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The Risk of Pancreatic Cancer in Families with Lynch Syndrome

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Abstract

Context—Lynch syndrome is an inherited cause of colorectal cancer caused by mutations of DNA mismatch repair genes. A number of extracolonic tumors have been associated with the disorder, including pancreatic cancer. However, the risk of pancreatic cancer in Lynch Syndrome is uncertain and not quantified.

Objective—To estimate pancreatic cancer risk in families with germline mismatch repair gene mutations.

Design, Setting, Patients—Cancer histories of probands and their relatives were evaluated in mismatch repair gene mutation carriers in the familial cancer registries of the Dana-Farber Cancer Institute and University of Michigan Comprehensive Cancer Center. Families enrolled prior to the study start date (June 2008) were eligible. Age-specific cumulative risks and hazard ratio

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estimates of pancreatic cancer risk were calculated and compared to the general population using modified segregation analysis, with correction for ascertainment.

Main Outcome Measures—Age-specific cumulative risks and hazard ratio estimates of pancreatic cancer risk

Results—Data on 6,342 individuals from 147 families with mismatch repair gene mutations were analyzed: 21% of families (31/147) reported at least one case of pancreatic cancer. Forty-seven pancreatic cancers were reported (21 male, 26 female) with no gender-related difference in age of diagnosis: 51.5 years v. 56.5 years for men and women respectively. The cumulative risk of pancreatic cancer in these families with gene mutations was 1.3% (95% CI: 0.31, 2.32) up to age 50 years and 3.7% (95% CI: 1.45, 5.88) up to age 70 years which represents an 8.6-fold increase (95% CI: 4.7, 15.7) compared to the general population.

Conclusions—Among 147 families with germline mismatch repair gene mutations, the risk of pancreatic cancer was increased compared to the U.S. population. Individuals with mismatch repair gene mutations and a family history of pancreatic cancer are appropriate to include in studies to further define the risk of pre-malignant and malignant pancreatic neoplasms and potential benefits and limitations of surveillance.

Introduction

Pancreatic cancer is the fourth leading cause of cancer deaths in the U.S.¹ Though most cases are thought to be sporadic, data suggest up to 10% of ductal adenocarcinomas may be due to an inherited predisposition based on familial clustering.^{2,3} For most pancreatic cancer kindreds, the causative gene has not been identified. In a subset of families, pancreatic cancer may be an integral tumor in a number of familial cancer syndromes with established germline mutations. These conditions include Peutz-Jeghers Syndrome (cumulative lifetime risk of 36%),^{4,5} Familial Atypical Multiple Mole Melanoma Syndrome (lifetime risk = 17%),⁶ Hereditary Breast/Ovarian Cancer Syndrome (lifetime risks = 1.2% and 2.1%, for *BRCA1* and *BRCA2* carriers, respectively),^{7,8} Hereditary Pancreatitis (lifetime risk = 40%)⁹ and the newly described Familial Pancreatic Cancer due to mutations in *PALB2* (risk not specified).¹⁰

Pancreatic cancer has also been observed in Lynch Syndrome, an autosomal dominant condition caused by defects in the mismatch repair (MMR) genes, *MLH1*, *MSH2*, *MSH6* or *PMS2*. Colorectal cancer (CRC) and endometrial cancer are the most common cancers in this condition, with other specific neoplasms also occurring more frequently than in the general population. Evidence to include pancreatic cancer in the Lynch Syndrome cancer spectrum has been difficult to interpret and an elevated risk has not been convincing.^{11–16} Most studies examining cancer risk in Lynch Syndrome have been from families with a strong history of early-onset CRCs. This lends itself to a number of problems including the overestimation of age-specific cumulative risks of component tumors due to ascertainment bias. Additionally, many published data report on a small number of cases with incomplete testing of the full pedigree. Analyses performed exclusively on observed genotypes lack power to accurately estimate uncommon events, such as cancers with lower prevalence in a given syndrome.

The goal of our study was to quantify the risk of pancreatic cancer in families with an identified pathogenic MMR gene mutation. We have used analytic tools that correct for ascertainment and provide genotype data on subjects whose mutation status is unknown.

