *et al.*, 2002) have reduced the geographical variation in genotype distribution across Argentina. Given that most immigrants who settled in Argentina during the 19th and 20th Centuries were European and most giardial infections found in Europeans are now of genotype B, it is perhaps not surprising that genotype B is

now so common in Argentina. More than 60% of the 803 isolates of *G. intestinalis* from Europeans recently investigated by Sprong *et al.* (2009) and 57% of the 108 isolates from Spaniards studied by Sahagún *et al.* (2008) were of genotype B. It is, however, unclear how immigration from areas where genotype B predominates could explain the apparent absence of genotype AII from the rural children investigated in the present study. Pelayo *et al.* (2008) found genotypes A and B to be equally common among children from a rural Cuban community and Gelanew *et al.* (2007) found genotype A to predominate in rural communities in Ethiopia.

In the present study, three children were each found infected with two genotypes (i.e. AII and B). The fact that the three attended the same school may indicate that they were exposed to the same source of infection. Mixed infections reflect the complex circulation of the parasites in the environment, and potential exposure to sources of multiple infection, such as sewage (Hopkins *et al.*, 1999; Lalle *et al.*, 2005; Gelanew *et al.*, 2007).

In the present study, the relative frequencies of the B and AII genotypes (among children with giardial infection) appeared to be unaffected by most environmental and socio-cultural conditions, such as the quality of the housing and sanitation, level of overcrowding, frequency of flooding and the presence/absence of household dogs. Kohli *et al.* (2008) found no statistically significant links between the relative frequencies of infecting genotypes in Brazilian children and any of the epidemiological variables that they investigated. Only the source of household water appeared to affect the relative frequencies of the B and AII genotypes significantly in the present study, perhaps indicating that there is more transmission of genotype B from well water than of genotype AII.

Person-to-person transmission of genotype AII may explain why, in the present study, compared with their older counterparts, the preschool children (with their relatively low standards of personal hygiene and high levels of mouth-hand contact) who were *Giardia*-positive were significantly more likely to be carrying this genotype. Pet-human transmission of any genotype may also be important in Argentina and merits further investigation. Although no significant link was observed, in the present study, between household ownership of a pet dog and the relative frequencies of the B and AII genotypes, it remains possible that other types of pet or livestock are involved in transmission.

In this study, the cyst loads in faecal material were similar whatever the infecting genotype. Although Hussein *et al.* (2009) found that people infected with genotype B excreted greater numbers of giardial cysts than those infected with genotype AII, their subjects were all patients who were seeking medical treatment for abdominal problems (not, as in the present study, a mixture of children with and without intestinal symptoms).

In the present study, the children found infected with genotype AII were significantly younger than those found infected

with genotype B (Fig. 1). This observation matches that made earlier, in Argentina, by Minvielle (2005). When Kohli *et al.* (2008) carried out a diachronic analysis of 47 children infected with *Giardia*, they concluded that all the children had been (re)infected with genotype B, irrespective of the initial infecting genotype. Genotype-B infection in older children might be a consequence of various factors: the genotype is endemic in schools, it causes more persistent infections than genotype AII, and/or it is more resistant to anti-giardiasis drugs.

Infection with *Giardia* may often be asymptomatic. In this study, the frequency of symptomatic illness (presumably symptomatic giardiasis) tended to decrease with age, probably as the result of acquired immunity (Hughes and Kelly, 2006). Pezzani (2007) found that, in rural Argentina, most intestinal parasites cause more symptoms in young children than in older children and adults. Although, in the present study, infection with genotype AII appeared to be no more or less likely to be symptomatic than infection with genotype B, genotype-B infection did appear to be significantly linked to abdominal pain. Other studies have reported a significant association between genotype A and the presence of symptoms (Read *et al.*, 2002; Aydin *et al.*, 2004; Haque *et al.*, 2005; Sahagún *et al.*, 2008) and other genotype-linked differences in clinical symptoms (Homan and Mank, 2001; Gelanew *et al.*, 2007).

Diarrhoea was reported for <20% of the *Giardia*-infected children investigated in the present study and was not significantly associated with the infecting genotype in any age-group. These results are very similar to those of Read *et al.* (2002), who studied Dutch children aged <5 years, and those of Haque *et al.* (2005), who genotyped the *Giardia* cysts excreted by patients in Bangladesh, only 29% of whom had diarrhoea.

