

## NIH Public Access

Author Manuscript

*Curr Genet Med Rep*. Author manuscript; available in PMC 2015 June 01.

### Published in final edited form as:

Curr Genet Med Rep. 2014 June 1; 2(2): 75-84. doi:10.1007/s40142-014-0035-9.

### **Rare Variants and Transcriptomics in Alzheimer disease**

### Crystal Humphries<sup>1,2</sup> and Martin A. Kohli<sup>2</sup>

<sup>1</sup>Department of Human Genetics, John T. Macdonald Foundation, University of Miami, Miller School of Medicine, 1501 NW 10th Avenue (BRB-531), Miami, FL 33136, USA

<sup>2</sup>John P. Hussman Institute for Human Genomics (HIHG), University of Miami, Miller School of Medicine, 1501 NW 10th Avenue (BRB-531), Miami, FL 33136, USA

### Abstract

Alzheimer disease (AD) is the most common dementia in the elderly, still without effective treatment. Early-onset AD (EOAD) is caused by mutations in the genes *APP*, *PSEN1* and *PSEN2*. Genome-wide association studies have identified >20 late-onset AD (LOAD) susceptibility genes with common variants of small risk, with the exception of *APOE*. We review rare susceptibility variants in LOAD with larger effects that have been recently identified in the EOAD gene *APP* and the newly discovered AD genes *TREM2* and *PLD3*. Human genetic studies now consistently support the amyloid hypothesis of AD for both EOAD and LOAD. Moreover, they identified biological processes that overlap with human transcriptomics studies in AD across different tissues, such as inflammation, cytoskeletal organization, synaptic functions, etc. Transcriptomic profiles of pre-symptomatic AD-associated variant carriers already reflect specific molecular mechanisms reminiscent to those of AD patients. This might provide an avenue for personalized medicine.

### Keywords

transcriptomics; Alzheimer disease; dementia; elderly; APP; PLD3; PSEN1; PSEN2; TREM2; personalized medicine; late-onset Alzheimer disease (LOAD)

### Introduction

### Alzheimer Disease

Alzheimer disease (AD) is the most common form of dementia in the elderly, accounting for up to 75% of all dementia patients. Currently, there are more than 25 million people worldwide with AD and their numbers are anticipated to double every 20 years due to the

Conflict of Interest

### MA Kolin declares no connets of interest.

### Human and Animal Rights and Informed Consent

All studies by the authors involving animal and/or human subjects were performed after approval by the appropriate institutional review boards. When required, written informed consent was obtained from all participants.

Corresponding author: Martin A. Kohli, Ph.D., Post-doctoral associate at the John P. Hussman Institute for Human Genomics, Miller School of Medicine, University of Miami, 1501 NW 10th Avenue (BRB-531), Miami, FL 33136, phone: 305-243-1740, mkohli@med.miami.edu, URL: http://www.hihg.org.

C Humphries declares no conflicts of interest. MA Kohli declares no conflicts of interest.

Humphries and Kohli

expected demographic shift towards higher age. Disease onset of AD is usually over 70 years of age. However, age-specific prevalence of about 4% at age 65 increases exponentially with age and exceeds 20% at age 90 [1]. AD is clinically characterized by a slow but progressive impairment in memory, executive function, language, and other areas of cognition leading to a loss of social and occupational functions [2]. The pathological hallmarks of AD are extracellular amyloid  $\beta$  (A $\beta$ ) plaques and intracellular neurofibrillary tangles (NFT) that accumulate over time in the aging brain. According to the predominant amyloid hypothesis of AD, aggregation of A $\beta$  initiates a pathogenic cascade that eventually leads to inflammation, loss of neurons and synapses and brain atrophy [3, 4].

### **Genetics of Alzheimer Disease**

The heritability of late-onset AD (LOAD) has been estimated to be between 58% and 79% and can be regarded as the proportion of disease vulnerability explained by heritable genetic factors. The remaining risk for LOAD has been attributed to environmental factors, for instance, exposure to vascular risk (e.g. obesity) and possibly beneficial psychosocial factors such as high education and physical exercise [1, 5]. Besides LOAD, there are early-onset forms of AD (EOAD) with disease onset prior to age 60 that represent up to 1% of all AD cases at an average prevalence of about 65 EOAD cases per 100,000 individuals [6, 7]. Approximately 60% of EOAD cases have several relatives also affected by AD and 13% of EOAD cases occur in families which are in concordance with autosomal-dominant inheritance of EOAD over at least three generations (ADEOAD) [6].

In the 1990's, genetic linkage and subsequent positional cloning studies in large, multigenerational ADEOAD families led to the discovery of the first causative AD mutations in the genes encoding the amyloid precursor protein (*APP*) [8, 9], presenilin 1 (*PSEN1*) [10, 11] and presenilin 2 (*PSEN2*) [12, 13]. Meanwhile, around 24 mutations in *APP*, 185 in *PSEN1* and 13 in *PSEN2* have been reported to be pathogenic for AD [14] (see [15] for review on clinical aspects of genetic variants in AD). Most of these extremely rare, familial variants are single amino acid substitutions and show dominant, fully penetrant cosegregation with ADEOAD. The majority of pathogenic variants in full-length APP are located near the N-terminal  $\beta$ -secretase and the C-terminal  $\gamma$ -secretase proteolytic cleavage site of amyloidogenic A $\beta$  fragments of APP that are endogenously produced by cells [15]. The presenilin proteins are the catalytic units of  $\gamma$ -secretase complexes that are involved in the proteolytic cleavage of APP to produce A $\beta$  fragments. Mutations in the presenilins most often alter the proteins' catalytic properties in a way that increase the absolute or relative production of most amyloidogenic A $\beta_{42}$  fragments [16]. Thus, genetic and biochemical evidence for the predominant amyloid hypothesis of AD is convincing [17].

Early genetic linkage and genetic association studies also led to the identification of the two most prominent genetic risk variants for LOAD in exon 4 of the apolipoprotein E (*APOE*) gene; both in terms of their high population frequencies and large effect on LOAD risk [18, 19]. Depending on the population, the *APOE*  $\varepsilon 4$  LOAD risk allele typically occurs in about 15–20% of individuals. Heterozygous carriers ( $\varepsilon 3/\varepsilon 4$ ) for the *APOE*  $\varepsilon 4$  risk allele are 3-fold and homozygous carriers ( $\varepsilon 4/\varepsilon 4$ ) up to 15-fold more likely to suffer from LOAD compared to individuals with the predominant *APOE*  $\varepsilon 3/\varepsilon 3$  genotype. Homozygous  $\varepsilon 4$  carriers reach