Methods

Selection and Description of Participants

A total of 147 families with deleterious mutations in *MLH1* (GenBank: NM_000249), *MSH2* (GenBank: NM_000251), and *MSH6* (GenBank: NM_000179) were eligible for inclusion at the start of the study in June 2008. Families were identified from hereditary CRC registries at Dana-Farber Cancer Institute (DFCI; n=80) and University of Michigan Cancer Center (UMCC; n=67). Families presenting to our cancer genetics programs are either by self-referral or health care provider referral and are enrolled on the basis of multiple cases of CRC, CRC diagnosis at a young age, or familial association of CRC with other Lynch Syndrome-associated tumors. Patients presenting for evaluation (proband) are routinely enrolled in the registries using institutional review board-approved protocols, and personal and family cancer histories and demographic data are obtained from the proband and participating relatives. Written informed consent is provided by probands for the confirmation of cancer diagnoses and deaths by review of medical records, pathology reports, or death certificates. Clinical information is updated periodically through follow-up clinic visits or telephone encounters. For this study, we selected patients with documented deleterious MMR gene mutations who were identified prior to June 2008. Analysis of MMR germline mutations in families was performed using standard molecular techniques for full gene sequencing and conducted on either the family member with CRC (or other Lynch Syndrome-associated cancer) or an “at-risk” first- or second-degree relative. Reports of pancreatic cancer were confirmed either by pathology report or death certificate.

Mutation Analysis

Mutation Analysis Technique: DNA from white blood cells was extracted and purified from the sample of blood provided by each proband, amplified by polymerase chain reaction, and directly sequenced in forward and reverse directions. For the *MLH1* gene, approximately 2300 base pairs were sequenced, comprising 19 exons and approximately 560 adjacent noncoding intronic base pairs. For the *MSH2* gene, approximately 2800 base pairs were sequenced, comprising 16 exons and approximately 470 adjacent noncoding intronic base pairs. For the *MSH6* gene, approximately 4080 base pairs were sequenced, comprising 10 exons and approximately 290 adjacent non-coding intronic base pairs. The non-coding intronic regions of *MLH1*, *MSH2* and *MSH6* that are analyzed by sequence analysis do not extend more than 20 base pairs proximal to the 5' end and 10 base pairs distal to the 3' end of each exon. The *MLH1* and *MSH2* genes are tested for large rearrangements that are not detected by sequence analysis. All coding exons of *MLH1* and *MSH2* and their respective promoters are examined for evidence of deletions and duplications by quantitative multiplexed endpoint PCR analysis.

Statistical Analysis

We used the information on diagnoses of pancreatic cancer in relatives of probands to estimate age-specific pancreatic cancer incidences in MMR mutation carriers by maximum likelihood, using a technique called modified segregation analysis. The method was implemented in MENDEL (v3.3.5).^{17,18} Information on genotype in relatives was included whenever available. However, mutation status was unknown for many family members (Table 1). Despite missing genotypes, these individuals do contribute important information to the analysis. The segregation analysis implemented by Mendel automatically handles missing genotype information by maximizing the marginal likelihood by summing over all possible genotype configurations in a family.¹⁹ Relatives were assumed to be followed from 20 years of age and to be censored at the age at diagnosis of pancreatic cancer, at the age of death, at the age at last follow-up, or at age 70 years, whichever occurred earlier. For individuals with missing age information the age was imputed based on relationship with proband, age of proband, deceased status at last follow-up (dead or alive). We also carried out a sensitivity analysis without imputing the age information to ensure that the age imputation did not artificially inflate estimates of penetrance and relative risk.

Our study included MMR carrier families ascertained through multiple individuals with CRC. Therefore the database potentially includes a greater representation of families with multiple CRC cases and mutation-positive probands than would be identified in population-based studies. Unless appropriate statistical methods are used, this type of ascertainment (a form of selection bias) can lead to overestimation of age-specific cumulative risks of pancreatic cancer. Using a conditional likelihood allows one to correct for ascertainment bias by modeling the probability of observed disease phenotypes and genotypes of the pedigrees conditional on ascertainment. This requires a model for the ascertainment probabilities, and we present results where a family was ascertained because of phenotype and genotype status of the proband and multiple affected first-degree relatives (FDRs) with CRC. This conditioning strategy was chosen based on the typical referral pattern of families to our cancer genetics clinics, emphasizing CRC as the primary reason for the referral. We also carried out additional sensitivity analyses with different alternative ascertainment mechanisms. Results are presented in the supplementary material and provided at: http://www.sph.umich.edu/bhramar/public_html/software/supplementary.doc.

