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Effects of Single Cell Protein Replacing Fish Meal in Diet on Growth Performance, Nutrient Digestibility and Intestinal Morphology in Weaned Pigs

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ABSTRACT: Three experiments were conducted to evaluate the ME value, standardized ileal digestibility (SID) of amino acids (AA) of fish meal, and the effects of single cell protein (Prosin and Protide) replacing fish meal in diet on growth performance, nutrient digestibility and intestinal morphology in weaned piglets. In Exp. 1, twenty-four barrows with initial BW of 30.8±2.6 kg were allotted to one of four dietary treatments. Diet 1 contained corn as the only energy source. The other three diets replaced 20% of the corn in diet 1 with one of the three protein feeds (fish meal, Prosin and Protide), and the DE and ME contents were determined by difference. In Exp. 2, eight barrows (initial BW of 25.6±3.2 kg) were fitted with ileal T-cannulas and allotted to a replicated 4×4 Latin square design. Three cornstarch-based diets were formulated using each of the protein feeds as the sole source of AA. A nitrogen-free diet was also formulated to measure endogenous losses of AA. In Exp. 3, one hundred and eighty piglets (initial BW of 7.95±1.59 kg) weaned at 28±2 d were blocked by weight and assigned to one of five treatments for a 28-d growth performance study, each treatment was fed to six pens with six pigs (three barrows and three gilts) per pen. The five treatments consisted of the control group (CON), which was a corn-soybean meal diet containing 5% fish meal, and the other four treatments, which replaced a set amount of fish meal with either Prosin (2.5% or 5%) or Protide (2.5% or 5%). The diets were formulated to provide same nutrient levels. The results showed that on a DM basis, both of the DE and ME contents were lower in Prosin and Protide than that of fish meal (p<0.05). The SID of CP and all essential AA were greater in fish meal than in Prosin and Protide (p<0.05). The pigs fed CON diet had greater weight gain and lower feed conversion rate (FCR) than pigs fed 5% Prosin and 5% Protide diets (p<0.05). The digestibility of CP was greater in pigs fed CON, 2.5% Prosin and 2.5% Protide diets than the pigs fed 5% Prosin and 5% Protide diets (p<0.05). Villus height in jejunum and ileum, and villus height to crypt depth ratio in the jejunum were higher (p<0.05) in pigs fed CON, 2.5% Prosin and 2.5% Protide diets compared with the 5% Prosin and 5% Protide diets. Pigs fed CON diet had greater villus height to crypt depth ratio in the ileum than the pigs fed 5% Prosin and 5% Protide diets (p<0.05). In conclusion, although Prosin and Protide contained lower ME content and SID of AA than fish meal, Prosin and Protide replacing 50% of fish meal in diet with identical nutrient levels could obtain similar performance, nutrient digestibility and intestinal morphology in weaned pigs. (Key Words: Single-cell Protein, Fish Meal, Performance, Nutrient Digestibility, Intestinal Morphology, Weaned Pigs)

INTRODUCTION

Single-cell protein (SCP) refers to the dried cells of microorganisms such as yeasts, bacteria, fungi and microalgae, grown in large-scale culture systems for use as protein sources in human food or animal feed (Zepka et al., 2010; Wang et al., 2013). Compared to fish meal, most sources of SCP (from bacteria and yeast) have similar lysine (Lys), methionine (Met) and cysteine (Cys) content, and a

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higher proportion of tryptophan (Trp) and threonine (Thr) (Skrede et al., 1998). There are not only proteins, but also contributes free amino acids, lipids, carbohydrates, vitamins and minerals in SCP (Anupama and Ravindra, 2000).

Production of SCP using waste materials as substrate, provides an economically feasible protein source that is useful in animal feed and products for human consumption as it often meets dietary requirements for protein (Kuhad et al., 1997). Algae, fungi and bacteria are the chief sources of microbial protein that can be utilized as SCP (Anupama and Ravindra, 2000). In general, high production rates and protein yields as well as ease of production control make

SCP more attractive as a protein source compared with conventional plant and animal origins (Paraskevopoulou et al., 2003).

The SCP can be a protein source that offers a potential alternative to fish meal, meat and bone meal, soybean meal, and other protein sources in animal nutrition (Hellwing et al., 2007b). Certain amounts of traditional protein sources can be substituted by SCP without impairment to the performance of pigs (Braude et al., 1977; Hanssen and Farstad, 1980). Previous studies showed that up to 40% 20% (chickens), and 50% (pigs) of dietary N could be replaced by SCP without affecting nitrogen retention, heat production, and energy retention (Hellwing et al., 2005; 2006; 2007a).

Prosin and Protide are two kinds of SCP, by-products of the production of lysine and nucleotide fermented from cornstarch by *Corynebacterium glutamicum*. As new sources of protein, information regarding their nutritional value and effects on pig performance are limited. Therefore, the objectives of the current experiment were to determine the ME content as well as standardized ileal digestibility (SID) of AA in Prosin and Protide, based on the ME and SID of AA values, to evaluate the effects of Prosin and Protide replacing 50% or 100% of fish meal in diet with identical nutrient levels on growth performance, nutrient digestibility and intestinal morphology in weaned piglets.

MATERIALS AND METHODS

The experimental protocols were approved by the Institutional Animal Care and Use Committee (IACUC) of China Agricultural University (Beijing, China). The fish meal used in this study was commercial supplied (Pesquera Diamante S. A., Peru), and Prosin and Protide were supplied by CJ Co. Ltd., (Liaocheng, China). Briefly, after pure L-Lysine or nucleotide was removed from the *Corynebacterium glutamicum* fermentation medium, the live bacteria was killed in high temperature, then the left medium was dried to produce Prosin and Protide. The analyzed chemical composition of Prosin and Protide is shown in Table 1.