close to complete penetrance at age 90 and older. Moreover, the APOE & risk allele is also associated with EOAD, an earlier disease onset of AD and the rarer APOE \varepsilon2 allele is protective for AD [20, 21]. Multiple functions of the APOE protein have been suggested in the pathogenesis of AD. There is strong evidence for differential effects of APOE isoforms on A $\beta$  aggregation and clearance. Additionally proposed mechanisms of APOE with regard to AD involve neurotoxicity, tau phosphorylation, synaptic plasticity and neuroinflammation [22]. The advent in high content genotype chip array technology enabled genome-wide association studies (GWAS) in large cohorts of unrelated LOAD cases and unaffected aged controls. To date, large GWAS consortia in LOAD have identified >20 non-APOE loci that show association with LOAD [23, 24]. Unlike APOE, these loci confer low individual, but reproducible risk to LOAD with odds ratios (OR) between 1.08 and 1.30. Several of these LOAD susceptibility genes can be functionally linked to the pathways of APP and protein tau, are enriched for immune response and inflammation and involve cell migration, lipid transport, endocytosis, hippocampal synaptic function, cytoskeletal function, axonal transport, regulation of gene expression and post-translational modification of proteins, and microglial and myeloid cell function. The population-attributable fraction (PAF) of APOE on risk for LOAD is at about 30%, while each single GWAS-identified locus contributes with individual PAFs between 1.0-8.0% to risk or protection of LOAD [24].

The genetics of sporadic LOAD is consistent with the amyloid hypothesis of AD, but seem to go far beyond the immediate APP pathway and are therefore complex. Moreover, the yet established susceptibility loci for LOAD do not completely explain the overall heritability of LOAD.

### Rare variants and insights from transcriptomics

GWAS successfully identified many susceptibility loci in LOAD, which is important for the better understanding of LOAD etiology. However, with the exception of APOE, GWASidentified associations confer small risk to LOAD and GWAS loci defining SNPs most often merely represent genetic markers correlated to nearby functional risk variants that remain to be revealed. Since the discovery of rare disease susceptibility variants is challenging due to their low abundance, resulting in low statistical power to detect association with disease in a genome-wide screening approach, most LOAD GWAS so far only considered common variants with minor allele frequencies (MAF) above 5% in the general population. Therefore, missing heritability in AD can be explained by low-frequency (5%>MAF>1%) and rare variants (MAF<1%) [25]. The first rare susceptibility variants for LOAD have been identified in the last two years and they show intermediate to large effects, some even comparable to APOE [26]. In the first part of this review, we focus on rare variant associations with LOAD that reached study-wide significance with independent replication in a chronological order of publication. In contrast to GWAS-identified common risk variants, pathogenic ADEOAD mutations and emerging rare susceptibility variants in LOAD are most often protein coding and therefore the actual functional variants. This opens an avenue of functional studies that can assess the molecular consequences of diseaseassociated variants in AD. The second part of this review concentrates on wholetranscriptomics studies from human post-mortem brain and peripheral blood cells. We compare biological processes with differential gene expression in AD to pathways revealed

by genetic studies and summarize first studies on transcriptional profiles typical to AD risk allele carriers.

### Genetics of rare variants in Alzheimer disease

### Rare variants in previously identified AD genes

**The rare APP p.A673T variant is protective from LOAD**—Next generation sequencing methods now allow for rapid and deep sequencing of many human genomes. Cruchaga et al. sequenced the coding region of ADEOAD genes in 440 probands of LOAD families and showed an overall overrepresentation of rare protein sequence changing variants in the genes *APP*, *PSEN1 and 2* when compared to 12,500 population controls, while describing known EAOD mutations and novel, likely pathogenic variants in *APP* (table 1) [27].

Johnsson et al. took advantage of whole-genome sequence data from 1,795 Icelander to search for low-frequency variants in the *APP* gene [28]. The genotypes of variants found at least twice were then imputed (*in silico* genotyped) into 71,700 Icelanders that had high-density SNP array genotypes available in order to test for association with LOAD. The rare *APP p.A673T* substitution (rs63750847) was found to confer a large protective effect from LOAD (OR=4.2–7.5, depending on age and cognitive status of control groups) and also showed reduced cognitive decline among elderly subjects without a diagnosis of AD. The population frequencies of *APP p.A673T* in Northern Europe are around 0.4% (MAF), whereas the protective allele seems to be even rarer in the US (MAF<0.01%). Functionally, *p.A673T* is located adjacent to the aspartyl protease  $\beta$ -site in APP and results in a reduced  $\beta$ -cleavage efficiency of the aspartyl protease  $\beta$ -site cleaving enzyme 1 (BACE1) and thereby in an about 40% reduction of A $\beta$  fragments, as the authors showed *in vitro*. Of importance, the strong protective effect of *APP p.A673T* serves as a proof of principle that reducing  $\beta$ -cleavage of APP may protect from AD.

### Discovery of new LOAD genes through rare variants

**Rare variants in TREM2, e.g. p.R47H confer risk to LOAD**—Two independent groups reported back-to-back a highly significant association with LOAD for the rare substitution *p.R47H* (rs75932628, MAF=0.3%) in the gene encoding the triggering receptor expressed on myeloid cells 2 (*TREM2*) [26, 29]. This variant already reached genome-wide significant association (p<5e-08) with LOAD in the large Icelandic study samples of Jonsson et al. described above, with subsequent replication in several LOAD cohorts of European descent. Interestingly, the rare allele of *p.R47H* confers similarly high risk to LOAD (overall OR=2.90) as the common *APOE &*4 allele and comparably reduces age of LOAD onset also by about three years per risk allele copy. In reminiscence of Jonsson et al.'s finding that the protective *APP p.A673T* substitution reduces cognitive decline in elderly controls, *TREM2 p.R47H* accelerates cognitive decline in aged controls [26]. Guerreiro et al. [29] independently found highly significant association of *p.R47H* in large European LOAD GWA study samples, while also showing an overall significant accumulation of rare variants in exon 2 of the *TREM2* gene in LOAD cases versus controls including additional variants such as *p.D87N*. Interestingly, they found that three variants

Humphries and Kohli

(Q33X, Y38C, and T66M) in the recessive state cause the rare Nasu-Hakola disease and related forms of early-onset dementia distinct from AD [29]. TREM2 is a membrane protein that forms a receptor signaling complex with the TYRO protein tyrosine kinase binding protein (TYROBP) and is involved in macrophage activation and inflammation. In the brain, *TREM2* is mainly expressed in microglia of white matter. *TREM2* expression and microglial phagocytosis of cell debris and amyloid concomitantly increases with the accumulation of A $\beta$  plaques in transgenic mice models of AD that carry pathogenic ADEAOD mutations in a human copy of the *APP* gene [26, 29]. Thus, loss-of-function variants in *TREM2* may interfere with A $\beta$  clearance and anti-inflammatory responses and thereby increasing the risk for AD.