The age-specific relative risks of pancreatic cancer in carriers were obtained using a proportional hazards model. We estimated the age specific log hazard ratio (HR) parameters for two age intervals <50 and ≥ 50 years. In all analyses, cancer incidences in noncarriers were assumed to follow the population cohort-specific rate as obtained through the Surveillance Epidemiology and End Results (SEER) 13 database (<http://seer.cancer.gov>). Cumulative risk (ie. penetrance) and 95% confidence intervals (CI) were calculated from the cumulative incidence. Details of the statistical methods are provided in the supplementary materials.

Results

A total of 6,342 individuals were included in the analysis: 147 probands, 1,017 FDRs, and 5,178 other relatives from the same side of the affected family with a greater than first-

degree relation (Table 1). The corresponding proportions of families carrying a mutation in one of three MMR genes were: 55/147 (37.4%) in *MLH1*, 81/147 (55.1%) in *MSH2*, and 11/147 (7.5%) in *MSH6*. The distribution of gene-specific mutations among both institutions is also shown in Table 1. Overall, a pathogenic gene mutation was detected in 302/6342 (8.2%) individuals and the number of subjects genotyped were similar across the two centers (individuals genotyped: 200 at UMCC v 232 at DFCI). 130 of the 432 individuals genotyped did not have an identified MMR gene mutation.

Description of pancreatic cancer cases

Forty-seven cases of pancreatic cancer were reported among 31 families. Eighteen families reported one case of pancreatic cancer, 10 families reported two cases of pancreatic cancer, and three families reported three cases of pancreatic cancer. Thirty-one pancreatic cancer cases were reported in families with an *MSH2* gene mutation, 13 cases were in families with an *MLH1* mutation, and 3 with an *MSH6* mutation. Of the 13 families with more than one case of pancreatic cancer, 62% (n=8) had a mutation in the *MSH2* gene compared to 30.8% (n=4) and 7.7% (n=1) in the *MLH1* and *MSH6* genes, respectively.

Forty-five percent (21/47) of pancreatic cancer cases were reported among men, who had a median age of 51.5 years at time of diagnosis (range: 19–85 years). There was no gender-related difference in age of pancreatic cancer diagnosis: 51.5 years v. 56.5 years for men and women respectively. However, 50% of men with pancreatic cancer were less than 50 years old at time of diagnosis compared to only 22% of women diagnosed at age less than 50 years (Table 2).

Of the 47 cases of pancreatic cancer, we were able to verify diagnoses with additional records in 3/23 from UMCC, and 7/24 from DFCI. In 20/23 of UMCC cases and 17/24 of DFCI cases, the diagnosis was based on family report alone. Pathology review was available for 2/23 UMCC cases, and 4/24 from DFCI, whereas other documentation (clinical report, death certificate) was obtained in 1/23 UMCC cases and 3 of the DFCI families. We were unable to verify the remaining pancreatic cancer cases due to patient confidentiality issues. We did not have permission to contact the next-of-kin of the reported cases of pancreatic cancer who were often biologically unrelated to the proband.

Estimates of Age-specific Cumulative Risk and Hazard Ratios

Among the 147 families with identified pathogenic MMR gene mutations, approximately 4% were diagnosed with pancreatic cancer by the age of 70 years. The increase in risk was more pronounced after age 40 years. The estimated decade-specific cumulative risks of pancreatic cancer for carriers of any of the three MMR as compared to the general population are presented in Table 3 and Figure 1.