Animals and experimental design

Experiment 1: Twenty four barrows (Duroc×Landrace× Large White) with initial BW of 30.8±2.6 kg were used to determine the DE and ME contents of fish meal, Prosin and Protide. Pigs were allotted to one of four experimental treatments, each treatment was fed to six pens with one pig per pen. The basal diet containing corn as the only energy source (96.31% corn and 3.69% minerals and vitamins) and this diet was used to determine the DE and ME content of corn. In the remaining three diets, 20% of the corn was substituted with fish meal, Prosin and Protide respectively

Table 1. Analyzed nutrient composition of fish meal and single-cell proteins (Prosin and Protide) (as-fed basis)

Item (%)	Fish meal ¹	Prosin ²	Protide ²
DM	88.83	94.09	95.99
Gross energy (kcal/kg)	4,710	4,784	5,164
CP	66.03	70.03	57.53
EE	1.72	8.23	7.44
Ca	3.42	0.04	0.01
P	2.42	0.39	1.63
Essential AA			
Arginine	3.49	3.28	2.61
Histidine	2.34	1.26	0.96
Isoleucine	2.69	2.45	2.35
Leucine	4.54	4.23	4.28
Lysine	4.97	8.23	1.97
Methionine	1.63	0.98	1.47
Phenylalanine	2.50	2.15	2.49
Threonine	2.62	2.86	2.61
Tryptophan	0.65	0.58	0.73
Valine	2.93	3.34	3.55
Nonessential AA			
Alanine	3.83	5.31	5.35
Asparagine	5.63	5.44	5.57
Cystine	0.84	0.32	0.26
Glutamine	6.92	8.05	8.35
Glycine	3.82	2.61	2.81
Proline	2.54	1.87	1.83
Serine	2.16	2.18	2.37
Tyrosine	1.88	1.40	1.13

¹ Fish meal (Commercial supply).

at the expense of corn, the energy values were determined by differences on the assumption that their contributions were additive (Woodworth et al., 2001). The diets were shown in Table 2.

Pigs were fed with two equal meals twice daily at 8:00 h and 17:00 h and had free access to water throughout the trial. Daily feed allowance was equivalent to 4% of body weight and was fed in mash form. Each feeding period consisted of 5 d for diet adaptation, followed by 5 d of total collection of feces and urine. In the morning meal on d 6 and 9, 1 g of chromic oxide was mixed into the meal and used as a fecal marker. Feces collection from each pig was initiated with the first appearance of the marker in the feces after d 6 and collection ceased when the marker first appeared in the feces after d 9 (Petersen and Stein, 2006). Feces were collected twice daily, weighed, composited for each pig, and stored at -20°C; Urine was collected in a bucket placed under the metabolic crate containing 50 mL of 6 N HCl as a preservative, urine collection was stored at -20°C until subsequent analysis (Adeola and Bajjalieh, 1997).

² Prosin and Protide (CJ Co. Ltd., Liaocheng, China).

Table 2. Ingredient composition of experimental diets containing fish meal and single-cell proteins (Prosin and Protide) (as-fed basis), Exp. 1

Ingredient (%)	Corn	Fish meal ¹	Prosin ²	Protide ²	
Corn	96.31	76.31	76.31	76.31	
Fish meal (66.03% CP)	-	20.00	-	-	
Prosin (70.03% CP)	-	-	20.00	-	
Protide (57.53% CP)	-	-	-	20.00	
Limestone	0.66	0.66	0.66	0.66	
Dicalcium phosphate	1.68	1.68	1.68	1.68	
Salt	0.35	0.35	0.35	0.35	
Vitamin-mineral premix ³	1.00	1.00	1.00	1.00	

¹Fish meal (Commercial supply).

Experiment 2: Eight barrows (Duroc×Landrace×Large White) with a BW of 25.6±3.2 kg were used to compare the SID of AA among fish meal, Prosin and Protide. Each pig was surgically fitted with a T-cannula at the distal ileum according to the method described by Stein et al. (1998). After surgery, pigs were transferred to individual pens (1.8×0.8 m) in a temperature-controlled room (22°C) where they were allowed to recover for 10 d. A standard diet and water were provided ad libitum during this time (Cervantes-Pahm and Stein, 2010). Pigs were then randomly assigned to a replicated 4×4 Latin square design. Three diets were formulated to contain each protein ingredient as the only protein and AA source. A N-free diet was also formulated to determine the basal endogenous losses of CP and AA. The three protein-containing diets were formulated to provide the same amount of CP (12%) in each diet. 0.30% chromic oxide were added to all diets as an indigestible marker (Table 3).

Each feeding period consisted of a 5-d adaptation period followed by 2 d for collection of ileal digesta. The BW of each pig was recorded at the beginning and the end of each period. Pigs were fed the diets at three times their maintenance energy requirement, based on their BW at the beginning of each period. Feed allowance was divided into two equal meals and fed at 8:00 and 17:00 each day. Ileal digesta were collected continuously for 12 h from 8:00 to 20:00 h on d 6 and 7 (Stein et al., 1998). A plastic bag was attached to the cannula using a cable tie to collect the digesta. The bags were collected and replaced using a new one at least once every 20 min. Ileal digesta were stored on ice during the 12-h collection period. After each day's collection, digesta were frozen at -20°C to limit microbial activity and N loss until the end of collection. At the end of

Table 3. Ingredient composition of experimental diets containing fish meal and single-cell proteins (Prosin and Protide) (as-fed basis), Exp. 2