Rare variants in PLD3, e.g. p.V232M confer risk to LOAD—Cruchaga et al. [30] applied whole-exome sequencing in multiple multiplex families with a high burden of LOAD and identified the rare missense variant p.V232M (rs145999145, MAF=0.4%) in the phospholipase D3 (PLD3) gene that co-segregated with disease in two independent families. Subsequent genotyping of p.V232M in several large LOAD case-control cohorts of European descent (>11,000 individuals) confirmed the LOAD risk conferring character of the variant allele with high significance and intermediate effect size (OR=2.1). This casecontrol association resulted in a large effect when only familial LOAD cases were compared against controls (OR=3.4). Moreover, the risk allele also showed association with an earlier onset of LOAD. The authors then sequenced the coding region of PLD3 in more than 2,000 LOAD cases and as many controls of European descent and found a genome-wide significant gene-based association with LOAD and several rare variants in the PLD3 gene (OR=2.6). Nominally significant single-variant associations with intermediate to large effect could be shown for PLD3 p.M6R and the synonymous splice site variant p.A442A. This gene-based association of rare PLD3 variants being overrepresented in LOAD cases was replicated in an African American case-control cohort as well as the single-variant association for p.A442A. PLD3 expression is high in several AD-relevant brain regions in healthy controls, but reduced in neurons of LOAD patients. PLD3 overexpression and knockdown experiments in cell cultures revealed that high PLD3 expression correlates with lower extracellular A $\beta$  levels and that PLD3 protein can be co-immunoprecipitated with APP. Thus, PLD3 protein is likely protective against AD through its role in APP trafficking [30].

### Transcriptomics in Alzheimer disease

EOAD and LOAD have similar clinical manifestations and pathological features [31]. This suggests that similar cellular and biological processes are disrupted in both forms of Alzheimer disease. This notion is also supported by genetics of AD, most recently complemented by rare variant associations as mentioned above. Genome-wide gene transcription is a measurable intermediate proxy of how the genomic sequence gives rise to the altered protein formation that eventually triggers disease and reflects disease progression and cellular coping mechanisms to pathological changes. There are two primary means of measuring genome-wide transcription: Microarray technology and massively parallel RNA sequencing (RNA-seq). While microarrays use hybridization to measure known transcripts, the recent advent of RNA-seq allows for measurement of known and novel transcripts,

including alternatively spliced transcripts. Both of these technologies have been used to shed light on gene transcriptional changes related to AD pathology.

### **Profiling in Post-Mortem Brain Tissue**

Since 2005, at least twenty-five studies have been published examining postmortem human brain tissue. These studies primarily focused their efforts on tissue from the frontal cortex, hippocampus, and temporal lobe because these regions are most affected by AD pathology, essentially  $A\beta$  plaques and NFTs [32–50]. These studies differ in their findings on which individual genes are significantly altered between AD patients and healthy controls; however, several biological processes are consistently indicated in AD. The most recent and largest studies describe several hundreds of differentially expressed genes in AD after correction for multiple testing [50, 51]. Although post-mortem studies might also reflect pathological processes do overlap with recent genetic findings (table 2).

**Processes with increased expression**—Overall gene expression in post-mortem brain tissue of AD patients is generally lowered compared to controls. However, gene expression in following biological processes is up-regulated in brains of AD subjects. Inflammation has been associated with AD since the 1980s. Several studies demonstrated that genes related to inflammation have increased expression in AD across several brain regions [32, 43, 46, 49, 50, 52, 53]. It is still unknown whether inflammation is the culprit, the result, or a secondary response of AD; however, it is important to note that five (CR1, CD33, HLA-DRB5-DRB1, INPP5D, MEF2C) of the 20 LOAD GWAS-identified genes are involved in inflammation [24] (table 2). Increased calcium signaling was also observed across many transcriptome studies [36, 48, 49, 54]. Studies in neurons and mice expressing human APP and presenilin genes harboring ADEOAD mutations also show altered calcium signaling and calcium storage in the endoplasmic reticulum (ER) as well as synaptic dysfunction and loss of dendritic spines [54]. Further, cellular processes involved in mitochondrial and metabolic functions were shown to have increased gene expression in several transcriptome studies [42, 44, 49]. Mitochondrial function is impaired by APP, Aβ and presenilins [55]. Moreover, PET scans report a decrease in resting-state brain glucose metabolism and metabolic failure in AD brains [56]. The expression of genes related to cytoskeletal architecture is also increased in AD; this is consistent with the tau hypothesis of AD [57]. Microtubules are a major component of the cytoskeleton and the formation of NFTs in AD increasingly depletes microtubules by hyper phosphorylated and misfolded tau [58]. Another cytoskeletal process increased in AD patients is the formation of cofilin-actin rods along axons and dendrites, which results in cellular disruption. Cofilin-actin rods are known to form in response to heat shock, osmotic pressure, and ATP rundown within the hippocampus and frontal cortex of AD patients [59].

**Processes with decreased expression**—Synaptic related processes are decreased in AD [36, 48, 49, 54]. While A $\beta$  plaques and oligomers indirectly destroy synapses, aggregation of NFTs in neurons results in apoptosis, and inadvertently destroys synapses [60, 61]. Normal neurons remain in the G<sub>0</sub> phase; however, most AD neurons re-enter the cell cycle into the G<sub>1</sub> phase. This departure from the normal cell cycle in neurons results in

axonal defects. These findings are congruent with the decrease in synaptic related processes and the finding that negative regulation of cell cycle processes in AD is decreased [50, 62]. Moreover, signal transduction is also decreased in AD [63], particularly insulin signaling [64–66]. Lastly, genes involved in myelination are also decreased in LOAD [50, 67]. This finding corresponds to studies demonstrating that brain regions with the most myelination are the most vulnerable to AD pathology and that A $\beta$  plaques form retroactively to the developmental progression of myelination in the brain [68].

### **Profiling in Peripheral Tissue**

Studies have attempted to find expression profiles specific to AD in peripheral blood leukocytes to serve as biomarkers. Apoptosis is increased in peripheral blood cells (PBCs) of patients with AD [69]. Chemokine and cytokine signaling processes, which are both heavily involved in inflammation, also show increased expression in AD [53, 70, 71]. This is in line with a general increase in inflammation in response to apoptosis [72, 73]. Moreover, increased expression of inflammatory genes in PBCs has been associated with dementia, and is thought to be triggered by progressing AD pathology [74]. Another response to inflammation observed in AD is increased expression of TGF- $\beta$  [75–77]. Decreasing the expression of TGF- $\beta$  within innate immune cells mitigates AD symptoms in mice, such as an increase in spatial memory and A $\beta$  phagocytosis [78].