There was a near 9-fold increase in risk of developing pancreatic cancer among families with pathogenic MMR gene mutations compared to the general population (HR 8.6, 95% CI: 4.7, 15.7). Table 3 also depicts the HRs for mutation carriers stratified by age (greater or less than 50 years). The estimated relative risk of pancreatic cancer was higher before age 50 years (HR 30.5 for ages 20–49 years, 95% CI: 14.2, 65.7) and then decreased with

increasing age (HR 5.1 for ages 50–70 years, 95% CI: 2.2, 11.8). The absolute cumulative risk of developing pancreatic cancer in MMR gene mutation carriers at age 50 years was 1.31% (95% CI: 0.31%, 2.32%) and 3.68% (95% CI: 1.45%, 5.88%) at age 70 years. These risks are significantly higher than the population-based cumulative age-specific incidences as reported in SEER 13, which are 0.04% and 0.52% for ages 50 and 70 years respectively.

MLH1 and *MSH2* mutation carriers had a similar increase in risk of developing pancreatic cancer compared to the general population. For carriers of mutations in *MLH1*, the overall HR was estimated at 7.5 (95% CI: 2.4, 23.0) compared to 10.9 (95% CI: 5.5, 21.9) for *MSH2* carriers. Given the small number of pancreatic cancer cases among *MSH6* carriers, risk estimates were not calculated for these mutation carriers. The hazard ratios were obtained from a proportional hazard model with a single log(HR) parameter across all ages. The two parameter model could not be fitted due to lack of data strength in each gene category.

Discussion

Among 147 families with germline MMR gene mutations, we found that the risk of developing pancreatic cancer was increased compared to the general population. The cumulative risk of developing pancreatic cancer was 3.68% by age 70 years, with cases among Lynch syndrome families occurring at an earlier age than sporadic cases.

The statistical methods used in this study afford many advantages.^{20, 21} Segregation analysis accounts for relatives who have undergone mutation analysis and those who have not. The probability of being a mutation carrier is calculated for all relatives whose mutation status is unknown and used to estimate the overall cumulative risk of cancer among family members with a germline alteration. Segregation analysis also minimizes ascertainment bias by conditioning the analysis on available phenotypic information provided for individual pedigrees. We chose *a priori*, to exclude from the risk estimate calculation all pancreatic cancers in probands and their FDRs with CRC. This yields a conservative estimate for pancreatic cancer risk and best corrects for how patients were ascertained at both centers. Data from the two registries provides a large sample of families who have undergone mutation analysis with identified MMR gene mutations.

Henry Lynch first reported pancreatic cancer in adenocarcinoma-prone families over 40 years ago²² and additional reports have described Lynch Syndrome families with pancreatic cancer.^{11–16} Pancreaticobiliary cancers are often included in the spectrum of Lynch Syndrome-associated malignancies, but data on the prevalence and risk of developing pancreatic cancer has been conflicting. A limitation of most existing data is that risk estimates were derived from families ascertained for a strong history limited to CRC.^{11,12,14} Additionally, studies that have not found an increased risk of pancreatic cancer were from homogeneous populations that have a preponderance of founder mutations and possibly a limited spectrum of tumors.¹²

However, emerging data support that pancreatic tumors are a component of Lynch Syndrome. Medullary carcinomas of the pancreas are a distinct variant of pancreatic

adenocarcinoma associated with microsatellite instability (MSI), loss of protein expression of MMR genes, family history of cancer in a FDR and germline MMR gene mutation.^{23,24} We recently reported on a known *MSH2* gene mutation carrier who developed a intraductal papillary mucinous neoplasm (IPMN) of the pancreas; the lesion showed MSI and loss of expression of *MSH2* and *MSH6* proteins.²⁵ A number of small studies suggest that MSI in sporadic pancreatic cancers offers a survival benefit similar to that observed in other Lynch Syndrome tumors. To determine if long-term survival in pancreatic cancer was attributed to defective MMR, Maple et al. ascertained its prevalence in pancreatic cancer patients who survived 3 or more years after surgery.²⁶ The data suggest that immunohistochemistry (IHC) for pancreatic cancer is both sensitive and specific for the MSI phenotype. A study of 130 families with MMR mutations reported 22 pancreatic cancers with many early-onset cases;²⁷ however, lifetime risks of developing pancreatic cancer were not calculated.