To conclude, this is the first report on the *Giardia* genotypes in the provinces of Mendoza and Chaco. Genotype B was



predominant in the children of these provinces and in Buenos Aires province. There was a significant association between genotype B and both residence in a rural area and the use of well water. Other environmental and socio-cultural conditions had no apparent effect on the infecting genotype of *Giardia*. Children infected with genotype AII were significantly younger than those infected with genotype B but symptomatic giardiasis was detected at similar frequency among children infected with each genotype. There was a statistically significant association between genotype-B infection and abdominal pain. Diarrhoea was found in <20% of the children infected with *Giardia* and was not significantly associated with the infecting genotype in any age-group. Future research to expand our knowledge on the relationships between *Giardia* genotype and symptoms in children should include studies on larger groups of children with symptomatic and asymptomatic infection and on children from other regions of Argentina.

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## REFERENCES

- Ajjampur, S. R., Sanzaran, P., Kannan, A., Sathyakumar, K., Sarkar, R., Gladstone, B. & Kang, G. (2009). *Giardia duodenalis* assemblages associated with diarrhea in children in South India identified by PCR-RFLP. *American Journal Tropical Medicine and Hygiene*, **80**, 16–19.
- Amar, C. F., Dear, P. H., Pedraza-Diaz, S., Looker, N., Linnane, E. & McLauchlin, J. (2002). Sensitive PCR-restriction fragment length polymorphism assay for detection and genotyping of *Giardia duodenalis* in human feces. *Journal of Clinical Microbiology*, **40**, 446–452.
- Aydin, A. F., Besirbellioglu, B. A., Avci, I. Y., Tanyuksel, M., Araz, E. & Pahsa, A. (2004). Classification of *Giardia duodenalis* parasites in Turkey into Groups A and B using restriction fragment length polymorphism. *Diagnostic Microbiology and Infectious Diseases*, **50**, 147–151.
- Basualdo, J. A., Cordoba, M. A., de Luca, M. M., Ciarmela, M. L., Pezzani, B. C., Grenovero, M. S. & Minvielle, M. C. (2007). Intestinal parasitoses and environmental factors in a rural population of Argentina, 2002–2003. *Revista do Instituto de Medicina Tropical de São Paulo*, **49**, 251–255.
- Bertrand, I., Albertini, L. & Schwartzbrod, J. (2005). Comparison of two target genes for detection and genotyping of *Giardia lamblia* in human feces by PCR and PCR-restriction fragment length polymorphism. *Journal of Microbiology*, **43**, 5940–5944.
- Cacciò, S. M. & Ryan, U. (2008). Molecular epidemiology of giardiasis. *Molecular and Biochemical Parasitology*, **160**, 75–80.
- Cacciò, S., Thompson, R. C. A., McLauchlin, J. & Smith, H. V. (2005). Unravelling *Cryptosporidium* and *Giardia* epidemiology. *Trends in Parasitology*, **21**, 430–437.
- Eligio-García, L., Cortes-Campos, A., Cota-Guajardo, S., Gaxiola, S. & Jiménez-Cardoso, E. (2008). Frequency of *Giardia intestinalis* assemblages isolated from dogs and humans in a community from Culiacan, Sinaloa, Mexico using beta-giardin restriction gene. *Veterinary Parasitology*, **156**, 205–209.
- Gamboa, M., Navone, G., Orden, A., Torres, M., Castro, L. & Oyhenart, E. (2011). Socio-environmental conditions, intestinal parasitic infections and nutritional status in children from a suburban neighborhood of La Plata, Argentina. *Acta Tropica*, in press.
- Gelanew, T., Lalle, M., Hailu, A., Pozio, E. & Cacciò S. (2007). Molecular characterization of human isolates of *Giardia duodenalis* from Ethiopia. *Acta Tropica*, **102**, 92–99.
- Graczyk, T. K., Bosco-Nizeyi, J., Ssebide, B., Thompson, R. C. A., Read, C. & Cranfield, M. R. (2002). Anthropozoonotic *Giardia duodenalis* genotype (assemblage) A infections in habitats of free-ranging human-habituated gorillas, Uganda. *Journal of Parasitology*, **88**, 905–909.
- Haque, R., Roy, S., Kabir, M., Stroup, S. E., Mondal, D. & Hout E. R. (2005). *Giardia* assemblage A infection and diarrhea in Bangladesh. *Journal of Infectious Diseases*, **192**, 2171–2173.
- Homan, W. L. & Mank, T. G. (2001). Human giardiasis: genotype linked differences in clinical symptomatology. *International Journal for Parasitology*, **31**, 822–826.
- Hopkins, R. M., Constantine, C. C., Groth, D. A., Wetherall, J. D., Reynoldson, J. A. & Thompson, R. C. A. (1999). PCR-based DNA fingerprinting of *Giardia duodenalis* isolates using the intergenic rDNA spacer. *Parasitology*, **118**, 531–539.