Most profiles that examined peripheral blood leukocytes in AD observed overall decreases in expression. In contrast to transcriptional profiles in brain tissue, peripheral blood leukocytes profiles show decreased expression of genes involved in cell structure related processes in AD [75, 79]. This finding has been explained by increased apoptosis observed in peripheral blood of AD patients. Similar to brain transcriptome studies, cellular signaling, lipid rafts, and cholesterol related processes are decreased in AD [77]. Two proteins responsible for lipid transport, APOE and APOJ (alias CLU) are genetically associated with AD, and the LOAD associated alleles of both *APOE* and *CLU* result in a decrease of lipid transportation [24, 51]. Moreover, lipid transport is reduced in patients with AD [76]. Additionally, AD patients had decreased expression of ATP-binding cassette transporters (ABC transporters), transporters which utilize ATP and carry out different processes within the cell. Seven ABC transporters have been directly linked to AD through functional studies or GWAS including *ABCA7* [80]. The cellular processes altered in the blood parallel those disrupted in the brain. PBC gene expression profiling in AD points to processes that are disrupted across the body and can potentially serve as a biomarker for AD.

### Single-Cell profiling

A $\beta$  plaques are extracellular, whereas NFTs are intracellular deposits typical to AD pathology. According to the tau hypothesis of AD, abnormal hyper-phosphorylation of the microtubule-associated protein tau (*MAPT*) leads to neurotoxic aggregates of tau, the formation of intracellular NFTs, the disintegration of microtubules, the collapse of neuronal transport and finally cell death [57]. To understand the transcriptional responses of neurons affected by intracellular NFTs, several studies applied single-cell transcriptomic profiling. Similar to post-mortem brain transcriptomics, there was an overall decrease of gene expression within neurons of AD patients with versus without NFTs [35, 62, 81]. Genes

Humphries and Kohli

involved in cell cycle, cell signaling, cytoskeleton, mitochondria, and metabolism were decreased in AD [35, 62, 81]. In addition, most cellular processes with increased gene expression in post-mortem brain studies also show increased expression within neurons affected by NFTs, such as inflammation and mitochondrial dysfunction [35, 62, 81]. Intriguingly, an increase in vesicle-mediated transport in singular neurons was observed prior to NFT development [62]. Defects in axonal transport were also observed in neurons prior to NFT formation. This increase in vesicle-mediated transport might be explained by tau oligomer toxicity prior to NFT formation, but might also be mediated by concomitant  $A\beta$  toxicity or neuroinflammation.

### Impact of AD-related variants on transcriptional profiles

To understand how human genetic variants impact AD pathogenesis, transcriptomics have been applied by either utilizing humanized transgenic cell and animal models or AD patientderived cells with and without variant allele status. Nagasaka et al. demonstrated that single causative ADEAOD mutations (*APP p.K595N/M596L*, *p.E693G* and *PSEN1 p.H163Y*) significantly impact transcriptional profiles [82]. The authors compared transcriptomic profiles of cultured fibroblasts from AD patients carrying an ADEOAD mutation with profiles from unaffected siblings that were non-carriers. While the levels of *APP* and *PSEN1* were comparable between fibroblasts of mutation carriers and non-carriers, up to 200 genes were differentially expressed between the groups, but showed similar profiles among AD affected mutation carriers.

Transcriptional profiles of *APOE* & AD risk allele carriers differed greatly when compared to non-risk allele carriers [49, 52]. Xu et al [49] compared hippocampal gene expression of AD patients with the *APOE* & 4/& genotype versus patients with the *APOE*  $\otimes$  / $\otimes$  genotype and found increased gene expression in processes such as cell growth, protein modification and RNA binding/editing; whereas gene expression was lowered in stress response, ER-Golgi transport, and mitochondrial oxidative phosphorylation. Expression differences were also observed between *APOE* & carriers and non-*APOE* & carriers when examining expression profiles in subjects with mild cognitive impairment (MCI) [52]. Similar to the Xu et al. study, genes involved in MHC class II protein complex, cell–matrix adhesion and cell growth had increased expression in *APOE* & carriers, whereas genes involved in processes such as mitochondrial electron transport, microtubule, synaptic and nucleosome assembly were down regulated [52].

Using post-mortem brain tissue, Rhinn et al [51] constructed gene expression networks to examine how APOE alleles influence gene network interactions in AD patients and healthy controls with different *APOE* risk genotypes. Interestingly, transcriptional profiles of nondemented *APOE*  $\note$  AD risk allele carriers already most resembled that of subjects with a diagnosis of LOAD when compared to patients with neurological diseases other than AD. Transfection of N2a-APP cells with human *APOE*  $\note$  alleles increased A $\beta_{40}$  and A $\beta_{42}$  levels, but did not with *APOE*  $\note$  alleles. Moreover, their analyses identified six genes (*RNF219, SV2A, HDLBP, ROGDI, CALU* and *PTK2B*) that exclusively interacted with the *APOE*  $\note$  allele, but not with the other *APOE* alleles. Importantly, knockdown of these genes

in *APOE*  $\mathcal{E}4$  allele transfected cells resulted in decreased A $\beta_{40}$  and A $\beta_{42}$  levels and had no effect on A $\beta$  levels in cells transfected with alternative *APOE* alleles.

### Conclusions

Three different study approaches so far led to the successful identification of rare variants in LOAD: (1) large-scale sequencing of autosomal-dominant early-onset Alzheimer disease (ADEOAD) genes in a case-control design with subsequent association testing in several even larger case-control cohorts followed by functional studies, (2) an analogous unbiased, genome-wide sequencing approach, and (3) a combined approach of sequencing and cosegregation analyses in several families enriched for LOAD also in conjunction with subsequent large association and molecular studies. Importantly and for the first time, strategy 1 showed genetic association between LOAD and a variant in the ADEOAD gene APP. Noteworthy, the strong protective effect of the newly discovered rare APP p.A673T variant serves as a proof of principle that reducing  $\beta$ -cleavage of APP may protect from AD. Strategy 2 and 3 led to the identification of completely novel LOAD susceptibility genes (TREM2, PLD3) that were not implicated by common GWAS variants. Given the complex nature of LOAD, it is likely that additional, yet unknown rare variant associations also with intermediate to large risk to or protection from LOAD will be revealed in the near future. Furthermore, biological processes defined by genes with causative mutations, rare and common susceptibility variants in AD overlap with processes indicated by human wholetranscriptomics studies in AD examining post-mortem brain, peripheral blood cells (PBC) and single neurons. Genes commonly up regulated in AD involve processes such as mitochondrial function, inflammation, calcium signaling and cytoskeletal organization, whereas gene expression in synaptic functions and signal transduction is reduced in AD. PBC transcriptional profiles reflect those obtained from brain tissues of AD patients. Thus, PBC profiling could become a practical biomarker for AD diagnosis and disease progression monitoring [83]. Moreover, transcriptome profiles of pre-symptomatic and AD affected pathogenic AD mutation or APOE risk allele carriers both reflect transcriptional changes reminiscent to those of LOAD patients. As more rare variants will be related to AD, transcriptional profiling in AD variant carriers will likely give variant specific insight into molecular mechanisms involved in LOAD pathogenesis which might provide an avenue for personalized medicine.