Ingredient (%)	Nitrogen- Fish free meal ¹		Prosin ²	Protide ²	
Cornstarch	70.04	67.90	66.46	63.28	
Sucrose	18.00	10.00	10.00	10.00	
Solka floc ³	4.00	-	-	-	
Fish meal (66.03% CP)	-	18.11	-	-	
Prosin (70.03% CP)	-	-	17.13	-	
Protide (57.53% CP)	-	-	-	20.86	
Soybean oil	3.00	2.00	2.00	2.00	
Limestone	0.38	-	0.56	1.37	
Dicalcium phosphate	3.03	0.84	2.70	1.34	
Salt	0.35	0.35	0.35	0.35	
Magnesium oxide	0.10	-	-	-	
Potassium carbonate	0.30	-	-	-	
Chromic oxide	0.30	0.30	0.30	0.30	
Vitamin-mineral premix ⁴	0.50	0.50	0.50	0.50	

¹ Fish meal (Commercial supply).

each collection day, ileal samples were thawed, pooled by pig, and a subsample was taken for lyophilization (Karr-Lilienthal et al., 2004).

Experiment 3: A total of 180 crossbred pigs (Duroc× Landrace×Large White; initial BW of 7.95±1.59 kg) weaned at 28±2 d of age were allocated to one of five dietary treatments on the basis of BW and gender in a randomized complete block design. Each treatment was fed to six pens with six pigs (three barrows and three gilts) per pen. The five treatments were as follows: the control group (CON) was a corn-soybean meal diet contained 5% fish meal; the other four treatments replaced fish meal with Prosin (2.5% or 5%) or Protide (2.5% or 5%), respectively. The diets were formulated to supply identical amounts of ME and SID lysine, methionine, threonine and tryptophan. The SID lysine and ME contents of fish meal, Prosin and Protide and the ME content of corn were obtained according to the data obtained in Exp. 1 and Exp. 2. The ME contents of whey powder (3,300 kcal/kg), and soybean oil (8,400 kcal/kg) were obtained from NRC (1998). The SID lysine contents of corn and whey powder were determined previously using pigs surgically equipped with a T-cannula in the distal ileum by our laboratory (data not published). The diets were shown in Table 4. The pigs were housed in 2×3 m raised pens equipped with a mesh floor.

² Prosin and Protide (CJ Co. Ltd., Liaocheng, China).

 $^{^3}$ Premix supplied per kg diet: vitamin A, 12,000 IU; vitamin D₃, 1,500 IU; vitamin E, 44.1 IU; menadione, 4.0 mg; riboflavin, 5.22 mg; pantothenic acid, 20.0 mg; niacin, 26.0 mg; vitamin B₁₂, 0.01 mg; manganese, 35.0 mg; iron, 95.0 mg; zinc, 90.0 mg; copper, 16.5 mg; iodine, 0.30 mg; selenium, 0.30 mg.

² Prosin and Protide (CJ Co. Ltd., Liaocheng, China).

³ Made by Chemical Reagents Company (Beijing, China).

 $^{^4}$ Premix supplied per kg diet: vitamin A, 11,000 IU; vitamin D₃, 1,500 IU; vitamin E, 44.1 IU; menadione, 4.0 mg; riboflavin, 5.22 mg; pantothenic acid, 20.0 mg; niacin, 26.0 mg; vitamin B₁₂, 0.01 mg; manganese, 35.0 mg; iron, 100.0 mg; zinc, 90.0 mg; copper, 16.5 mg; iodine, 0.30 mg; selenium, 0.30 mg.

Table 4. Ingredient composition of experimental diets containing fish meal and single-cell proteins (Prosin and Protide) (as-fed basis), Exp. 3

Ingredients (%)	CON ¹	2.5% Prosin ²	2.5% Prosin ²	2.5% Protide ²	5% Protide ²	
Corn	61.54	60.88	60.23	60.31	59.06	
Soybean meal	21.59	21.61	21.64	22.10	22.64	
Fish meal (66.03% CP)	5.00	2.50	-	2.50	-	
Prosin (70.03% CP)	-	2.50	5.00	-	-	
Protide (57.53% CP)	-	-	-	2.50	5.00	
Whey powder	6.00	6.00	6.00	6.00	6.00	
Soybean oil	2.14	2.47	2.79	2.52	2.88	
Dicalcium phosphate	1.26	1.56	1.88	1.38	1.48	
Limestone	0.69	0.76	0.80	0.86	1.03	
Chromic oxide	0.25	0.25	0.25	0.25	0.25	
Salt	0.35	0.35	0.35	0.35	0.35	
L-Lys-HCl (78%)	0.42	0.35	0.28	0.48	0.56	
MHA^3	0.04	0.05	0.06	0.03	0.04	
L-threonine	0.17	0.17	0.17	0.17	0.17	
L-tryptophan	0.05	0.05	0.05	0.05	0.04	
Premix ⁴	0.50	0.50	0.50	0.50	0.50	
Nutrient levels, calculated (%)						
ME (kcal/kg)	3,265	3,265	3,265	3,265	3,265	
CP, analyzed	19.46	19.48	19.51	19.48	19.46	
Ca, analyzed	0.78	0.77	0.79	0.78	0.76	
P, analyzed	0.62	0.65	0.62	0.64	0.65	
SID lysine	1.20	1.20	1.20	1.20	1.20	
SID methionine	0.32	0.32	0.32	0.32	0.32	
SID threonine	0.74	0.74	0.74	0.74	0.74	
SID tryptophan	0.22	0.22	0.22	0.22	0.22	

¹ CON = Fish meal (Commercial supply). ² Prosin and Protide (CJ Co. Ltd., Liaocheng, China).