- Hughes, S. & Kelly, P. (2006). Interactions of malnutrition and immune impairment, with specific reference to immunity against parasites. *Parasite Immunology*, **28**, 577–588.
- Hussein, A. I., Yamaguchi, T., Nakamoto, K., Iseki, M. & Tokoro, M. (2009). Multiple-subgenotype infections of *Giardia intestinalis* detected in Palestinian clinical cases using a subcloning approach. *Parasitology International*, **58**, 258–262.
- Kohli, A., Bushen, O. Y., Pinkerton, R. C., Houpt, E., Newman, R. D., Sears, C. L., Lima, A. M. & Guerrant, R. L. (2008). *Giardia duodenalis* assemblage, clinical presentation and markers of intestinal inflammation in Brazilian children. *Transactions of the Royal Society of Tropical Medicine and Hygiene*, **102**, 718–725.
- Lalle, M., Pozio, E., Capelli, G., Bruschi, F., Crotti, D. & Cacciò, M. (2005). Genetic heterogeneity at the beta-giardin locus among human and animal isolates of *Giardia duodenalis* and identification of potentially zoonotic subgenotypes. *International Journal for Parasitology*, **35**, 207–213.
- Lebbad, M., Ankarkev, J., Tellez, A., Leiva, B., Andersson, J. O. & Svard, S. (2008). Dominance of *Giardia* assemblage B in León, Nicaragua. *Acta Tropica*, **106**, 44–53.
- López Santorno, M. S., Garraza, M., Gamboa, M. I., Zonta, M. L. & Navone, G. T. (2006). Enteroparasitosis en la población infantil de Fuerte Esperanza, Chaco, Argentina. *Acta Bioquímica Clínica Latinoamericana*, **3**, (Suppl. 3), 235.
- Mahdy, A. K., Surin, J., Wan, K. L., Mohd-Adnan, A., Al-Mekhlafi, M. S. & Lim, Y. A. L. (2009). *Giardia intestinalis* genotypes: risk factors and correlation with clinical symptoms. *Acta Tropica*, **112**, 67–70.
- Milano, A., Oscherov, E., Palladino, A. & Bar, A. (2007). Enteroparasitosis infantil en un área urbana del nordeste argentino. *Medicina (Buenos Aires)*, **67**, 238–242.
- Minvielle, M. C. (2005). *Detección de genotipos A (I y II) y B por PCR-RFLP de Giardia lamblia en heces humanas y animales de La Plata y su área de influencia*. MSc thesis, Universidad Nacional de San Martín, San Martín, Argentina.
- Minvielle, M. C., Pezzani, B. C., Córdoba, M. A., De Luca, M. M., Apezteguia, M. C. & Basualdo, J. A. (2004). Epidemiological survey of *Giardia* spp. and *Blastocystis hominis* in an Argentinian rural community. *Korean Journal of Parasitology*, **42**, 121–127.
- Minvielle, M., Molina, N., Polverino, D. & Basualdo, J. (2008). First genotyping of *Giardia lamblia* from human and animal feces in Argentina, South America. *Memórias do Instituto Oswaldo Cruz*, **103**, 98–103.
- Molina, N., Polverino, D., Minvielle, M. & Basualdo, J. (2007). PCR amplification of triosephosphate isomerase gene of *Giardia lamblia* in formalin-fixed feces. *Revista Latinoamericana de Microbiología*, **49**, 6–11.
- Paintlia, A. S., Descoteaux, S., Spencer, B., Chakraborti, A., Mahajan, R. C., Ganguly, N. K. & Samuelson, J. (1998) *Giardia lamblia* groups A and B among young adults in India. *Clinical Infectious Diseases*, **26**, 190–191.
- Pelayo, L., Nuñez, F. A., Rojas, L., Furuset Hansen, E., Gjerde, B., Wilke, H., Mulder, B. & Robertson, L. (2008). *Giardia* infections in Cuban children: the genotypes circulating in a rural population. *Annals of Tropical Medicine and Parasitology*, **102**, 585–595.
- Pérez-Cordón, G., Cordova Paz Soldán, O., Vargas Vásquez, F., Velasco Soto, J., Sempere Bordes, L., Sánchez Moreno, M. & Rosales, M. (2008). Prevalence of enteroparasites and genotyping of *Giardia lamblia* in Peruvian children. *Parasitology Research*, **103**, 459–465.
- Pezzani, B. C. (2007). *Estrategias de control de las parasitosis intestinales en una comunidad rural*. PhD thesis, Facultad de Ciencias Naturales y Museo, Universidad Nacional de La Plata, La Plata, Argentina.
- Ponce-Macotela, M., Martínez-Gordillo, M. N., Bermúdez-Cruz, R. M., Salazar-Schettino, P. M., Ortega-Pierres, G. & Ey, P. L. (2002). Unusual prevalence of the *Giardia intestinalis* A-II subtype amongst isolates from humans and domestic animals in Mexico. *International Journal for Parasitology*, **32**, 1201–1202.
- Ratanapo, S., Mungthin, M., Soontrapa, S., Faithed, C., Siripattanapipong, S., Rangsin, R., Naaglor, T., Piyaraj, P., Taamasri, P. & Leelayoova, S. (2008). Multiple modes of transmission of giardiasis in primary schoolchildren of a rural community, Thailand. *American Journal of Tropical Medicine and Hygiene*, **78**, 611–615.
- Read, C., Walters, J., Robertson, I. D. & Thompson, R. C. A. (2002). Correlation between genotype of *Giardia duodenalis* and diarrhoea. *International Journal for Parasitology*, **32**, 229–231.
- Robertson, L. J., Hanevik, K., Escobedo, A. A., Mørch, K. & Langeland, N. (2009). Giardiasis — why do the symptoms sometimes never stop? *Trends in Parasitology*, **26**, 78–82.
- Sahagún, J., Clavel, A., Goñi, P., Seral, C., Llorente, M. T., Castillo, F. J., Capilla, S., Arias, A. & Gómez-Lus, R. (2008). Correlation between the presence of symptoms and the *Giardia duodenalis* genotype. *European Journal of Clinical Microbiology and Infectious Diseases*, **27**, 81–83.
- Salomón, C., Tonelli, R., Borremans, C., Bertello, D., De Jong, L., Jofré, C., Enriquez, V., Carrizo, L. & Costamagna, S. (2007). Prevalencia de parásitos intestinales en niños de la ciudad de Mendoza, Argentina. *Parasitología Latinoamericana*, **62**, 49–53.
- Souza, S. L. P., Gennari, S. M., Richtzenhain, L. J., Pena, H. F. J., Funada, M. R., Cortez, A., Gregori, F. & Soares, R. M. (2007). Molecular identification of *Giardia duodenalis* isolates from humans, dogs, cats and