### Acknowledgments

The authors like to thank Dr. Margaret Pericak-Vance and Dr. John Gilbert from the John P. Hussman Institute for Human Genomics (HIHG) at the Dr. John T. Macdonald Foundation Department of Human Genetics of the Miller School of Medicine at the University of Miami for their support and funding as well as the NIA for the grant 1R01AG027944 and the Alzheimer's Association for the grant IIRG09133827.

### References

Papers of particular interest, published recently, have been highlighted as:

- Of importance;
- •• Of major importance

- Qiu C, Kivipelto M, von Strauss E. Epidemiology of Alzheimer's disease: occurrence, determinants, and strategies toward intervention. Dialogues Clin Neurosci. 2009; 11(2):111–28. [PubMed: 19585947]
- Castellani RJ, Rolston RK, Smith MA. Alzheimer disease. Dis Mon. 2010; 56(9):484–546. [PubMed: 20831921]
- 3. Hardy J, Selkoe DJ. The amyloid hypothesis of Alzheimer's disease: progress and problems on the road to therapeutics. Science. 2002; 297(5580):353–6. [PubMed: 12130773]
- 4. Braak H, Braak E. Neuropathological stageing of Alzheimer-related changes. Acta Neuropathol. 1991; 82(4):239–59. [PubMed: 1759558]
- Gatz M, Reynolds CA, Fratiglioni L, Johansson B, Mortimer JA, Berg S, et al. Role of genes and environments for explaining Alzheimer disease. Arch Gen Psychiatry. 2006; 63(2):168–74. [PubMed: 16461860]
- Campion D, Dumanchin C, Hannequin D, Dubois B, Belliard S, Puel M, et al. Early-onset autosomal dominant Alzheimer disease: prevalence, genetic heterogeneity, and mutation spectrum. Am J Hum Genet. 1999; 65(3):664–70. [PubMed: 10441572]
- Kokmen E, Beard CM, Offord KP, Kurland LT. Prevalence of medically diagnosed dementia in a defined United States population: Rochester, Minnesota, January 1, 1975. Neurology. 1989; 39(6): 773–6. [PubMed: 2725870]
- Goate A, Chartier-Harlin MC, Mullan M, Brown J, Crawford F, Fidani L, et al. Segregation of a missense mutation in the amyloid precursor protein gene with familial Alzheimer's disease. Nature. 1991; 349(6311):704–6. [PubMed: 1671712]
- St George-Hyslop PH, Tanzi RE, Polinsky RJ, Haines JL, Nee L, Watkins PC, et al. The genetic defect causing familial Alzheimer's disease maps on chromosome 21. Science. 1987; 235(4791): 885–90. [PubMed: 2880399]
- Schellenberg GD, Bird TD, Wijsman EM, Orr HT, Anderson L, Nemens E, et al. Genetic linkage evidence for a familial Alzheimer's disease locus on chromosome 14. Science. 1992; 258(5082): 668–71. [PubMed: 1411576]
- Sherrington R, Rogaev EI, Liang Y, Rogaeva EA, Levesque G, Ikeda M, et al. Cloning of a gene bearing missense mutations in early-onset familial Alzheimer's disease. Nature. 1995; 375(6534): 754–60. [PubMed: 7596406]
- Levy-Lahad E, Wijsman EM, Nemens E, Anderson L, Goddard KA, Weber JL, et al. A familial Alzheimer's disease locus on chromosome 1. Science. 1995; 269(5226):970–3. [PubMed: 7638621]
- Rogaev EI, Sherrington R, Rogaeva EA, Levesque G, Ikeda M, Liang Y, et al. Familial Alzheimer's disease in kindreds with missense mutations in a gene on chromosome 1 related to the Alzheimer's disease type 3 gene. Nature. 1995; 376(6543):775–8. [PubMed: 7651536]
- 14\*\*. Cruts M, Theuns J, Van Broeckhoven C. Locus-specific mutation databases for neurodegenerative brain diseases. Hum Mutat. 2012; 33(9):1340–4. Alzheimer Disease & Frontotemporal Dementia Mutation Database: www.molgen.ua.ac.be/ADMutations. [PubMed: 22581678]
- 15\*\*. Ringman JM, Coppola G. New genes and new insights from old genes: update on Alzheimer disease. Continuum (Minneap Minn). 2013; 19(2 Dementia):358–71. Review on clinical aspects of genetic variants in Alzheimer disease. [PubMed: 23558482]
- De Strooper B, Saftig P, Craessaerts K, Vanderstichele H, Guhde G, Annaert W, et al. Deficiency of presenilin-1 inhibits the normal cleavage of amyloid precursor protein. Nature. 1998; 391(6665):387–90. [PubMed: 9450754]
- Goate A, Hardy J. Twenty years of Alzheimer's disease-causing mutations. J Neurochem. 2012; 120 (Suppl 1):3–8. [PubMed: 22122678]
- Pericak-Vance MA, Bebout JL, Gaskell PC Jr, Yamaoka LH, Hung WY, Alberts MJ, et al. Linkage studies in familial Alzheimer disease: evidence for chromosome 19 linkage. Am J Hum Genet. 1991; 48(6):1034–50. [PubMed: 2035524]
- Corder EH, Saunders AM, Strittmatter WJ, Schmechel DE, Gaskell PC, Small GW, et al. Gene dose of apolipoprotein E type 4 allele and the risk of Alzheimer's disease in late onset families. Science. 1993; 261(5123):921–3. [PubMed: 8346443]