Our study has several limitations. As in most Lynch Syndrome registries our families were ascertained through an affected proband with a classic Lynch Syndrome tumor, notably CRC. To reduce ascertainment bias, our analysis of pancreatic cancer cases specifically excluded these probands, as well as any FDRs who also had CRC. We also relied in large part on the probands' report of pancreatic cancers in their families which may be inaccurate.²⁸⁻³¹ The majority of pancreatic cancer patients were deceased and not available for mutation analysis. Therefore it is not possible to accurately determine those cases that may have been sporadic and whose inclusion in the analysis may have overestimated risk. Recall bias may also occur when family members without a personal history of cancer are under-reported by the proband. Although family structure was completely enumerated for a three-generation pedigree at both sites by certified genetic counselors during the retrospective review and construction of the pedigrees, it is possible the selective expansion of branches of the family with cancer might lead to this type of recall bias. In our study, this bias may be present but is likely minimal for two reasons. First, pedigree structures are routinely verified by multiple relatives who undergo genetic evaluation, increasing the chances of completely enumerating all affected and unaffected members. Second, since the majority of relatives with pancreatic cancer were identified in FDRs and second-degree relatives, the standardized construction of three-generation pedigrees is likely to reduce the magnitude of this potential recall bias. Another potential limitation reflects our choice to impute missing ages. However, this was done conservatively and is unlikely to inflate risk as shown in the supplemental sensitivity analyses. Despite these potential limitations, the elevated risk of pancreatic cancer was similar at both study sites, lending credence to the results.

Our findings have implications regarding the care of patients and families with a known MMR gene mutation. Information on cancer risk is important in planning cancer prevention and determining the efficacy of proposed prevention strategies.³²⁻³⁸ Several screening trials aimed at identifying early pancreatic neoplasia through radiographic and endoscopic imaging are currently underway in patients with genetic syndromes associated with high incidence of pancreatic cancer.^{32,33} MMR gene mutation carriers with a family history of pancreatic cancer may need to be screened in a similar manner to these individuals. Ongoing screening programs will provide information on the risks and benefits of early detection of

pancreatic neoplasms, and allow further study on the spectrum of pancreatic lesions in Lynch Syndrome families.

In summary, families with an identified pathogenic MMR gene mutation have an increased lifetime risk of developing pancreatic compared to the general U.S. population. Further studies are necessary in individuals with Lynch Syndrome to further define the risk of pre-malignant and malignant pancreatic neoplasms and the potential benefits and limitations of surveillance. Pancreatic cancer is a clinically relevant component of Lynch Syndrome and quantifying this risk for gene carriers should be incorporated into clinical management.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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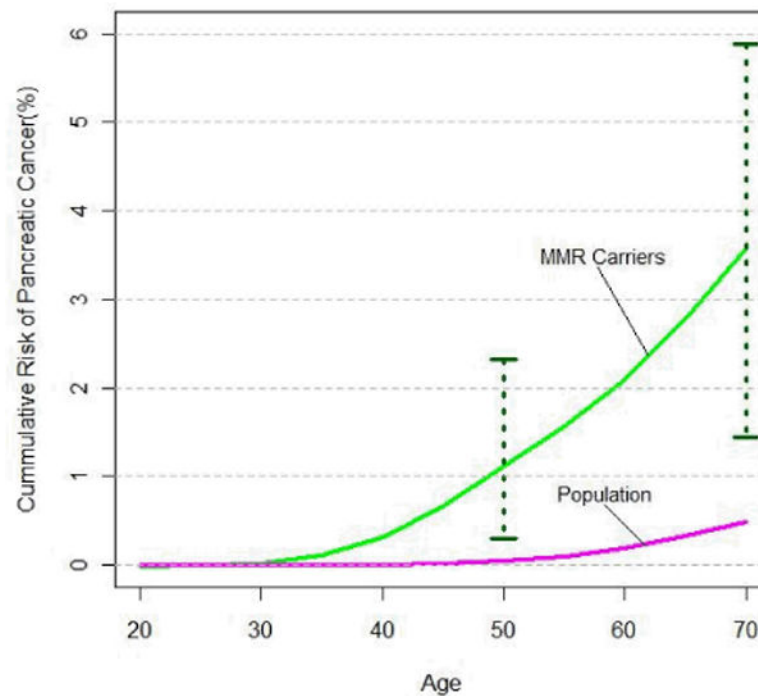


