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Product-to-Parent Reversion of Trenbolone: Unrecognized Risks for Endocrine Disruption

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Abstract

Trenbolone acetate (TBA) is a high-value steroidal growth promoter often administered to beef cattle, whose metabolites are potent endocrine-disrupting compounds. We performed laboratory and field phototransformation experiments to assess the fate of TBA metabolites and their photoproducts. Unexpectedly, we observed that the rapid photohydration of TBA metabolites is reversible under conditions representative of those in surface waters (pH 7, 25°C). This product-to-parent reversion mechanism results in diurnal cycling and substantial regeneration of TBA metabolites at rates that are strongly temperature- and pH-dependent. Photoproducts can also react to produce structural analogs of TBA metabolites. These reactions also occur in structurally similar steroids, including human pharmaceuticals, which suggests that predictive fate models and regulatory risk assessment paradigms must account for transformation products of high-risk environmental contaminants such as endocrine-disrupting steroids.

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Supplementary Materials

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Materials and Methods

Fig. S1 to S9

Table S1

References (41, 42)

Humans discharge a multitude of bioactive organic contaminants into receiving waters that adversely affect aquatic organisms (1–3). Risk assessment approaches for regulating these contaminants often are simplistic, typically assuming that if degradation occurs, the associated ecological risk greatly decreases. However, there is growing sentiment that some environmental transformation reactions result in minimal mitigation of risk, forming products that retain bioactive moieties, exhibit greater toxicity, or affect different biological end points (4, 5).

The androgenic steroid trenbolone acetate [TBA; 17 β -(acetyloxy)estra-4,9,11-trien-3-one] is an anabolic growth promoter implanted in over 20 million cattle annually (6, 7), with annual revenue attributable to its use likely exceeding \$1 billion (8). Given the extensive use of TBA, its dominant metabolite [17 α -trenbolone (17 α -TBOH)] and other known metabolites [17 β -trenbolone (17 β -TBOH) and trendione (TBO)] can be widespread in agricultural environments (9, 10). 17 α -TBOH and 17 β -TBOH are potent endocrine-disrupting compounds, with concentrations as low as 10 to 30 ng/liter causing skewed sex ratios and reduced fecundity in fish (7, 11). Nevertheless, observations of TBA metabolite occurrence have not yet translated to concern, because they are believed to exhibit limited persistence in receiving waters. For example, manufacturer studies used for regulatory approvals specifically point to limited ecosystem risks of TBA metabolites due to rapid photodegradation (12).

Accordingly, we previously identified the structure and bioactivity of TBA metabolite photoproducts, showing that 17 β -TBOH undergoes rapid photohydration (13) to yield 12,17-dihydroxy-estra-5(10),9(11)-diene-3-one (or simply 12-hydroxy-17 β -TBOH) (14, 15). An analogous C12-hydroxylated photoproduct was identified for TBO (15), whereas 17 α -TBOH yields both major C12 [12-hydroxy-17 α -TBOH (15)] and minor C5 (5-hydroxy-17 α -TBOH) hydroxylated photoproducts (supplementary text and figs. S1 and S2). Because these photoproduct mixtures retained bioactivity (15), we focused on photoproduct stability across a range of conditions representative of light (day) and dark (night) surface waters. When photoproduct transformation was observed, we used liquid chromatography high-resolution tandem mass spectroscopy (LC-HRMS/MS) and nuclear magnetic resonance (NMR) to characterize the resulting product structures and the potential for persistent bioactivity in surface waters (16).

Simulated day-night cycling experiments conducted over 72 hours at pH 7, 25°C revealed unexpected persistence of TBA metabolites (Fig. 1A). For example, 17 α -TBOH decay during irradiation was consistently followed by concentration rebound during subsequent dark periods, with equivalent rates of photodecay and dark regeneration for each diurnal cycle. Both NMR (figs. S3 to S10) and LC-HRMS/MS confirmed that 17 α -TBOH was regenerated in the dark. This dynamic behavior appears to be linked to the thermal (i.e., nonphotochemical) instability of C5 and C12 hydroxylated photoproducts, which decayed concurrently with 17 α -TBOH regrowth.