- 20. Pericak-Vance MA, Haines JL. Genetic susceptibility to Alzheimer disease. Trends Genet. 1995; 11(12):504–8. [PubMed: 8533168]
- 21. Farrer LA, Cupples LA, Haines JL, Hyman B, Kukull WA, Mayeux R, et al. Effects of age, sex, and ethnicity on the association between apolipoprotein E genotype and Alzheimer disease. A meta-analysis. APOE and Alzheimer Disease Meta Analysis Consortium. JAMA. 1997; 278(16): 1349–56. [PubMed: 9343467]
- Kim J, Basak JM, Holtzman DM. The role of apolipoprotein E in Alzheimer's disease. Neuron. 2009; 63(3):287–303. [PubMed: 19679070]
- 23. Bettens K, Sleegers K, Van Broeckhoven C. Genetic insights in Alzheimer's disease. Lancet Neurol. 2013; 12(1):92–104. [PubMed: 23237904]
- 24\*\*. Lambert JC, Ibrahim-Verbaas CA, Harold D, Naj AC, Sims R, Bellenguez C, et al. Metaanalysis of 74,046 individuals identifies 11 new susceptibility loci for Alzheimer's disease. Nat Genet. 2013; 45(12):1452–8. Latest GWAS on late-onset Alzheimer disease by the International Genomics of Alzheimer Disease Project (IGAP). [PubMed: 24162737]
- Manolio TA, Collins FS, Cox NJ, Goldstein DB, Hindorff LA, Hunter DJ, et al. Finding the missing heritability of complex diseases. Nature. 2009; 461(7265):747–53. [PubMed: 19812666]
- 26\*\*. Jonsson T, Stefansson H, Steinberg S, Jonsdottir I, Jonsson PV, Snaedal J, et al. Variant of TREM2 associated with the risk of Alzheimer's disease. N Engl J Med. 2013; 368(2):107–16. Identification of TREM2 as a novel LOAD susceptibility gene. The rare variant p.R47H confers similarly large risk to late-onset AD as the APOE ε4 allele. [PubMed: 23150908]
- 27\*\*. Cruchaga C, Haller G, Chakraverty S, Mayo K, Vallania FL, Mitra RD, et al. Rare variants in APP, PSEN1 and PSEN2 increase risk for AD in late-onset Alzheimer's disease families. PLoS One. 2012; 7(2):e31039. ADEOAD genes (APP, PSEN1 and 2) overall show an overrepresentation of rare coding variants in late-onset Alzheimer disease subjects. [PubMed: 22312439]
- 28\*\*. Jonsson T, Atwal JK, Steinberg S, Snaedal J, Jonsson PV, Bjornsson S, et al. A mutation in APP protects against Alzheimer's disease and age-related cognitive decline. Nature. 2012; 488(7409): 96–9. Identification of the rare APP p.A673T substitution that protects from AD with large effect size through reduced Aβ-cleavage. [PubMed: 22801501]
- 29\*\*. Guerreiro R, Wojtas A, Bras J, Carrasquillo M, Rogaeva E, Majounie E, et al. TREM2 variants in Alzheimer's disease. N Engl J Med. 2013; 368(2):117–27. Identification of TREM2 as a novel LOAD susceptibility gene. The rare variant p.R47H confers similarly large risk to late-onset AD as the APOE ε4 allele. [PubMed: 23150934]
- 30\*\*. Cruchaga C, Karch CM, Jin SC, Benitez BA, Cai Y, Guerreiro R, et al. Rare coding variants in the phospholipase D3 gene confer risk for Alzheimer's disease. Nature. 2013 Identification of PLD3 as a novel LOAD susceptibility gene in a combined approach of sequencing of LOAD families, large cohort genotyping and gene-based sequencing association analyses.
- Mullan M, Crawford F, Axelman K, Houlden H, Lilius L, Winblad B, et al. A pathogenic mutation for probable Alzheimer's disease in the APP gene at the N–terminus of β–amyloid. Nature genetics. 1992; 1(5):345–7. [PubMed: 1302033]
- 32. Avramopoulos D, Szymanski M, Wang R, Bassett S. Gene expression reveals overlap between normal aging and Alzheimer's disease genes. Neurobiol Aging. 2011; 32(12):2319, e27–34. [PubMed: 20570407]
- Brooks WM, Lynch PJ, Ingle CC, Hatton A, Emson PC, Faull RL, et al. Gene expression profiles of metabolic enzyme transcripts in Alzheimer's disease. Brain Res. 2007; 1127(1):127–35. [PubMed: 17109828]
- 34. Cao K, Chen-Plotkin AS, Plotkin JB, Wang LS. Age-correlated gene expression in normal and neurodegenerative human brain tissues. PLoS One. 2010; 5(9)
- Dunckley T, Beach TG, Ramsey KE, Grover A, Mastroeni D, Walker DG, et al. Gene expression correlates of neurofibrillary tangles in Alzheimer's disease. Neurobiol Aging. 2006; 27(10):1359– 71. [PubMed: 16242812]
- 36. Emilsson L, Saetre P, Jazin E. Alzheimer's disease: mRNA expression profiles of multiple patients show alterations of genes involved with calcium signaling. Neurobiol Dis. 2006; 21(3):618–25. [PubMed: 16257224]

- Haroutunian V, Katsel P, Schmeidler J. Transcriptional vulnerability of brain regions in Alzheimer's disease and dementia. Neurobiol Aging. 2009; 30(4):561–73. [PubMed: 17845826]
- Horesh Y, Katsel P, Haroutunian V, Domany E. Gene expression signature is shared by patients with Alzheimer's disease and schizophrenia at the superior temporal gyrus. Eur J Neurol. 2011; 18(3):410–24. [PubMed: 20695885]
- Katsel P, Li C, Haroutunian V. Gene expression alterations in the sphingolipid metabolism pathways during progression of dementia and Alzheimer's disease: a shift toward ceramide accumulation at the earliest recognizable stages of Alzheimer's disease? Neurochem Res. 2007; 32(4–5):845–56. [PubMed: 17342407]
- 40. Katsel P, Tan W, Haroutunian V. Gain in brain immunity in the oldest-old differentiates cognitively normal from demented individuals. PLoS One. 2009; 4(10):e7642. [PubMed: 19865478]
- 41. Lukiw WJ. Gene expression profiling in fetal, aged, and Alzheimer hippocampus: a continuum of stress-related signaling. Neurochem Res. 2004; 29(6):1287–97. [PubMed: 15176485]
- Mills JD, Nalpathamkalam T, Jacobs HI, Janitz C, Merico D, Hu P, et al. RNA-Seq analysis of the parietal cortex in Alzheimer's disease reveals alternatively spliced isoforms related to lipid metabolism. Neurosci Lett. 2013; 536:90–5. [PubMed: 23305720]
- Parachikova A, Agadjanyan MG, Cribbs DH, Blurton-Jones M, Perreau V, Rogers J, et al. Inflammatory changes parallel the early stages of Alzheimer disease. Neurobiol Aging. 2007; 28(12):1821–33. [PubMed: 17052803]
- 44. Silva AR, Grinberg LT, Farfel JM, Diniz BS, Lima LA, Silva PJ, et al. Transcriptional alterations related to neuropathology and clinical manifestation of Alzheimer's disease. PLoS One. 2012; 7(11):e48751. [PubMed: 23144955]
- 45. Tan MG, Chua WT, Esiri MM, Smith AD, Vinters HV, Lai MK. Genome wide profiling of altered gene expression in the neocortex of Alzheimer's disease. J Neurosci Res. 2010; 88(6):1157–69. [PubMed: 19937809]
- Tollervey JR, Wang Z, Hortobagyi T, Witten JT, Zarnack K, Kayikci M, et al. Analysis of alternative splicing associated with aging and neurodegeneration in the human brain. Genome Res. 2011; 21(10):1572–82. [PubMed: 21846794]
- Weeraratna AT, Kalehua A, Deleon I, Bertak D, Maher G, Wade MS, et al. Alterations in immunological and neurological gene expression patterns in Alzheimer's disease tissues. Exp Cell Res. 2007; 313(3):450–61. [PubMed: 17188679]
- 48. Xu PT, Li YJ, Qin XJ, Kroner C, Green-Odlum A, Xu H, et al. A SAGE study of apolipoprotein E3/3, E3/4 and E4/4 allele-specific gene expression in hippocampus in Alzheimer disease. Mol Cell Neurosci. 2007; 36(3):313–31. [PubMed: 17822919]
- Xu PT, Li YJ, Qin XJ, Scherzer CR, Xu H, Schmechel DE, et al. Differences in apolipoprotein E3/3 and E4/4 allele-specific gene expression in hippocampus in Alzheimer disease. Neurobiol Dis. 2006; 21(2):256–75. [PubMed: 16198584]
- 50\*\*. Zhang B, Gaiteri C, Bodea LG, Wang Z, McElwee J, Podtelezhnikov AA, et al. Integrated systems approach identifies genetic nodes and networks in late-onset Alzheimer's disease. Cell. 2013; 153(3):707–20. Most recent transcriptome analyses in Alzheimer disease. Network analyses identify hub genes on biological processes disrupted in Alzheimer disease. [PubMed: 23622250]
- 51\*\*. Rhinn H, Fujita R, Qiang L, Cheng R, Lee JH, Abeliovich A. Integrative genomics identifies APOE epsilon4 effectors in Alzheimer's disease. Nature. 2013; 500(7460):45–50. Identifies a handful of genes responsible for increased A $\beta$  levels in carriers of the common APOE  $\epsilon$ 4 risk allele for late-onset AD. [PubMed: 23883936]
- 52. Bossers K, Wirz KT, Meerhoff GF, Essing AH, van Dongen JW, Houba P, et al. Concerted changes in transcripts in the prefrontal cortex precede neuropathology in Alzheimer's disease. Brain. 2010; 133(Pt 12):3699–723. [PubMed: 20889584]
- Maes OC, Xu S, Yu B, Chertkow HM, Wang E, Schipper HM. Transcriptional profiling of Alzheimer blood mononuclear cells by microarray. Neurobiol Aging. 2007; 28(12):1795–809. [PubMed: 16979800]