Figure 1. Age-specific cumulative risk of pancreatic cancer in families with pathogenic mutations in *MLH1*, *MSH2* or *MSH6* genes

MMR Carriers=families with mismatch repair gene carriers (*MLH1*, *MSH2* or *MSH6*)

The penetrance curves in Figure 1 were generated by plotting the age-specific cumulative risks of pancreatic cancer (as presented in Table 3) for a set of discrete ages from 20 to 70 years at five-year intervals and then applying a smoothing spline function. The 95% Confidence Intervals corresponding to the age-specific cumulative risk of pancreatic cancer for MMR carrier families were also plotted for ages 50 to 70 years. Population estimates of age-specific cumulative risks of pancreatic cancer are given by pancreatic cancer incidence rates reported in Surveillance Epidemiology and End Results (SEER) 13, from 1992–2005 (<http://seer.cancer.gov>).

Table 1

Characteristics of study population and distribution of mutations by gene

	UMCC	DFCI	Total
Number of probands*	67	80	147
Number of FDR	459	558	1017
Total number of individuals	2660	3682	6342
Number of males	1395	1900	3295
Number of females	1265	1782	3047
(Male: Female)	1:1	1:1	1:1
Number of subjects genotyped	200	232	432
Number of mutation positive subjects	144	158	302
Mutated Gene**			
<i>MLH1</i>	18 (26.9)	37 (46.3)	55 (37.4)
<i>MSH2</i>	42 (62.7)	39 (48.8)	81 (55.1)
<i>MSH6</i>	7 (10.4)	4 (5.0)	11 (7.5)

UMCC=University of Michigan Cancer Center, DFCI=Dana-Farber Cancer Institute, MMR= mismatch repair; FDR=first-degree relative.

* proband=index patient per family presenting for genetic evaluation

** n (% of total families with MMR gene mutations)

Table 2

Age of diagnosis for pancreatic cancer cases stratified by gender.

Center	Total		Men				Women			
	# Pancreatic Cancer cases		# of cases (# cases with known ages at diagnosis)	Proportion of cancers diagnosed < age 50* (%)	Median age at diagnosis (range)*	# of cases (# cases with known ages at diagnosis)	Proportion of cancers diagnosed < age 50* (%)	Median age at diagnosis (range)*	Proportion of cancers diagnosed < age 50* (%)	Median age at diagnosis (range)*
UMCC	23		13 (8)	6/8 (75.0)	42.5 (19–75)	10 (6)	1/6 (16.7)	55 (36–73)		
DFCI	24		8 (8)	2/8 (25.0)	65 (40–85)	16 (12)	3/12 (25.0)	62 (27–79)		
Combined	47		21 (16)	8/8 (50.0)	51.5 (19–85)	26 (18)	4/18 (22.2)	56.5 (27–79)		

UMCC=University of Michigan Cancer Center, DFCI=Dana-Farber Cancer Institute

* Calculated from cases with known ages at diagnosis of pancreatic cancer

Table 3

Age-specific cumulative risk of pancreatic cancer*

Age	Cumulative Risk: Population** %	Cumulative Risk: Families with MMR gene mutations % (95% CI)	Hazard Ratio (95% CI)
20	0	0	30.5 (14.2, 65.7) <i>ages 20-49</i>
30	0	0.03	
40	0.01	0.23	
50	0.04	1.31 (0.31, 2.32)	
60	0.18	1.98	5.1 (2.2, 11.8) <i>ages 50-70</i>
70	0.52	3.68 (1.45, 5.88)	

MMR= Mismatch repair, CI= confidence interval

* Two age-specific hazard ratios in proportional hazards model (<50 years, ≥ 50 years), corrected for ascertainment by conditioning on genotype and phenotype of proband and phenotype of all colorectal cancer affected first-degree relatives

** Surveillance Epidemiology and End Results 1992-2005 (<http://seer.cancer.gov>)