The temperature was maintained between 26 and 30°C. All pigs had free access to feed and water throughout the 4-wk feeding trial.

Pigs and feeders were weighed at the beginning and end of the experiment to determine weight gain, feed intake and feed conversion rate (FCR). From d 26 to 28 of the experiment, approximately 50 g of feces were collected daily from each pen and the samples were stored at -20°C. The three day collection of feces was thawed, pooled by pen and then oven dried at 60°C for 72 h (Li et al., 2012). All samples were ground to pass through a 1.0 mm screen before analysis. On d 28, two pigs from each pen were electronically stunned and killed, and tissue samples from the small intestine were collected immediately. The middle sections of the duodenum, jejunum, and ileum were aseptically isolated, flushed with a 0.9% salt solution, fixed with 10% formaldehyde-phosphate buffer, and kept at 4°C for microscopic assessment of mucosal morphology (Liu et al., 2010).

Analytical assays

Analyses for DM, CP, Ca, and total phosphorus in diets and feces were conducted according to the methods of AOAC (2000). Gross energy was determined using an automatic adiabatic oxygen bomb calorimeter (Parr 1281 Automatic Energy Analyzer, Moline, IL). The chromium content in the diets and digesta was measured using an Atomic Absorption Spectrophotometer (Hitachi Z-5000 Automatic Absorption Spectrophotometer, Tokyo, Japan) according to the procedure of Williams et al. (1962).

The AA content in the diets and digesta were assayed using ion-exchange chromatography with an Automatic Amino Acid Analyzer (L-8800 Hitachi Automatic Amino Acid Analyzer, Tokyo, Japan) after hydrolyzing with 6 mol/L HCl at 110°C for 24 h. Cystine was determined as cysteic acid and methinonine as methionine sulfone after preoxidation with performic acid and pre-column derivation using phenylisothiocyanate (L-8800 Hitachi Automatic Amino Acid Analyzer, Tokyo, Japan). Tryptophan was determined after hydrolyzing with 4 mol/L NaOH at 110°C

³ DL-methionine hydroxy analogue (84%) provided by Novus International, St. Louis, MO.

⁴ Premix supplied per kg diet: vitamin A, 11,500 IU; vitamin D₃, 1,500 IU; vitamin E, 44.1 IU; menadione, 4.0 mg; riboflavin, 5.22 mg; pantothenic acid, 20.0 mg; niacin, 26.0 mg; vitamin B₁₂, 0.01 mg; manganese, 35.0 mg; iron, 110.0 mg; zinc, 95.0 mg; copper, 16 mg; iodine, 0.30 mg; selenium, 0.30.

Table 5. The DE and ME contents in fish meal and single-cell proteins (Prosin and Protide) (as-fed basis), Exp. 11

Item	Fish meal ²	Prosin ³	Protide ³	SEM	p-value
DM basis					
DE in ingredients (kcal/kg)	$3,603^{a}$	3,234 ^b	3,156 ^b	35.34	< 0.001
ME in ingredients (kcal/kg)	3,143 ^a	2,878 ^b	2,777 ^b	38.16	< 0.001
Fed basis					
DE in ingredient (kcal/kg)	3,243 ^a	3,047 ^b	3,029 ^b	39.63	0.008
ME in ingredient (kcal/kg)	$2,829^{a}$	$2,712^{ab}$	2,665 ^b	42.77	0.031

¹Data are the means of eight pigs (initial BW of 30.8±2.6 kg). ²Fish meal (Commercial supply). ³Prosin and Protide (CJ Co. Ltd., Liaocheng, China).

for 22 h using phenylisothiocyanate (Model 76337, Agilent Technologics, Waldbronn, Germany).

Histological measurements were conducted according to the procedures described by Shen et al. (2009). Briefly, formalin-fixed intestinal samples were prepared using paraffin embedding techniques. Samples were sectioned at a 6 μ m thickness and stained with hematoxylin and eosin. Villus height and crypt depth were then measured at 40× magnification with a microscope (CK40, Olympus Optical Company, Shenzhen, China). At least 10 well-oriented and intact villi were selected from each pig to measure crypt depth and villus height.

Statistical analysis

All data were analyzed using SAS (SAS inst. Inc., Cary, NC). In Exp. 1 and Exp. 2, data were analyzed using the Proc-Mixed procedure of SAS with each replicate as the experimental unit. The statistical model for the DE, ME, and SID of AA values had treatment as a fixed effect, period and pig as random effects. In Exp. 3, all data were analyzed as a randomized complete block design with pen as the experimental unit using the GLM procedure. Results were expressed as least squares means and standard error of the mean (SEM). Statistical significance was taken at p<0.05.

RESULTS

Chemical characteristics

The DM content in Prosin and Protide was higher than that of fish meal (Table 1). Prosin had higher CP than fish meal, but CP was lowest in Protide. Fish meal contained higher Ca and P. For essential AA, Prosin (8.23%) contained higher lysine content than fish meal (4.97%) and Protide (1.97%), but fish meal (1.63%) and Protide (1.47%) contained higher methionine than Prosin (0.98%); The concentrations of other essential AA were similar among different protein sources, except for histidine, which was higher in fish meal (2.34%) than those of Prosin (1.26%) and Protide (0.96%).

Experiment 1: On DM basis, both of the DE and ME contents were lower in Prosin (3,234 and 2,878 kcal/kg) and Protide (3,156 and 2,777 kcal/kg) than that of fish meal

(3,603 and 3,143 kcal/kg) (p<0.05; Table 5). On a fed basis, the DE content for fish meal (3,243 kcal/kg) was greater than those of Prosin (3,047 kcal/kg) and Protide (3,029 kcal/kg) (p<0.05); The fish meal (2,829 kcal/kg) had higher ME content than Protide (2,665 kcal/kg) (p<0.05), but the ME content in Prosin (2,712 kcal/kg) did not differ from fish meal or Protide.