- 54. Woods NK, Padmanabhan J. Neuronal calcium signaling and Alzheimer's disease. Adv Exp Med Biol. 2012; 740:1193–217. [PubMed: 22453989]
- Begley JG, Duan W, Chan S, Duff K, Mattson MP. Altered calcium homeostasis and mitochondrial dysfunction in cortical synaptic compartments of presenilin-1 mutant mice. J Neurochem. 1999; 72(3):1030–9. [PubMed: 10037474]
- Mosconi L. Brain glucose metabolism in the early and specific diagnosis of Alzheimer's disease. FDG-PET studies in MCI and AD. Eur J Nucl Med Mol Imaging. 2005; 32(4):486–510. [PubMed: 15747152]
- 57. Iqbal K, del Alonso AC, Chen S, Chohan MO, El-Akkad E, Gong CX, et al. Tau pathology in Alzheimer disease and other tauopathies. Biochim Biophys Acta. 2005; 1739(2–3):198–210. [PubMed: 15615638]
- Bamburg JR, Bloom GS. Cytoskeletal pathologies of Alzheimer disease. Cell Motil Cytoskeleton. 2009; 66(8):635–49. [PubMed: 19479823]
- Huang TY, Minamide LS, Bamburg JR, Bokoch GM. Chronophin mediates an ATP-sensing mechanism for cofilin dephosphorylation and neuronal cofilin-actin rod formation. Dev Cell. 2008; 15(5):691–703. [PubMed: 19000834]
- Serrano-Pozo A, Mielke ML, Gomez-Isla T, Betensky RA, Growdon JH, Frosch MP, et al. Reactive glia not only associates with plaques but also parallels tangles in Alzheimer's disease. Am J Pathol. 2011; 179(3):1373–84. [PubMed: 21777559]
- Sheng M, Sabatini BL, Sudhof TC. Synapses and Alzheimer's disease. Cold Spring Harb Perspect Biol. 2012; 4(5)
- Williams C, Mehrian Shai R, Wu Y, Hsu YH, Sitzer T, Spann B, et al. Transcriptome analysis of synaptoneurosomes identifies neuroplasticity genes overexpressed in incipient Alzheimer's disease. PLoS One. 2009; 4(3):e4936. [PubMed: 19295912]
- 63. Ruggiero R, Kale A, Thomas B, Baker NE. Mitosis in neurons: Roughex and APC/C maintain cell cycle exit to prevent cytokinetic and axonal defects in Drosophila photoreceptor neurons. PLoS Genet. 2012; 8(11):e1003049. [PubMed: 23209426]
- 64. Liu Y, Liu F, Grundke-Iqbal I, Iqbal K, Gong CX. Deficient brain insulin signalling pathway in Alzheimer's disease and diabetes. J Pathol. 2011; 225(1):54–62. [PubMed: 21598254]
- 65. Lee HK, Kumar P, Fu Q, Rosen KM, Querfurth HW. The insulin/Akt signaling pathway is targeted by intracellular beta-amyloid. Mol Biol Cell. 2009; 20(5):1533–44. [PubMed: 19144826]
- 66. Moloney AM, Griffin RJ, Timmons S, O'Connor R, Ravid R, O'Neill C. Defects in IGF-1 receptor, insulin receptor and IRS-1/2 in Alzheimer's disease indicate possible resistance to IGF-1 and insulin signalling. Neurobiol Aging. 2010; 31(2):224–43. [PubMed: 18479783]
- Blalock EM, Geddes JW, Chen KC, Porter NM, Markesbery WR, Landfield PW. Incipient Alzheimer's disease: microarray correlation analyses reveal major transcriptional and tumor suppressor responses. Proc Natl Acad Sci U S A. 2004; 101(7):2173–8. [PubMed: 14769913]
- Bartzokis G. Age-related myelin breakdown: a developmental model of cognitive decline and Alzheimer's disease. Neurobiol Aging. 2004; 25(1):5–18. author reply 49–62. [PubMed: 14675724]
- Leuner K, Schutt T, Kurz C, Eckert SH, Schiller C, Occhipinti A, et al. Mitochondrion-derived reactive oxygen species lead to enhanced amyloid beta formation. Antioxid Redox Signal. 2012; 16(12):1421–33. [PubMed: 22229260]
- Calciano MA, Zhou W, Snyder PJ, Einstein R. Drug treatment of Alzheimer's disease patients leads to expression changes in peripheral blood cells. Alzheimers Dement. 2010; 6(5):386–93. [PubMed: 20185375]
- 71. Fehlbaum-Beurdeley P, Sol O, Desire L, Touchon J, Dantoine T, Vercelletto M, et al. Validation of AclarusDx, a blood-based transcriptomic signature for the diagnosis of Alzheimer's disease. J Alzheimers Dis. 2012; 32(1):169–81. [PubMed: 22785402]
- Ankarcrona M, Winblad B. Biomarkers for apoptosis in Alzheimer's disease. Int J Geriatr Psychiatry. 2005; 20(2):101–5. [PubMed: 15660410]
- Bergman M, Salman H, Beloosesky Y, Djaldetti M, Bessler H. Are peripheral blood cells from patients with Alzheimer disease more sensitive to apoptotic stimuli? Alzheimer Dis Assoc Disord. 2002; 16(3):156–60. [PubMed: 12218646]