- cattle from the state of Sao Paulo, Brazil, by sequence analysis of fragments of glutamate dehydrogenase (*gdh*) coding gene. *Veterinary Parasitology*, **149**, 258–264.
- Sprong, H., Cacciò, S. M. & van der Giessen, J. W. (2009). Identification of zoonotic genotypes of *Giardia duodenalis*. *PLoS Neglected Tropical Diseases*, **3**, e558.
- Thompson, R. C. A. (2004). The zoonotic significance and molecular epidemiology of *Giardia* and giardiasis. *Veterinary Parasitology*, **126**, 15–35.
- Van Keulen, H., Macechko, P. T., Wade, S., Schaaf, S., Wallis, P. M. & Erlandsen, S. L. (2002). Presence of human *Giardia* in domestic, farm and wild animals, and environmental samples suggest a zoonotic potential for giardiasis. *Veterinary Parasitology*, **108**, 97–107.
- Volotao, A. C., Costa-Macedo, L. M., Haddad, F. S. M., Brandão, A., Peralta, J. M. & Fernandes, O. (2007). Genotyping of *Giardia duodenalis* from human and animal samples from Brazil using beta-giardin gene: a phylogenetic analysis. *Acta Tropica*, **102**, 10–19.
- Yason, J. A. & Rivera, W. L. (2007). Genotyping of *Giardia duodenalis* isolates among residents of slum area in Manila, Philippines. *Parasitology Research*, **101**, 681–687.