A noteworthy feature of both 5- and 12-hydroxy-17 α -TBOH is their allylic alcohol moiety (Fig. 1A). Steroidal allylic alcohols are prone to dehydration but typically under more aggressive conditions (e.g., low pH and high temperature) (17). However, our data suggest

that 5- and 12-hydroxy-17 α -TBOH dehydrate under ambient conditions, with the driving force being regeneration of the conjugated trienone system. This coupled photohydration-dehydration mechanism results in net reversibility of 17 α -TBOH photolysis, a product-to-parent reversion mechanism also observed for other TBA metabolites (Fig. 1B). 17 β -TBOH and TBO exhibited reduced regrowth during 12-hour dark periods (~1% of initial mass) relative to 17 α -TBOH (~15%), which we attribute to their photoproducts being more susceptible to concurrent phototransformation and hydroxylation rather than dehydration (supplementary text and fig. S11).

We also simulated TBA metabolite transport from the photic zone of surface water into darker regions (such as lake hypolimnia, hyporheic zones, or benthic sediments) by photolyzing TBA metabolite solutions to >99% transformation (pH 7, 6 hours irradiation) and then storing the resulting photoproduct mixtures in the dark for 5 days at 25°C. Consistent with the diurnal cycling experiments, 17 α -TBOH exhibited substantial reversion, with over 60% of its initial mass recovered after 120 hours, whereas 17 β -TBOH and TBO yielded 10% recovery over this period (Fig. 2A).

The rates and extents of product-to-parent reversion were highly dependent on solution conditions, with some promoting rapid and near-complete TBA metabolite regeneration. For example, $88 \pm 3\%$ of the initial 17 α -TBOH mass was regenerated nearly instantaneously when photoproduct mixtures were acidified to pH 2, with slightly lower recoveries ($66 \pm 5\%$) when raised to pH 12 (Fig. 2A). Reversion of 17 β -TBOH and TBO was also acid-catalyzed ($65 \pm 7\%$ and $32 \pm 4\%$ recovery, respectively), but their regrowth was more limited at pH 12. We believe that these recoveries at pH 2 probably reflect the maximum photoproduct mass available for reversion.

The reversion of 17 α -TBOH was also enhanced at pH 9 and pH 5, at least initially, relative to pH 7 (Fig. 2B). Thus, acid- and base-catalyzed reversion will probably result in higher 17 α -TBOH concentrations in mildly acidic or alkaline waters. In fact, the initial rate of reversion at pH 5 is fast enough to slow 17 α -TBOH phototransformation relative to the rate observed at neutral pH (supplementary text and fig. S12). In manufacturer regulatory studies, slower rates of 17 α -TBOH phototransformation at pH 5 relative to pH 7 were attributed, we believe incorrectly, to variations in solar irradiance from cloudy weather experienced during data collection at pH 5 (12).

Reversion is also temperature-dependent. At pH 7, rates of 17 α -TBOH regrowth increased nearly 30-fold from 5° to 35°C (Fig. 2C), a temperature range representative of seasonal variations. Faster reversion rates at 35°C ultimately allowed more complete 17 α -TBOH recovery in the dark (30% over 12 hours and 75% over 60 hours). Accordingly, we expect photoproduct-to-parent reversion to be most prominent in warm sunlit waters, whereas colder winterlike conditions should promote photoproduct stability by slowing dehydration.

Accounting for these effects, we can now present a more complete characterization of TBA metabolite fate in surface waters. Consistent with established pathways for allylic sterols (17), we propose that dehydration at moderately acidic to low pH proceeds via unimolecular (E1) elimination involving the formation of a resonance-stabilized carbocation (Fig. 3).

Parent regeneration from this carbocation intermediate may be accompanied by concurrent transformations, including structural rearrangements and/or reaction with naturally abundant nucleophiles, to yield complex mixtures of isomers, structural analogs, and substituted derivatives of TBA metabolites with uncharacterized bioactivity. For example, while monitoring TBO photo-product reversion at pH 5, we observed evidence of photoproduct interconversion and formation of a presumed structural analog that coelutes with TBO (supplementary text and figs. S13 to S18). These phenomena were not observed at neutral or basic pH, which is consistent with the proposed acid-promoted carbocation intermediate. At neutral and basic pH, dehydration is more likely to proceed via enol or enolate formation in parallel to reactions yielding higher-order hydroxylated products.