Experiment 2: The SID of CP (85.87%) was greater in fish meal than in Prosin (80.25%) and Protide (81.03%) (p<0.05) (Table 6). For essential AA, the SID for all AA in fish meal were greater than in Prosin and Protide (p<0.05), but no significant differences were found between Prosin

Table 6. SID of CP and AA in fish meal and single-cell proteins (Prosin and Protide), Exp.2¹

Item	Fish	Prosin ³	Protide ³	CEM	m volue
Item	$meal^2$	PIOSIII	Protide	SEM	p-value
CP (%)	85.87 ^a	80.25 ^b	81.03 ^b	1.52	0.031
Essential AA (%))				
Arginine	93.50^{a}	85.97 ^b	87.24 ^b	0.91	0.019
Histidine	84.86^{a}	78.07^{b}	78.31 ^b	0.51	0.023
Isoleucine	88.36 ^a	80.65^{b}	81.27^{b}	1.14	0.030
Leucine	88.92^{a}	79.09^{b}	75.86^{b}	1.55	0.019
Lysine	87.37 ^a	79.04^{b}	81.59 ^b	0.60	0.005
Methionine	89.99 ^a	84.24^{b}	83.35 ^b	0.95	0.030
Phenylalanine	89.65 ^a	83.95 ^b	81.52 ^b	0.98	0.021
Threonine	85.78 ^a	78.73^{b}	78.14^{b}	0.84	0.031
Tryptophan	87.38 ^a	81.86^{b}	82.99^{b}	0.35	0.003
Valine	86.88 ^a	79.31 ^b	78.78^{b}	0.82	0.010
Nonessential AA	(%)				
Alanine	88.56 ^a	82.00^{b}	77.99 ^c	0.66	
Asparagine	80.91 ^a	73.94 ^b	76.11 ^b	0.68	0.004
Cystine	77.68	72.89	73.52	1.39	0.012
Glutamine	82.43 ^a	77.70^{b}	76.22^{b}	0.68	0.163
Glycine	84.00^{a}	78.06^{b}	76.93^{b}	1.05	0.016
Proline	113.59 ^c	140.20^{a}	124.58 ^b	1.09	0.033
Serine	82.53 ^a	78.09^{b}	69.81 ^c	0.63	0.001
Tyrosine	87.74 ^a	76.96 ^b	78.72 ^b	0.98	0.002

¹ Data are the means of eight pigs (initial BW of 25.6±3.2 kg) used in a replicated 4×4 Latin square design.

 $^{^{\}rm a,\,b}$ Means in the same row with different superscripts are significantly different (p<0.05).

² Fish meal (Commercial supply).

³ Prosin and Protide (CJ Co. Ltd., Liaocheng, China).

a,b,c Means in the same row with different superscripts are significantly different (p<0.05).

Table 7. Effects of single-cell proteins (Prosin and Protide) replacing fish meal in diet on growth performance of weaned piglets, Exp. 3¹

Item —	Fish meal (%)	Prosin	3(%)	Protid	e ³ (%)	SEM	n voluo
	5	2.5	5	2.5	5	SEM	p-value
ADG (g/d)	370 ^a	354 ^{ab}	339 ^{bc}	363 ^{ab}	343 ^{bc}	9.12	0.019
ADFI (g/d)	528	518	538	539	531	12.63	0.876
FCR	1.44 ^c	1.47 ^{bc}	1.59 ^{ab}	1.49 ^{bc}	1.56 ^{ab}	0.05	0.018

A total of 180 pigs (initial BW of 7.95±1.59 kg and 28±2 d of age) were allotted to provide six pigs per pen and six pens per treatment.

Table 8. Effects of single-cell proteins (Prosin and Protide) replacing fish meal in diet on nutrient digestibility of weaned piglets, Exp. 3¹

Item (%)	Fish meal ² (%)	Prosir	Prosin ³ (%)		Protide ³ (%)		1
item (%)	5	2.5 5		2.5	5	SEM	p-value
DM	86.51	85.30	84.60	85.95	85.54	0.60	0.356
Gross energy	86.79	86.77	87.00	86.16	86.67	0.64	0.817
CP	85.15 ^a	83.74 ^a	81.28 ^b	84.17 ^a	81.17 ^b	0.73	< 0.001
P	59.89	59.70	58.72	59.99	59.37	1.28	0.944
Ca	67.11	67.88	66.98	67.36	67.24	1.14	0.994

A total of 180 pigs (initial BW of 7.95±1.59 kg and 28±2 d of age) were allotted to provide six pigs per pen and six pens per treatment.

and Protide. For nonessential AA, the SID of most AA was greater in fish meal than in Prosin and Protide, except for cystine and proline (p<0.05). Prosin and Protide had greater SID of proline than fish meal (p<0.05), but no significant differences were found in cystine among fish meal, Prosin and Protide.

Experiment 3: The pigs fed CON diet contained 5% fish meal had greater weight gain than pigs fed 5% Prosin and 5% Protide diets (p<0.05) (Table 7), but no differences were observed in weight gain among pigs fed CON, 2.5% Prosin or 2.5% Protide diets. Pigs fed the CON diet had lower FCR than pigs fed 5% Prosin and Protide diets (p<0.05). The

ADFI was not different among treatments.

The digestibility of CP was greater in pigs fed CON, 2.5% Prosin and 2.5% Protide diets than the pigs fed 5% Prosin and 5% Protide diets (p<0.05) (Table 8), but no differences were observed among pigs fed CON, 2.5% Prosin or 2.5% Protide diets. The digestibility of DM, gross energy, Ca and P was not affected by dietary protein sources.