- 74. Metti AL, Cauley JA. How predictive of dementia are peripheral inflammatory markers in the elderly? Neurodegener Dis Manag. 2012; 2(6):609–22. [PubMed: 23441140]
- 75. Booij BB, Lindahl T, Wetterberg P, Skaane NV, Saebo S, Feten G, et al. A gene expression pattern in blood for the early detection of Alzheimer's disease. J Alzheimers Dis. 2011; 23(1):109–19. [PubMed: 20930264]
- 76. Chen KD, Chang PT, Ping YH, Lee HC, Yeh CW, Wang PN. Gene expression profiling of peripheral blood leukocytes identifies and validates ABCB1 as a novel biomarker for Alzheimer's disease. Neurobiol Dis. 2011; 43(3):698–705. [PubMed: 21669286]
- 77. Fehlbaum-Beurdeley P, Jarrige-Le Prado AC, Pallares D, Carriere J, Guihal C, Soucaille C, et al. Toward an Alzheimer's disease diagnosis via high-resolution blood gene expression. Alzheimers Dement. 2010; 6(1):25–38. [PubMed: 20129318]
- 78. Town T, Laouar Y, Pittenger C, Mori T, Szekely CA, Tan J, et al. Blocking TGF-beta-Smad2/3 innate immune signaling mitigates Alzheimer-like pathology. Nat Med. 2008; 14(6):681–7. [PubMed: 18516051]
- Kalman J, Kitajka K, Pakaski M, Zvara A, Juhasz A, Vincze G, et al. Gene expression profile analysis of lymphocytes from Alzheimer's patients. Psychiatric genetics. 2005; 15(1):1–6. [PubMed: 15722950]
- Wolf A, Bauer B, Hartz AM. ABC Transporters and the Alzheimer's Disease Enigma. Front Psychiatry. 2012; 3:54. [PubMed: 22675311]
- Ginsberg SD, Che S, Wuu J, Counts SE, Mufson EJ. Down regulation of trk but not p75NTR gene expression in single cholinergic basal forebrain neurons mark the progression of Alzheimer's disease. J Neurochem. 2006; 97(2):475–87. [PubMed: 16539663]
- Nagasaka Y, Dillner K, Ebise H, Teramoto R, Nakagawa H, Lilius L, et al. A unique gene expression signature discriminates familial Alzheimer's disease mutation carriers from their wildtype siblings. Proc Natl Acad Sci U S A. 2005; 102(41):14854–9. [PubMed: 16199521]
- Humpel C. Identifying and validating biomarkers for Alzheimer's disease. Trends Biotechnol. 2011; 29(1):26–32. [PubMed: 20971518]

## Table 1 Overview of rare variants associated with late-onset Alzheimer disease (LOAD)

association in one study and remaining variants are likely pathogenic according to co-segregation in LOAD families. Minor allele frequencies (MAF) are Variants in bold achieved study-wide significance and were replicated in an independent study sample. Variants in italic obtained nominal significant reported as were given by the cited studies and reflect frequencies in populations of European descent.

Gene	Amino acid change	dbSNP rs#	Odds ratio	MAF (%)	PolyPhen2 prediction	Reference
APP	A673T N660Y	rs63750847 -	<b>4.2</b> * _	<b>0.4</b> <0.01	<b>benign</b> probably damaging	[28]
PLD3	<b>V232M</b> <i>M6R</i> A442A (38)	rs145999145	<b>2.1</b> 7.7 2.3	<b>0.4</b> <0.1 1.6	probably damaging benign	[30]
TREM2	<b>R47H</b> <i>D87N</i> Q33Ter <sup>~</sup> Y38C <sup>~</sup> T66M <sup>~</sup>	<b>rs75932628</b> <i>rs142232675</i> rs104894002 - rs201258663	2.9	<b>0.3</b> 0.1 0.01 <0.08 <0.03	probably damaging probably damaging probably damaging probably damaging	[26, 29] [29] [29] [29]

protective from AD

Curr Genet Med Rep. Author manuscript; available in PMC 2015 June 01.

 $\tilde{c}$ ause frontotemporal dementia–like syndrome or Nasu-Hakola disease in homozygous state

ss variant affects splice site

Table 2

# Overview of biological processes altered in Alzheimer disease (AD) from transcriptome studies since 2005

Listed beside each process are studied tissues, direction of change and genes involved in the same process implicated to AD by Genome-Wide Association Studies (GWAS).

Humphries and Kohli

Processes	Tissues		Direction	GWAS genes	References
Inflammation/Immune System	1 Brain Tir 2 PBCs 3 Single-co	ssue ell	Increased	BINI CD33 CRI EPHAI HLA-DRBI INPP5D MEF2C MS4A6A	<ol> <li>[32, 43, 46, 49, 50, 52, 53]</li> <li>[53, 70, 71].</li> <li>[35, 45, 62, 70, 75, 81]</li> </ol>
Calcium Signaling	Brain Tissue		Increased		[36, 48, 49, 54]
Mitochondrial and Metabolic functions	1 Brain Ti 2 PBCs 3 Single-C	ssue	Increased		1 [42, 44, 49] 2 [38] 3 [35, 62, 81]
Cytoskeleton related processes	1 Post-Mo 2 PBCs 3 Single-C	ottem Brain Cell	Increased	CASS4 CD2AP CELF1 NME8 SORL1	1 [32, 49, 52] 2 [53, 75, 79].
Cell-cycle processes	1 Brain T 3 Single-	lissue cell	Decreased		1 [50, 62] 3 [35, 62, 81]
Synaptic Transmission	1 Brain Ti 2 PBCs 3 Single-or	ssue ell	Decreased	PICALM CASS4	1 [32, 36-44, 52, 62] 2 [53] 3 [75]
Signal Transduction	1 Brain Ti 2 PBCs 3 Single-or	ssue ell	Decreased	SORLI CD33 CD2AP	<ol> <li>[42,48,49,52,63-66].</li> <li>[75,79].</li> <li>[35, 62, 81]</li> </ol>
Cholesterol Related Processes	1 Brain Ti 2 PBCs 3 Single-co	ssue ell	Decreased	ABCA7 APOE CLU SORLI	1 [42] 2 [35] 3 [76, 77]

Curr Genet Med Rep. Author manuscript; available in PMC 2015 June 01.

Page 16

Processes	Tissues	Direction	<b>GWAS</b> genes	References
TGFß related signaling	PBCs	Increased		[70-77]
ABC transporters	PBCs	Decreased	ABCA7	[76, 80]
Myelination	Brain tissue	Decreased	SLC24A4	[50, 67]
Vesicle-mediated transport	<ol> <li>Brain Tissue</li> <li>Single-Cell</li> </ol>	Increased	CASS4 CELF1 NME8	1 [42] 2 [62]