Beyond model laboratory systems, we have also observed reversion in more complex aquatic matrices, including Iowa River water (Fig. 4), as well as in experiments using environmentally relevant TBA metabolite concentrations (for example, ~420 ng/liter 17 β -TBOH; fig. S19). To explore reversion in agricultural receiving waters with competing attenuation pathways (such as sorption and biodegradation), we dosed a small collection pond on a rangeland with manure from TBA-implanted cattle (supplementary text). After overnight leaching increased 17 α -TBOH concentrations in the pond to 6.1 ng/liter, concentrations decreased by ~50% during the day, consistent with phototransformation. At sunset, a large (31-liter) sample was collected and processed so that subsamples of this baseline sample could be subjected to different storage conditions (Fig. 4). Laboratory storage at 35°C significantly increased 17 α -TBOH concentrations (from 7 to 20 ng/liter; Games-Howell post-hoc test, $P = 0.003$), whereas samples stored at 1°C were statistically identical to the baseline sample (Games-Howell post-hoc test, $P = 0.65$). However, when the temperature of these 1°C samples was subsequently raised after 24 hours to 35°C, 17 α -TBOH concentrations statistically increased to 15 ng/liter (Games-Howell post-hoc test, $P = 0.005$). These data are consistent with expected trends for product-to-parent reversion dynamics under actual field conditions. Incidentally, we also observed similar decay and regrowth characteristics for a major uncharacterized, nontarget compound, probably steroidal, in our gas chromatography–tandem mass spectrometry chromatograms, whose occurrence was highly correlated to known TBA metabolites (figs. S20 to S22).

Although growth-promoting steroids provide indisputable economic and environmental advantages (such as reduced land use, nutrient loads, and greenhouse gas emissions) for animal agriculture (18), their use should not compromise environmental health. Contrary to prevailing assumptions of limited environmental persistence, product-to-parent reversion results in the enhanced persistence of TBA metabolites via a dynamic exposure regime that defies current fate models and ecotoxicology study designs. Reversion also provides a route to novel steroidal isomers, structural analogs, and derivatives, or “environmental designer steroids,” with as-yet-uncharacterized properties and risks. Temperature- and pH-dependent reversion rates also imply substantial uncertainty for nearly all existing TBA metabolite occurrence data. Collectively, these possibilities suggest the unrecognized occurrence of TBA metabolites and/or structurally similar, bioactive transformation products that may contribute to otherwise unexplained observations of endocrine disruption in agriculturally affected receiving waters (19, 20). For example, although chemical analyses often cannot

identify causative agents, a recent study indicated widespread (35% of tested waters) androgenic activity in U.S. waters (21).

We also recently observed photoproduct-to-parent reversion for dienogest, a potent progestin used as an oral contraceptive (22), and for dienedione, an illicit anabolic steroid marketed as a “bodybuilding supplement” (fig. S23). Therefore, it appears that other pharmaceutical steroids, namely those with dienone and trienone moieties, also may exhibit enhanced environmental persistence and pose greater ecological risks than currently realized. In fact, although this work focused solely on phototransformation, we believe that the propensity for reversion is more generally linked to the conjugated enone moiety of TBA metabolites and these other pharmaceuticals. Ultimately, such observations of product-to-parent reversion illustrate that comprehensive assessment of environmental transformation products should be prioritized for high-risk contaminants such as endocrine-disrupting steroids. Further, the use of TBA or similar steroids that are equally prone to environmental transformations that preserve bioactive moieties may need to be reevaluated in favor of more-sustainable pharmaceutical technologies designed to ensure ecosystem protection.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