No dietary effects were found in crypt depth of the duodenum, jejunum and ileum among treatments (Table 9). Villus height in jejunum and ileum, and villus height to crypt depth ratio in the jejunum were higher (p<0.05) in pigs fed the CON, 2.5% Prosin and 2.5% Protide diets

Table 9. Effects of single-cell proteins (Prosin and Protide) replacing fish meal in diet on intestinal morphology of weaned piglets, Exp. 3¹

Itam	Fish meal ² (%)	Prosir	n ³ (%)	Protid	e ³ (%)	CEM	
Item	5	2.5	5	2.5	5	SEM	p-value
Duodenum							
Villus height (μm)	424	425	429	412	426	7.02	0.569
Crypt depth (µm)	204	196	200	203	197	4.20	0.533
Villus height to crypt depth ratio	2.08	2.17	2.15	2.03	2.17	0.04	0.236
Jejunum							
Villus height (μm)	359 ^a	352 ^a	323 ^b	348 ^a	321 ^b	7.16	0.001
Crypt depth (µm)	183	182	180	183	187	3.13	0.906
Villus height to crypt depth ratio	1.97 ^a	1.95 ^a	1.77 ^b	1.91 ^a	1.72 ^b	0.05	0.002
Ileum							
Villus height (μm)	486 ^a	478 ^a	415 ^b	477 ^a	420^{b}	7.59	0.001
Crypt depth (µm)	190	205	200	203	193	7.14	0.746
Villus height to crypt depth ratio	2.56^{a}	2.34^{ab}	2.09^{c}	2.39^{ab}	2.18bc	0.08	0.001

A total of 180 pigs (initial BW of 7.95±1.59 kg and 28±2 d of age) were allotted to provide six pigs per pen and six pens per treatment.

² Fish meal (Commercial supply). ³ Prosin and Protide (CJ Co. Ltd., Liaocheng, China).

^{a,b,c} Means in the same row with different superscripts are significantly different (p<0.05).

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^{a,b,c}Means in the same row with different superscripts are significantly different (p<0.05).

compared with the 5% Prosin and 5% Protide diets. Pigs fed CON diet had greater villus height to crypt depth ratio in the ileum than the pigs fed 5% Prosin and 5% Protide diets (p<0.05), but villus height to crypt depth ration of the CON group was not different from the pigs fed 2.5% Prosin and 2.5% Protide diets.

DISCUSSION

The concentrations of CP and AA in fish meal were similar to those reported by NRC (1998) and Kim and Easter (2001). The CP and AA profile of Prosin was similar to that shown earlier for SCP (Aas et al., 2006), but Prosin contained higher level of lysine, alanine and glutamine, and lower level of methionine and tyrosine. Additionally, the Protide had similar AA profile with Prosin, but it contained lower level of CP, lysine and histidine, and higher level of methionine and tryptophan than that of Prosin. The differences between Prosin and Protide may mainly be because the Prosin and Protide are byproducts of lysine and nucleotide production, respectively, and the SCP produced by different processing technology may cause different nutritional properties which may also influence its utilization by animals (Hellwing et al., 2007a, b). The results of nutritional composition also showed that most essential AA in Prosin and Protide was similar with fish meal, except for lysine and methionine, with Prosin having higher lysine content than fish meal and Protide, but fish meal and Protide contained higher methionine than Prosin. This indicated that the Prosin used in current study contained similar nutritional composition with fish meal, but the AA profile of Prosin is less balanced than fish meal.

From the results in Exp. 1, we can see that the DE and ME contents of fish meal both as fed and DM basis measured in this experiment were slightly greater than the values reported by NRC (1998). Although the GE content of fish meal was lower than Prosin and Protide, the DE and ME contents of fish meal as DM basis were significant higher than Prosin and Protide. This reflected that the energy utilization of fish meal is higher than Prosin and Protide, but no significant differences were found in ME or DE contents between Prosin and Protide. As far as we know, data on DE and ME contents of SCP were limited. Therefore, the values obtained in the present study for Prosin and Protide could reasonably be used to formulate diets in pig production.

The results of Exp. 2 showed that the SID of CP was 85.78%, and the SID of essential AA in fish meal ranged from 84.86% for histidine and arginine to 93.5%, which was similar to the SID values reported by Urbaityte et al. (2009) and Cervantes-Pahm and Stein (2010). The SID of CP and essential AA obtained for Prosin and Protide ranged

from 75.86% for leucine of Protide to 87.24% for arginine of Protide, which were close to the values for SCP obtained in growing pigs by Wang et al. (2013), but Prosin and Protide used in current study had greater SID of arginine, lower SID of lysine. The SID of CP and essential AA of Prosin and Protide were significant lower than fish meal, which indicated that although most essential AA in Prosin and Protide was similar with fish meal (results in Table 1), the AA utilization was worse than fish meal. Skrede et al. (1998) suggested that N digestibility was influenced by digestibility of nucleic acids and other non-protein constituents in SCP. The lower digestibility for SCP may result from its higher non-protein nitrogen (e.g., nucleotides content), which is poorly used by swine (Wang et al., 2013). SCP contains approximately 10% nucleic acids, mostly as RNA (Skrede et al., 1998), the RNA and DNA contents are much higher than those found in fish meal, for instance (Greife, 1984). Therefore, the high nucleic acids contents may be the reason for decreased AA digestibility of Prosin and Protide.