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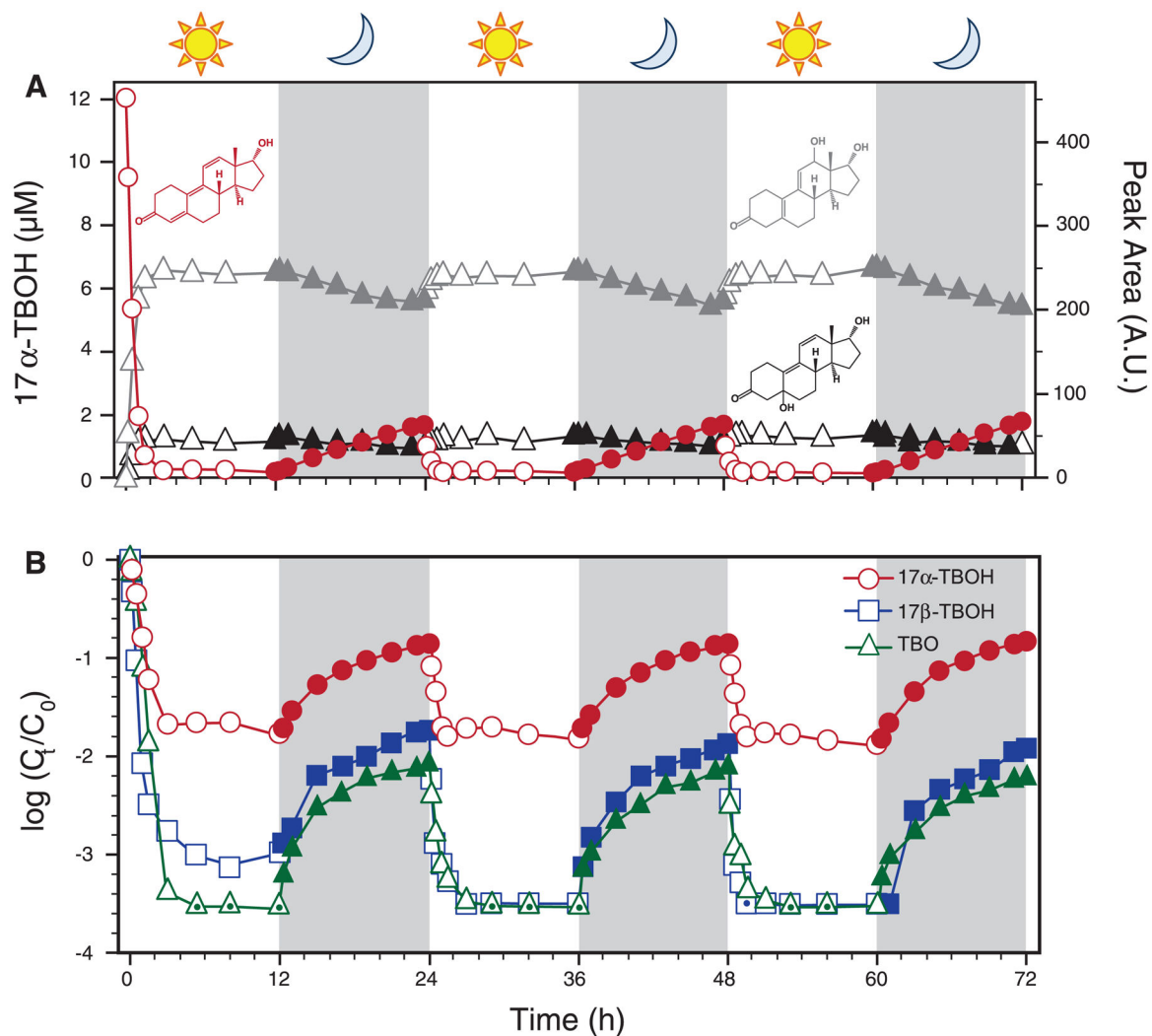


Fig. 1. Diurnal cycling of TBA metabolites

(A) Day-night cycling of 17 α -TBOH (red), 12-hydroxy-17 α -TBOH (gray), and 5-hydroxy-17 α -TBOH (black) at pH 7, 25°C. Data for 17 α -TBOH are presented as aqueous concentration (left axis), whereas peak areas from spectrophotometric absorbance [wavelength (λ) = 350 nm for 17 α -TBOH; λ = 254 nm for photoproducts] are also presented (right axis). (B) Corresponding data for 17 β -TBOH and TBO, where symbols with a center dot indicate values below our detection limit. In these cases, data points correspond to this limit (\sim 3 nM) for 17 β -TBOH and TBO. C_t/C_0 , time-dependent concentration normalized to initial concentration; h, hours; A.U., arbitrary units.