In Exp. 3, the results of performance showed that the full replacement of fish meal protein by Prosin and Protide significantly decreased the performance of weaned piglets, even though all the nutrients were kept consistent among diets. This was consistent with the previous result from Ø verland et al. (2004) who suggested that replacing soybean meal with 5%, 10%, or 15% SCP in diets containing identical ME, available lysine, methionine and threonine reduced the growth performance of pigs linearly during the piglet period, but had no adverse effect on growing-finishing or overall period growth performance of pig. Furthermore, Ø verland et al. (2001) reported that feeding diets in which the SCP replaced 50% of the lysine in soybean meal (low SCP diets containing either 30 or 52 g/kg SCP) did not affect growth performance during the growing period. This indicated that more amounts of protein can be replaced by SCP in growing or finishing pigs' diet than that of weaned piglets' diet; an inclusion of a certain level of SCP in weaned piglets' diet is feasible, because the results obtained from our study showed that 50% replacement of fish meal protein with Prosin or Protide in diets containing identical nutrient levels did not affect the performance of weaned piglets.

The reason for decreased performance in pigs fed the 5% Prosin or 5% Protide diet may be partly due to the decreased digestibility of CP observed in our study, but it was similar among the pigs fed 5% fish meal, 2.5% Prosin and 2.5% Protide diets. The higher inclusion of Prosin and Protide in diet could decrease the digestibility of CP. This was consistent with the study in grower-finisher pigs observed by Zhao et al. (2012) who suggested that the nitrogen metabolism in growing-finishing pigs was

unaffected when 25% of dietary soybean protein was replaced by SCP, but the nitrogen digestibility of 50% replacement group was 5% lower during the growing period and 3% lower during the finishing period than that of CON group. Aas et al. (2006) also showed that the apparent digestibility of nitrogen, sum of amino acids decreased as dietary SCP levels increased.

The morphological changes in the intestine would be followed by a change in nutrient digestion and absorption (Pluske et al., 1997), therefore, the villus height to crypt depth ratio of small intestine would affect the nutrient digestibility and absorption capacity (Montagne et al., 2003). In the current study, the digestibility of CP was decreased significantly when replaced 100% of fish meal with Prosin or Protide in weaned pig's diets. We also observed that the villus height to crypt depth ratio in the jejunum was significantly decreased in the pigs fed 5% Prosin and 5% Protide diets compared with the pigs fed CON, 2.5% Prosin and 2.5% Protide diets. But pigs fed CON diet had a greater villus height to crypt depth ratio in ileum than the pigs fed 5% Prosin and 5% Protide diets. This demonstrated that full replacement of fish meal with Prosin or Protide in diet decreased villus height and villus height to crypt depth ratio in the jejunum and ileum, resulted in lower digestibility of CP, and reduced the growth performance of weaned piglets.

Our findings suggest that although Prosin and Protide contained lower ME content and SID of AA than fish meal, Prosin and Protide replacing 50% of fish meal in diet with identical nutrient levels could obtain similar performance, nutrient digestibility and intestinal morphology in weaned pigs.

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REFERENCES

- Aas, T. S., B. Grisdale-Helland, B. F. Terjesen, and S. J. Helland. 2006. Improved growth and nutrient utilisation in Atlantic salmon (*Salmo salar*) fed diets containing a bacterial protein meal. Aquaculture 259:365-376.
- Adeola, O. and N. L. Bajjalieh. 1997. Energy concentration of high-oil corn varieties for pigs. J. Anim. Sci. 75:430-436.
- Anupama, P. and L. Ravindra, 2000. Value-added food: single cell protein. Biotechnol. Adv. 18:459-479.
- AOAC. 2000. Official methods of analysis, 17th ed. Association of

- Official Analytical Chemists, Arlington, VA.
- Berge, G. M., G. Baeverf jord, A. Skrede, and T. Storebakken. 2005. Bacterial protein grown on natural gas as protein source in diets for Atlantic salmon, Salmo salar, in saltwater. Aquaculture 244:233-240.
- Braude, R., Z. D. Hosking, K. D. MitchellI, S. Plonka, and I. E. Sambrook. 1977. Pruteen, a new source of protein for growing pigs. I. Metabolic experiment: Utilization of nitrogen. Livest. Prod. Sci. 4:79-89.
- Cervantes-Pahm, S. K. and H. H. Stein. 2010. Ileal digestibility of amino acids in conventional, fermented, and enzyme-treated soybean meal and in soy protein isolate, fish meal, and casein fed to weanling pig. J. Anim. Sci. 88:2674-2683.
- Greife, H. A. 1984. Die Nukleinsäuren ein gesundheitlicher Risikofaktor beim Einsatz von "Single-Cell Protein" in der Tierernährung? (Teil 1). Kraftfutter. 67:412-414.
- Hanssen, J. T. and L. Farstad. 1980. Effects of feeding large amounts of "Pruteen" and "Toprina" on some biological parameters in growing finishing pigs. Acta Agric. Scand. 30:74-80.
- Hellwing, A. L. F., A. H. Tauson, Ø. Ahlstrøm, and A. Skrede. 2005. Nitrogen and energy balance in growing mink (Mustela vison) fed different levels of bacterial protein meal produced with natural gas. Arch. Anim. Nutr. 59:335-352.
- Hellwing, A. L. F., A. H. Tauson, and A. Skrede. 2006. Effect of bacterial protein meal on protein and energy metabolism in growing chickens. Arch. Anim. Nutr. 60:365-381.
- Hellwing, A. L. F., A. H. Tauson, A. Skrede, and N. P. Kjos. 2007a. Bacterial protein meal- effects on protein and energy metabolism in pigs. Animal 1:45-54.
- Hellwing, A. L. F., A. H. Tauson, A. Skrede, N. P. Kjos, and Ø. Ahlstrøm. 2007b. Bacterial protein meal in diets for pigs and minks: Comparative studies on protein turnover rate and urinary excretion of purine base derivatives. Arch. Anim. Nutr. 61:425-443.
- Karr-Lilienthal, L. K., N. R. Merchen, C. M. Grieshop, M. A. Flahaven, D. C. Mahan, N. D. Fastinger, M. Watts, and G. C. Fahey, Jr. 2004. Ileal amino acid digestibilities by pigs fed soybean meals from five major soybean-producing countries. J. Anim. Sci. 82:3198-3209.
- Kim, S. W. and R. A. Easter. 2001. Nutritional value of fish meals in the diets for young pigs. J. Anim. Sci. 79:1829-1839.
- Kuhad, R. C., A. Singh, K. K. Tripathi, R. K. Saxena, and K. E. L. Eriksson. 1997. Microorganisms as an alternative source of protein. Nutr. Rev. 55:65-75.
- Li, P. F., X. S. Piao, Y. J. Ru, X. Han, L. F. Xue, and H. Y. Zhang. 2012. Effects of adding essential oil to the diet of weaned pigs on performance, nutrient utilization, immune response and intestinal health. Asian-Aust. J. Anim. Sci. 25:1617-1626.
- Liu, P., X. S. Piao, P. A. Thacker, Z. K. Zeng, P. F. Li, D. Wang, and S. W. Kim. 2010. Chito-oligosaccharide reduces diarrhea incidence and attenuates the immune response of weaned pigs challenged with *Escherichia coli* k88. J. Anim. Sci. 88:3871-3879.
- Montagne, L., J. R. Pluske, and D. J. Hampson. 2003. A review of interactions between dietary fibre and the intestinal mucosa, and their consequences on digestive health in young nonruminant animals. Anim. Feed Sci. Technol. 108:95-117.