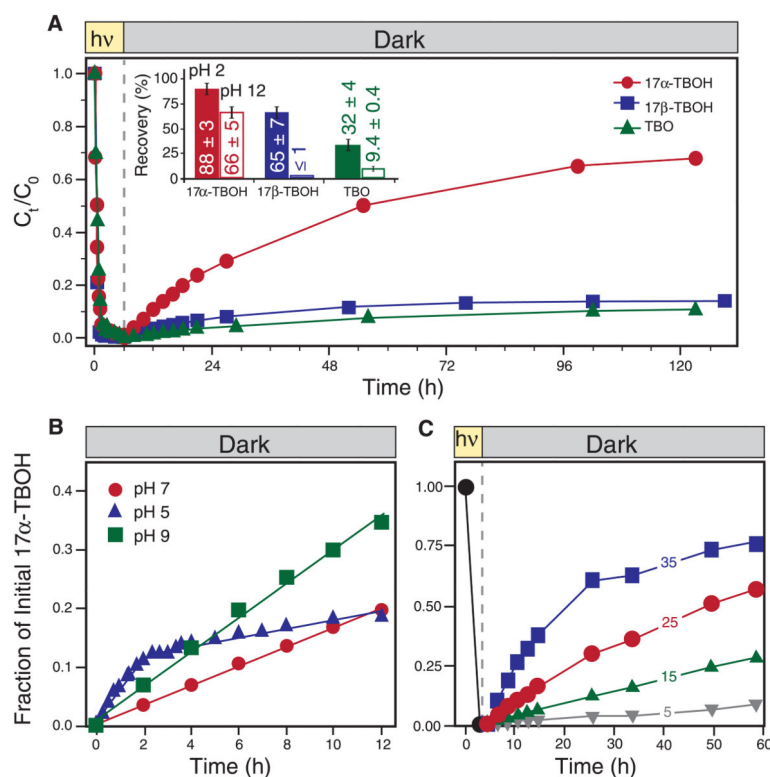


Fig. 2. Dark regrowth of TBA metabolites and the influence of pH and temperature on reversion (A) Regeneration of 17 α -TBOH, 17 β -TBOH, and TBO in photoproduct mixtures stored in the dark (pH 7, 25°C). hv, light energy. The inset reports the percent recovery (mean and standard deviation of triplicate analyses) when photoproduct mixtures were adjusted to pH 2 or pH 12. Also shown is the regeneration of 17 α -TBOH as a function of (B) pH (5, 7, and 9 at 25°C) and (C) temperature (5, 15, 25, and 35°C at pH 7) from photoproduct mixtures. In (B), the initial rate of regrowth obtained from linear regression analysis was enhanced at pH 9 (0.3 $\mu\text{M hour}^{-1}$) and, at least initially [time (t) < 2 hours], at pH 5 (0.6 $\mu\text{M hour}^{-1}$) relative to pH 7 (0.17 $\mu\text{M hour}^{-1}$). The slower rate observed over longer time scales at pH 5 (0.07 $\mu\text{M hour}^{-1}$) probably reflects differences in the dehydration rates of 5- and 12-hydroxy-17 α -TBOH.

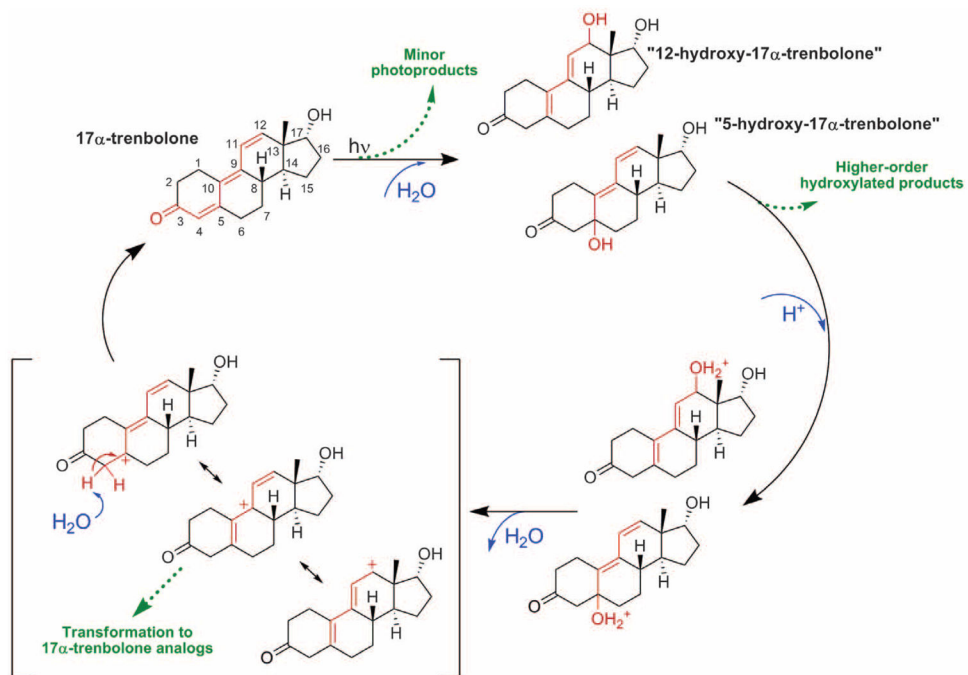


Fig. 3. Proposed acid-catalyzed reversible photohydration of 17 α -TBOH

Dashed green arrows indicate pathways that break the product-to-parent reversion cycle.

Red highlights detail the changes in the triene structure during photolysis and subsequent dehydration.

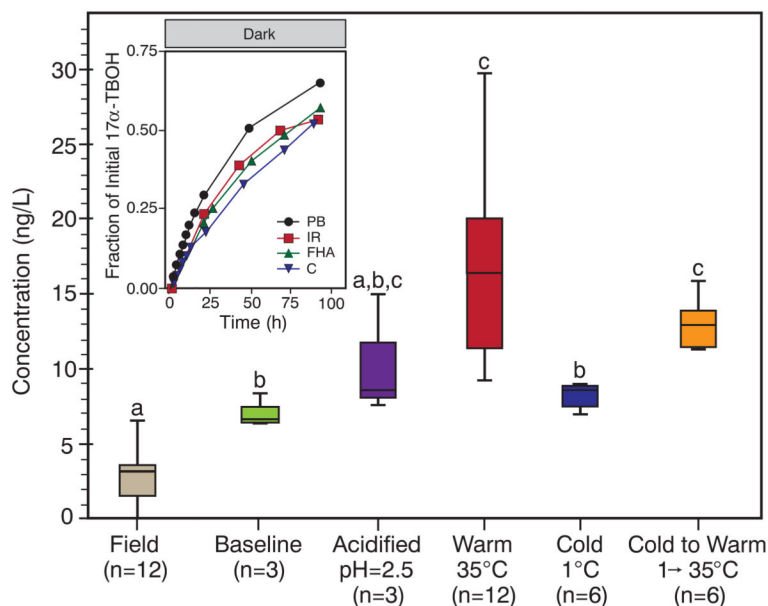


Fig. 4. 17 α -TBOH dynamics in a field mesocosm and complex aquatic matrices
 After dosing the collection pond (at 22:00), field samples ($n = 4$, in triplicate) were analyzed over 17 hours. At sunset (15:00), the baseline sample was processed and split into subsamples, which were either acidified to pH 2.5, incubated [warm, temperature (T) = 35°C], refrigerated (cold, $T = 1^\circ\text{C}$), or refrigerated then incubated after 24 hours (cold to warm, $T = 1^\circ \rightarrow 35^\circ\text{C}$). Group means are different [one-way analysis of variance: $F(5,37) = 17.5$, $P < 0.001$] if groups do not share the same letter as other groups (Games-Howell post-hoc test: $P < 0.05$). Lines, boxes, and whiskers represent median, interquartile, and minimum and maximum values, respectively. The inset shows regeneration of 17 α -TBOH in photoproduct mixtures (dark, 25°C) in solutions of 10 mg/liter Fluka Humic Acid (FHA), 250 mg/liter of bicarbonate as CaCO₃ (C), and Iowa River water (IR; turbidity, 15.1 nephelometric turbidity units; alkalinity, 196 mg/liter as CaCO₃; total hardness, 270 mg/liter as CaCO₃; total dissolved organic carbon, 16.6 mg/liter; pH 8.2.). Provided for comparison are data from a phosphate buffer (PB) system. Unless noted, all systems were at pH 7. The rate of reversion is less in Iowa River water than would be expected for its alkaline pH, probably because of the slight inhibition observed in model systems with Fluka Humic Acid and bicarbonate.