- NRC. 1998. Nutrient requirements of swine, 10th ed. National Academy Press, Washington, DC.
- Ø verland, M., S. Skrede, and T. Matre. 2001. Bacterial protein grown on natural gas as feed for pigs. Acta. Agric. Scand. A. Anim. Sci. 51:97-106.
- Ø verland, M., N. P. Kjos, and A. Skrede. 2004. Effect of bacterial protein meal grown on natural gas on growth performance and carcass traits of pigs. Ital. J. Anim. Sci. 3:323-336.
- Paraskevopoulou, A., I. Athanasiadis, M. Kanellaki, A. Bekatorou, G. Blekas, and V. Kiosseogloua. 2003. Functional properties of single cell protein produced by kefir microflora. Food Res. Int. 36:431-438.
- Petersen, G. I. and H. H. Stein. 2006. Novel procedure for estimating endogenous losses and measuring apparent and true digestibility of phosphorus by growing pigs. J. Anim. Sci. 84:2126-2132.
- Pluske, J. R., D. J. Hampson, and I. H. Williams. 1997. Factors influencing the structure and function of the small intestine in the weaned pig: A review. Livest. Prod. Sci. 51:215-236.
- SAS. 1999. SAS user's guide: Statistics (Version 8.01). SAS Inst. Inc., Cary, NC, USA.
- Shen, Y. B., X. S. Piao, S. W. Kim, L. Wang, P. Liu, I. Yoon, and Y. G. Zhen. 2009. Effects of yeast culture supplementation on growth performance, intestinal health, and immune response of nursery pigs. J. Anim. Sci. 87:2614-2624.
- Skrede, A., T. Storebeakken, G. M. Berge, O. Herstad, K. G. Aarstad, and F. Sundstøl. 1998. Digestibility of bacterial protein (BioProtein) grown on natural gas in mink, pigs, chicken and Atlantic salmon. Anim. Feed Sci. Technol. 76:103-116.

- Stein, H. H., C. F. Shipley, and R. A. Easter. 1998. Technical note: A technique for inserting a T-cannula into the distal ileum of pregnant sows. J. Anim. Sci. 76:1433-1436.
- Urbaityte, R., R. Mosenthin, M. Eklund, H. Piepho, N. Sauer, and M. Rademacher. 2009. Standardised ileal crude protein and amino acid digestibilities in protein supplements for piglets. Arch. Anim. Nutr. 63:356-378.
- Wang, J. P., J. D. Kim, J. E. Kim, and I. H. Kim. 2013. Amino acid digestibility of single cell protein from *Corynebacterium* ammoniagenes in growing pigs. Anim. Feed Sci. Technol. 180:111-114.
- Williams, C. H., D. J. David, and O. Iismaa. 1962. The determination of chromic oxide in faeces sample by atomic absorption spectrophotometry. J. Agric. Sci. 59:381-385.
- Woodworth, J. C., M. D. Tokach, R. D. Goodband, J. L. Nelssen, P. R. O' Quinn, D. A. Knabe, and N. W. Said. 2001. Apparent ileal digestibility of amino acids and the digestible and metabolizable energy content of dry extruded-expelled soybean meal and its effects on growth performance of pigs. J. Anim. Sci. 79:1280-1287.
- Zepka, L. Q., E. Jacob-Lopes, R. Goldbeck, L. A. Souza-Soares, and M. I. Queiroz. 2010. Nutritional evaluation of single-cell protein produced by *Aphanothece microscopica Nägeli*. Bioresour. Technol. 101:7107-7111.
- Zhao, Y., B. Yu, X. B. Mao, J. He, Z. Q. Huang, Q. Mao, and D. W. Chen. 2012. Effect of dietary bacterial lysine by-product meal supplementation on growth performance and excretion of purine base derivatives in growing-finishing pig. Livest. Sci. 149:18-24.